



Impact of interventions on mosquitoes resting behaviour and species composition in Lugeye village in Magu district, Northwestern Tanzania

^{1,2}Eliningaya J. Kweka¹, ¹Humphrey D. Mazigo

^{1,2}Department of Medical Parasitology and Entomology, Catholic University of Health and Allied Sciences, P.O. Box 1464, Mwanza, Tanzania

¹Tanzania Plant Health and Pesticides Authority, Pesticides Bioefficacy section, P.O. Box 3024, Arusha, Tanzania

Abstract

Background: Understanding the impact of intervention tools on vector behaviour, host preference, resting, and infectivity rates is paramount in malaria control planning. Magu district was one of the districts in lake zone regions in northwestern Tanzania covered with indoor residual spray and long-lasting insecticidal nets in the main malaria control campaign. After interventions, this study evaluated the mosquito's host preference and resting behaviour in Lugeye village in rainy and dry seasons.

Methods: Mosquitoes were collected both indoors and outdoors using the rest pots. The collection was done in both rainy and dry seasons. The samples were collected bi-weekly for three months each season.

Results: A total of 254 mosquitoes were collected in both dry and rainy seasons, indoors and outdoors. The most abundant species were *Anopheles funestus* s.s. and *An.arabiensis*. Most blood meals from bovines rested outdoors, while human blood meal sources rested outdoors. Sporozoite-positive mosquitoes were found only during the dry season.

Conclusion: This study's findings have shown that implementing IRS and LLIN interventions has led to a species shift from *An.gambiae* s.s. to *An.funestus* and *An.arabiensis*. The inclusion of vector insecticide resistance information can be of paramount importance in appropriate intervention tool selection.

Keywords: Bloodmeals, *An.funestus*, *An.arabiensis*, outdoor, indoor

Introduction

Malaria is still one of the public health challenges in sub-Saharan Africa (WHO, 2023). In sub-Saharan Africa, four countries have been reported contributing the highest malaria cases globally within the continent (Nigeria (26.8%), the Democratic Republic of the Congo (12.3%), Uganda (5.1%) and Mozambique (4.2%)) and other four countries contributing more than half of malaria death globally are, Nigeria (31.1%), the Democratic Republic of the Congo (11.6%), Niger (5.6%) and the United Republic of Tanzania (4.4%) (WHO, 2023). The malaria-related mortality has been decreasing from 25 in 2000 to 10 in 2019 deaths per 100,000 populations at risk (WHO, 2020). All these achievements have been attained due to the implementation of sensitive malaria diagnostic tools, the prescription of appropriate anti-malarial drugs and effective vector control tools (WHO, 2020). The effective vector control has been implemented widely using long-lasting insecticidal nets (LLINs), Indoor residual spray (IRS) and in very limited application of Larval Source Management (LSM) (Derua et al., 2019; Diouf et al., 2020; Tusting et al., 2013; Zhou et al., 2020).

* Corresponding author: Eliningaya J. Kweka E-mails: pat.kweka@gmail.com



Tanzania's main efficient malaria vectors are the sibling species of *Anopheles gambiae* and *An. funestus* group (Kabula et al., 2011; Kweka et al., 2008a; Kweka et al., 2020). Among the *An. gambiae* sibling species, *An. arabiensis*, *An. gambiae* s.s. and *An. merus* have been reported to be vectors in Tanzania (Kweka et al., 2008a; Kweka et al., 2020; Kweka et al., 2008b). In *An. funestus* group, Tanzania's most efficient recorded vectors are *An. funestus* s.s (Kweka et al., 2008a; Kweka et al., 2020), *An. parensis*(Kweka et al., 2008a), *An. rivulorum* (Kweka et al., 2008a; Kweka et al., 2020) and *An. lesoni* (Kweka et al., 2020).

Since the wide community coverage with LLINs in 2000, the vector population has been shrinking across the country, and vector species have shifted. (Bayoh et al., 2010; Kitau et al., 2012) Due to the wide coverage of indoor surfaces with insecticides, either LLINs or IRS, mosquitoes have opted to forfeit the benefits of LLINs and IRS by feeding and resting outdoors. (Russell et al., 2011) or developing insecticide resistance for progeny survivorship (Kreppel et al., 2020; Kulkarni et al., 2006; Mahande et al., 2012; Mbepera et al., 2017; Nnko et al., 2017). The increased proportion of outdoor vectors leading to outdoor residual malaria transmission has been witnessed in different areas with high malaria transmission (Russell et al., 2011). Insecticide resistance in vector populations has been found to exist in all classes of insecticides used for public health vector control. (Kabula et al., 2014; Matowo et al., 2014).

