

## Original Research Article

# LncRNA gas5 regulates granulosa cell apoptosis and viability following radiation by X-ray through sponging miR-205-5p and Wnt/ $\beta$ -catenin signaling pathway in granulosa cell tumor of ovary

Yan Li<sup>1</sup>, Xing Ma<sup>2</sup>, Jun Li<sup>1</sup>, Saifei He<sup>2</sup>, Juhua Zhuang<sup>2</sup>, Guoyu Wang<sup>2</sup>, Ying Ye<sup>2\*</sup>, Wei Xia<sup>2\*</sup>

<sup>1</sup>Department of Obstetrics, Shanghai First Maternity and Infant Hospital, Tongji University School of Medicine, <sup>2</sup>Department of Nuclear Medicine, The Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine, PR China

\*For correspondence: **Email:** [weixia1911@163.com](mailto:weixia1911@163.com), **Tel:** +86-13277726355

Sent for review: 03 Mar 2020

Revised accepted: 26 Mar 2020

### Abstract

**Purpose:** The study explored the role of lncRNA gas5 in ovarian granulosa cells exposed to X-ray in granulosa cell tumor of ovary (GCTO).

**Methods:** Exposed the KGN cell line (KALANG, Beijing, China) to X-ray to mimic the radiotherapy for GCSO patients in vitro, cell viability was checked by CCK8 assays. RT-qPCR detected the RNA expression of apoptosis-related genes while Western Blot for biomarkers in wnt/ $\beta$ -catenin signaling. Differential expressions of lncRNA gas5 were examined after cells exposed to X ray for 0, 24, 48hs. We over expressed gas5 and assessed resultant cell viabilities, apoptosis and signaling. The sponging between gas5 and miR-205-5p was verified through Luciferase Assay. CCK8, RT-qPCR and Western Blot were applied for investigations into the correlation between miR-205-5p and cell viability and apoptosis after miR-205-5p augmentation. Similarly, the interactions between the gas5 and miR-205-5p were assessed after co-transfection of miR-205-5p mimics and oe-gas5. Last, wnt inhibitor was used to study the role of signaling pathway in KGN cells.

**Results:** Exposure of KGN to X-ray reduced cell viabilities and increased apoptosis. Gas5 had reduced expression in cells while miR-205-5p increased. Gas5 upregulation could protect the cells from apoptosis and add to the cell viability and activation of wnt/ $\beta$ -catenin signaling. lncRNA gas5 targeted miR-205-5p and miR-205-5p mimics could counteract functions of up-regulated lncRNA gas5, regulating Wnt/ $\beta$ -catenin signaling pathway. Inactivation in Wnt/ $\beta$ -catenin could suppress cell viability.

**Conclusions:** lncRNA gas5 regulated the cell apoptosis and viability after cellular radiation, which might be a potential therapeutic target to combine into radiotherapy for GCTO patients in clinical stage.

**Keywords:** Ovary, proliferation, apoptosis, lncRNA gas5, x-ray

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

Tropical Journal of Pharmaceutical Research indexed by Science Citation Index (SciSearch), Scopus, International Pharmaceutical Abstract, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

## INTRODUCTION

Ovarian follicle is one of the most important structures among female organs which plays an important role in regulating progressions of

proliferation, differentiation and apoptosis in granulosa cell tumor of ovary (GCTO). After primordial follicles are activated and grow into primary follicles and secondary follicles, granulosa cells begin to proliferate and form

layers [1]. Granular cell layer is a composition of fast-changing ovarian follicles. Before oocyte and theca cells, granulosa cells are often observed in follicular initial cells that undergo apoptosis in atretic follicles, which indicate that they could be initiator of follicular atresia [2,3]. In addition, granulosa cells can secrete several factors such as gonadal steroids, endocrine hormones and growth factors, which are important for their growth and survival [4].

X-rays are types of electromagnetic radiation which are present in the outer space and are well known for their ability to penetrate human tissues [6]. They are frequently used in the diagnosis and treatment of diseases, leading to pathological damage to tissues [5]. Among cell organelles, mitochondrion is the most sensitive to ionizing radiation [6]. Impaired mitochondrial function is an important index of oxidative damage [7]. With the advances in technology, damages from radiation, especially their effects on reproductive system, are receiving growing attention. Researches have also proved that oxidative stress is an essential pathological factor in the infertility of both males and females [8]. lncRNAs are non-coding RNAs which have more than 200 nucleotides in length [9]. These RNAs can regulate tumor growth through different mechanisms [10]. Researches have also proven that lncRNAs are important in the control of radio resistance of cancers [11]. When exposed to X-ray, expression of lncRNA-p21 increased, inhibiting  $\beta$ -catenin signaling and inducing apoptosis, leading to increase in sensitivity of CRC to radiation [12]. Up-regulated lncRNA ANRIL could increase resistance of cancers to radiation through suppressing apoptosis and inducing proliferation. Furthermore, its function in regulating tumors is mediated through negative controlling of miR-125a, which is a kind of tumor suppressor [13]. Though, noncoding RNAs played important roles in tumor cell growth and apoptosis as well as resistance to radiation. Currently, there are few research related to gas5 in GCTO.

