

Influence of Seasonality and Bathymetry on Decapod Crustacean Community Structure in Malindi - Ungwana Bay, Kenya

Collins K. Ndoro^{1,2}, Boaz Kaunda-Arara², Cosmas N. Munga³ and Renison Ruwa⁴

¹Fisheries Department, Ministry of Agriculture, Livestock and Fisheries, PO Box 90423-80100, Mombasa, Kenya; ²University of Eldoret, Department of Fisheries and Aquatic Sciences, PO Box 1125-30100, Eldoret, Kenya; ³Technical University of Mombasa, Department of Environment and Health Sciences, Marine Sciences Section, PO Box 90420 – 80100, Mombasa, Kenya; ⁴Kenya Marine and Fisheries Research Institute, PO Box 81651-80100, Mombasa, Kenya.

Keywords: Decapoda, Penaeidae, abundance, monsoons, fisheries management, Kenya

Abstract — Decapod crustaceans support both artisanal and semi-industrial fisheries in the Western Indian Ocean (WIO) region. Despite their commercial value, data on their community structure is lacking in most of the region but are important for stock management. This study provides information on seasonal and bathymetric variation in decapod crustacean community structure in Malindi-Ungwana Bay, a biodiversity rich ecosystem in coastal Kenya. Samples were collected in the northeast (NEM) and southeast (SEM) monsoon seasons during an experimental bottom trawling survey in 2011. A total of 43 transects covering an estimated area of 1 873 km² were trawled in four depth zones (0-10, 10-20, 20-40 and 40-100 m) in both seasons. Twenty species of decapod crustaceans belonging to the Penaeidae, Portunidae, Calappidae, Majidae, Matutidae, Palinuridae and Scyllaridae were harvested. Overall crustacean biomass was higher in the SEM than the NEM. Penaeid prawns were numerically the most abundant in both the NEM (89.3%) and SEM (85.3%) seasons, *Fenneropenaeus indicus* being the most abundant in the NEM (58%) and SEM (42%). nMDS plots revealed separation of crustacean assemblages between depth zones but not the seasons. Two-way crossed ANOSIM indicated significant difference in species composition between the depth zones but not the seasons, with higher species diversity in the shallower depth strata. Canonical Correspondence Analysis revealed that temperature, salinity, Secchi depth and dissolved oxygen influence the bathymetric distribution of species in the bay. Recommendations are made that these factors be taken into consideration in the management of the crustacean fishery in the bay.

INTRODUCTION

Marine decapod crustaceans account for nearly 6.9% of worldwide fin and shellfish landings by weight (FAO, 2011) and these landings have been on the upward trend in recent decades (FAO, 2008, 2012). The rise in decapod crustacean catches is mostly attributable to increased effort as a result of dwindling fish stocks worldwide (Jackson *et al.*, 2001; Worm *et al.*, 2006; FAO, 2010). Crustacean stocks are, however, increasingly being threatened with overfishing (FAO, 2012), mostly because of the global shift in targeting these resources.

Fishing pressure can cause shifts in species community structure and ecosystem function (Jennings and Kaiser, 1998) with trophodynamic consequences (Leibold, 1996). Several abiotic and biotic factors may further influence the distribution patterns and community structure of marine benthic communities like crustaceans. Such abiotic factors include depth profile (Fannelli *et al.*, 2007; Munoz, *et al.*, 2008), salinity gradient (Gillett, 2008), substratum type (Lavrado *et al.*, 2000) and rainfall patterns (Teikwa and Mgaya, 2003). Important biotic factors comprise environmental productivity (Follesa *et al.*, 2009) and biological interactions (Jackson, *et al.*, 2001).

A limited number of studies have been conducted on decapod crustaceans in the Western Indian Ocean (WIO) compared to the temperate latitudes. Most of the WIO studies have concentrated on species distribution (Mutagyera, 1984; Munga *et al.*, 2012), the functional biology of single species (Wakwabi and Jaccarini, 1993; Wakwabi, 1996; Teikwa and Mgaya, 2003), and stock assessments and fisheries (Groeneveld and Melville-Smith, 1995; Groeneveld, 2000; Mwatha, 2005). Decapod crustaceans form an important link between lower and higher trophic levels and studies on factors that affect their community structure may offer useful information on ecosystem function at the local scale (Papiol *et al.*, 2012). In ecosystems that are heavily fished and influenced by environmental

variability, such as the expansive Malindi-Ungwana Bay in Kenya, information on variability in a resource's community structure is useful in assessing spatio-temporal drivers of assemblages. This study therefore aimed to provide information on the environmental correlates of crustacean community structure in the Malindi-Ungwana Bay.

The bay is the most productive nearshore ecosystem in coastal Kenya (Nzioka, 1981; Mutagyera, 1984) and has thus been the focus of various trawling expeditions. Recent resource-use conflict and a fishing ban in the bay (Munga *et al.*, 2012) requires that scientific information be available to support management.

METHODS

Study site and survey design

The study was carried out within Malindi-Ungwana Bay on the northern coast of Kenya (Fig. 1). The bay lies between 2° 30'S - 3° 30'S and 40° 000'E - 41° 000'E. It is the only known trawlable ground on the Kenyan coast, extends along ~200 km of coastline and has a continental shelf ranging from 15-60 km in width, with an estimated fishing ground of 35 300 km² (Iversen, 1984; Mueni, 2006). The Athi and Tana Rivers (Fig. 1) discharge an estimated 6 000 million m³ of freshwater and 3 million tonnes of sediment annually into the bay (Tychsen, 2006), which is affected by the monsoons that prevail on the Kenyan coast (McClanahan, 1988). Briefly, the northeast monsoon season (NEM, November–March) is a period of calm seas, elevated sea surface temperatures and higher salinities, while the southeast monsoon (SEM, April–October) is characterised by rough seas, cool weather, lower salinities and higher plankton productivity. The influence of this seasonality on community structure of crustaceans in coastal East Africa is not well documented.

