

Original Research Article

In silico model and design of novel 5 α -reductase inhibitors for treatment of benign prostatic hyperplasia

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Abstract

Purpose: To carry out *in silico* design of 5 α -reductase inhibitors and study their potential for use in the treatment of benign prostate hyperplasia (BPH).

Methods: *In silico* molecular docking simulation-based virtual screening of NCI Diversity Set-II containing 1880 diverse ligands was performed against human 5 α -reductase for identification of potential lead molecules. The pharmacological properties and toxicity of the lead compounds were determined using software, Marvin Sketch and OSIRIS online programs, respectively.

Results: Three compounds: ZINC13099050, ZINC01569237 and ZINC17995347_2 showed potent inhibition of 5 α -reductase enzyme protein and good pharmacokinetic properties without any serious toxic effects.

Conclusion: The selected lead molecules are promising inhibitors of 5 α -reductase. They are recommended for further structure-based development of drugs for the treatment of BPH.

Keywords: Benign prostate hyperplasia, 5 α -Reductase, Docking, Virtual-screening, Ligand, Drug design

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INTRODUCTION

Benign prostate hyperplasia (BPH) is characterised by progressive enlargement of the prostate gland and obstruction of urinary flow by compression of the urethra in elderly men. It results in increased irritability of the detrusor muscle due to chronic obstruction of the urinary tract [1]. The usual symptoms observed in men suffering from BPH are weak urinary stream, urinary urgency, incomplete emptying of the urinary bladder, as well as difficulty in initiating urination, nocturia or dribbling [2]. It was

observed in a prevalence survey that more than 42 % of men above 50 years of age have experienced symptoms of BPH, and only one third of this population actually seek treatment for BPH [3,4]. Treatment is not required for patients with mild symptoms of the disease, while the patients with moderate-to-severe symptoms may initially require medical treatment or surgical therapy later [1,2]. Blockage of alpha receptor results in relaxation of the urethral sphincter which helps in emptying the urinary bladder. Vascular smooth muscle relaxation and lowering of blood pressure are enhanced by non-selective

alpha blockers, while no effect is observed on resting blood pressure by selective alpha blockers [3,4].

Catheter insertion is required for emptying the urinary bladder in acute urinary retention caused by BPH. Fortunately, only about 1 % men with BPH suffer from the problem of urinary retention annually [1,2]. The use of several categories of medication such as decongestants, general anesthesia, anti-cholinergics, opioids, benzodiazepines, and calcium channel blockers triggers urinary retention in BPH patients [3,4]. Clinical benefits are usually noticeable 2 to 4 weeks after commencement of BPH treatment. The alpha blockers used for the treatment of BPH only help in controlling the symptoms of the disease: they do not affect the pathological basis of the disease since they neither reduce the risk of urinary retention nor the need for surgical intervention [2,3]. A common side effect observed in the use of alpha blockers is orthostatic hypotension. The alpha-blocker drugs are highly contraindicated with drugs used for erectile dysfunction due to increased risk of orthostatic hypotension [1-4].

Dihydrotestosterone and other androgenic hormones stimulate prostate growth. The conversion of testosterone to dihydrotestosterone is catalysed by 5-alpha reductase, and its inhibition results in reduction in the size of the prostate. The use of 5-alpha reductase inhibitor drugs has resulted in improvement in the control of symptoms of benign prostate hyperplasia. A minimum of six months therapy is required for complete treatment of BPH. Unlike alpha blockers, 5-alpha reductase inhibitors lead to low risk of urinary retention and reduced need for surgical intervention. Thus, there is need for development of newer drugs with selective modes of action and fewer side effects, for the treatment of BPH.

EXPERIMENTAL

Sequence selection

The primary sequence of human 5 α -reductase was retrieved from the NCBI Genbank (Accession Number: AAP36172.1). The 5 α -reductase enzyme protein sequence was retrieved from Genbank in FASTA format and used for generating its two and three-dimensional models [5].

Secondary structure prediction

The secondary structure of 5 α -reductase enzyme protein was predicted with FASTA sequences

using GOR IV online server. The GOR IV server predicts protein secondary structure from the given protein sequence with respect to annotation of the protein function and its structure [6].