In this work, functions of lncRNA gas5 and miR-205-5p would be measured to figure out mechanisms in regulating reproduction in female.

## EXPERIMENTAL

### Cell culture

Human ovarian granulosa cell line KGN was purchased (KALANG, Beijing, China). The KGN cells were incubated in RPMI-1640 medium contained 20% fetal bovine serum (FBS), 100  $\mu$ mol/ml penicillin and streptomycin. After

incubation, cells in log phase were collected and exposed to 10GY X-ray for 0h, 24h and 48h before harvest. The radiation was generated using Philips RT250 (Kimtron, USA)

### Cell transfection

The pcDNA3.1 plasmid (4 $\mu$ l) (Invitrogen™, USA) was applied to clone the full-length sequence lncRNA gas5, constructing a pcDNA3.1-gas5 and an empty plasmid worked as a control. miR-205-5p mimics, mimics NC, miR-205-5p inhibitor and NC inhibitor were used for transfection adopting Lipofectamine™ 3000 Transfection Reagent (Thermo Fisher, USA). Hence, we achieved differential gas5 and miR-205-5p expressions in cells that had been exposed to radiation for 24h and 48h before.

### RT-qPCR

Total RNAs were extracted from the cells using Trizol reagent (Beyotime, Shanghai, China) according to manufacturer's instructions. Then 20 $\mu$ l of TaqMan™ Reverse Transcription Reagents (Invitrogen™, USA) was applied to reverse the RNAs to cDNAs. Next, 20 $\mu$ l SYBR Green qPCR Mix (Beyotime, Shanghai, China) was used for PCR quantitation. Conditions of PCR: pre-denaturation, 95°C, 5min; denaturation, 95°C, 30s; annealing, 55°C, 30s; extension, 72°C, 30s, 40 cycles. T100™ Thermal Cycler (Bole, Shanghai, China) was used to analyze the results. The RNA primers for gas5, miR-205-5p, GAPDH, U6, Bcl-2, Bcl-xl and Caspas-3 were used. The relative expressions of these genes above were detected in cells with exposure to X-Ray for 0 (normal cells); 24h and 48h with or without transfection.  $2^{-\Delta\Delta Ct}$  methods were applied.

### CCK-8

We collected the cells after 0, 24 and 48h exposure of 10GY X-ray. Then cells were seeded into 96 well plate with  $1 \times 10^5$  cells per well and incubated at 37°C, 5% CO<sub>2</sub>. 10 $\mu$ l of CCK-8 solution was added into the plate at 24h, 48h and 72h and the optical density (OD) values of cells were quantified at 450nm using microplate reader (Thermo Fisher, USA). Similarly, the 24, 48h groups after transfection were selected to undergo cell viability assays in order to research the correlations between the gene expression and viabilities.

### Luciferase report assay

A putative binding was predicted on Starbase (<http://starbase.sysu.edu.cn/agoClipRN>)

A.php?source=lncRNA). KGN cells were co-transfected with 25ng lncRNA gas5 wt or lncRNA gas5 mut with 20ul mimics NC or miR-205-5p mimics through Lipofectamine™ 3000 Transfection Reagent (Thermo Fisher, USA). After 48 hours, the luciferase reporter assay system (Promega, USA) was used to analyze luciferase activity [14].

### Western blot

Cells exposed to X-ray were washed with PBS twice then the cells were lysed with 250ul RIPA reagent (Beyotime, Shanghai, China) for 20min. After that, the supernatant liquid was collected after centrifugation at 1000 rpm for 5min. Total proteins were quantified using BCA assay kit (Beyotime, Shanghai, China)[15]. Next, 40µg of total proteins were separated by SDS-PAGE and transferred into PVDF membranes after which the membranes were blocked with 8% non-fat milk powder at room temperature for at least 2h[16]. This was followed by the addition of primary antibodies and incubation of the membranes at 4°C overnight. Primary antibodies adopted were as follows: Anti-Wnt3a antibody (1 µg/ml, ab28472), Anti-beta Catenin antibody (1:5000, ab32572) and anti-GAPDH (1:1500; ab181602). Then membranes were rinsed and Goat Anti-Mouse IgG H&L (HRP) (1:800; ab205719) and Goat Anti-Rabbit IgG H&L (HRP) (1:800; ab205718) were added and the mixture was incubated for 1h at room temperature. Pierce™ ECL Plus Western Blotting Substrate (Thermo Fisher, USA) was used for development and gray values of proteins were measured with GAPDH as the internal reference.

