

Oregano essential oil in the diet of broilers: performance, carcass characteristics, and blood parameters

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Abstract

Efficacy of the phyto-genic feed additive oregano essential oil (OEO) (*Origanum vulgare* L.) was assessed as an alternative to an antibiotic as a growth promoter (AGP) in broiler diets. Three hundred male broiler chicks were assigned to five treatments, which consisted of diets with different levels of OEO (300, 600, and 900 mg/kg of diet), a negative control, and a positive control. Broiler performance was evaluated from 1 to 21 and 1 to 39 days old. From 1 to 39 days old, the broilers of the negative control group presented lower feed intake than those fed OEO300. No significant effect was observed on weight gain. The greatest feed conversion ratio occurred in the positive control group. Broilers treated with OEO300 had greater carcass yield than those in the positive control group. The haemogram, leukogram, and heterophil/lymphocyte ratio were positively influenced by OEO300. Red blood cell and leukocyte counts increased in a dose-dependent manner in broilers fed OEO-supplemented diets, while broilers in the positive control group had the lowest levels of haematocrit, mean corpuscular haemoglobin concentration, and plasma protein. Differential leukometry revealed lymphocyte numbers were increased with OEO treatment and reduced in the positive control group. For hepatic and renal metabolism, the broilers in the positive control group exhibited the greatest serum activities of aspartate aminotransferase, alanine aminotransferase, gamma-glutamyl transferase, and alkaline phosphatase. Although more studies on its efficacy are needed, OEO at 300 mg/kg may be used as a phyto-genic additive in broiler diets, especially those without AGP.

Keywords: antibiotic as growth promoter, immune system, phyto-genic feed additive.

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Introduction

The production of secondary compounds that are essential for plant survival and preservation depends on genetic and environmental factors. Many of these compounds elicit different reactions in living organisms and are therefore called active principles. Consequently, these plants are characterized as being medicinal, since they exhibit therapeutic activities owing to the presence of one or more active principles (Martins *et al.*, 2003). These plants can be added to animal diets as phyto-genic feed additives after being dried and milled or as essential oils (Jacela *et al.*, 2010; Madhupriya *et al.*, 2018). The use of alternative compounds that act as zootechnical additives, ensuring productive efficiency, has received significant attention, especially after the publication by the European Union of Regulation 1831/2003, which, as of January 2006, restricted the use of all antibiotic and chemotherapeutic agents as performance enhancers in animal production.

The main components of oregano essential oil (OEO) (*Origanum vulgare* L.) are carvacrol and thymol, which together represent up to 85% of its composition (Burt, 2004). Oregano essential oil has been associated with antifungal (Bedoya-Serna *et al.*, 2018), antibacterial, and antioxidant activity in vitro (Gandra *et al.*, 2013) and in vivo (Zanini *et al.*, 2013; Ri *et al.*, 2017), stimulation of enzyme secretion (He *et al.*, 2017), and modulation of microbial colonization of the gastrointestinal tract and intestinal morphology (Dias

et al., 2015; He *et al.*, 2017). However, reports of its effects on the performance and carcass characteristics of broilers have been inconsistent, with positive effects on these characteristics (Mathlouthi *et al.*, 2012; Peng *et al.*, 2016) as opposed to no effects (Alp *et al.*, 2012; Koyama *et al.*, 2014) being detected.

Phytogenic feed additives may have beneficial effects on the immunomodulatory activity and anti-inflammatory potential of broilers (Mohiti-Asli & Guanaatparast-Rashti, 2017). Therefore, analyses of blood parameters such as the erythrogram, leukogram, and serum biochemistry are important tools to evaluate the health status and detect pathophysiological phenomena and abnormalities of poultry.

Thus, the objective of this work was to evaluate the effects of OEO on the performance, carcass characteristics, and blood parameters of broilers.

Materials and Methods

All animal procedures were approved by the Board of Ethics and Animal Experimentation of the Federal Rural University of Rio de Janeiro (Protocol no. 010285).