Model building and evaluation

The three-dimensional structure of the enzyme protein was modelled using SWISS MODEL online homology model program. The three-dimensional structure model of the protein generated with the SWISSMODEL server depends on the quality of the BLAST sequence alignment and the template structure used. The USFC-Chimera and Swiss PDB Viewer (SPDBV) softwares were used for graphical representation of the protein model, while its evaluation and validation were carried out on the RAMPAGE server through generation of Ramachandran plot [7-9].

Prediction of ligand binding site in protein

The ligand binding site of human 5 α -reductase enzyme was predicted using SiteHound-web server. The potential ligand binding site was identified with SITEHOUND algorithm through the recognition of certain regions characterized by favourable non-bonded interactions with a chemical probe [10].

Preparation of receptor protein

The tertiary model of 5 α -reductase enzyme was prepared for molecular docking simulation by addition of polar hydrogens and Kolmann's charge, and assigning of AutoDock-4 (AD4) atom types. The format of the tertiary protein model was converted from *.pdb to *.pdbqt using AutoDock [11,18].

Selection of chemical libraries

A total of 1880 diverse molecules present in the NCI "Diversity Set II molecular library were virtually screened to identify possible lead compounds. All the ligands used in virtual screening followed the Lipinsky's rule of five, i.e., ClogP < 5; H bond donor < 5; H bond acceptor < 10 and molecular weight < 500, for good pharmacokinetic properties [12].

Grid box formation

Table I shows the grid box points of the receptor protein which were utilized for all docking runs. The grid box was placed in the centre of the ligand-binding site to ensure that all the extended conformations of ligand fitted into it.

Table I: Coordinates of grid box for the three proteins

Protein	x-D	y-D	z-D	Spacing (Å)	x center	y center	z center
Model	40	40	40	0.386	-19.188	-2.153	33.528

Preparation of grid maps

The map files for different atom types in ligands and receptor viz: A, HD, F, I, Br, N, P, S, Cl, SA, NA, C and OA. were prepared by running Autogrid utility of the AutoDock suite. These map files were used in AutoDock for carrying out molecular docking simulations [13,18].

Docking parameters

Lamarckian genetic algorithm (LGA) is the primary conformational search approach in AutoDock [13]. A population was created for various possible conformations of the ligand, and mutations and exchange of conformational parameters of the ligand molecule took place to compete in a manner similar to that of biological evolution in successive generations. This was followed by the final selection of individual ligand with minimum binding energy. The individual conformation searched for local minima and local conformational space. Then, these information were passed to later generations. Semi-empirical free energy force field was used to predict the free energies associated with the binding of small molecules to macromolecular targets. The force field was based on a comprehensive thermodynamic model that allowed intramolecular energy incorporations into the predicted binding free energy by evaluating energies for the bound and unbound states. A docking parameter file was prepared for each ligand using the following parameters: GA runs = 150; maximum number of evaluations (short) = 250000; maximum number of generations = 27000; GA runs 10, and degree of gene mutation = 0.02 [18].

Virtual screening process

The files necessary for performing virtual screening process were prepared using Raccoon software. Raccoon is a graphical user interface for AutoDock-based virtual screening of ligand libraries against a macromolecule. It is used to split molecular ligand library file with multiple number of ligands so as to convert them into the pdbqt format required by AutoDock, and filter unwanted ligands using some common criteria such as Lipinski's rules, fragment-like "rule of 3" and drug-likeness. The input file is validated at every step by evaluating for the presence of non-

standard atom types and ensuring that parameters, input filenames, and grid maps have coherent formats [13].

Results analysis of virtual screening

After the screening, a script summarize_results.py from Scripps Research Institute was utilized to sort the binding energies of the docked ligands and select the best hits. The results obtained from molecular docking simulation were evaluated on the basis of hydrophobic and polar interactions between ligand and the binding residues present in the active ligand binding site of the macromolecule. Usually, the empirical range of the free binding energy should be in the range of -5 to -15 kcal/mole. The binding affinity was calculated from Eq 1.

$$K_i = e^{[(\Delta G)/(RT)]} \dots\dots\dots (1)$$

where ΔG is change in Gibbs free energy upon binding, R is universal gas constant, and T is absolute temperature.

Prediction of physicochemical properties

The various physicochemical properties of the virtually screened ligands such as molecular weight, calculated partition coefficient (ClogP), two-dimensional polar surface area (2D-PSA), number of hydrogen bond donors, and acceptor sites were calculated using software Marvin Sketch [17].