In Lake Zone regions, including the study areas, Magu District, with 91.8% coverage between 2015 to 2017 were sprayed with Actellic 300CS (Primiphos methyl) (Kakilla et al., 2020; Mashauri et al., 2017). Insecticide resistance has been reported to occur in this region among vector species (Kakilla et al., 2020; Kisinza et al., 2017; Philbert et al., 2017) The occurrence of insecticide resistance threatens the use of tools with insecticides. This study assessed the impact of the intervention tools implemented in the study area on species composition, feeding and resting behavior, and infective rates among vector populations in rainy and dry seasons.

Material and Methods

Study Site

This study was conducted in the Lugeye village (02.332159S, 33.150529E) in Magu District, Mwanza Region. Mwanza is among the Lake Zone regions with high malaria prevalence in Tanzania. They are highly inhabited by peasants who produce maize, paddy, cotton, and vegetables in small-scale farming. The study area has two rain seasons: the long rain season starts from October to December, while the short rain season starts from February to April. This district had full coverage of IRS using Primiphos methyl insecticides and LLINs (Kakilla et al., 2020; Mashauri et al., 2017).

Mosquitoes Collection and Identification

Mosquitoes were collected using two methods during the study. Firstly, the Center for Disease Control (CDC) light trap (model 512, John W. Hock Company, Gainesville, FL) was operated as elaborated by a previous study for the collection of indoor host-seeking mosquitoes (Lines et al., 1991; Shiff et al., 1995). Secondly, outdoor resting mosquitoes were collected using pots previously utilized for the purpose (Odiere et al., 2007; van den Bijllaardt et al., 2009). Twenty houses were used, each house trapping mosquitoes both indoors and outdoors over the same night. The collection was done for one month in the rainy and dry seasons. The mosquitoes collected were identified morphologically in the field using the key developed by Gillies and Coetzee (Gillies and Coetzee, 1987). The collected *An. gambiae* s.l. mosquitoes were identified at species level using the method developed by Scott and others (Scott, Brogdon and Collins, 1993), while the *An. funestus* sibling species were identified using a

method developed by Koekemoer and others (Koekemoer et al., 2002). Mosquitoes were separated by seasons collected.

Host blood meal identification.

The blood-fed mosquitoes collected indoors and outdoors were prepared by smearing the abdomen in the Whatman filter paper No.1 (Bray, Gill and Killick-Kendrick, 1984). They were labelled by place and date of collection. The blood meal source host was identified using the Enzymes-linked immunosorbent assay (ELISA) protocol. (Beier et al., 1988). The study tested four hosts as possible blood meal sources: bovine, goat, dog, and human.

Sporozoite rates

The collected mosquitoes of both *An. gambiae* s.l. and *An. funestus* group, the head and thorax were taken and subjected to the ELISA protocol developed by Wirtz et al. 1987 (Wirtz et al., 1987). The specimen was considered positive when the cut-off value was similar to or above the positive control.

Data Analysis

The data analysis was done using IBM SPSS Version 26 (IBM Corp., Armonk, NY, USA). The proportion of mosquitoes collected outdoors and indoors was compared using the Chi-square test. The comparison by seasons was done using the Chi-square test. The comparison was regarded to have significance when the P-value was less than 5%.

Results

Mosquitoes collection and species identification

A total of 254 mosquitoes were collected indoors and outdoors for both seasons. In the rainy season, eight (8) (100%) mosquitoes were sampled, and all were identified as *An. lesoni*. In dry season 246 mosquitoes were collected, 96 (39.02) were *An. arabiensis*, 129 (52.44%). *An. funestus* s.s., 1 (0.41%) *An. constani* and 20(8.13%) specimens were not identified (Figure 1). The *An. funestus* s.s. abundance was statistically significantly higher in the dry season than in the rainy season ($C^2 = 68.59, P < 0.001$, Figure 1). The abundance of *An. arabiensis* was statistically abounding in the dry season and then in the rainy season ($C^2 = 45.13, P < 0.001$, Figure 1). *An. lesoni* population was higher in the rainy season than in the dry season, which was found to be statistically significant ($C^2=192.08, P < 0.001$, Figure 1).