Three hundred one-day-old male Cobb 500 chicks were used. They were stratified by bodyweight (43.69 g) and housed in metal battery cages with three levels. Each level was divided into two blocks, and each cage (0.90 m long × 0.85 m wide × 0.40 m high) housed 10 broilers. Each cage was provided with a cup-type infant drinking fountain and a plate-type feeder, which were changed to drinkers and feeders at 15 days old.

The chicks had been vaccinated against Marek's disease, fowlpox, Gumboro, and Newcastle disease at the hatchery. Feed rations and water were provided ad libitum throughout the experimental period.

In the first week, the broilers were maintained under constant lighting. In the second week, lighting was provided for 23 hours, while from days 18 to 28 lighting was provided for 16 hours. From 28 days old onwards, the broilers were maintained under natural lighting.

The diets (Table 1) that were provided to the broilers were isoproteic and isocaloric with a starter diet being provided from day 1 to 21 and a grower diet being fed thereafter (days 22 to 39). Both diets were formulated to meet the nutritional requirements of the poultry, according to the recommendations of the Brazilian Poultry and Swine Table (Rostagno *et al.*, 2005). The additives that constituted the treatments were included in the reference diet instead of inert kaolin material.

The treatments consisted of the reference diet (RD) without antimicrobials (negative control); RD with 10 mg colistin/kg feed (positive control); RD with 300 mg OEO/kg feed (OEO300); RD with 600 mg OEO/kg feed (OEO600); and RD with 900 mg OEO/kg feed (OEO900).

The OEO was microencapsulated (Givaudan do Brasil Ltd, SP, Brazil), which involves introducing the oil in an organic matrix to preserve it until it is ingested because of the volatile properties of essential oils. The analytical composition of the oil was 67% carvacrol, 12% p-cymene, 6% γ-terpinene, 5% thymol, 3.4% β-caryophyllene, 2% linalool, 2% myrcene, and 1% α-pinene (Givaudan do Brasil Ltd, São Paulo, Brazil).

To stimulate a microbiological challenge in the broilers, two breeding experiments were carried out before the experiment was initiated, without a sanitary rest and in a box with wood shavings. The broilers that were bred in this bed received no antimicrobials. After natural drying, a sample of this bed was sent to the Laboratory of Microbiology of the Veterinary Institute of UFRRJ for analysis. The total bacterial count was 45×10^5 CFU/g, using the PCA (plate count agar) method. The total coliform count was 22×10^4 CFU/g using the MacConkey method. Salmonella-Shigella agar (ASS) analysis was also performed and the presence of *Citrobacter freundii* was detected. The bed was sieved and 4 g/(bird-day) were mixed in the feed at 10, 12, 14, 16, 18, and 20 days old.

The performance parameters evaluated were feed intake, weight gain, viability, and feed conversion in the periods from 1 to 21 days old and 1 to 39 days old. The productive efficiency index (PEI) was calculated using the following formula:

$$PEI = \frac{\text{bodyweight(kg)} \times \text{viability}}{\text{age in days} \times \text{feed conversion}} \times 100$$

At 39 days old, the broilers were fasted for 12 hours immediately prior to euthanasia, which occurred at day 40. To evaluate carcass characteristics, two broilers that were close to the average weight of the group were removed from each replicate, totalling 12 broilers per treatment.

The broilers were euthanized by cervical dislocation, with posterior bleeding in the jugular vein, scalded, plucked, and eviscerated. Subsequently, the carcasses were weighed and stored in plastic bags, which were sealed, labelled, and refrigerated at 4 °C for 24 hours. After cooling, the carcasses and their cuts were weighed.

Carcass yield was calculated based on the chilled carcass weight (gutted and without heads and feet) and live weight after fasting. Cut yields were calculated based on the weight of the chilled cuts and chilled carcass.

