

Prevalence of Trypanosomosis in Small Ruminants at Slaughter in Jos, Plateau State, Nigeria

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ABSTRACT

The prevalence of *Trypanosoma* infection in indigenous breeds of small ruminants at slaughter in Jos was studied from 680 blood samples collected from 300 sheep and 380 goats from December 1999 to May 2000. 8(2.67%) sheep and 4(1.05%) goats were positive for *trypanosoma* infection. *Trypanosoma vivax* and *T. congolense* were respectively responsible for 75% and 25% of the infection in each animal specie. It is, therefore suggested that the epidemiology of trypanosomosis in small ruminants be taken more seriously and the animals be subjected to strict veterinary supervision.

Key words: Trypanosomosis, sheep, goat, abattoir, Jos, Nigeria

INTRODUCTION

Trypanosomosis is caused by *trypanosomes* which are blood parasites of Vertebrates (Ukoli, 1984). It is a disease of man and animals of economic importance in tropical areas of the world and remains one of the major diseases limiting growth of livestock industry in Nigeria and indeed tropical Africa (Leefflang, 1978). The disease is transmitted by tsetse-fly vector (Duggan, 1962) either cyclically (Stephen, 1970) or mechanically (Thomas and Lanborn, 1934; Dixon *et al.*, 1971) although other less important modes of transmission exist. Trypanosomosis occurs wherever tsetse flies are prevalent but may also be transmitted mechanically by other haematophagous flies (Weitz, 1970).

Laboratory and field study of animal trypanosomosis have tended to concentrate on cattle (Murray *et al.*, 1977; Leefflang 1978) since the disease in small ruminants is viewed as unimportant especially in West Africa (Kalu *et al.*, 1986). However, available evidence on the epidemiology of the disease in Nigeria, (Krammer, 1966; Joshua and Ige, 1982; Kalu and Agu, 1984; Daniel *et al.*, 1994; Kalu and Lawani 1996) and other parts of Africa (Griffin and Allonby, 1979; Ahmadu *et al.*, 2002) indicated that not only are infection rates high, serious losses result from naturally acquired caprine and ovine infection. Records on the study of trypanosomosis in small ruminants in Nigeria are few and disjointed (Joshua and Ige, 1982; Kalu and Magaji, 1986; Fakae and Chiejina, 1993, Adah *et al.*, 1993; Daniel *et al.*, 1994; Kalu and Lawani, 1996; Dadah *et al.*, 1997; Akinwale *et al.*, 2006).

This study is therefore aimed at investigating the prevalence of trypanosomosis among sheep and goats at slaughter in Jos central abattoir and documenting the most prevalent *Trypanosoma* specie in the study area. Abattoir sampling though associated with low incidence rate still provide an idea on the disease situation.

MATERIALS AND METHODS

Animals

Most of the animals sampled are not reared in Jos but are purchased from various parts of northern Nigeria especially Borno, Yobe, Sokoto and Kano states. They are indigenous breeds of sheep and goats, which are never

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held in Jos for more than two weeks before they are slaughtered.

Sample collection

Five millilitres of blood was collected once a week between December 1999 and May 2000 from each animal at slaughter into clean Bijou bottles containing ethylene diamine tetra-acetic acid (EDTA) as anticoagulant. The bottles were gently tilted to ensure proper mixing of blood and anticoagulant. They were labeled and kept in cooler containing ice packs and transported to the laboratory for processing.

Data analysis

The blood samples were analyzed for *trypanosomes* at the Parasitology Department of the National Veterinary Research Institute Vom, Plateau state, Nigeria using the haematocrit centrifugation technique (HCT), wet blood film (WBF), thin blood film and thick blood film (Woo, 1970; Kalu *et al.*, 1986). Identification of *Trypanosoma* specie was done using morphological differentiation of parasites on Giemsa-stained films as described by Murray *et al.* (1977).

Sample analysis

Data generated were analyzed using descriptive statistics such as percentage and frequency (Gomez and Gomez, 1984)

RESULTS

The prevalence of *Trypanosoma* species in sheep at slaughter within the study period is presented in Table 1. A total of three hundred (300) samples were examined out of which 8(2.67%) were positive for trypanosomosis, this represents 6(75%) and 2(25%) of *Trypanosoma vivax* and *T. congolense*, respectively. The prevalence in goats within the same period is presented in Table 2. Three hundred and eighty (380) blood samples were taken out of which 4(1.05%) were positive for trypanosomosis 3(75%) were *Trypanosoma vivax* while 1(25%) was *T. congolense*.