ADME profiling and toxicity prediction of lead compounds

The toxic effects of the virtually screened lead molecules were predicted using OSIRIS online program. The presence of major toxic effects such as mutagenicity, tumorigenicity, irritant effects and reproductive effects in the lead molecules were predicted on the basis of functional groups present in their chemical structures. This program was also used to calculate drug-likeness score and Drug Score of the lead molecules on the basis of their physicochemical properties [14].

RESULTS

Primary structure

The 5 α -reductase enzyme protein sequence was retrieved from the NCBI GenBank. The primary protein sequence of 5 α -reductase in FASTA format is shown in Figure 1.

```
MATATGVAERLLAALAYLQCAVGCVAFARNRQTNSVYGRHALPSHRLRVPARAAMVWQELPSLALPLYQ
YASESAPRLRSAPNCILLAMFLVHYGHRCILYFPLMRGGKPMPLLACTMAIMFCTCNGYLQSRVLSHCAV
YADDVWTDPRFLIGFGLWLTGMLNIHSDHILNLRKPGDTGYKIPRGGFEEVYTAANYFGEIMEWCGYA
LASWSVQGAFAFFTFCLSGRAKEHHEWYLRKFEYFKFRKIIIPFLF
```

Figure 1: FASTA format of 5 α -reductase enzyme protein sequence

Secondary structure model

The two-dimensional secondary structure model of 5 α -reductase enzyme protein predicted by online server GOR IV is shown in Figure 2. The secondary structure predicted by GOR IV server came out in two outputs forms, one of which is an eye-friendly protein sequence, while the other is in the form of rows present in the predicted secondary structure. The probability values for each secondary structure at each amino acid position are also indicated. The secondary structure was predicted on the basis of the highest probable compatibility with respect to predicted helix segment of a minimum of four residues, and a predicted extended segment of at least two residues.

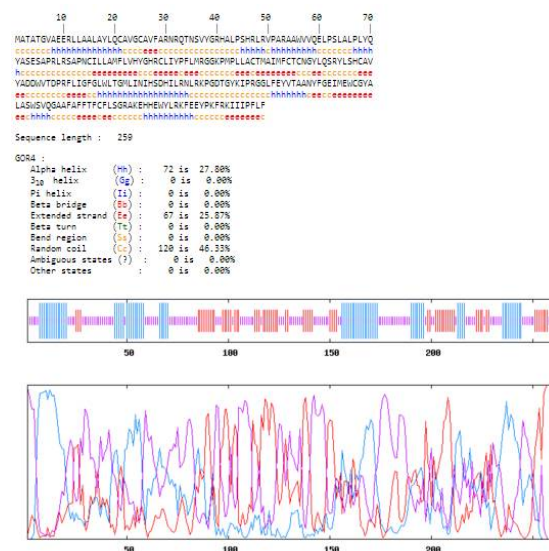


Figure 2: Two-dimensional structure of 5 α -reductase enzyme predicted with GOR IV online server. (H = helix, E = extended or beta strand, C = coil)

Tertiary protein model

The tertiary structure of 5 α -reductase enzyme protein was predicted using homology model with

templates of A chain of protein models 4QUV and 4A2N, while the protein energies of J and N chains of 3J2W model were minimized. The three-dimensional model of 5 α -reductase enzyme protein predicted with SWISSMODEL Server is shown in Figure 3.

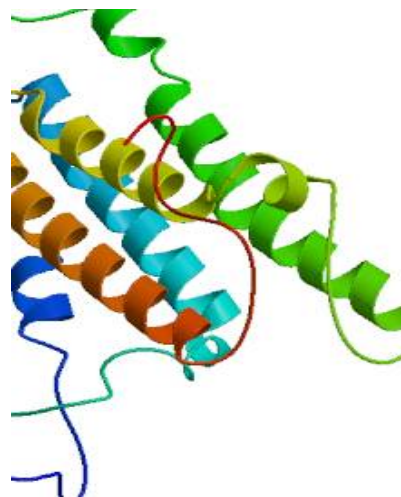


Figure 3: Three-dimensional structure of 5 α -reductase enzyme modelled using SWISSMODEL online server

Protein model evaluation

The three-dimensional model of 5 α -reductase enzyme protein was evaluated using RamPage server through generation of Ramachandran plot for the predicted model prepared using SWISSMODEL Server. The Ramachandran plot obtained with RamPage online server displayed all the torsion angles of the protein model. Usually, residue names are displayed on pointing at any location in the plot. This helps to identify the presence of any residues with bad torsion angle in the protein model.