### Statistical analysis

Data were displayed by mean±SD and analyzed through SPSS 19.0 (IBM, USA). All experiments were repeated three times. T-test was used to compare the two groups. P<0.05 was considered significant.

## RESULTS

### X-ray suppressed cell proliferation and promoted apoptosis

Compared to normal cells, cells that were radiated had lower level of cell viabilities (Figure

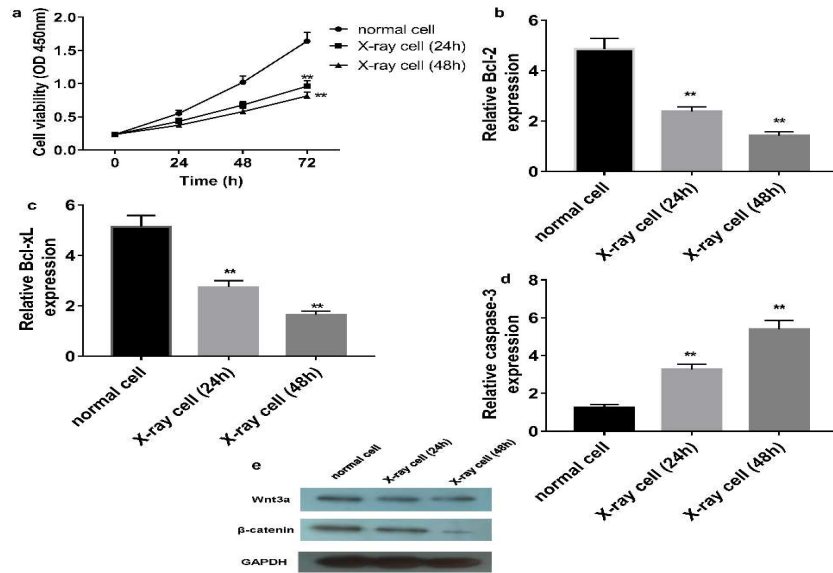
1A) which reduced gradually with time. The X-ray radiated cells had lower expressions of anti-apoptosis genes (Bcl-2 and Bcl-xL) and higher level of caspase-3, suggesting that irradiation promoted apoptosis (Figure 1B-D). Expressions of proteins examined by Western Blot indicated that X-ray could reduce expression level of both Wnt and β-catenin (Figure 1E).

### lncRNA gas5 expressed lower and promoted proliferation in X-ray exposed cells and inhibited apoptosis

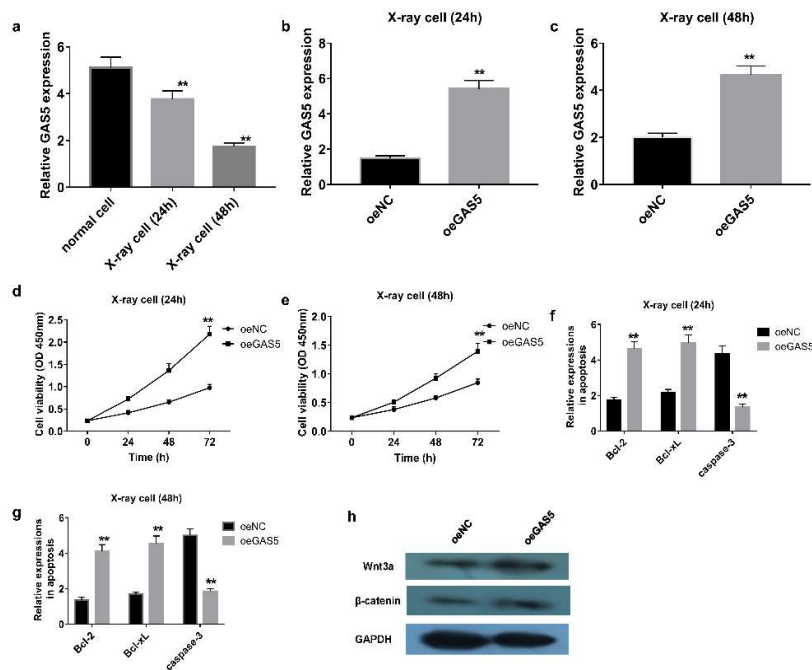
Expressions of lncRNA gas5 were detected in cells exposed to X-ray respectively for 0h, 24h and 48 hr. Compared to normal cells (0h), lncRNA gas5 was expressed lower in X-ray treated cells (Figure 2A). The expression of lncRNA gas5 decreased significantly as the exposure time increased (Figure 2B, C). We upregulated the lncRNA gas5 in cells with 24h and 48h exposure to X-Ray and CCK8 results showed that increased lncRNA gas5 could promote viabilities (Figure 2D). Moreover, cell viabilities were higher in cells that were pretreated with radiation for 24h compared to those for 48h (Figure 2E). On the other hand, overexpression of lncRNA gas5 could up regulate expressions of Bcl-2 and Bcl-xL but inhibited expression of caspase-3 (Figure 2F, G). Expressions of wnt3a and beta-catenin were increased by overexpressed lncRNA gas5 (Figure 2H).