The data were collected during two trawl surveys in the bay using the FV Vega, a medium-sized Kenyan prawn trawler. The first survey was conducted during 22 January - 4 February 2011 (NEM), the second between

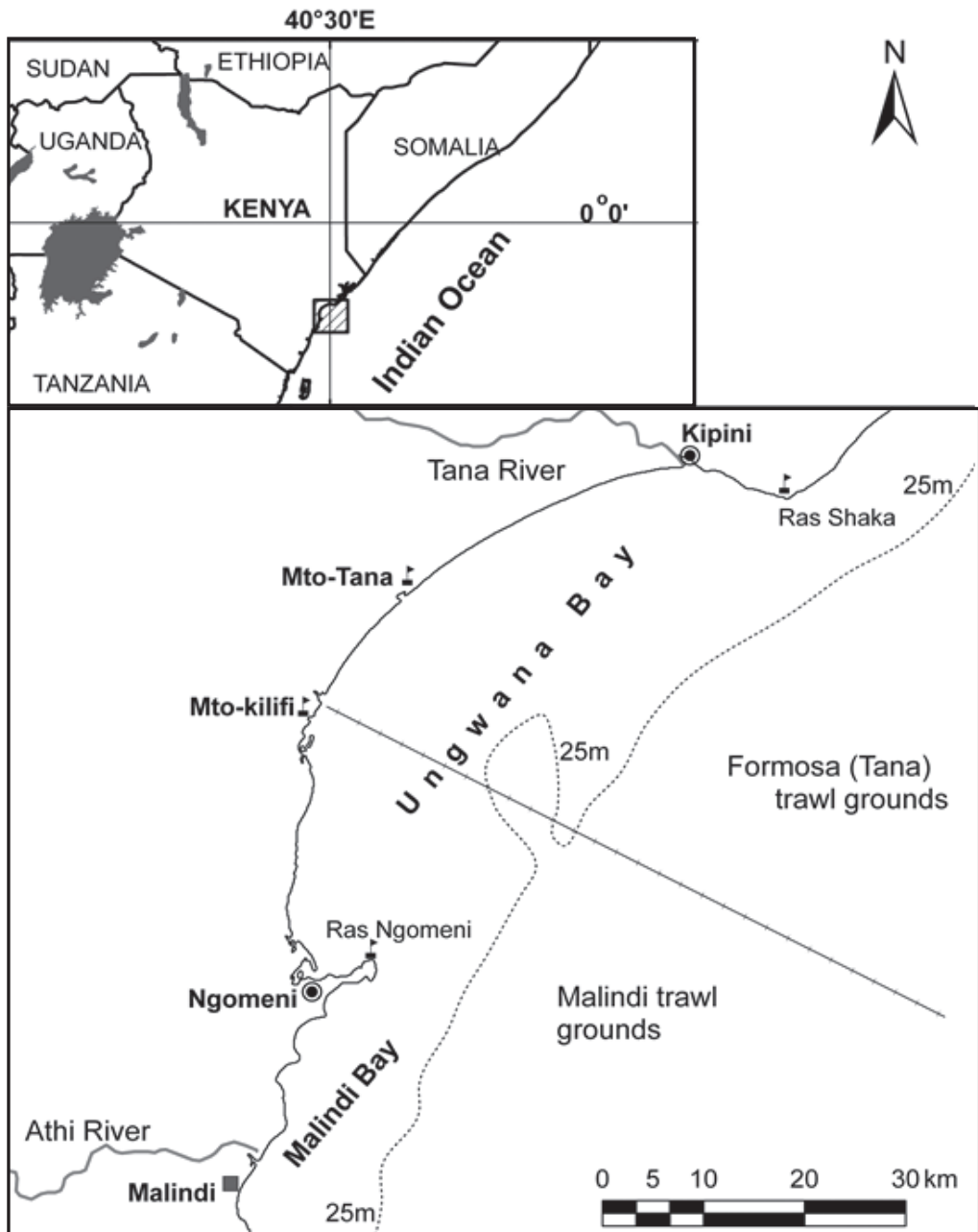


Figure 1. Map of the Malindi–Ungwana Bay area showing the discharge point of the Athi and Tana Rivers into the bay and the demarcation of the Formosa and Malindi commercial fishing (Source: Munga *et al.*, 2012).

22 May - 4 June 2011 (SEM). The trawler was fitted with two outrigger trawl nets made of nylon, comprising wings, a net body and cod-end with a total length of 44.3 m. The wings were 19.1 m long with a mesh size of 45 mm;

the net body was also 19.1 m long but with a mesh size of 70.4 mm, and the cod-end was 6.1 m long with a mesh size of 45 mm. The nets each had a 22.5 m head rope length, a 25.4 m foot rope and a 28 m restraining chain

between the two trawl doors. They were lowered simultaneously by hydraulic winch and the trawl start time recorded when the nets reached the sea floor, while the trawl end time was recorded when the vessel started retracting the nets.

A total of 43 trawl transects were surveyed within the bay during a 13-day period in each season. The surveys were stratified by depth into four zones: 0-10 m (zone 1); 10-20 m (zone 2); 20-40 m (zone 3) and 40-100 m (zone 4). The depth zone intervals were not uniform because the shelf steepens rapidly a few miles from the shore.

The percentage area of each depth zone within the trawlable area was used to determine the proportion of sampling time apportioned to each zone in each season, given that a maximum of four trawls could be conducted per day. The four depth zones were 471, 803, 468 and 133 km² in area and were allocated 8 (zone 1), 17 (zone 2), 13 (zone 3) and 5 (zone 4) trawls, respectively, totalling 43 trawls in each seasonal survey. The trawl transects ran parallel to the shoreline to remain within the respective depth zone as much as possible; shallow areas, coral and rocky areas were avoided. The geographical coordinates of the start and end point of each trawl transect were determined using a GPS. Trawling was done at a speed of 2.5-3.0 knots during the day between 06:00-18:00 h and each trawl lasted an hour. The same transects were trawled during the NEM and SEM.

Sampling methods

Data on environmental variables were recorded at the start of each trawl during the NEM. A bottom water sample was collected using a Niskin bottle and dissolved oxygen, temperature and salinity measured using a digital meter. Water transparency was measured from the side of the boat using a Secchi disc. The depths of each transect position were measured using an echo sounder. The net was hauled onto the deck at the end of each trawl and the decapod crustaceans sampled using a protocol that depended on the catch size. When the catch

was small and manageable (e.g. the total catch could be worked within an hour), the total haul was processed and sorted into the various crustacean groups. These were identified to species following identification keys by the FAO (1984) and De Grave and Franssen (2011) for prawns, and Stephenson (1948), Branch *et al.* (2007), Ng *et al.* (2008) and Richmond (2011) for crabs. Species were weighed to the nearest 0.1 g, and carapace lengths and widths measured to the nearest 1 mm using a Vernier caliper. Catches that were too large (requiring over an hour to work) were sub-sampled. All large specimens in the haul were first removed, identified and weighed individually to the nearest gram. The remaining catch was turned to achieve uniform mixing and then sub-divided into portions (sub-samples) of approximately equal size, one of which (b, kg) was randomly selected for analysis. The total weight (a, kg) of the other portions was also recorded. These weights were later added to that of the large specimens to determine the weight of the total haul. The weight of each species in the haul was estimated by multiplying their weight in the sub-sample (b, kg) by a raising factor (RF): RF= (a + b) / b.