Binding site prediction

The amino acid residues Arg176, Ile185, Pro186, Arg187, Phe191, Glu192, Tyr193, Val194, Thr195, His237 and Tyr240 were predicted as the binding residues present in the active site of the modelled human 5 α -reductase enzyme protein.

Virtual screening

The potential ligands were selected through analysis of the ligand-protein interactions for top ranking pose of each ligand, and interactions of docked compounds were visualized. The molecular docking results of top five molecules selected in AutoDock-based virtual screening of human 5 α -reductase enzyme protein with NCI Diversity set-II ligand library having 1880 small ligand molecules, are shown in Table 2.

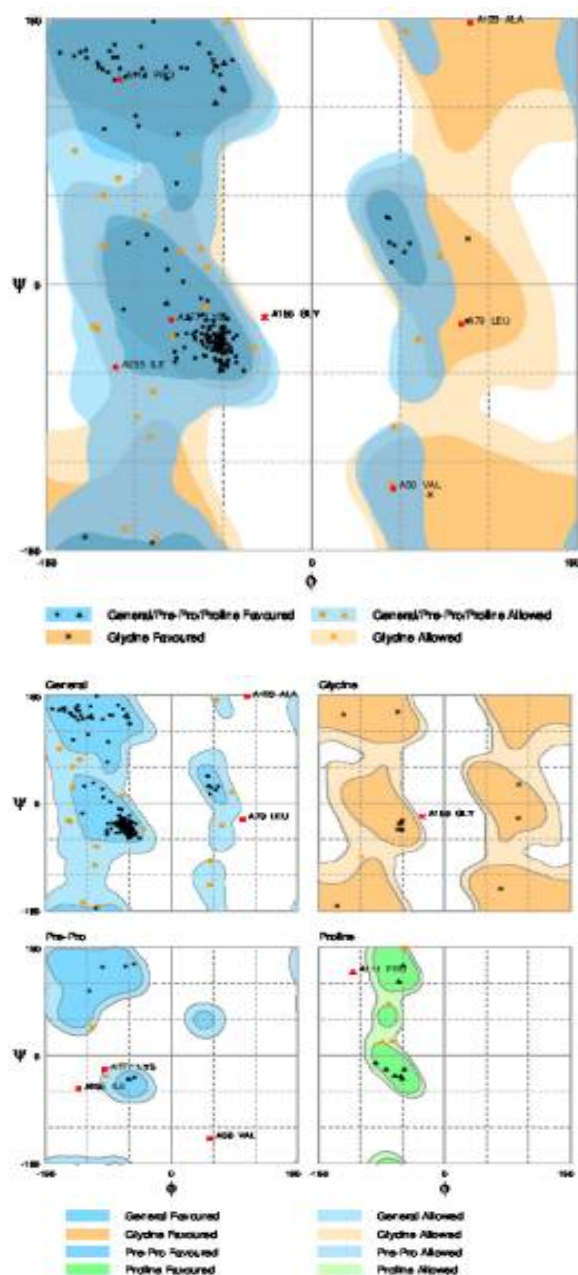


Figure 4: Ramachandran plot for 5 α -reductase enzyme protein model. The Ramachandran plot indicated that 84.9 % of residues had ϕ and ψ angles in the favoured region, while 11.8 % of residues had ϕ and ψ angles in the allowed regions

Physicochemical properties of lead compounds

The best five lead molecules selected on the basis of lowest binding energy after performing molecular docking simulation-based virtual screening of NCI Diversity set-II ligand library were used for determination/calculation of important physicochemical properties, with Marvin Sketch software and Osiris online tool. These properties were partition coefficient (cLogP), two-dimensional polar surface area (2D

PSA), molecular weight, as well as hydrogen bond donor (HBD) and hydrogen bond acceptor (HBA) sites. The physicochemical properties of the five best lead molecules for human 5 α -reductase enzyme protein are shown in Table 3.