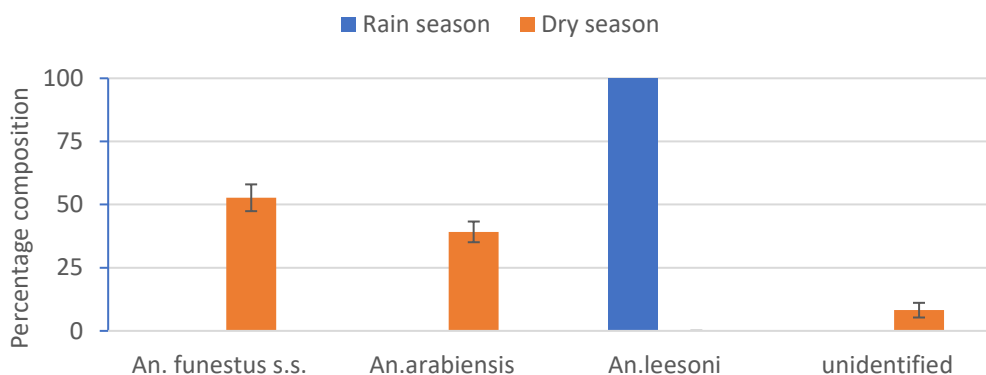


Figure 1: The species composition of mosquitoes collected in all seasons.

Blood meal host identification

A total of 115 blood meal samples were analysed. Twelve (12) samples were collected in dry season while one hundred and three (103) were sampled in rainy season. In rainy season, 6(50%) were from Bovine, 5(41.67%) from Human, 1(8.33) from dog while none was found from goats (Figure 2). In dry season 103 samples of blood were collected. Twelve (12) (11.65%) were unidentified, 24(23.30%) were from bovine, 1(0.97%) was from Dog, 5(4.85%) were from Goat while 61(59.22%) were from Human (Figure 2). The comparison of blood meals source by season was found to be statistically significant for three host species. Bovine caught most outdoor ($\chi^2=15.73$, $P<0.001$), Human caught most indoor ($C^2=5.78$ $P=0.016$), Dog ($C^2=5.70$ $P=0.017$) and Goad ($C^2=2.75$, $P=0.097$) (Figure 2). The comparison of blood meal by site of mosquitoes collection (outdoor/indoor) found that, for the three host species, there was statistically significant different results between Indoor and outdoor. In Bovine ($C^2=70.19$, $P<0.001$), Human ($C^2=85.62$, $P<0.001$), Dog ($C^2=7.79$, $P<0.005$) and Goat ($C^2=0.12$, $P<0.733$) (Figure 3).