Data analysis

Biomass estimates of penaeid prawns were calculated using the Swept Area Method (Sparre & Venema, 1998). The distance trawled (D) per transect was estimated in units of nautical miles (nm) as:

$$D = 60 * \text{Sqrt}((\text{Lat1} - \text{Lat2})^2 + (\text{Lon1} - \text{Lon2})^2 * \cos^2(0.5 * (\text{Lat1} + \text{Lat2}))) \dots\dots\dots(1)$$

Where:

- Lat1= Latitude at start of haul (degrees)
- Lat2 = Latitude at end of haul (degrees)
- Lon1 = Longitude at start of haul (degrees)
- Lon2 = Longitude at end of haul (degrees)

The estimated trawl distance was then multiplied by the length of the head rope (22.5 m) to get the trawled area (A, nm²), with a correction factor of 0.5 for the net configuration (Pauly, 1980):

$$\text{Swept Area (A)} = D * 22.5 * 0.5 \dots\dots\dots(2)$$

The catch densities (converted to kg.km^{-2}) of species were then derived for each haul, while species biomass (B, Kg) in each haul was derived as:

$$B = \text{density} \times A / X1 \dots\dots\dots(3)$$

Where X1 is sampling proportion of crustaceans present in the swept area ($X1 = 1$, assuming all the crustaceans were fully accessible to the trawl).

Multivariate non-metric multi-dimensional scaling (MDS) was used to describe the community structure by depth zones and season (NEM and SEM) based on the Bray-Curtis similarity index (Clarke and Warwick, 2001). Two-way ANOSIM was used to test for differences in community structure with depth zones and seasons as factors, while two-way SIMPER analysis identified which crustacean species contributed most to the dissimilarity. The resulting R-values provided a measure of variation between samples, ranging from -1 to 1. Values tending to zero indicated little difference in species composition between depths/seasons, while values tending to +1 inferred differing composition (Clarke & Warwick, 2001). Canonical Correspondence Analysis (CCA) was used to analyse the influence of environmental factors (depth, temperature, dissolved oxygen, Secchi depth and salinity) on species distribution only during the NEM. The statistical analyses were performed using PRIMER version 6.

Two-way ANOVA (on $\log(x+1)$ transformed data) was used to test for the effect of seasons and depth on overall crustacean biomass, species richness and diversity in the bay. Taxonomic richness (S) and the Shannon-Wiener diversity index (H') were used to describe the community structure. As S reflected the total number of species of crustaceans in a stratum, the mean richness was calculated according to the depth zones and season. H' was calculated following Magurran (1988):

$$H' = - \sum_i p_i \log(p_i)$$

Where p_i is the proportion of the total count arising from the i th species in the stratum or season.

RESULTS

Species composition, abundance and catch rates

Totals of 767 and 1 808 crustaceans were sampled during the NEM and SEM, respectively, including 20 decapod crustacean species belonging to seven families. During the NEM, 11 species were harvested, mostly penaeid prawns (5 species), but relatively more portunids (9 species) made up the 19 species were caught during SEM (Table 1). Among the penaeid species, *Fenneropenaeus indicus* had the highest mean abundance (individuals. $\text{km}^{-2} \pm \text{SE}$) of $8\,318 \pm 4\,132$, followed by *Metapeneus monoceros* ($1\,489 \pm 689$), *Penaeus semisulcatus* ($1\,069 \pm 415$) and *Penaeus monodon* ($1\,008 \pm 439$) (Table 1). The other crustaceans were recorded in low numbers of <400 individuals. km^{-2} (Table 1). Penaeid prawns were numerically the most abundant both in the NEM and SEM (89.7% and 85.3 %, respectively), with *F. indicus* making up the greatest proportion of the catch (NEM, 57.6%; SEM, 41.5%). The other penaeids occurred in variable proportions (Table 1). The Portunidae made up 9.4% and 12.3% of the numbers caught in the NEM and SEM, respectively, while the Palinuridae were very low in abundance in both seasons (Table 1).

Higher mean catch rates ($\text{kg.km}^{-2} \pm \text{SE}$) were recorded in depth zone 1 (64.2 ± 18.7 kg.km^{-2}) during the NEM, with catch rates of 15.7 ± 5.5 and 17.9 ± 9.5 kg.km^{-2} being obtained in zones 2 and 3, respectively, during this season. During the SEM, catch rates of 67.14 ± 18.6 and 56.6 ± 18.1 kg.km^{-2} were obtained from depth zones 1 and 2, respectively, while zones 3 and 4 yielded lower rates of 14.25 ± 5.8 and 1.31 ± 1.1 kg.km^{-2} , respectively. Two-way ANOVA revealed that depth had a significant effect on the crustacean harvest ($F = 3.89$; $df = 2, 130$; $P = 0.022$). This was not the case with season ($F=0.014$; $df= 1, 130$; $P=0.95$), or the interaction of season and depth ($F = 0.57$; $df = 2,130$; $P = 0.54$).

Table 1. Mean abundance (individuals.Km² ± SE) and percentage composition of decapod crustaceans caught in trawls during the northeast (NEM) and southeast monsoons (SEM) in Malindi-Ungwana Bay. Values without standard errors indicate that species appeared in only one transect; (-) indicates absence of catch.

Species	%	NEM	%	SEM
Penaeidae				
<i>Fenneropenaeus indicus</i>	57.6	8318 ± 4132	41.5	12151 ± 3329
<i>Marsupenaeus japonicus</i>	0.9	149 ± 35	0.6	418 ± 135
<i>Metapenaeus monoceros</i>	12.5	1489 ± 688.8	27.0	6790 ± 1580
<i>Penaeus monodon</i>	8.9	1008 ± 439.3	5.9	1233 ± 294
<i>Penaeus semisulcatus</i>	9.8	1069 ± 414.6	10.3	3777 ± 1784
Portunidae				
<i>Portunus sanguinolentus</i>	9.3	347± 156.1	9.7	331 ±106
<i>Thalamita crenata</i>	0.1	19	0.05	20
<i>Charybdis feriata</i>	-	-	0.6	38 ±7
<i>Charybdis hellerii</i>	-	-	0.8	144 ± 124
<i>Charybdis natator</i>	-	-	0.2	81
<i>Charybdis smithii</i>	-	-	0.5	172
<i>Podophthalmus vigil</i>	-	-	0.05	21
<i>Portunus pelegicus</i>	-	-	0.05	21
<i>Scylla serrata</i>	-	-	0.3	33 ± 13
Calappidae				
<i>Calappa calappa</i>	0.3	40	-	-
<i>Calappa</i> sp.	0.1	21	0.4	168
Matutidae				
<i>Ashtoret lunaris</i>	0.7	97	1.0	91 ± 17
Majidae				
<i>Majid</i> sp.	-	-	0.2	31 ± 11
Scyllidae				
<i>Thenus orientalis</i>	0.4	20 ± 0.4	0.7	58 ± 24
Palinuridae				
<i>Panulirus ornatus</i>	0.3	-	0.1	21± 0.3

A greater harvest of prawns (the most abundant crustaceans in the bay) was obtained from depth zone 1 (78.7 ± 22.6 kg.km⁻²) during the NEM. Depth zones 2 and 3 yielded near equal catches of 20.4±7.0 and 20.9±11 kg.km⁻², respectively. During the SEM, depth zones 1 and 2 yielded near equal catches of prawns of 112.4 ± 30.3 and 103 ± 31.01 kg.km⁻², respectively, higher than those recorded during the NEM. One-way ANOVA showed no significant difference in prawn catch rates during the NEM (F = 2.14; df = 2, 36; P = 0.13) or SEM (F=0.93; df = 2, 43; P = 0.40) in the three shallower depth zones. No prawns were harvested in depth zone 4 (40-100 m) in either season.