Table 1. Prevalence of trypanosome infection among sheep at slaughter at the Jos Central Abattoir, Nigeria

| Months | No. examined | No. positive | Prevalence rate (%) | <i>Trypanosoma</i> species | |
|----------|--------------|--------------|---------------------|----------------------------|----------------------|
| | | | | <i>T. vivax</i> | <i>T. congolense</i> |
| December | 50 | - | - | - | - |
| January | 50 | - | - | - | - |
| February | 50 | - | - | - | - |
| March | 50 | 1 | 2.00 | 1 | - |
| April | 50 | 2 | 4.00 | 1 | 1 |
| May | 50 | 5 | 10.00 | 4 | 1 |
| Total | 300 | 8 | 2.67 | 6 (75%) | 2 (25%) |

Table 2. Prevalence of *trypanosome* infection among Goats at slaughter at the Jos Central abattoir, Nigeria

| Months | No. examined | No. positive | Prevalence rate (%) | <i>Trypanosoma</i> species | |
|----------|--------------|--------------|---------------------|----------------------------|----------------------|
| | | | | <i>T. vivax</i> | <i>T. congolense</i> |
| December | 50 | - | - | - | - |
| January | 70 | - | - | - | - |
| February | 70 | - | - | - | - |
| March | 50 | 1 | 2.00 | 1 | - |
| April | 70 | 1 | 1.43 | - | 1 |
| May | 70 | 2 | 2.86 | 2 | - |
| Total | 380 | 4 | 1.05 | 3 (75%) | 1 (25%) |

DISCUSSION AND CONCLUSION

The result of this study showed a prevalence of 2.67% and 1.05% in sheep and goats respectively. This is similar to the reports of Daniel *et al.* (1994) in Gombe State and that of Kalu and Lawani (1996) in Kano State as well as Dadah *et al.* (1997) in Plateau State but lower than the earlier report of Joshua and Ige (1982) in goats in Plateau State, all in Northern Nigeria probably due to the period of study. The presence of *Trypanosoma vivax* and *T. congolense* further confirmed that this species occur in sheep and goats as earlier reported (Joshua and Ige, 1982; Kalu and Magaji, 1986; Dadah *et al.*, 1997). This study shows that trypanosomosis was first encountered in March which probably resulted from the onset of the rains since higher prevalence of the disease have been reported to be associated with the rainy season (Griffin and Allonby, 1979; Kalu and Lawani, 1996). The highest prevalence was recorded in the month of May which is similar to the reports of Joshua and Ige (1982). *Trypanosoma vivax* was the dominant trypanosome species encountered in this study, it accounted for 75% of all trypanosomosis recorded. This may be attributed to the fact that *T. vivax* can be transmitted in the absence of tsetse fly by haematophagous flies.

It is therefore suggested that the epidemiology of trypanosomosis in small ruminants be taken more seriously despite the low prevalence rate recorded. Apart from this, sheep and goats should be subjected to veterinary supervision and be included in the chemotherapeutic and prophylactic campaign programmes of government and herd owners.