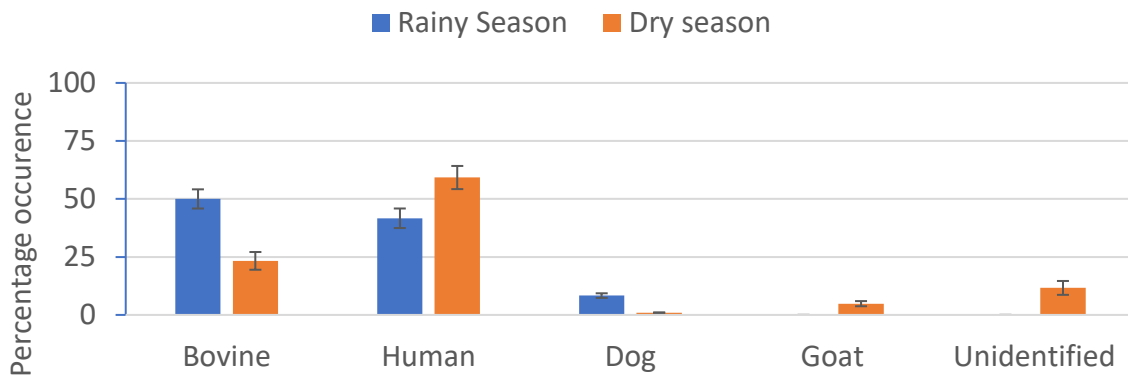


Figure 2: The bloodmeal analysis in seasonality from different hosts

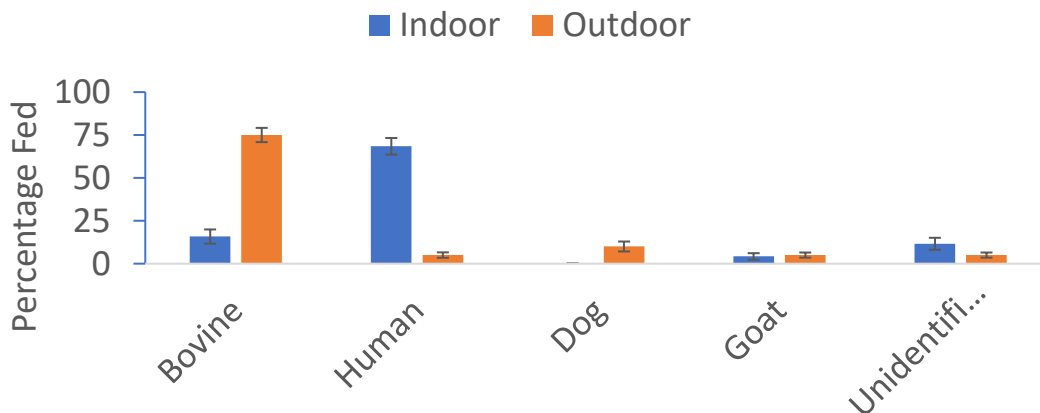


Figure 3: The bloodmeal analysis by resting position of vectors from different hosts.



Sporozoite rates

In the dry season, all eight (8) mosquitoes collected were found to be circumsporozoite protein-negative. In the rainy season, among 246 mosquitoes tested, 10 (4.07%) were found to be circumsporozoite protein positive. Among those tested positive, 6(60%) were *An. arabiensis*, and 4 (40%) were *An. funestus* s.s. All sporozoite-positive mosquitoes were collected in the dry season.

Discussion

The findings of this study have shown that malaria vector abundance has been influenced by seasonality in the population of *An. funestus* s.s. dominating during dry season while and *An. arabiensis* dominating the rainy season. The similar species composition was revealed by previous study conducted in a similar area (Kakilla et al., 2020). Also, in other areas of Tanzania, there has been predominance of *An. arabiensis* and *An. funestus* sibling species in the recent past (Kweka et al., 2008a; Kweka et al., 2020; Kweka et al., 2008b; Lwetoijera et al., 2014).

This confirms that, by far, *An. arabiensis* and *An. funestus* are the major malaria vectors in mainland Tanzania (Kweka et al., 2008a; Kweka et al., 2020; Kweka et al., 2008b; Lwetoijera et al., 2014). In this study site, there is a high population shift of the malaria vectors; in a study conducted in the district before the mass intervention of IRS and LLINs, *An. gambiae* s.s. had an upper hand over *An. arabiensis* (Kisinja et al., 2017) This study revealed that the population of *An. gambiae* s.s. has been diminished and replaced by *An. arabiensis* and *An. funestus* s.s.

These vectors recently have shown high tolerance to different classes of insecticides used in LLINs and IRS (Kabula et al., 2014; Kakilla et al., 2020; Kisinja et al., 2017; Matowo et al., 2014; Mbepera et al., 2017; Nnko et al., 2017). Different vector species have shown to have different mechanisms to tolerate insecticides toxicity such as biochemical and behavioral resistance (Kulkarni et al., 2006; Kweka et al., 2020; Matowo et al., 2014; Yewhalaw and Kweka, 2016) These mechanisms have enhanced the survivorship of the vector population and transmitted malaria across different ecological areas despite intensive interventions.

The vector abundance was found to be higher in the dry season than the rainy season; this agrees with the ecological studies of the species, which found that in rainy season habitat washing is higher in habitats with shallow water and stable in large water bodies such as in swamps were *An. funestus* and *An. gambiae* s.l. breed most (Kweka et al., 2012). In this study, the *An. arabiensis* and *An. funestus* were abundant in the dry season than in the rainy season. That was found to agree with the ecological conditions of the vector habitats which gain temperature to enhance larval growth and shorten the life cycle in the dry season (Mala et al., 2011). The study site has stable habitats, which become more productive in the dry season than in the rainy season.

A similar scenario has been found by previous studies in different area across Africa for having more vectors in the dry season than in rainy season (Fillinger et al., 2009; Kweka et al., 2011; Kweka et al., 2012). The results of the current study of having high population of vectors in dry season is contrary to what was found in other malaria endemic countries where the population was bottlenecked during the season (Dao et al., 2014) while in Brazil a study indicated reduced adult survivorship when the temperature is increased in the field (Chu et al., 2020). It's also known that, during the rainy season



the movements of mosquitoes are restricted due to rain (Roiz et al., 2010). This might have contributed to the observed scenario in this study.