ADME and toxicity profiles of lead compounds

The pharmacokinetic parameters of the five lead compounds were determined with respect to absorption, distribution, metabolism, excretion and toxicity, using online server Osiris Molecular Property Explorer. Three of the five selected lead compounds i.e. ZINC13099050, ZINC01569237 and ZINC17995347_2 had good pharmacokinetic profiles, with very little or no toxic effects. The five selected lead molecules were also evaluated for various parameters such as drug-likeness and Drug Score values. The three molecules, i.e., ZINC13099050, ZINC01569237 and ZINC17995347_2 had acceptable levels of drug-likeness and Drug Score values. The ADME and toxicity results of the best lead molecules obtained after virtual screening are shown in Figure 5, Figure 6, Figure 7, Figure 8 and Figure 9.

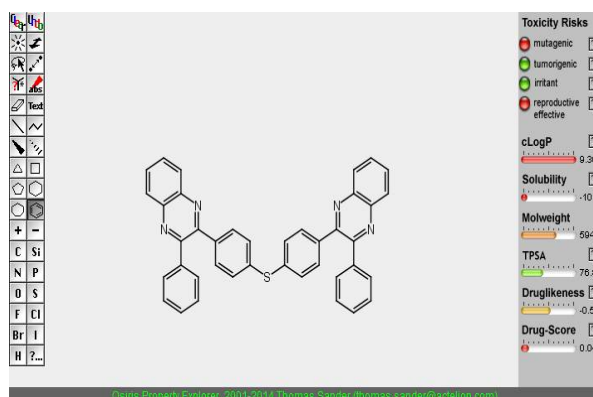
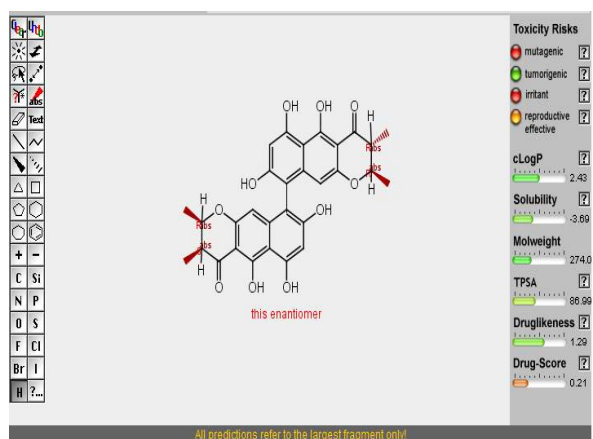
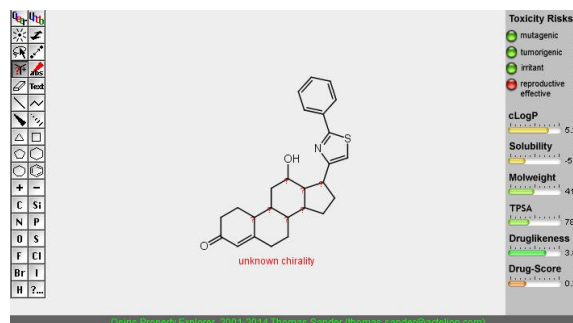
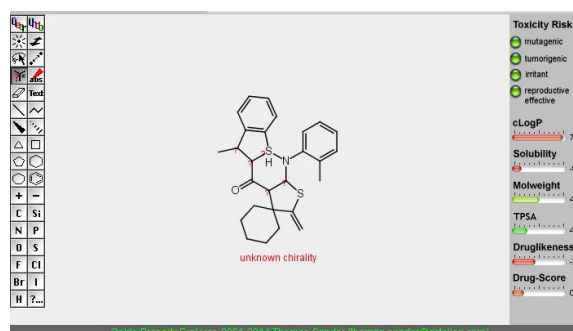
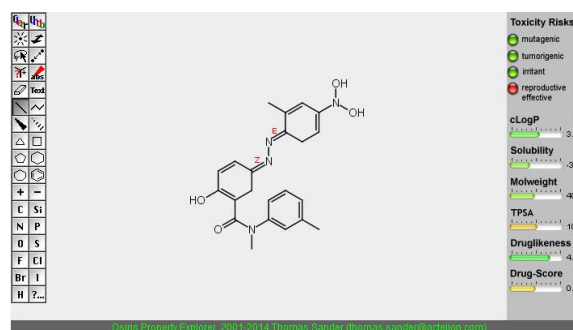
Table 2: Binding energies and affinities of top five hits from NCI Diversity Set II after virtual screening of human 5 α -reductase enzyme