Table 1 Dietary ingredients and calculated nutrient composition of basal starter and grower diets to which oregano essential oil or the antibiotic colistin sulfate were added (% , as-fed basis)

Ingredient (%)	Starter (1 to 21 days old)	Grower (22 to 39 days old)
Maize	56,189	58,997
Soybean meal	36,140	32,567
Soybean oil	3,020	3,993
Dicalcium phosphate	1,834	1,695
Limestone	0,880	0,837
Salt	0,492	0,471
Inert (kaolin)	0,800	0,800
DL-Methionine	0,238	0,228
L-lysine HCl	0,161	0,176
Vitamin mix ¹	0,100	0,100
Mineral mix ²	0,050	0,050
L-Threonine	0,036	0,036
Cloreto de colina	0,050	0,040
BHT	0,010	0,010
Calculated composition		
Crude protein (%)	20,7900	19,4100
Metabolizable energy (Mcal/kg)	3,0000	3,1000
Calcium (%)	0,8840	0,8240
Available phosphorus (%)	0,4420	0,4110
Sodium (%)	0,2140	0,2050
Digestible amino acids (%)		
Arginine	1,3588	1,2545
Lysine	1,1641	1,0908
Methionine + Cysteine	0,8199	0,7799
Methionine	0,5329	0,5070
Threonine	0,7520	0,7031
Tryptophan	0,2361	0,2178

¹ Provided per kg of product: vit. A: 7.500.000 UI; vit. D₃: 2.500.000 UI; vit. E: 18.000 mg; vit. K₃: 1.200 mg; vit. B₁: 1.500 mg; vit. B₂: 5.500 mg; vit. B₆: 2.000 mg; vit. B₁₂: 12.500 mcg; biotin: 67 mg; pantothenic acid: 10 g; niacin: 35 g;

² Provided per kg of product: Mn: 120 g; Cu: 13 g; Fe: 60 g; Zn: 100 g; I: 2500 mg; Se: 500 mg

Blood samples from two broilers per experimental unit, totalling 60 broilers, were collected at 40 days old for haematological analysis. Samples for haemogram and leukogram analyses were packed in tubes containing ethylenediaminetetraacetic (EDTA) anticoagulant. Samples for biochemical analyses were conditioned in tubes without anticoagulant. The samples were transported under refrigeration and sent to the Laboratory of Clinical Pathology of the Veterinary Institute of the Federal Rural University of Rio de Janeiro for analysis.

The total plasma protein level was determined by refractometry (Coles, 1984). The level of haemoglobin was assayed by the oxy-haemoglobin method with the samples being centrifuged at 1000 × g for 10 min to determine the optical density using a Klett Summerson® colorimeter and with the value being corrected for the haemoglobin unit (Campbell & Dein, 1984). Haematocrit was determined by the micro-haematocrit method, according to the methodology described by Jain (1993). Red blood cell and total leukocyte counts were performed in a haemocytometer using Natt & Herrick's solution (Natt & Herrick, 1951). The blood was diluted 1/100 and 1/200 for the leukocyte and red cell counts, respectively. Leukocytes from the central 1 mm³ quadrant (25 subdivisions of the improved central haemocytometer reticulum) were

counted and the red cells were counted in this same quadrant in $1/5 \text{ mm}^3$ (five subdivisions of the improved central haemocytometer reticulum). The correction factors for total leukocyte and erythrocyte counts were the cell counts multiplied by 1000 and 10,000, respectively, taking into account the area and height of the chamber as well as the dilution. For the differential leukocyte count, blood smears were stained with Giemsa to determine relative values and subsequently the absolute values for lymphocytes, heterophils, and monocytes. The mean corpuscular volume (MCV) and mean corpuscular haemoglobin concentration (MCHC) were calculated using the following standardized formulas (Wintrobe, 1933; Haney *et al.*, 1992):

$$\text{MCV (fL)} = \frac{\text{haematocrit(\%)} \times 10}{\text{RBC}(\mu\text{L}) \times 10^{-6}}$$

$$\text{MCHC(\%)} = \frac{\text{haemoglobin(g/dL)} \times 100}{\text{haematocrit(\%)}}$$