The findings of this study also have shown that bovines and humans were highly preferred hosts while dogs and goats were the least. According to highly abundant species *An. arabiensis* (zoophilic species) and *An. funestus* s.s. (anthropophilic species) which prefers bovine and humans, respectively, most similar preference was observed in previous studies (Kibret et al., 2017). In both seasons, the higher blood meal was found to be from humans and animals due to vector preference of vectors and accessibility of the hosts (Mahande et al., 2007). The high access to human blood by vectors was worrying as the community had high coverage IRS and LLINs interventions. This high blood meal access from human might be attributed by the insecticide tolerance level within the vector population (Kakilla et al., 2020; Kisinza et al., 2017). Similar has been found that insecticide tolerance enhances the vectors to access human bloodmeals regardless of intervention covering the population (Glunt et al., 2018).

The findings on sporozoite rates have shown that in this study, the infected mosquitoes were found during the dry season, while in the rainy season, none contained sporozoite protein. The dry season has been found to have no shelters outdoors to hide during the day and, therefore, resting mostly indoors. Resting indoors increases the human-vector contact risks. In a previous study it was revealed that adult mosquitoes cannot tolerate high temperatures, therefore during the day they have to hide under the shaded area which include human shelters and cowsheds (Faye et al., 1997; Magombedze, Ferguson and Ghani, 2018; Mayagaya et al., 2015) The increase in sporozoite rates in *An. funestus* and *An. arabiensis* characterizes the malaria transmission efficiency played by the two species in the lake zone.

Conclusion

The findings of this study have shown that *An. arabiensis* and *An. funestus* are the main malaria vectors in the study site, with high abundance and infectivity in the dry season. Assessing their insecticide resistance can generate complementary information for designing effective control programmes.

Declarations

Ethics approval and consent to participate: The Catholic University of Health and Allied Sciences gave the ethical approval.

Consent for publication: Not applicable

Availability of data and materials: All data used in this study will be available upon request from the corresponding author

Competing interests: Authors declared to have no competing interests

Funding: This study received financial support from the Tanzania Plant Health and Pesticides Authority.

Authors' contributions: EJK conceptualized and designed the study, and HDM coordinated field activities. EJK and HDM also conducted data analysis and manuscript writing. Both have endorsed the submission of this manuscript.



Acknowledgements: The authors wish to express their appreciation for the help rendered by NIMR and Mwanza Center field assistants during mosquito collection. The head of Household consented to the study being conducted in their residences. Ms. Lucy Kisima is appreciated for providing literature access to the TPHPA library.

References

- Bayoh, M.N., Mathias, D.K., Odiere, M.R., Mutuku, F.M., Kamau, L., Gimnig, J.E., Vulule, J.M., Hawley, W.A., Hamel, M.J., Walker, E.D., 2010. *Anopheles gambiae*: historical population decline associated with regional distribution of insecticide-treated bed nets in western Nyanza Province, Kenya. *Malaria journal* 9, 62.
- Beier, J.C., Perkins, P.V., Wirtz, R.A., Koros, J., Diggs, D., Gargan, T.P., 2nd, Koech, D.K., 1988. Bloodmeal identification by direct enzyme-linked immunosorbent assay (ELISA), tested on *Anopheles* (Diptera: Culicidae) in Kenya. *Journal of medical entomology* 25, 9-16.
- Bray, R., Gill, G., Killick-Kendrick, R., 1984. Current and possible future techniques for the identification of blood meals of vector haematophagous arthropods. World Health Organization, Geneva.
- Chu, V.M., Sallum, M.A.M., Moore, T.E., Emerson, K.J., Schlichting, C.D., Conn, J.E., 2020. Evidence for family-level variation of phenotypic traits in response to temperature of Brazilian *Nyssorhynchus darlingi*. *Parasites & vectors* 13, 55.
- Dao, A., Yaro, A.S., Diallo, M., Timbiné, S., Huestis, D.L., Kassogué, Y., Traoré, A.I., Sanogo, Z.L., Samaké, D., Lehmann, T., 2014. Signatures of aestivation and migration in Sahelian malaria mosquito populations. *Nature* 516, 387-390.
- Derua, Y.A., Kweka, E.J., Kisinza, W.N., Githeko, A.K., Mosha, F.W., 2019. Bacterial larvicides used for malaria vector control in sub-Saharan Africa: review of their effectiveness and operational feasibility. *Parasites & vectors* 12, 426.
- Diouf, E.H., Niang, E.H.A., Samb, B., Diagne, C.T., Diouf, M., Konaté, A., Dia, I., Faye, O., Konaté, L., 2020. Multiple insecticide resistance target sites in adult field strains of *An. gambiae* (s.l.) from southeastern Senegal. *Parasites & vectors* 13, 567.
- Faye, O., Konate, L., Mouchet, J., Fontenille, D., Sy, N., Hebrard, G., Herve, J.P., 1997. Indoor resting by outdoor biting females of *Anopheles gambiae* complex (Diptera:Culicidae) in the Sahel of northern Senegal. *Journal of medical entomology* 34, 285-289.
- Fillinger, U., Sombroek, H., Majambere, S., van Loon, E., Takken, W., Lindsay, S.W., 2009. Identifying the most productive breeding sites for malaria mosquitoes in The Gambia. *Malaria journal* 8, 62.
- Gillies, M., Coetzee, M., 1987. A supplement to the Anophelinae of Africa South of the Sahara. *Publ S Afr Inst Med Res, JOHANNESBURG*
- Glunt, K.D., Coetzee, M., Huijben, S., Koffi, A.A., Lynch, P.A., N'Guessan, R., Oumbouke, W.A., Sternberg, E.D., Thomas, M.B., 2018. Empirical and theoretical investigation into the potential impacts of insecticide resistance on the effectiveness of insecticide-treated bed nets. *Evolutionary applications* 11, 431-441.
- Kabula, B., Derua, Y.A., Tungui, P., Massue, D.J., Sambu, E., Stanley, G., Mosha, F.W., Kisinza, W.N., 2011. Malaria entomological profile in Tanzania from 1950 to 2010: a review of mosquito distribution, vectorial capacity and insecticide resistance. *Tanzania journal of health research* 13, 319-331.
- Kabula, B., Kisinza, W., Tungu, P., Ndege, C., Batengana, B., Kollo, D., Malima, R., Kafuko, J., Mohamed, M., Magesa, S., 2014. Co-occurrence and distribution of East (L1014S) and West (L1014F) African