Compound ID	Chemical structure	Binding energy (kcal/mole)
ZINC05492794		-11.42
ZINC17465979		-10.33
ZINC13099050		-10.32
ZINC01569237		-10.16
ZINC17995347_2		-10.11

Table 3: Physicochemical properties of 5 virtually-screened lead molecules

S/no.	Compound ID	Mol wt	ClogP	2D PSA (Å ²)	HBA	HBD
1	ZINC05492794	594	9.36	76.86	5	0
2	ZINC17465979	274	2.43	86.99	5	1
3	ZINC13099050	419	5.29	78.43	5	1
4	ZINC01569237	449	7.24	45.61	5	0
5	ZINC17995347_2	408	3.04	108.90	8	3

Three out of the five best lead molecules ZINC13099050, ZINC01569237 and ZINC17995347_2 showed promising and potent inhibition of 5 α -reductase as well as good pharmacokinetic properties, and they had no toxic effects. Moreover, the three lead molecules had very good drug-likeness and Drug Score values. The ADME and toxicity results of all the lead molecules obtained by virtual screening are shown in Figures 5 - 9. The remaining lead molecules showed poor pharmacokinetic profiles, and exhibited some major toxic effects such as mutagenicity, tumorigenicity and adverse reproductive effects.

**Figure 5:** ADME-T profiling and toxicity prediction of the virtually-screened lead molecule ZINC05492794**Figure 6:** ADME-T Profiling and toxicity prediction of the virtually-screened lead molecule ZINC17465979**Figure 7:** ADME-T Profiling and toxicity prediction of the virtually-screened lead molecule ZINC13099050**Figure 8:** ADME-T Profiling and toxicity prediction of the virtually-screened lead molecule ZINC01569237**Figure 9:** ADME-T Profiling and toxicity prediction of the virtually-screened lead molecule ZINC17995347_2

DISCUSSION

The protein sequence of human 5 α -reductase enzyme obtained from NCBI was used to predict its secondary structure on the basis of local information using two networks i.e. sequence-to-structure network and structure-to-structure network. The secondary structure model of the 5 α -reductase enzyme was predicted using

FASTA sequence with online GOR-IV program. The results revealed that random coil (46.33 %) dominated the secondary structure elements, while alpha helices (27.80 %) and extended strand (25.87 %) were present at nearly equal levels.

The three-dimensional model obtained through homology modelling was evaluated using the Ramachandran plot, which indicated that 84.9 % of residues had ϕ and ψ angles in the favoured region, while 11.8 % of residues had ϕ and ψ angles in the allowed regions. The other properties observed in the protein model such as bond angle, bond length and torsion angles were in the range of values expected for a naturally folded protein.

The physicochemical properties of the five best lead molecules for human 5 α -reductase enzyme protein showed that three of them had good enzyme inhibition properties and excellent pharmacokinetic profiles, without any major toxic effects such as mutagenicity, tumorigenicity, irritant effects or reproductive effects.

CONCLUSION

A three-dimensional model of the protein sequence of human 5 α -reductase has been successfully built with homology modelling technique using online SWISSMODEL server. The predicted model has been validated using Ramachandran plot from RamPage online server. The ligand binding site present in the modelled protein has been identified using SiteHound-web server. The identified ligand binding site was further utilized for the molecular docking simulation-based *in silico* virtual screening of 1880 diverse ligands in Diversity Set II released by the National Cancer Institute (NCI), using AutoDock. Three compounds ZINC13099050, ZINC01569237 and ZINC17995347_2 show promising results with potent inhibition of 5 α -reductase and good pharmacokinetic properties, without the presence of any serious toxic effects. Thus, these molecules are promising lead compounds for further structure-based discovery of novel drugs for the treatment of BPH.

DECLARATIONS

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directly or indirectly helped in the completion of this research.

Conflict of interest

No conflict of interest is associated with this work.

Contribution of authors

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. All authors contributed equally to this work. Jian-sheng Wang and Yi Zhang designed the frame work for performing this research. Yi Zhang and Yu-juan Zhang performed all the molecular docking simulation as well as virtual screening process. The pharmacokinetic profiling as well as toxicity evaluation is performed by Jian-sheng Wang and Long-feng Chen. The writing contribution is equally shared by the authors.