Bathymetric and seasonal distribution of species

Among the penaeid prawns, *F. indicus* and *M. japonicus* were restricted to depth zones 1 and 2 during both the NEM and SEM (Table 2). *M. monoceros* occurred in depth zones 1-3 during both seasons, but higher numbers were harvested in depth zones 1 and 2 during SEM. *P. monodon* and *P. semisulcatus* occurred in depth zones 1-3 during the SEM, with *P. semisulcatus* harvested in higher numbers in zones 1 and 2 during SEM (Table 2). Amongst the portunid crabs, only *Portunus sanguinolentus* was harvested in considerable numbers in both

Table 2. Seasonal distribution and abundance (individuals.Km² ± SE) of decapod crustaceans within depth zones during a) the northeast monsoon (NEM) and b) southeast monsoon (SEM) in Malindi-Ungwana Bay. Values without standard errors indicate that species appeared in only one transect; (-) indicates absence of catch.

Species	Depth Strata			
	Zone 1 (0-10 m)	Zone 2 (10-20 m)	Zone 3 (20-40 m)	Zone 4 (40-100 m)
a) NEM				
<i>Fenneropenaeus indicus</i>	11265 ± 5383	458 ± 386	-	-
<i>Marsupenaeus japonicus</i>	248	116 ± 16	-	-
<i>Metapenaeus monoceros</i>	1918 ± 953	735	367 ± 320	-
<i>Penaeus monodon</i>	1239 ± 539	199 ± 74	-	-
<i>Penaeus semisulcatus</i>	-	564 ± 544	1405 ± 577	-
<i>Portunus sanguinolentus</i>	576 ± 201	117 ± 12	-	-
<i>Thalamita crenata</i>	19	-	-	-
<i>Charybdis feriata</i>	-	-	-	-
<i>Charybdis hellerii</i>	-	-	-	-
<i>Charybdis natator</i>	-	-	-	-
<i>Charybdis smithii</i>	-	-	-	-
<i>Podophthalmus vigil</i>	-	-	-	-
<i>Portunus pelagicus</i>	-	-	-	-
<i>Scylla serrata</i>	-	-	-	-
<i>Calappa calappa</i>	40	-	-	-
<i>Calappa</i> sp.	-	21	-	-
<i>Ashtoret lunaris</i>	97	-	-	-
<i>Majidae</i> sp.	-	-	-	-
<i>Thenus orientalis</i>	20	21	20	-
<i>Panulirus ornatus</i>	-	-	-	-
b) SEM				
<i>Fenneropenaeus indicus</i>	15437 ± 4217	8208 ± 5210	-	-
<i>Marsupenaeus japonicus</i>	520 ± 294	350 ± 161	-	-
<i>Metapenaeus monoceros</i>	7003 ± 2046	7024 ± 3141	4342	-
<i>Penaeus monodon</i>	1127 ± 428.2	1799 ± 250	632	-
<i>Penaeus semisulcatus</i>	1799 ± 250	6680 ± 3144	461 ± 234.0	-
<i>Portunus sanguinolentus</i>	486 ± 168	176 ± 59	21	-
<i>Thalamita crenata</i>	20	-	-	-
<i>Charybdis feriata</i>	41 ± 1.0	41 ± 12	21	-
<i>Charybdis hellerii</i>	-	20	-	268
<i>Charybdis natator</i>	-	81	-	-
<i>Charybdis smithii</i>	-	-	-	172
<i>Podophthalmus vigil</i>	-	-	21	-
<i>Portunus pelagicus</i>	-	21	-	-
<i>Scylla serrata</i>	20	39 ± 19	-	-
<i>Calappa calappa</i>	-	-	-	-
<i>Calappa</i> sp.	-	-	167	-
<i>Ashtoret lunaris</i>	90 ± 24	98	-	-
<i>Majidae</i> sp.	20	42	-	-
<i>Thenus orientalis</i>	20 ± 0.2	54 ± 34	145	-
<i>Panulirus ornatus</i>	20	21	-	-

seasons, and mainly at shallower depths (Table 2). Amongst the Palinuridae (lobsters), *Thenus orientalis* was harvested in depth zones 1-3 in both seasons, while *Panulirus ornatus* was caught in depth zones 1 and 2 during the SEM.

MDS plots revealed that different crustacean assemblages were found in the depth zones but not the seasons (Fig. 2). Two-way crossed ANOSIM indicated that the difference in these assemblages between

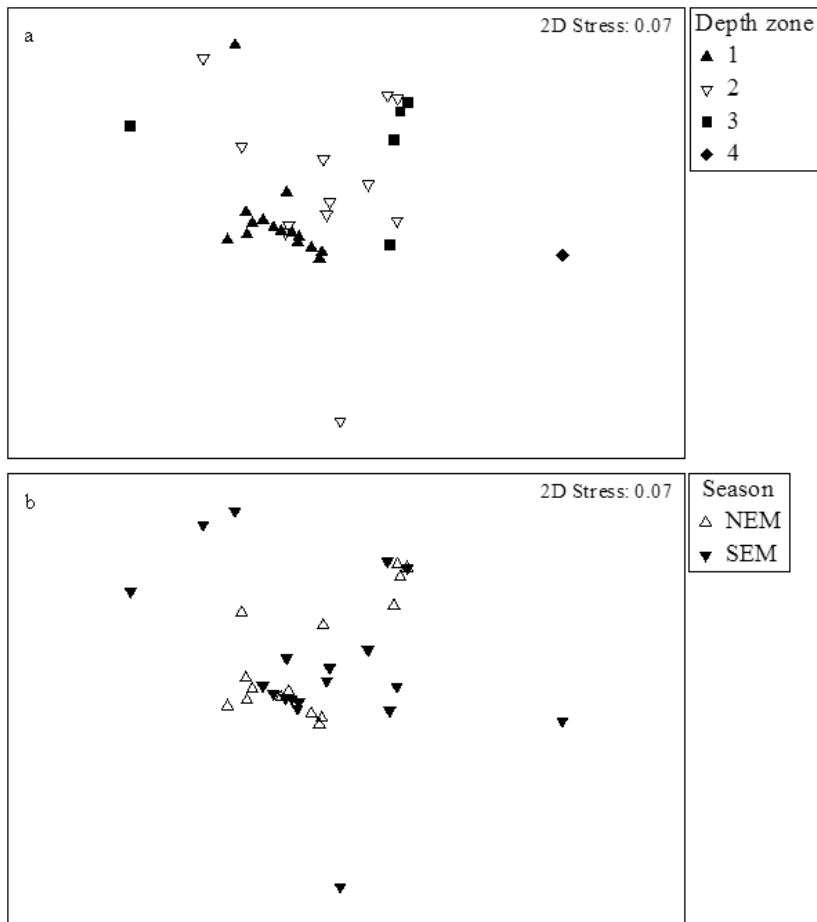


Fig. 2. Non-metric MDS plots showing decapod crustacean abundance according to a) depth zones and b) season in Malindi-Ungwana Bay, Kenya, based on combined northeast (NEM) and southeast monsoon (SEM) data.