- knock-down resistance in *Anopheles gambiae* sensu lato population of Tanzania. *Tropical medicine & international health* : TM & IH 19, 331-341.
- Kakilla, C., Manjurano, A., Nelwin, K., Martin, J., Mashauri, F., Kinung'hi, S.M., Lyimo, E., Mangalu, D., Bernard, L., Iwuchukwu, N., Mwalimu, D., Serbantez, N., Greer, G., George, K., Oxborough, R.M., Magesa, S.M., 2020. Malaria vector species composition and entomological indices following indoor residual spraying in regions bordering Lake Victoria, Tanzania. *Malaria journal* 19, 383.
- Kibret, S., Wilson, G.G., Ryder, D., Tekie, H., Petros, B., 2017. Malaria impact of large dams at different eco-epidemiological settings in Ethiopia. *Tropical medicine and health* 45, 4.
- Kisizza, W.N., Nkya, T.E., Kabula, B., Overgaard, H.J., Massue, D.J., Mageni, Z., Greer, G., Kaspar, N., Mohamed, M., Reithinger, R., Moore, S., Lorenz, L.M., Magesa, S., 2017. Multiple insecticide resistance in *Anopheles gambiae* from Tanzania: a major concern for malaria vector control. *Malaria journal* 16, 439.
- Kitau, J., Oxborough, R.M., Tungu, P.K., Matowo, J., Malima, R.C., Magesa, S.M., Bruce, J., Mosha, F.W., Rowland, M.W., 2012. Species shifts in the *Anopheles gambiae* complex: do LLINs successfully control *Anopheles arabiensis*? *PloS one* 7, e31481.
- Koekemoer, L.L., Kamau, L., Hunt, R.H., Coetzee, M., 2002. A cocktail polymerase chain reaction assay to identify members of the *Anopheles funestus* (Diptera: Culicidae) group. *The American journal of tropical medicine and hygiene* 66, 804-811.
- Kreppel, K.S., Viana, M., Main, B.J., Johnson, P.C.D., Govella, N.J., Lee, Y., Maliti, D., Meza, F.C., Lanzaro, G.C., Ferguson, H.M., 2020. Emergence of behavioural avoidance strategies of malaria vectors in areas of high LLIN coverage in Tanzania. *Scientific reports* 10, 14527.
- Kulkarni, M.A., Rowland, M., Alifrangis, M., Mosha, F.W., Matowo, J., Malima, R., Peter, J., Kweka, E., Lyimo, I., Magesa, S., Salanti, A., Rau, M.E., Drakeley, C., 2006. Occurrence of the leucine-to-phenylalanine knockdown resistance (kdr) mutation in *Anopheles arabiensis* populations in Tanzania, detected by a simplified high-throughput SSOP-ELISA method. *Malaria journal* 5, 56.
- Kweka, E.J., Mahande, A.M., Nkya, W.M., Assenga, C., Lyatuu, E.E., Nyale, E., Mosha, F.W., Mwakalinga, S.B., Temu, E.A., 2008a. Vector species composition and malaria infectivity rates in Mkuzi, Muheza District, north-eastern Tanzania. *Tanzania journal of health research* 10, 46-49.
- Kweka, E.J., Mazigo, H.D., Lyaruu, L.J., Mause, E.A., Venter, N., Mahande, A.M., Coetzee, M., 2020. Anopheline Mosquito Species Composition, Kdr Mutation Frequency, and Parasite Infectivity Status in Northern Tanzania. *Journal of medical entomology* 57, 933-938.
- Kweka, E.J., Nkya, W.M., Mahande, A.M., Assenga, C., Mosha, F.W., Lyatuu, E.E., Massenga, C.P., Nyale, E.M., Mwakalinga, S.B., Lowassa, A., 2008b. Mosquito abundance, bed net coverage and other factors associated with variations in sporozoite infectivity rates in four villages of rural Tanzania. *Malaria journal* 7, 59.
- Kweka, E.J., Zhou, G., Lee, M.C., Gilbreath, T.M., 3rd, Mosha, F., Munga, S., Githeko, A.K., Yan, G., 2011. Evaluation of two methods of estimating larval habitat productivity in western Kenya highlands. *Parasites & vectors* 4, 110.
- Kweka, E.J., Zhou, G., Munga, S., Lee, M.C., Atieli, H.E., Nyindo, M., Githeko, A.K., Yan, G., 2012. Anopheline larval habitats seasonality and species distribution: a prerequisite for effective targeted larval habitats control programmes. *PloS one* 7, e52084.
- Lines, J.D., Curtis, C.F., Wilkes, T.J., Njunwa, K.J., 1991. Monitoring human-biting mosquitoes (Diptera: Culicidae) in Tanzania with light-traps hung beside mosquito nets. *Bulletin of Entomological Research* 81, 77-84.