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REFERENCES

1. McConnell JD, Roehrborn CG, Bautista OM, Andriole Jr GL, Dixon CM, Kusek JW, Lopor H, McVary KT, Nyberg Jr LM, Clarke HS, Crawford ED. The long-term effect of doxazosin, finasteride, and combination therapy on the clinical progression of benign prostatic hyperplasia. *New Engl J Med* 2003; 349(25): 2387-2398.
2. Simpson RJ. Benign prostatic hyperplasia. *Br J Gen Pract.* 1997; 47(417): 235-240.
3. Paolone DR. Benign prostatic hyperplasia. *Clinics in geriatric medicine* 2010; 26(2): 223-239.
4. Dipiro JT, Talbert RL, Yee GC, Matzke GR, Wells BG, Posey LM. *Pharmacotherapy 3rd. A pathophysiologic approach. The McGraw-Hill Companies, Inc. New York;* 2008.
5. Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW. *GenBank. Nuc Acid Res.* 2013; 41(D1): D36-D42.

6. Garnier J, Gibrat JF, Robson B, Doolittle R. GOR secondary structure prediction method version IV. *Methods Enzymol* 1996; 266: 540-553.
7. Arnold K, Bordoli L, Kopp J, Schwede T. The SWISS-MODEL workspace: a web-based environment for protein structure homology modelling. *Bioinformatics*. 2006; 22(2): 195-201.
8. Bordoli L, Kiefer F, Arnold K, Benkert P, Battey J, Schwede T. Protein structure homology modeling using SWISS-MODEL workspace. *Nature protocols*. 2009; 4(1): 1-3.
9. Lovell SC, Davis IW, Arendall WB, de Bakker PI, Word JM, Prisant MG, Richardson JS, Richardson DC. Structure validation by C α geometry: ϕ , ψ and C β deviation. *Proteins: Structure, Function, and Bioinformatics*. 2003; 50(3): 437-450.
10. Hernandez M, Ghersi D, Sanchez R. SITEHOUND-web: a server for ligand binding site identification in protein structures. *Nuc Acids Res*. 2009; 37(suppl 2): W413-W416.
11. Mujwar S, Pardasani KR. Prediction of Riboswitch as a Potential Drug Target for Infectious Diseases: An In-silico Case Study of Anthrax. *J Med Imaging & Health Infor*. 2015; 5(1): 7-16.
12. Irwin JJ, Sterling T, Mysinger MM, Bolstad ES, Coleman RG. ZINC: a free tool to discover chemistry for biology. *Journal of chemical information and modeling*. 2012; 52(7): 1757-1768.
13. Forli S, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS and Olson AJ. Raccoon Autodock Preparation Tool; The Scripps Research Institute; *J Comput Chem*. 2009; 30(16); 2785-2791.
14. Thomas Sander, Actelion Pharmaceuticals Ltd., Gewerbestrasse 16, 4123 Allschwil, Switzerland, Email: thomas.sander@actelion.com.
15. Mujwar S, Pardasani KR. Riboswitch as a target for *Streptococcus pneumoniae*. *Onl J Bioinform*. 2012; 13(2): 285-313.
16. Lakshmi PT, Sathish kumar R, Annamalai A. In Silico analysis of binding efficiency of Quercetin with cancer targets. *Adv Biomed Pharm*. 2015; 2(4): 169-174.
17. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev*. 2012; 64: 4-17.
18. Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS, Olson AJ. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J Comput Chem*. 2009; 30(16): 2785-2791.
19. Yachdav G, Kloppmann E, Kajan L, Hecht M, Goldberg T, Hamp T, Hönigschmid P, Schafferhans A, Roos M, Bernhofer M, Richter L. PredictProtein—an open resource for online prediction of protein structural and functional features. *Nuc Acid Res*. 2014; 42: W337-W343.
20. Mujwar S, Pardasani KR. Prediction of riboswitch as a potential drug target and design of its optimal inhibitors for *Mycobacterium tuberculosis*. *Int J Comput Biol Drug Des* 2015; 8(4): 326-347.
21. Kaur A, Mujwar S, Adlakha N. In-silico analysis of riboswitch of *Nocardia farcinica* for design of its inhibitors and pharmacophores. *Int J Comput Biol Drug Des*. 2016; 9(3): 261-276.
22. Soni N, Pardasani KR, Mujwar S. In-silico Analysis Of Dietary Agents As Anticancer Inhibitors Of Insulin Like Growth Factor 1 Receptor (IGF1R). *Int J Pharm Pharm Sci*. 2015; 7(9): 191-196.