the depth zones were significant but again not the seasons ($R = 0.410$; $p = 0.002$ and $R = -0.029$; $p = 0.598$ respectively). A pairwise ANOSIM comparison test further confirmed the significance of the differences in the assemblages between depth zones 1 and 2, and between 1 and 3 ($P < 0.05$ in all cases; Table 3). A greater abundance of *F. indicus* in depth zone 1, and *P. semisulcatus* and *Charybdis natator* in depth zone 2, was responsible for the difference between these depth zones (two-way SIMPER: Table 4). Similarly, a greater abundance of *F. indicus* in zone 1 and *P. semisulcatus* and *Calappa pelii* in depth zone 3 (two-way SIMPER: Table 5) was responsible for the difference between these depth zones.

Species richness and diversity

Depth zones 1 and 2 were more diverse than zones 3 and 4, evidenced by the results on species richness (S) and the Shannon-Wiener diversity index (H') (Fig. 3a). Also, species diversity was higher in the SEM than NEM (Fig. 3b). Results of two-way ANOVA yielded significant differences in crustacean species richness ($\text{nos. transect}^{-1}$) between depth zones ($df = 2$; $F = 3.651$; $P = 0.039$). However, the test revealed no significant difference between seasons or the interaction of depth zone and season ($df = 1$; $F = 1.872$; $P = 0.182$ and $df = 2$; $F = 0.196$; $P = 0.823$, respectively). A post hoc Fisher LSD

Table 3. Pair-wise ANOSIM comparison of differences in crustacean composition between depth zones in Malindi-Ungwana Bay based on abundance (%). Significance at $P \leq 0.05$ in bold.

Depth zones	R statistic	p-value	Possible permutations	Actual permutations	Number \geq observed
1, 2	0.232	0.013	3185325	999	12
1, 3	0.806	0.001	54450	999	0
1, 4	0.952	0.125	8	8	1
2, 3	0.083	0.271	17325	999	270
2, 4	0.638	0.111	9	9	1
3, 4	0.5	0.400	5	5	2

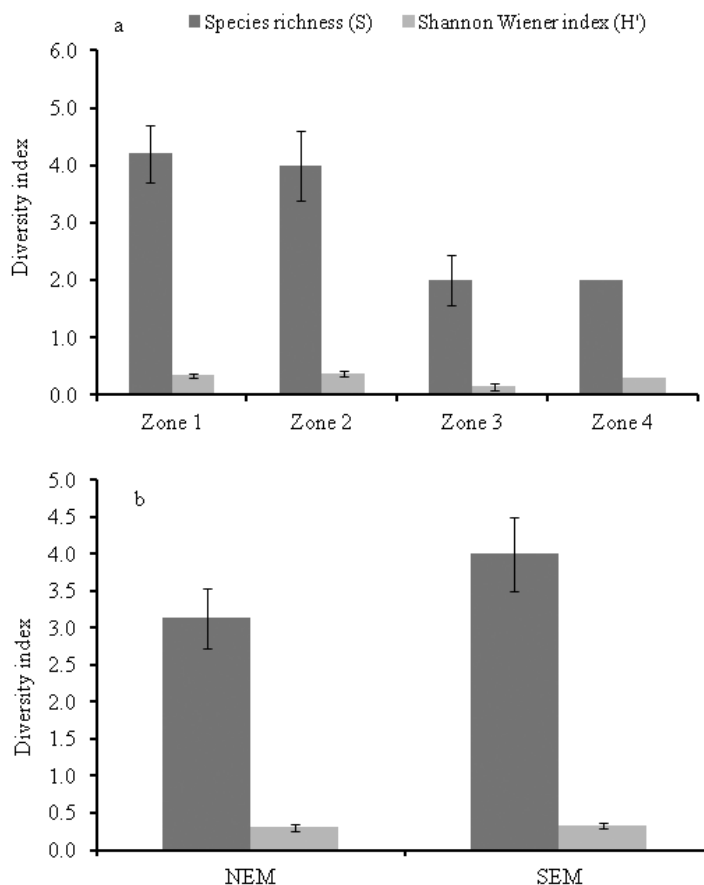


Fig. 3. Bathymetric variation of diversity measures ($S \pm SE$ and $H' \pm SE$) of decapod crustacean species according to a) depth and b) season in Malindi-Ungwana Bay, Kenya.

test confirmed that depth zone 2 differed significantly from depth zone 3 in both the NEM and SEM, and depth zone 3 differed significantly from depth zone 1 in the SEM ($P < 0.05$ in all cases).

Environmental measurements and species associations

No significant differences were encountered in temperature, salinity, dissolved inorganic nutrients (phosphates and nitrates), chlorophyll-*a* or biological oxygen demand

Table 4. Two-Way SIMPER Analysis of decapod crustacean species contribution to dissimilarity in terms of abundance (%) between depth zones 1 and 2 in Malindi-Ungwana Bay. The average dissimilarity was 65.3%, notable contributors to this being highlighted in bold.