- Lwetoijera, D.W., Harris, C., Kiware, S.S., Dongus, S., Devine, G.J., McCall, P.J., Majambere, S., 2014. Increasing role of *Anopheles funestus* and *Anopheles arabiensis* in malaria transmission in the Kilombero Valley, Tanzania. *Malaria journal* 13, 331-331.
- Magombedze, G., Ferguson, N.M., Ghani, A.C., 2018. A trade-off between dry season survival longevity and wet season high net reproduction can explain the persistence of *Anopheles* mosquitoes. *Parasites & vectors* 11, 576.
- Mahande, A., Mosha, F., Mahande, J., Kweka, E., 2007. Feeding and resting behaviour of malaria vector, *Anopheles arabiensis* with reference to zooprophyllaxis. *Malaria journal* 6, 100.
- Mahande, A.M., Dufour, I., Matias, J.R., Kweka, E.J., 2012. Knockdown Resistance, rdl Alleles, and the Annual Entomological Inoculation Rate of Wild Mosquito Populations from Lower Moshi, Northern Tanzania. *Journal of global infectious diseases* 4, 114-119.
- Mala, A.O., Irungu, L.W., Shililu, J.I., Muturi, E.J., Mbogo, C.C., Njagi, J.K., Githure, J.I., 2011. Dry season ecology of *Anopheles gambiae* complex mosquitoes at larval habitats in two traditionally semi-arid villages in Baringo, Kenya. *Parasites & vectors* 4, 25.
- Mashauri, F.M., Manjurano, A., Kinung'hi, S., Martine, J., Lyimo, E., Kishamawe, C., Ndege, C., Ramsan, M.M., Chan, A., Mwalimu, C.D., Changalucha, J., Magesa, S., 2017. Indoor residual spraying with micro-encapsulated pirimiphos-methyl (Actellic® 300CS) against malaria vectors in the Lake Victoria basin, Tanzania. *PLoS one* 12, e0176982.
- Matowo, J., Jones, C.M., Kabula, B., Ranson, H., Steen, K., Mosha, F., Rowland, M., Weetman, D., 2014. Genetic basis of pyrethroid resistance in a population of *Anopheles arabiensis*, the primary malaria vector in Lower Moshi, north-eastern Tanzania. *Parasites & vectors* 7, 274.
- Mayagaya, V.S., Nkwengulila, G., Lyimo, I.N., Kihonda, J., Mtambala, H., Ngonyani, H., Russell, T.L., Ferguson, H.M., 2015. The impact of livestock on the abundance, resting behaviour and sporozoite rate of malaria vectors in southern Tanzania. *Malaria journal* 14, 17.
- Mbepera, S., Nkwengulila, G., Peter, R., Mause, E.A., Mahande, A.M., Coetzee, M., Kweka, E.J., 2017. The influence of age on insecticide susceptibility of *Anopheles arabiensis* during dry and rainy seasons in rice irrigation schemes of Northern Tanzania. *Malaria journal* 16, 364.
- Nnko, E.J., Kihamia, C., Tenu, F., Premji, Z., Kweka, E.J., 2017. Insecticide use pattern and phenotypic susceptibility of *Anopheles gambiae* sensu lato to commonly used insecticides in Lower Moshi, northern Tanzania. *BMC research notes* 10, 443.
- Odiere, M., Bayoh, M.N., Gimnig, J., Vulule, J., Irungu, L., Walker, E., 2007. Sampling outdoor, resting *Anopheles gambiae* and other mosquitoes (Diptera: Culicidae) in western Kenya with clay pots. *Journal of medical entomology* 44, 14-22.
- Philbert, A., Lyantagaye, S.L., Pradel, G., Ngwa, C.J., Nkwengulila, G., 2017. Pyrethroids and DDT tolerance of *Anopheles gambiae* s.l. from Sengerema District, an area of intensive pesticide usage in north-western Tanzania. *Tropical medicine & international health : TM & IH* 22, 388-398.
- Roiz, D., Rosà, R., Arnoldi, D., Rizzoli, A., 2010. Effects of temperature and rainfall on the activity and dynamics of host-seeking *Aedes albopictus* females in northern Italy. *Vector borne and zoonotic diseases (Larchmont, N.Y.)* 10, 811-816.
- Russell, T.L., Govella, N.J., Azizi, S., Drakeley, C.J., Kachur, S.P., Killeen, G.F., 2011. Increased proportions of outdoor feeding among residual malaria vector populations following increased use of insecticide-treated nets in rural Tanzania. *Malaria journal* 10, 80.
- Scott, J.A., Brogdon, W.G., Collins, F.H., 1993. Identification of single specimens of the *Anopheles gambiae* complex by the polymerase chain reaction. *The American journal of tropical medicine and hygiene* 49, 520-529.