After two hours at ambient temperature, the collected blood without anticoagulant was centrifuged at $400 \times g$ for 5 min to harvest the serum and then stored at $-20 \text{ }^\circ\text{C}$. Analyses of liver and renal function, including the activities of aspartate aminotransferase (AST) (K048, Bioclin, Brazil), alanine aminotransferase (ALT) (K049, Bioclin, Brazil), gamma-glutamyl transferase (GGT) (K080, Bioclin, Brazil), alkaline phosphatase (ALP) (K021, Bioclin, Brazil), creatinine (K067, Bioclin, Brazil), urea (K056, Bioclin, Brazil), and uric acid (K039, Bioclin, Brazil) were performed using a commercial kit and a Bio-2000 automatic spectrophotometer (Bioplus, São Paulo, Brazil), with automatic calibration and high performance reading. The assays were performed using the kinetic method, according to the manufacturer's instructions.

The experiment was conducted in a randomized block design with five treatments and six replicates, with five metal batteries composed of three levels, with each floor representing one block, totalling three blocks per battery and 30 assessments in total.

The results were submitted to analysis of variance, and means were compared using the Student-Newman-Keuls test at 5% ($P < 0.05$) using SISVAR version 5.3 software (Ferreira, 2011).

Results and Discussion

No effects of the treatments were observed ($P > 0.05$) for feed intake, weight gain, and feed conversion (Table 2) in the initial phase. The average feed consumption was greater than that suggested for the manual feeding of Cobb 500 broilers (Cobb-Vantress-Brasil, 2008), indicating that any health challenge to which the broilers were subjected with 5% bedding mixed in the diet did not affect the variables. The performance results in the initial phase are in agreement with those obtained by Kirkpinar *et al.* (2011) and Alp *et al.* (2012), who found no difference for feed intake, weight gain and feed conversion at day 21 in broilers administered 300 mg OEO/kg diet. Zanini *et al.* (2013) also observed no significant difference in weight gain, either in the initial phase or in the entire experimentation phase, when broiler feed was supplemented with OEO at 0.5 g/kg and 1 g/kg. Franciosini *et al.* (2016) reported that when feed was supplemented with a mixture of aqueous extracts of oregano and rosemary (2 g/kg), broilers presented a significant improvement in weight gain compared with the control treatment without the extracts.

From day 1 to day 39, chicks treated with the OEO300 diet displayed higher feed intake than those in the negative control group ($P < 0.05$). A similar result was obtained by Tekce & Gül (2016) when administering OEO (*Origanum syriacum*) to broilers. Hussein *et al.* (2015) stressed that even though phytogetic additives are regarded as palatability stimulants, further studies were needed to corroborate this.

Weight gain was not affected ($P > 0.05$) by increasing OEO levels. A similar result was reported by Alp *et al.* (2012) and Mohiti-Asli & Ghanaatparast-Rashti (2015), who observed no significant difference in weight gain in broilers supplemented with OEO at 300 mg/kg and 500 mg/kg of diet, respectively, compared with the control group.

At the end of the 1- to 39-day period, the lowest feed conversion ($P < 0.05$) was observed in the broilers from the positive control group ($P < 0.05$). Feed conversion in the broilers treated with the three OEO levels did not differ significantly from that of broilers in the negative control group. A similar result was obtained by Rizzo *et al.* (2010), who observed that broilers treated with antibiotics displayed a lower feed conversion than broilers administered a commercial mixture of oregano and cinnamon essential oils, as well as those of the negative control group. Essential oils can have an important impact on feed conversion and stabilization of microbial populations, and can also increase nutrient absorption (Alagawany *et al.*, 2018). The viability and PEI of the broilers in present research were not significantly affected by the treatments.