### miR-205-5p is the target of lncRNA gas5 and promoted viability in cells with pre-exposure to radiation

Starbase v2.0 predicted that miR-205-5p targeted lncRNA gas5 (Figure 3A). Therefore, luciferase reporter assay was applied to make sure of the binding sites between lncRNA gas5 and miR-205-5p, showing that the luciferase activity only decreased significantly in the group that was co-transfected with wild-type of gas5 and miR-205-5p mimics, which indicated that wild type of miR-205-5p could directly bind lncRNA gas5 (Figure 3B). miR-205-5p expression was higher in the group with X-ray exposure than untreated cells (Figure 3C). Thereafter, we knocked down miR-205-5p by transfecting miR-205-5p inhibitor into the cells with pre-exposure to radiation for 24h and 48h.



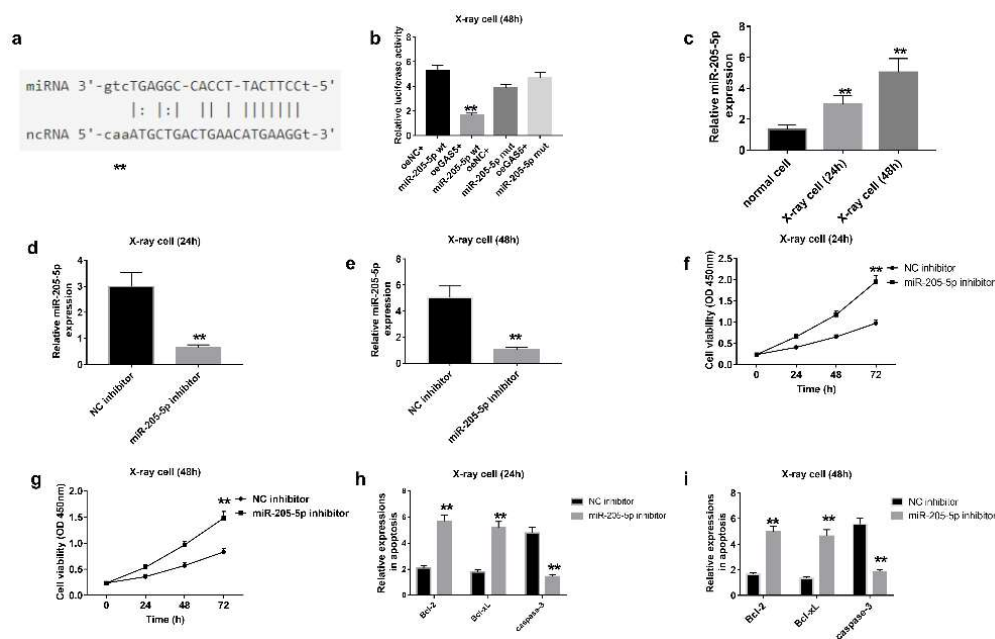
**Figure 1:** X-ray suppressed proliferation and accelerated apoptosis of KGN cells. A. Cell viabilities were detected through CCK-8,  $p < 0.05$ . B, C, D. RT-qPCR was applied to measure expressions of factors related to apoptosis,  $p < 0.05$ . E. western blot was used to evaluated expressions of proteins in Wnt/ $\beta$ -catenin signaling pathway,  $p < 0.05$ .



**Figure 2:** LncRNA gas5 expressed lower in X-ray treated cells with promoting proliferation and repressing apoptosis. A. Expressions of gas5 were analyzed through RT-qPCR,  $p < 0.05$ . B, C. overexpressed gas5 expressions were detected by RT-qPCR,  $P < 0.05$ . D, E. CCK-8 was applied to measure cell viabilities,  $p < 0.05$ . F, G. Apoptosis was validated through RT-qPCR,  $p < 0.05$ . H. Expressions of proteins were evaluated by western blot,  $p < 0.05$

Lower expression of miR-205-5p was detected (Figure 3D, E). Correspondingly, cell viabilities were measured, which disclosed that in cells with pre-exposure to radiation for both 24h and 48h groups, inhibited miR-205-5p could promote

viabilities of cells (Figure 3F, G). As for apoptosis, inhibition of miR-205-5p could activate Bcl-2 and Bcl-xL and silence caspase-3, signifying deterred cellular apoptosis (Figure 3H, I).