- Shiff, C.J., Minjas, J.N., Hall, T., Hunt, R.H., Lyimo, S., Davis, J.R., 1995. Malaria infection potential of anopheline mosquitoes sampled by light trapping indoors in coastal Tanzanian villages. *Medical and veterinary entomology* 9, 256-262.
- Tusting, L.S., Thwing, J., Sinclair, D., Fillinger, U., Gimnig, J., Bonner, K.E., Bottomley, C., Lindsay, S.W., 2013. Mosquito larval source management for controlling malaria. *The Cochrane database of systematic reviews* 2013, Cd008923.
- van den Bijllaardt, W., ter Braak, R., Shekalaghe, S., Otieno, S., Mahande, A., Sauerwein, R., Takken, W., Bousema, T., 2009. The suitability of clay pots for indoor sampling of mosquitoes in an arid area in northern Tanzania. *Acta tropica* 111, 197-199.
- WHO, 2020. World malaria report 2020: 20 years of global progress and challenges. World Health Organization, Geneva.
- WHO, 2023. World malaria report 2023. World Health Organization, Geneva.
- Wirtz, R.A., Zavala, F., Charoenvit, Y., Campbell, G.H., Burkot, T.R., Schneider, I., Esser, K.M., Beaudoin, R.L., Andre, R.G., 1987. Comparative testing of monoclonal antibodies against Plasmodium falciparum sporozoites for ELISA development. *Bull World Health Organ* 65, 39-45.
- Yewhalaw, D., Kweka, E.J., 2016. Insecticide resistance in East Africa—history, distribution and drawbacks on malaria vectors and disease control, in: Trdan, S. (Ed.), *Insecticides Resistance*. Rijeka: IntechOpen, pp. 189-215.
- Zhou, G., Lee, M.C., Atieli, H.E., Githure, J.I., Githeko, A.K., Kazura, J.W., Yan, G., 2020. Adaptive interventions for optimizing malaria control: an implementation study protocol for a block-cluster randomized, sequential multiple assignment trial. *Trials* 21, 665.