

Research Article

Comparative *in-vitro* studies on the efficacy of ivermectin against gastrointestinal sheep nematode

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Abstract

Purpose: This study was designed to evaluate the relative efficacy of various brands of ivermectin injection available for use in clinical veterinary practice in Nigeria.

Method: Ivermectin injections were evaluated by a larval development assay (LDVA), using the larvae of *Strongyles* (predominantly *Haemonchus contortus*) of sheep. The effect of standard solutions of the drug from the various brands on the transformation of L₁ to L₃ and survival of L₃ larvae was used to assess bioactivity. The 50% lethal concentration (LC₅₀) was determined from regression line obtained by probit transformation of the biological data. The LC₅₀ values for each of the brands were compared with that of the innovator brand (Ivomec Super[®]) for any significant difference.

Results: The LC₅₀ values obtained for the five brands varied widely. It ranges from 1.1±0.17 ng/ml for the innovator brand to 2.3±0.3, 3.0±0.3, 8.0±0.2 and 17.0±0.3 ng/ml for the other four brands. The biological assays performed on each of the five brands were of comparable precision. LC₅₀ for Ivomec super[®] was significantly different from those of the other four brands (Student's t test, p < 0.01).

Conclusion: The bioactivities of brands of ivermectin injections available in Nigeria are significantly different. This is a probable reason for the varied treatment response to various brands of ivermectin injection in veterinary practice in Nigeria. This justifies the need for drug regulatory bodies in Nigeria to ensure that ivermectin injections registered for use in Nigeria meets approved standards before the drugs are allowed to be imported into the country.

Key words: Bioactivity, ivermectin, sheep nematode

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Introduction

Ivermectin is an analogue of avermectin, which belongs to a family of 16-membered macrocyclic lactones. It is known to increase membrane permeability to chloride ions possibly as a result of their interaction with chloride ion channels¹. Its broad spectrum of activity and wide safety margin has made it the drug of choice for nematode and arthropod parasitism in cattle, sheep, goat, swine and horses². However, farmers and clinicians in Nigeria have observed wide variation in treatment response with different brands of Ivermectin injection against endo- and ecto-parasites of animals. This often results in poor helminth control and sometimes treatment failure, with the consequent economic loss.

The availability of different formulations of the injection from various manufacturers warrants the comparative evaluation of bioactivity of the various brands in Nigerian market. In this paper therefore, we describe the comparative anthelmintic activity of various brands of ivermectin injection against gastrointestinal nematode of sheep, using a larval development assay.

Materials and Methods

Nutritive medium

The nutritive medium was as described by Hubert and Kerboeuf⁴. It is composed of Earle's balance salt solution plus yeast extract diluted in saline solution (1 gm yeast extract in 90 ml of saline solution).

Preparation of ivermectin solution

The ivermectin injection brands investigated are Iverject[®] (Batch No. 349590, Special T. Product, UK), Ivomec Super[®] (Batch No. HN 00270, MSD, Meril, Netherlands), Kepromec[®] (Batch No. 0IJ15, Kepro B.V, Holland), Ivojec (Batch No. 2001.12, Sinochem Ningbo, China), and Ivermectin[®] (Batch No. 2531, Anupco, UK). Appropriate

aliquots of these drugs were taken from different stock aqueous solutions (1000 ng/ml, 100 ng/ml and 10 ng/ml) of drug prepared from the injection (apart from Ivomec super[®] which contains 10% clorsulon in addition to the ivermectin labeled content, the other brands of ivermectin injections contain 1% w/v solution of ivermectin in propylene glycol). The final concentration in the assay tubes were over the range of 0.2 – 60.0 ng/ml of ivermectin.

Nematode Egg Recovery Technique

The technique used was that previously described by Prichard³. Briefly, 10-15 g of faeces of sheep was suspended in water and cleared of organic debris by filtration through sieves (1mm and 100 μ m) and the eggs were collected on a 20 μ m sieve. These eggs were further cleared from organic debris by centrifugation in magnesium sulphate (density 1.10) for 5 min at 1000 *g*. The supernatant was filtered through 100 μ m and 60 μ m sieves and the eggs were washed in water and collected on a 20 μ m sieve.

The concentration of eggs was estimated in 50 μ l samples and adjusted to 100 – 120 eggs/ml. The egg suspension was diluted with the filtrate from the first step of egg extraction (described above) in order to provide rumen bacteria necessary for nematode larvae development. To avoid the proliferation of fungi, 5 μ g of amphotericin B was added per ml of egg suspension.

Larval development viability assay (LDVA)

Using a 5-ml test tube, 150 μ l of nutritive medium was added to 50 μ l of egg suspension containing approximately 100 eggs. Three replicates per concentration or water (control) were made. The tubes were covered and put in an incubator at 29 °C, for 48 hr to allow development of the parasite to first stage larvae. Appropriate aliquots of the drug was then added. The third stage larvae

were obtained seven days later. At this time, the parasite was counted by separating the larvae into two classes, living third stage larvae (L₃) and dead larvae. Larval development parameter is given by:

$$\frac{\text{No. of Living L}_3/\text{total no. of nematode in tubes with anthelmintic}}{\text{divided by no. of living L}_3/\text{total number of nematode in control tube (water)}}$$

Statistical analysis

Data from LDVA were transformed by probit transformation and plotted against the logarithm of concentration⁷. Probit transformation was performed to transform a typical sigmoid dose-response curve to a linear function. The concentration required to kill 50% of L₃ (LC₅₀) was calculated from this linear regression scale.