Species	Depth zone 1 Average abundance (%)	Depth zone 2 Average abundance (%)	Average dissimilarity	Contribution (%)
<i>Fenneropeneus indicus</i>	58.11	22.89	19.35	29.63
<i>Penaeus semisulcatus</i>	1.32	29.05	14.47	22.15
<i>Metapenaeus monoceros</i>	22.26	18.65	10.89	16.67
<i>Portunus sanguinolentus</i>	7.13	6.95	7.07	10.81
<i>Penaeus monodon</i>	9.43	5.45	4.43	6.78
<i>Charybdis natator</i>	0.00	6.67	3.18	4.87

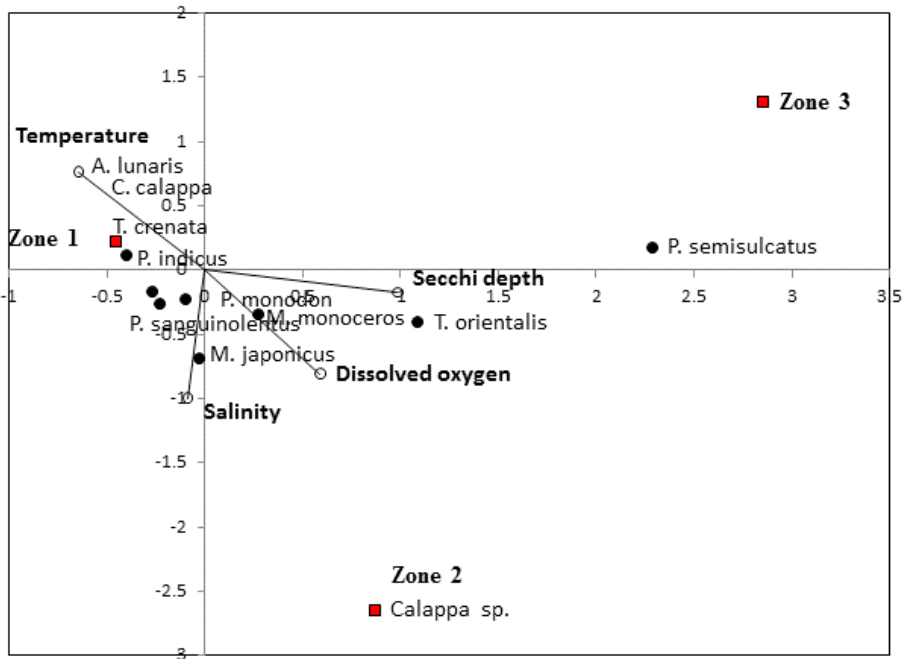


Fig. 4. Canonical Correspondence Analysis (CCA) showing the influence of physico-chemical factors on crustacean species abundance in the three shallower depth zones within Malindi-Ungwana Bay during the NEM. Species names are as shown on Table 1.

in bottom water samples collected from the different depth strata in the NEM (Table 6). Turbidity (Secchi depth) decreased with depth (0-10 m to 40-100 m); i.e. with increasing distance from the shore. Canonical Correspondence Analysis (CCA) of the influence of these environmental variables on crustacean abundance during the NEM are presented in Figure 4. Depth zones 2 and 3 are dissimilar from zone 1 on the x-axis,

the differences being attributable to water temperature in zone 1, and water clarity, dissolved oxygen and salinity in zones 2 and 3. Respective indicators of this are *Ashtoret lunaris* and *Calappa calappa* in zone 1, and *M. japonicus*, *P. monodon*, *M. monoceros*, *P. sanguinolentus* and *T. orientalis* in zone 2. The *Calappa* sp. was associated with depth zone 2 but apparently was not influenced in abundance by the selected environmental variables.

Table 5. Two-Way SIMPER Analysis of decapod crustacean species contribution to dissimilarity in terms of abundance (%) between depth zones 1 and 3 in Malindi-Ungwana Bay. The average dissimilarity was 91.7%, notable contributors to this being highlighted in bold.

Species	Depth zone 1	Depth zone 3		Contribution (%)
	Average abundance (%)	Average abundance (%)	Average dissimilarity (%)	
<i>Penaeus semisulcatus</i>	1.32	69.90	35.15	38.32
<i>Fenneropenaeus indicus</i>	58.11	0.00	28.59	31.16
<i>Metapenaeus monoceros</i>	22.26	14.28	12.25	13.35
<i>Calappa</i> sp.	0.00	10.39	4.90	5.34
<i>Penaeus monodon</i>	9.43	1.54	4.58	4.99

DISCUSSION

Higher abundances of crustaceans were harvested during this study in the oceanographically rougher SEM than the calmer NEM season using the same vessel. Although previous studies have yielded similar results (e.g. Mwatha, 2005; Munga *et al.*, 2012), the causes of this difference are unclear. Contributory factors may be higher recruitment to the fishery by the Penaeidae (the most abundant group), higher vulnerability to gear or higher environmental productivity during the SEM. Nonetheless, more studies are needed to determine the causal factors for seasonal variability in crustacean abundance in the Malindi-Ungwana Bay.

Significant differences in the relative abundance of penaeid prawns have been

found between dry and wet seasons elsewhere (Teikwa & Mgaya, 2003; de Freitas, 2011), suggesting the influence of rainfall and hence salinity on prawn catches. Rainfall is thought to initiate the migration of prawns offshore from estuaries, either by lowering salinities or simply the mechanical disturbance of run-off and of the bottom sediments (Meager *et al.*, 2003; de Freitas 2011). Juvenile prawns are known to move to offshore fisheries as a result of a reduction in inshore salinities during the rainy season (Staples and Vance, 1986). The SEM in coastal Kenya is typically a wet season and there is an increase in river discharge into the Malindi-Ungwana Bay during this season which introduces organic matter to the bay (Tychsen, 2006). This probably increases primary productivity in the bay, as it does in the Mediterranean

Table 6. Environmental variables (mean \pm SE) in the different depth zones measured during the northeast monsoon (NEM) in Malindi-Ungwana Bay, Kenya. Df = 3; p-values in bold are significant at $P < 0.05$.

Environmental variables	0-10 m	10-20 m	20-40 m	40-100 m	ANOVA	
					F	P
Temperature ($^{\circ}$ C)	27.7 \pm 0.2	27.2 \pm 0.3	27.7 \pm 0.2	27.3 \pm 0.2	1.000	0.408
Salinity	36.3 \pm 0.2	36.4 \pm 0.2	36.2 \pm 0.1	37.0 \pm 0.6	1.900	0.151
Secchi depth (m)	1.5 \pm 0.2	8.6 \pm 0.7	12.7 \pm 1.2	14.0 \pm 1.2	19.22	0.000
Dissolved Oxygen (mg/l)	5.5 \pm 0.1	5.7 \pm 0.0	5.4 \pm 0.1	5.7 \pm 0.2	3.050	0.043
Chlorophyll- <i>a</i> (μ g/l)	0.2 \pm 0.0	0.3 \pm 0.1	0.3 \pm 0.1	0.3 \pm 0.1	0.557	0.647
(Nitrate + Nitrite) - N (μ M)	1.8 \pm 0.4	1.3 \pm 0.1	1.2 \pm 0.2	0.8 \pm 0.2	1.084	0.370
Phosphates - P (μ M)	1.1 \pm 0.2	0.9 \pm 0.1	1.1 \pm 0.1	1.2 \pm 0.6	0.839	0.482
BOD _{5days} (mg/l)	4.7 \pm 0.2	4.6 \pm 0.1	4.1 \pm 0.2	3.5 \pm 0.1	5.885	0.003

(de Juan & Cartes, 2011). Such increased nutrient input into Malindi-Ungwana Bay would suggest that productivity-induced seasonal differences cause changes in crustacean abundance in the bay. This would suggest the need for seasonally structured management regulations for exploitation of its penaeid prawns.