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REFERENCES

- Adah, M. I., Otesile, E. B. and Joshua, R. A. (1993). Susceptibility of Nigeria West Africa dwarf and Red Sokoto goats to strains of *Trypanosoma congolense*. *Vet. Parasitol.* 47(3-4),; 177-188.
- Ahmadu, B., Lovelance, C. E. and Samui, K. L. (2002). A survey of trypanosomiasis in Zambian goats using haematocrit centrifuge technique and polymerase chain reaction. *J. S. Afr. Vet. Assoc.* 73(4), 224-226.
- Akinwale, O. P., Nock, I. H., Esiebo, K. A., Edeghere, H. U. and Olukosi, Y. A. (2006). Studies on the susceptibility of Sahel goats to experimental *Trypanosoma vivax*. *Vet. Parasitol.* 137(3-4), 210-213.
- Dadah, A. J., Duhlińska-Popova, D. D. Daniel, A. D. and Dede P. M. (1997). Trypanosomosis among sheep and goats at slaughter in Jos abattoir, Nigeria. *Rev. Elev. Med. Vet. Pays. Trop.* 50(3), 214-216.
- Daniel A. D., Joshua R. A., Kalejaiye, J. O. and Dada, A. J. (1994). Prevalence of trypanosomiasis in a region of Northern Nigeria. *Rev. Elev. Med. Vet. Pays. Trop.* 47(3) 295-297.
- Dixon J. B., Cull, R. S., Dunbur I. F., Greenhill R. T., Gruishan, C. G., Hill, M. A., Landeg, F. J. and Miller, W. M. (1971). Non cyclical transmission of trypanosomiasis in Uganda II. Experimental assessment of the survival time of *T. brucei*. *Vet. Rec.* 89, 233-235.
- Duggan, A. J. (1962). A survey of sleeping sickness in northern Nigeria from the earliest time to the present day. *Trans. Roy. Soc. Trop. Med. Hyg.* 56, 439.
- Fakae, B. B. and Chiejina, S. N. (1993). The prevalence concurrent Trypanosome and gastro intestinal nematode in West Africa dwarf goat and sheep. *Vet. Parasitol.* 49(2-4), 313-318.
- Gomez, K. A. and Gomez, A. A. (1984). *Statistical Procedure for Agricultural Research*, 2nd edition. Longman Singapore. 680pp.
- Griffin, L. and Allonby, E. W. (1979). Studies on the epidemiology of trypanosomiasis in sheep and goat in Kenya. *Trop. Anim. Hlth. Prod.* 11(3), 133-142.
- Joshua, R. A. and Ige, K. (1982): The incidence of Trypanosomiasis in Red Sokoto goats at slaughter. *Bull. Anim. Hlth. Prod. Afr.*, 30, 35-39.
- Kalu, A. U. and Agu, W. E. (1984). *Trypanosoma vivax* in goats, Preliminary report on the course of the disease in naturally acquired infection. *Proceedings of National Conference on the Diseases of Ruminants*, Vom, Nigeria. pp. 186-189.
- Kalu, A. U., Edeghere, H. U. and Lawani, F. A. (1986). Comparison of diagnostic techniques during subclinical single infection of trypanosomiasis in goats. *Vet. Parasitol.* 22,37-47.
- Kalu, A.U. and Lawani F. A. (1996). Observations on the epidemiology of ruminant trypanosomiasis in Kano State, Nigeria. *Rev Elev Med pays Trop.* 49(3), 213-217.
- Kalu, A. U. and Magaji, Y. (1986). An endemic focus on trypanosomiasis in Benue State, Nigeria. Nigerian Institute for Trypanosomiasis Research (NITR). Annual Report, 1986.
- Krammer, W. (1966). Incidence of trypanosomiasis in the West African dwarf goats and sheep in Nsukka, eastern Nigeria. *Bull. Epizoot. Dis. Afr.* 14, 423-428.
- Leefflang, P. (1978): Bovine trypanosomiasis in northern Nigeria. A contribution to the epidemiology, host specific

- and drug sensitivity of *T. vivax*. Unpublished Ph. D Thesis, University of Utrecht. 139 pp.
- Murray, M, Murray, P. K. and Meintryre, W. I. M. (1977). An improved parasitological technique for the detection of African trypanosomiasis. *Trans. Roy. Soc. Trop. Med. Hyg.* 71, 325-326.
- Stephen, L. E. (1970): Clinical manifestation of trypanosomiasis in livestock and other domestic animals. In: African trypanosomiasis (Muligan, H. W., ed.). George Allen and Union, London. pp. 774-794.
- Thomas, J. G. and Lanbom, W. A. (1934). Mechanical transmission of trypanosomiasis, leishmaniasis and yaws through biting haematogomous flies. *Br. Med. J.* 2,58.
- Ukoli, F. M. A. (1984). Introduction to parasitology in tropical Africa. Thompson Press, New Delhi, India. pp. 354-389.
- Weitz, B. G. F. (1970). Host of *Glossina*. In: African Trypanosomiasis (Muligan, H. W. Allen. G., eds.). Unwin Ltd., London. pp. 317-326.
- Woo, P. T. K. (1970):. Evaluation of haematocrit centrifuge and other techniques for the field diagnosis of trypanosomiasis and filariasis. *Acta. Trop.* 28, 298-303.