Table 2 Performance of broilers of broilers supplemented with different levels of oregano essential oil or antibiotic

Performance measures	Treatments ¹					CV (%)	P-value
	NC	PC	OEO (mg/kg)				
			300	600	900		
1 to 21 days old							
Feed consumption (g)	1290	1271	1286	1245	1231	4.61	0.346
Weight gain (g)	888	877	913	871	862	4.98	0.326
Feed conversion (g/g)	1.45	1.45	1.41	1.43	1.43	3.15	0.432
1 to 39 days old							
Feed consumption (g)	4275 ^b	4397 ^{ab}	4454 ^a	4349 ^{ab}	4402 ^{ab}	2.21	0.045
Weight gain (g)	2579	2531	2675	2620	2614	3.83	0.185
Feed conversion (g/g)	1.66 ^a	1.74 ^b	1.66 ^a	1.66 ^a	1.69 ^a	2.53	0.014
Viability (%)	98	93	100	97	93	6.51	0.284
productive efficiency index (%)	262	257	272	266	266	3.57	0.150

Row means with different letters differ statistically at $P < 0.05$ by the Student-Newman-Keuls (SNK) test

¹NC: negative control, PC: positive control (colistin sulfate), OEO: oregano essential oil

Treatment with OEO300 resulted in higher carcass yield ($P < 0.05$) compared with the positive control (Table 3). Peng *et al.* (2016) reported higher carcass yield in broilers treated with OEO at 300 and 600 mg/kg diet than in those from the positive control group.

The broilers treated with the OEO900 diet had higher levels ($P < 0.05$) of abdominal fat compared with those from the negative and positive control groups ((Table 3). The broilers treated with OEO300 and OEO600 presented intermediate values of abdominal fat compared with the other treatments.

Deposition of abdominal fat is an unfavourable characteristic since there is a strong and positive correlation between abdominal fat deposition and fat in the carcass, which depreciates the carcass (Gaya *et al.*, 2006). Unlike in this study, Calislar *et al.* (2009) observed no difference in the relative weight of abdominal fat of broilers administered 300, 500, and 700 ppm of an OEO-based commercial product. In a gene expression analysis of intraepithelial lymphocytes from broiler intestines, Kim *et al.* (2010) concluded that carvacrol, the main constituent of OEO, modulates lipid metabolism-associated gene expression. Lillehoj *et al.* (2011) observed that broilers treated with carvacrol displayed altered lipid, linoleic acid, and androgen and oestrogen metabolism. In agreement with these studies, Peng *et al.* (2016) observed that broilers treated with 600 mg OEO/kg had a lower percentage of abdominal fat than broilers treated with 300 mg oregano/kg diet and those in the negative and positive control groups. In contrast, Nieto *et al.* (2018) did not observe any difference in abdominal fat in broilers treated with essential oils from two oregano species.

The data presented in Table 3 reveal that there was no effect ($P > 0.05$) of the various treatments on the relative weights of the liver, gizzard, heart, bursa, and spleen. These results are in agreement with those of Corduk *et al.* (2013), who found no difference in the relative weight of the liver and gizzard with administration of 250 mg OEO/kg. Conversely, Koyama *et al.* (2014) observed increased bursa weight in broilers from the negative control group and those treated with a mixture of essential oils compared with broilers from the positive control group and those that received less varied essential oil mixtures. The bursa of Fabricius and thymus are the main immune organs in broilers. Maturation of B lymphocytes and production of antibodies occurs in the bursa of Fabricius (Lebien & Tedder, 2008), which is strategically located at the end of the gastrointestinal tract, allowing contact between cells of the immune system and antigens in the diet (Sharma, 1998). The higher the microbial load, the more active the immune system becomes (Thomke & Elwinger, 1998).