The penaeid prawns were mostly restricted to the shallow zones 1 and 2 (0-10 and 10-20 m) during both seasons. However, *Penaeus semisulcatus*, *Metapenaeus monoceros* and *P. monodon* had a wider bathymetric distribution that was more distinct during the SEM. This may be attributable to a higher tolerance of environmental variability by these species than the other penaeids. A similarly wide bathymetric distribution associated with temperature tolerance has been reported for *P. semisulcatus* in Kuwait (Ye *et al.*, 1999), for *M. monoceros* in the Iskenderum Bay (Can *et al.*, 2004) and in the Gulf of Antalya in Turkey (Yilmaz *et al.*, 2009), and for both species in Mozambique (de Freitas, 2011).

Changes in salinity and temperature probably contributed to high species richness in Malindi-Ungwana Bay during the SEM, with a peak in diversity in depth zones 1 and 2. Increased runoff during the SEM results in temperature and salinity stratification (Papiol *et al.*, 2012) and this may contribute to higher larval survival (due to differential thermal warming of strata) and, hence, high species diversity in the bay during this season. Other studies have found that crustacean diversity is affected by temperature, salinity and prey availability within estuaries and bays (Vance *et al.*, 1985; Meager *et al.* 2003; Papiol *et al.*, 2012).

The distribution of species is affected by a number of environmental variables such as temperature and productivity, which can affect both their maximum densities and the extreme limits in their distribution (Fanelli *et al.*, 2007). Wiener and Read (1982) observed that decapod crustacean communities show definite changes in structure (species abundance and composition) with season and depth. However, Munoz *et al.* (2008) found no seasonality in decapod communities in the West Mediterranean Sea due to spatial

interactions between assemblages. The presence of most of the species in depth zones 1 and 2 in this study may have been due to high primary productivity and other favourable environmental factors such as salinity, dissolved oxygen and substratum type. However, the study did not cover some of these parameters.

Although some studies (e.g. Fanelli *et al.*, 2007; Wiener & Read, 1982) have reported the influence of both seasonality and depth on crustacean distributions, this study, like others (e.g. Munoz *et al.*, 2008) found that seasonality had less influence on the crustacean assemblages than depth. Environmental variables directly related to depth, such as temperature and dissolved oxygen, may play an important role in structuring the assemblages. Canonical Correspondence Analysis indicated that temperature influenced the composition of species in depth zone 1 during the NEM, while dissolved oxygen, water clarity and salinity had a greater effect on species composition in depth zones 2 and 3. A lack of environmental parameters during the SEM precluded determination of the effects of these environmental variables during this season.

In conclusion, the study found that penaeid prawns were the most abundant group in all the depth zones and seasons in Malindi-Ungwana Bay. All crustacean groups occurred in higher abundance in the SEM than in the NEM. This variation was attributed to seasonal changes in oceanographic conditions and crustacean behaviour. Decapod crustacean assemblage structure in the bay appeared to be more influenced by depth than seasonality, and most of the biomass was found at shallower depths. The study also revealed higher crustacean species diversity in the bay during the SEM than the NEM in all depth zones. Taking these results into consideration, it is recommended that the spatial and temporal variation in crustacean community structure should be considered when developing a fisheries management plan for the bay, and that future studies should determine the parameters that influence the seasonal abundance in populations.

Acknowledgements – We are grateful to the South West Indian Ocean Fisheries Project (SWIOFP) crustacean scientific trawl team led by Dr Edward Kimani and team members (Julius Manyala, Rashid Kaka, Thomas Mkare, Captain Joseph Mwanthi, Dickson Odongo, Boaz Orembo and Joseph Kilonzi) and the *FV Vega* crew for their good team work. We thank the SWIOFP for funding this study through a fellowship to CKN. The Kenya Marine and Fisheries Research Institute (KMFRI) provided laboratory space and logistical support. The Kenyan Permanent Secretary, Ministry of Fisheries Development, and Director of Fisheries provided work leave to CKN.

References

- Branch GM, Beckley LE, Branch ML, Griffiths CL (2007) Two Oceans: A guide to the marine life of Southern Africa. Struik Publishers, South Africa, 368 pp
- Can MF, Mazlum Y, Demirci A, Akta M (2004) The catch composition and catch per unit of swept area (CPUE) of penaeid shrimps in the bottom trawls from İskenderun Bay, Turkey. Turkish Journal of Fisheries and Aquatic Sciences 4: 87-91
- Clark KR, Warwick RM (2001) Change in marine communities: An approach to statistical analysis and interpretation, 2nd edition. PRIMER-E: Plymouth
- de Freitas AJ (2011) The Penaeoidea of South Africa IV- The Family Penaeidae: Genus *Penaeus*. Oceanographic Research Institute Investigational Report (59): 1-125
- DeGrave S, Franssen CHJM (2011) *Carideorum catalogus*: The recent species of the dendrobranchiate, stenopodidean, procarididean and caridean shrimps (Crustacea: Decapoda). Zoologische Mededelingen, Leiden 85: 195-589
- de Juan S, Cartes JE (2011) Influence of environmental factors on the dynamics of macrobenthic crustaceans on soft-bottoms of the Ebro Delta continental shelf (northwestern Mediterranean). Scientia Marina 75: 691-700
- Fanelli E, Colloca F, Ardizzone G (2007) Decapod crustacean assemblages off the West coast of central Italy (western Mediterranean). Scientia Marina 71:19-28
- FAO (1984) FAO Species identification sheets for fishery purposes, Western Indian Ocean Fishing Area 51, Volume VI
- FAO (2008) The state of world fisheries and aquaculture. FAO Fisheries and Aquaculture Department, 176 pp
- FAO (2010) The state of world fisheries and aquaculture. FAO Fisheries and Aquaculture Department, 197 pp
- FAO (2011) Yearbook of fishery statistics summary tables. FAO, 120 pp
- FAO (2012) The state of world fisheries and aquaculture. FAO Fisheries and Aquaculture Department, 209 pp
- Follesa MC, Porcu C, Gastoni A, Mulas A, Sabatini A, Cau A (2009) Community structure of bathyal decapod crustaceans off South-Eastern Sardinian deep-waters (Central-Western Mediterranean). Marine Ecology 30: 188–199
- Gillett R (2008) Global study of shrimp fisheries, FAO fisheries technical paper 475. Food and Agriculture Organization of the United Nation, Rome. 331 pp
- Groeneveld JC (2000) Stock assessment, ecology and economics as criteria for choosing between trap and trawl fisheries for spiny lobster *Palinurus delagoae*. Fisheries Research 48: 141-155