**Figure 3:** MiR-205-5p was the direct target of gas 5 and promoted proliferation A. Starbase v2.0 was for finding predicted binding sites of gas5 and miRNAs. B. Luciferase report assay was used to determine binding conditions between gas5 and miR-205-5p,  $p < 0.05$ . C. Expressions of mi-205-5p was detected by RT-qPCR,  $p < 0.05$ . D, E. RT-qPCR was used to measure expressions of transfected miR-205-5p,  $p < 0.05$ . F, G. Cell viabilities with inhibited miR-205-5p and NC was validated by CCK-8,  $P < 0.05$ . H, I. RT-qPCR was for evaluating expressions of factors in apoptosis,  $p < 0.05$

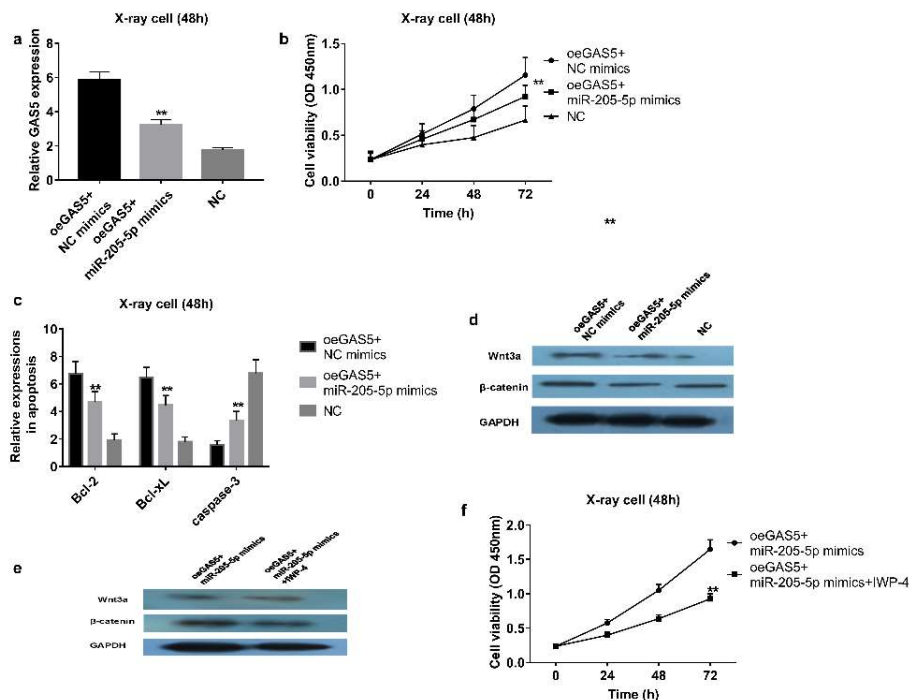
### LncRNA gas5 regulated cell progression by targeting miR-205-5p through Wnt/ $\beta$ -catenin signaling pathway

After assessment of the functions of lncRNA gas5 and miR-205-5p individually, interplays in between were further studied. First, when miR-205-5p was up-regulated, expression of lncRNA gas5 decreased (Figure 4A). lncRNA gas5 increased viabilities of cells while overexpressed miR-205-5p could partly reverse the promotive effect generated by upregulated gas5 (Figure 4B). After that, apoptosis was detected through expressions of Bcl-2, Bcl-xL and caspase-3. Up-regulated miR-205-5p could restore Bcl-2 and Bcl-xL and reverse the suppression of caspase-3 induced by lncRNA gas5 upregulation (Figure 4C). Moreover, expressions of Wnt 3a and  $\beta$ -catenin were both increased by upregulated lncRNA gas5 while miR-205-5p mimics could offset the boost (Figure 4D). In order to make sure that Wnt/ $\beta$ -catenin signaling pathway played a part in the cellular functions, IWP-4, a Wnt/ $\beta$ -catenin signaling pathway inhibitor was introduced, which could silence Wnt 3a and  $\beta$ -catenin so as to investigate the changes in cell viabilities brought by the inactivation of Wnt/ $\beta$ -catenin signaling (Figure 4E, F). The CCK8 results unveiled the reduced cell viabilities after the IWP-4 treatment, supporting the previous hypothesis that gas5 might regulate cell

progression by targeting miR-205-5p through Wnt/ $\beta$ -catenin signaling pathway

### DISCUSSION

The present research mainly dedicated to the exploration of the role of lncRNA gas5 in granulosa cell tumor of ovary after X-ray treatment in vitro. At first, the GCTO-like cell line KGN was acquired and treated with X-ray for 0, 24 and 48 hours to mimic the radiotherapy for GCTO patients in vitro. Cell viability decreased and apoptosis increased when the treated time increased. We found gas5 was silenced when the cells were radiated and upregulation of gas5 could promote the cell viability and inhibit apoptosis, suggesting that gas5 might deter the pro-apoptosis effect of X-ray. In addition, bioinformatics and Luciferase assays confirmed the targeted gene of lncRNA gas5, miR-205-5p. Therefore, we studied the role of miR-205-5p in cellular functions singly first and discovered that miR-205-5p expression increased as radiation was pre-treated; miR-205-5p inhibitor promoted the cellular viabilities and deterred apoptosis. Then we went further into interactions between gas5 and miR-205-5p in modulations of the radiation-exposed cell functions and came to a finding that miR-205-5p mimics could partially reverse the cellular changes in viabilities and apoptosis induced by gas5 upregulation.