Relative bioactivity of the various brands was determined by comparing the mean LC₅₀ of each of the other brands with the LC₅₀ for Ivomec Supe using a Student *t*-test (2-tailed) at 95% confidence interval. Probability (*p*) values < 0.01 was considered to be significant⁵.

Results and Discussion

A linear relationship was observed between larval survival and drug concentration for the various brands of ivermectin injection (Figure 1). The calculated LC₅₀ of Iverject, Ivomec super, Kepromec, Ivojec and Ivermectin were 2.3 ng/ml, 1.1 ng/ml, 8.0 ng/ml, 17.0 ng/ml and 3.0 ng/ml, respectively. The result of the comparison of the bioactivity of the various injection brands with Ivomec super is as shown in the Table 1.

The results show variation in the bioactivity (LC₅₀) of the brands of Ivermectin injection investigated, ranging from 1.1 ng/ml to 17 ng/ml. In a previous work, Ademola⁶ reported varying LC₅₀ (0.79-4.0 ng/ml) for Ivomec MSD against strongyles of sheep from different farms in Oyo State, Nigeria. In

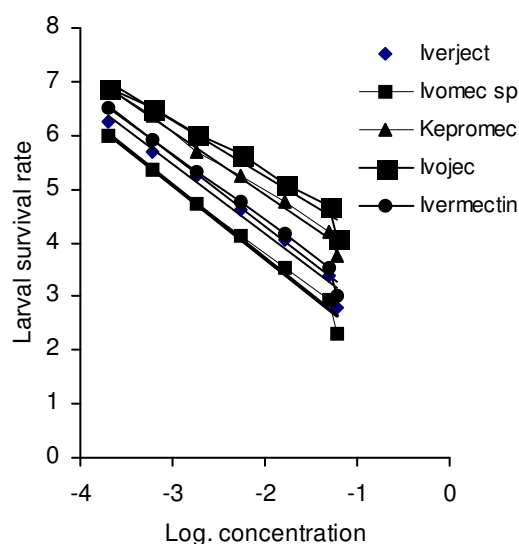


Figure 1: Linear relationship between mean values of live L₃ (on a probit) of strongyles following a 7 day incubation period in ivermectin injection brands and ivermectin concentration. Each point represents the mean of three replicates.

Table 1: Comparison of the bioactivity of ivermectin injection brands with Ivomec super.

Injection brands	LC ₅₀ (ng/ml)	p-value
1. Ivomec super	1.1±0.2*	
2. Iverject	2.3±0.3	< 0.01
3. Ivermectin	3.00±0.3	< 0.01
4. Kepromec	8.0±0.2	< 0.01
5. Ivojec	17.0±0.3	< 0.01

*Reference product

another report⁷, it was shown that Weybridge strain of *Haemonchus contortus* is more susceptible (LC₅₀ – 0.4 ng/ml) than the Australian strain (LC₅₀ – 8.9 ng/ml) to ivermectin (Ivomec MSD). An ivermectin resistant strain of *H. contortus* was also reported to have LC₅₀ of 8.0 ng/ml⁸.

The strongyles used for this present work are from a mixed infection naturally acquired by sheep, and were identified to be

predominantly *H. contortus*. Ivomec MSD is the innovator company's brand and it showed the highest activity ($LC_{50} = 1.1$ ng/ml). However, it contains clorsulon in addition to the ivermectin active ingredient. The latter is reported (in the package insert) to be specific anthelmintic for liver fluke. It is possible that this combination could enhance the activity of ivermectin against other nematodes.

Ivomec super was selected as the reference brand for estimating bioactivity of the other brands because it is the innovator brand. When compared with other products, the LC_{50} for Ivomec super was significantly different ($p < 0.01$). This implies that the bioactivity of this product is significantly different from this reference product. Results of this study could explain the variation in treatment response often reported by clinicians prescribing these different brands of ivermectin. Significant difference in the bioactivity of the brands is a reflection of the difference in the quality these drug products being marketed by different manufacturers. The biologic inequivalence of the different brands shown by this *in vitro* study could be due to differences in the formulation of the products or chemical compositions.

Conclusion

The results of this study show that the brands of ivermectin injection investigated are not bioequivalent. This could explain why different brands of ivermectin injection may show variation in treatment responses.

An investigation of the chemical equivalence of the brands by a physicochemical assay method is warranted in order to provide supportive data that could further explain the differences in the LC_{50} obtained. The results underscore the need for post market surveillance of drugs intended for veterinary use after initial registration by drug regulatory authorities.

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