- Groeneveld JC, Melville-Smith R (1995) Spatial and temporal variability in the multispecies crustacean trawl fishery along the East Coast of South Africa and Southern Mozambique, 1988-1993. *South Africa Journal of Marine Science* 15: 123-136
- Iversen SA (1984) Kenyan marine fish resources in waters deeper than 10 m investigated by R/V "Dr. Fridtjof Nansen." The Proceedings of NORAD-Kenya seminar to review the marine fish stocks and fisheries in Kenya, Mombasa, Kenya, 13-15 March 1984
- Jackson JBC, Kirby M X, Berger WH, Bjorndal KA, Botsford LW, Bourque BJ, Bradbury RH, Cooke R, Erlandson J, Estes JA, Hughes TP, Kidwell S, Lange CB, Lenihan HS, Pandolfi JM, Peterson CH, Steneck RS, Tegner MJ, Warner RR (2001) Historical overfishing and the recent collapse of coastal ecosystems. *Science* 293: 629-637
- Jennings S, Kaiser MJ (1998) The effects of fishing on marine ecosystems. *Advances in Marine Biology* 34: 201-351
- Lavrado HP, Falcao AP, Carvalho-Cunha P, Silva SHG (2000) Composition and distribution of Decapoda in Guanamara Bay, RJ. *Nauplius* 8: 15-23
- Leibold MA (1996) A graphical model of keystone predators in food webs: Trophic regulation of abundance, incidence and diversity patterns in communities. *The American Naturalist* 147: 784-812
- Magurran AE (1988) Ecological diversity and its measurement. Chapman and Hall, London, 172 pp
- McClanahan TR (1988) Seasonality in East Africa's coastal waters. *Marine Ecology Progress Series* 44: 191-199
- Meager JJ, Vance DJ, Williamson I, Loneragen NR (2003) Seasonal variation, and environmental influences on juvenile banana prawn (*Penaeus merguensis*) abundance in a subtropical estuary (Logan River) of eastern Australia. *Estuarine Coastal and Shelf Science*, 56: 1-8
- Mueni EM (2006) A trophic model for the Ungwana Bay Fishery, Kenya: Linking fisheries to ecosystem management. MSc Thesis, UNESCO-IHE Institute for Water Education, Delft, The Netherlands
- Munga C, Fulanda B, Manyala J, Kimani E, Ohtomi J, Vanreusel A (2012) Bottom shrimp trawling impacts on species distribution and fishery dynamics: Ungwana Bay fishery Kenya before and after the 2006 trawl ban. *Fisheries Science* 78: 209-219
- Munoz JEG, Manjón-Cabeza ME, Raso JEG (2008) Decapod crustacean assemblages from littoral bottoms of the Alborán Sea (Spain, west Mediterranean Sea): Spatial and temporal variability. *Scientia Marina* 72: 437-449
- Mutagyera WB (1984) Distribution of some deep water prawn and lobster species in Kenya's waters. Proceedings of NORAD-Kenya seminar to review the marine fish stocks and fisheries in Kenya, Mombasa, Kenya, 13-15 March 1984
- Mwatha, G. (2005). Stock assessment and population dynamics of penaeid prawns in the prawn trawling grounds of Malindi-Ungwana bay: The challenges of managing the prawn fishery in Kenya. WIOMSA MARG 1 Project report no: WIOMSA/MARG-I/2005 – 06

- Ng PKL, Guinot D, Davie PJF (2008) *Systema Brachyorum: Part I. An annotated checklist of extant brachyuran crabs of the world.* Raffles Bulletin of Zoology Suppl. 17: 1-286
- Nzioka RM (1981) The evaluation of marine fisheries resources of Kenya. Proceedings of the workshop of KMFRI on aquatic resources of Kenya, 32 pp
- Papiol V, Cartes JE, Fanelli E, Maynou F (2012) Influence of environmental variables on the spatio-temporal dynamics of benthic-pelagic assemblages in the middle slope of the Balearic Basin (NW Mediterranean). *Deep-Sea Research* 61: 84-99
- Pauly D (1980) A selection of simple methods for the assessment of tropical fish stocks. FAO Fisheries Circular No. 729, 54 pp
- Richmond MD (ed) (2011) Field guide to the seashores of Eastern Africa and the Western Indian Ocean Islands. SIDA, 464 pp
- Sparre P, Venema SC (1998) Introduction to tropical fish stock assessment: Part 1 manual; FAO Fisheries Technical Paper 306/1, 376 pp
- Staples J, Vance DJ (1986) Emigration of juvenile banana prawns *Penaeus merguensis* from a mangrove estuary and recruitment to offshore areas in the wet-dry tropics of the Gulf of Carpentaria, Australia. *Marine Ecology Progress Series* 27: 239-252
- Stephenson TA (1948) The constitution of the intertidal fauna and flora of South Africa. Part 3. *Annals of the Natal Museum* 11: 207-324
- Teikwa ED, Mgaya YD (2003) Abundance and reproductive biology of the penaeid prawns of Bagamoyo coastal waters, Tanzania. *Western Indian Ocean Journal of Marine Science* 2: 117-126
- Tychsen J (ed) (2006) *KenSea. Environmental sensitivity atlas for the coastal area of Kenya.* Geological Survey of Denmark and Greenland (GEUS), Copenhagen, 76 pp
- Vance DJ, Stapples DJ, Kerr JD (1985) Factors affecting year to year variation in the catch of banana prawns in the Gulf of Carpentaria, Australia. *Journal du Conseil International pour l'Exploration de la Mer* 47: 83-97
- Wakwabi EO, Jaccarini V (1993) The distribution and abundance of planktonic penaeid larvae in Tudor creek, Mombasa, Kenya. *Hydrobiologia* 264: 185-192
- Wakwabi EO (1996) Recruitment of giant (Jumbo) tiger prawn in the backwaters of Tudor Creek, Mombasa, Kenya. IOC/UNESCO/SAREC/WIOMSA MARG I report, Contract no. SC. 298010.5
- Wierner EL, Read TH (1982) Seasonal composition and abundance of decapod crustacean assemblages from the south Atlantic bight. *USA Bulletin of Marine Science* 32: 181-206
- Worm B, Barbier EB, Beumont N, Duffy JE, Folke C, Halpern BS, Jackson JBC, Lotze HK, Micheli F, Palumbi SR, Sala E, Selkoe KA, Stachowicz J, Watson R (2006) Impacts of biodiversity loss on ocean ecosystem services. *Science* 311: 787-790

- Ye Y, Mohammed HM, Bishop JM (1999)
Depth, salinity and temperature preferences of newly recruited penaeid shrimp in Kuwait waters. *Fisheries Oceanography* 8: 128-138
- Yilmaz ZS, Ozvarol AB, Ozvarol Y (2009)
Fisheries and shrimps economy, some biological properties of the shrimp *Metapenaeus monoceros* (Fabricus 1798) in the Gulf of Antalya (Turkey). *Journal of Animal and Veterinary Advances* 8: 2530-2536.