**Figure 4:** LncRNA gas5 regulated proliferation and apoptosis of X-ray treated cells through Wnt/ $\beta$ -catenin signaling pathway A. Relative expressions of gas5 was measured through RT-qPCR,  $p < 0.05$ . B. Cell viabilities were detected through CCK-8,  $P < 0.05$ . C. Apoptosis were evaluated by RT-qPCR,  $p < 0.05$ . D, E. Western blot was for analyzing expressions of proteins,  $P < 0.05$ . F: CCK-8 was used to measure cell viability,  $p < 0.05$

Granulosa cell tumors of ovary (GCTO) was a type of gynecological oncology which occurs rare [17]. The treatments for GCTO included radiotherapy[18]. Recent years, researches discovered that RNAs were involved with the GCTO progression. For instance, the presence of TRET C228T mutation tended to appear in recurrent tumors instead of primary ones in GCTO[19]. Silencing FOXO/PTEN in Granulosa cells were correlated with the progression of GCTO[20]. There have been some previous researches concerning lncRNAs and GCTO. It was disclosed that lnc LET could induce cell apoptosis and inhibit viabilities in GCTO-like cells; lncRNA SRA might promote cell growth and suppress viabilities in GCTO mice and lncRNA MEG3 promoted the cell viability and inhibit viabilities in GCTO mouse cells via p53/p66 signaling pathway[21-23].

LncRNA gas5 was widely reported to participate in the regulation of ovarian disorders[24, 25]. Yet, there is no literature relating to gas5 and GCTO. Therefore, we surveyed the role of gas5 in GCTO.

LncRNAs could be regulated by radiation and showed radioresistance in many kinds of cancer

cells[26, 27]. Researches have also discovered that lncRNAs transcripts were changed because of ultraviolet rays or ionizing radiation in peripheral blood mononuclear cell (PBMC)[28], thymocyte[29] and melanocyte[30]. LncRNA gas5 was discovered in various cancers to play a regulator in radiotherapy efficacy[31-35]. Therefore, we researched the role of gas5 in GCTO regarding radiation *in vitro*.

Wnt signaling pathway was first verified in 1982 and Wnt1 was the first gene of Wnt family[36], which is an important signaling pathway in regulating cell progression. Previous researches have proved that Wnt/ $\beta$ -catenin signaling pathway could activate expression of survivin, an anti-apoptosis gene, to increase radioresistance of progenitor cells of mammary glands[37]. We found in this research that wnt/ $\beta$ -catenin signaling changed as X-ray exposure, gas5 or miR-205-5p adjusted, giving a hint that the signaling pathway might be involved with the regulatory mechanism beneath the lncRNA gas5. Hence, we introduced the signaling pathway inhibitor, IWP-4, to silence the signaling and observed the corresponding changes in cell viability and results showed that resultant cellular viabilities were inhibited by IWP-4, indicating that

the lncRNA gas5/miR-205-5p axis might regulate the cell viability and apoptosis via wnt/ $\beta$ -catenin signaling in GCTO-radiation cellular model.

## CONCLUSION

lncRNA gas5 was expressed lower in GCTO-radiation cellular model and upregulation inhibited cell apoptosis and promoted cell proliferation by suppressing miR-205-5p and activating Wnt/ $\beta$ -catenin signaling, indicating that it could be a potential target to be considered combining with radiotherapy in GCTO. However, the present study requires more substantial animal and clinical studies for further validation.

## DECLARATIONS

### Acknowledgement

This research is funded by Key Specialty Construction Project of Pudong Health and Family Planning Commission of Shanghai (no. PWZzk2017-21) and also Science and Technology Development Fund of Shanghai Pudong New Area (No. PKJ2017-Y13)

### Conflict of interest

No conflict of interest is associated with this work.

### Contribution of authors

We declare that this work was done by the author(s) named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. All authors read and approved the manuscript for publication. Zaixue Jiang performed most experiments and took part in drafting the manuscript. Xingui Tian designed the work, analysed the data and participated in drafting the manuscript. Xiaomei Lu helped analyse the data. Baimao Zhong designed and supervised the work, and edited the final version of the manuscript. All authors read and approved the final version of the manuscript.

### Open Access

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative

(<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

## REFERENCES

1. Young, J.M. and A.S. McNeilly, *Theca: the forgotten cell of the ovarian follicle. Reproduction*, 2010; 140(4): 489-504.
2. Tilly J.L., Kowalski KI, Johnson AL, Hsueh AJ. *Involvement of apoptosis in ovarian follicular atresia and postovulatory regression. Endocrinology*, 1991; 129(5): 2799-801.
3. Inoue N, Matsuda F, Goto Y, Manabe N. *Role of cell-death ligand-receptor system of granulosa cells in selective follicular atresia in porcine ovary. J Reprod Dev*, 2011. 57(2): 169-75.
4. Matsuda F, Inoue N, Manabe N, Ohkura S. *Follicular growth and atresia in mammalian ovaries: regulation by survival and death of granulosa cells. J Reprod Dev*, 2012. 58(1): 44-50.
5. Goldschmidt, H. and W.K. Sherwin, *Reactions to ionizing radiation. J Am Acad Dermatol*, 1980. 3(6): p. 551-79.
6. YeC, Sun W. [Research progress in effect of electromagnetic radiation on mitochondrial function]. *Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi*, 2014. 32(2): 153-7.
7. Vayssier-Taussat M, Kreps SE, Adrie C, Dall'Ava J, Christiani D, Polla BS. *Mitochondrial membrane potential: a novel biomarker of oxidative environmental stress. Environ Health Perspect*, 2002. 110(3): 301-5.
8. Agarwal A, Aponte-Mellado A, Premkumar BJ, Shaman A, Gupta S. *The effects of oxidative stress on female reproduction: a review. Reprod Biol Endocrinol*, 2012. 10: 49.
9. Ernst C, Morton CC. *Identification and function of long non-coding RNA. Front Cell Neurosci*, 2013. 7: 168.
10. Gupta RA, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ et al. *Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. Nature*, 2010. 464(7291): 1071-6.
11. Skvortsov S, Jimenez CR, Knol JC, Eichberger P, Schiestl B, Debbage P, Skvortsova I, Lukas P. *Radioresistant head and neck squamous cell carcinoma cells: intracellular signaling, putative biomarkers for tumor recurrences and possible therapeutic targets. Radiother Oncol*, 2011. 101(1): 177-82.
12. Wang G, Li Z, Zhao Q, Zhu Y, Zhao C, Li X, Ma Z, Li X, Zhang Y. *LincRNA-p21 enhances the sensitivity of radiotherapy for human colorectal cancer by targeting the Wnt/ $\beta$ -catenin signaling pathway. Oncol Rep*, 2014. 31(4): 1839-45.
13. HuX, Jiang H, Jiang X. *Downregulation of lncRNA ANRIL inhibits proliferation, induces apoptosis, and enhances radiosensitivity in nasopharyngeal carcinoma cells through regulating miR-125a. Cancer Biol Ther*, 2017.

- 18(5): 331-338.
14. Hu HB, Chen Q, Ding SQ. LncRNA LINC01116 competes with miR-145 for the regulation of ESR1 expression in breast cancer. *Eur Rev Med Pharmacol Sci*, 2018. 22(7): 1987-1993.
  15. AnastasJN, Moon RT. WNT signalling pathways as therapeutic targets in cancer. *Nat Rev Cancer*, 2013. 13(1): 11-26.
  16. Zhang Y-X, Yuan J, Gao Z-M, Zhang Z-G. LncRNA TUC338 promotes invasion of lung cancer by activating MAPK pathway. *Eur Rev Med Pharmacol Sci*, 2018. 22(2): 443-449.
  17. Hillman RT, Celestino J, Terranova C, Beird HC, Gumbs C, Little L et al. KMT2D/MLL2 inactivation is associated with recurrence in adult-type granulosa cell tumors of the ovary. *Nat Commun*, 2018. 9(1): 2496.
  18. Wolf JK, Mullen J, Eife PJ, Burke TW, Levenback C, Gershenson DM. Radiation treatment of advanced or recurrent granulosa cell tumor of the ovary. *Gynecol Oncol*, 1999. 73(1): 35-41.
  19. Pilsworth JA, Cochrane DR, Xia Z, Aubert G, Färkkilä AEM, Horlings HM et al. TERT promoter mutation in adult granulosa cell tumor of the ovary. *Mod Pathol*, 2018. 31(7): p. 1107-1115.
  20. Liu, Z., et al., FOXO1/3 and PTEN Depletion in Granulosa Cells Promotes Ovarian Granulosa Cell Tumor Development. *Mol Endocrinol*, 2015. 29(7): 1006-24.
  21. Han Q, Zhang W, Meng J, Ma J, Li A. LncRNA-LET inhibits cell viability, migration and EMT while induces apoptosis by up-regulation of TIMP2 in human granulosa-like tumor cell line KGN. *Biomed Pharmacother*, 2018. 100: 250-256.
  22. Li Y, Wang H, Zhou D, Shuang T, Zhao H, Chen B. Up-Regulation of Long Noncoding RNA SRA Promotes Cell Growth, Inhibits Cell Apoptosis, and Induces Secretion of Estradiol and Progesterone in Ovarian Granular Cells of Mice. *Med Sci Monit*, 2018. 24: 2384-2390.
  23. Xiong Y, Liu T, Wang S, Chi H, Chen C, Zheng J. Cyclophosphamide promotes the proliferation inhibition of mouse ovarian granulosa cells and premature ovarian failure by activating the lncRNA-Meg3-p53-p66Shc pathway. *Gene*, 2017. 596: 1-8.
  24. Lin H, Xing W, Li Y, Xie Y, Tang X, Zhang Q. Downregulation of serum long noncoding RNA GAS5 may contribute to insulin resistance in PCOS patients. *Gynecol Endocrinol*, 2018. 34(9): 784-788.
  25. Long X, Song K, Hu H, Tian Q, Wang W, Dong Q, Yin X, Di W. Long non-coding RNA GAS5 inhibits DDP-resistance and tumor progression of epithelial ovarian cancer via GAS5-E2F4-PARP1-MAPK axis. *J Exp Clin Cancer Res*, 2019. 38(1): 345.
  26. Yang X, Xu H-X, Xu X-H, Ru G, Liu W, Zhu J-J. Knockdown of long non-coding RNA HOTAIR inhibits proliferation and invasiveness and improves radiosensitivity in colorectal cancer. *Oncol Rep*, 2016. 35(1): 479-87.
  27. Brodie S, Lee HK, Jiang W, Cazacu S, Xiang C. The novel long non-coding RNA TALNEC2, regulates tumor cell growth and the stemness and radiation response of glioma stem cells. *Oncotarget*, 2017; 8(19): 31785-31801.
  28. Beer L, Nemeč L, Wagner T, Ristl R, Altenburger LM, Ankersmit HJ, Mildner M. Ionizing radiation regulates long non-coding RNAs in human peripheral blood mononuclear cells. *J Radiat Res*, 2017; 58(2): 201-209.
  29. Gao H, Dong Z, Wei W, Shao L, Jin L, Lv Y, Zhao G, Jin S. Integrative analysis for the role of long non-coding RNAs in radiation-induced mouse thymocytes responses. *Acta Biochim Biophys Sin (Shanghai)*, 2017; 49(1): 51-61.
  30. Zeng Q, Wang Q, Chen X, Xia K, Tang J, Zhou X. Analysis of lncRNAs expression in UVB-induced stress responses of melanocytes. *J Dermatol Sci*, 2016; 81(1): 53-60.
  31. Chen L, Ren P, Zhang Y, Gong B, Yu D, Sun X. Long noncoding RNA GAS5 increases the radiosensitivity of A549 cells through interaction with the miR21/PTEN/Akt axis. *Oncol Rep*, 2020; 43(3): 897-907.
  32. Gao J, Liu L, Li G, Cai M, Tan C, Han X, Han L. LncRNA GAS5 confers the radio sensitivity of cervical cancer cells via regulating miR-106b/IER3 axis. *Int J Biol Macromol*, 2019; 126:994-1001.
  33. Lin J, Liu Z, Liao S, Li E, Wu X, Zeng W. Elevation of long non-coding RNA GAS5 and knockdown of microRNA-21 up-regulate RECK expression to enhance esophageal squamous cell carcinoma cell radiosensitivity after radiotherapy. *Genomics*, 2020; 112(3): 2173-2185.
  34. Xue Y, Ni T, Jiang Y, Li Y. Long Noncoding RNA GAS5 Inhibits Tumorigenesis and Enhances Radiosensitivity by Suppressing miR-135b Expression in Non-Small Cell Lung Cancer. *Oncol Res*, 2017; 25(8): 1305-1316.
  35. Yang J, Hao T, Sun J, Wei P, Zhang H. Long noncoding RNA GAS5 modulates alpha-Solanine-induced radiosensitivity by negatively regulating miR-18a in human prostate cancer cells. *Biomed Pharmacother*, 2019; 112: 108656.
  36. Nusse R, Varmus HE. Many tumors induced by the mouse mammary tumor virus contain a provirus integrated in the same region of the host genome. *Cell*, 1982; 31(1): 99-109.
  37. Chen MS, Woodward WA, Behbod F, Peddibhotla S, Alfaro MP, Buchholz TA, Rosen JM. Wnt/beta-catenin mediates radiation resistance of Sca1+ progenitors in an immortalized mammary gland cell line. *J Cell Sci*, 2007; 120(Pt 3): 468-77.