Table 3 Carcass, breast, leg, and thigh yield, and relative weights of abdominal fat and edible offal of broilers supplemented with different levels of oregano essential oil or antibiotic

Trait	Treatments ¹					CV (%)	P-value
	NC	PC	OEO (mg/kg)				
			300	600	900		
Yield (%)							
Carcass	69.27 ^{ab}	68.72 ^b	70.16 ^a	69.59 ^{ab}	70.01 ^{ab}	1.82	0.028
Breast	37.18	38.30	38.28	38.02	38.25	4.87	0.540
Leg	15.14	14.45	14.84	15.15	14.96	5.46	0.217
Thigh	16.98	16.78	16.50	16.64	16.19	6.00	0.383
Wing	11.18 ^{ab}	11.67 ^a	10.73 ^b	11.35 ^{ab}	11.12 ^{ab}	5.91	0.019
Back	19.58	18.90	19.70	18.83	19.52	5.88	0.197
Relative weight (%)							
Abdominal fat	1.10 ^a	1.13 ^a	1.37 ^{ab}	1.43 ^{ab}	1.63 ^b	32.07	0.020
Liver	2.50	2.62	2.43	2.41	2.62	15.6	0.542
Gizzard	1.80	1.88	1.67	1.81	1.83	13.09	0.260
Heart	0.73	0.74	0.69	0.67	0.74	13.71	0.332
Bursa	0.13	0.11	0.10	0.10	0.10	26.03	0.052
Spleen	0.18	0.19	0.18	0.17	0.16	27.82	0.637

Means with different letters in the same row differ statistically at $P < 0.05$ by the Student-Newman-Keuls (SNK) test

¹NC: negative control, PC: positive control (colistin sulfate), OEO: oregano essential oil

All haemogram-related blood parameters were influenced by treatments ($P < 0.05$) (Table 4), although the negative control values were within normality for broilers, according to Jain (1993), Cardoso & Tessari (2003), and Campbell (2007). According to these authors, blood values may be influenced by nutritional status, gender, age, season, trauma, breeding, and environmental stress. Therefore, a negative control that is submitted to the same experimental conditions is essential to evaluate these and other physiological parameters, and is considered more reliable than external reference values.

Table 4 Blood profile for broilers supplemented with different levels of oregano essential oil or antibiotic

Blood profile measures	Treatments ¹					CV (%)	P-value
	NC	PC	OEO (mg/kg)				
			300	600	900		
Red blood cells ($\mu\text{L} \times 10^6$)	2.28 ^a	2.27 ^a	2.43 ^b	2.68 ^{bc}	2.84 ^c	20.34	0.029
Haematocrit (%)	32.58 ^a	24.67 ^b	33.25 ^a	33.17 ^a	32.67 ^a	7.91	0.000
Haemoglobin (g/dL)	7.62 ^c	6.40 ^d	8.81 ^a	8.67 ^{ab}	8.25 ^b	7.08	0.000
Plasma proteins (g/dL)	4.50 ^a	3.68 ^b	4.67 ^a	4.38 ^a	4.25 ^a	11.43	0.000
Mean corpuscular volume (fL)	139.51 ^a	100.30 ^c	125.27 ^b	125.58 ^b	115.77 ^b	11.34	0.000
Mean corpuscular haemoglobin concentration (%)	32.63 ^a	26.10 ^b	33.09 ^a	32.98 ^a	29.20 ^a	12.12	0.000

Means with different letters in the same row differ statistically ($P < 0.05$) by the Student-Newman-Keuls (SNK) test

¹NC: negative control, PC: positive control (colistin sulfate), OEO: oregano essential oil

Compared with the negative and positive controls, supplementing the diet with OEO resulted in a marked increase ($P < 0.05$) in red blood cell and haemoglobin levels. The lowest haematocrit was recorded in the positive control ($P < 0.05$), with no difference being observed between treatments. According to Campbell (2007), the normal haematocrit of broilers can vary from 35% to 55%, representing a broad range and making this parameter too unreliable to correlate with the treatments and be used as isolated data.

The MCV was significantly lower in all the blood samples from broilers treated with OEO at various levels. However, these values were higher than those in broilers treated with the antibiotic colistin sulfate (positive control), and microcytosis was observed in OEO-treated broilers. Broilers from the positive control group had the lowest haematocrit, MCHC, and plasma protein levels ($P < 0.05$), which together could be

indicative of anaemia and liver dysfunction. These results corroborate those obtained by Tollba *et al.* (2010), who observed that haematocrit, haemoglobin, and plasma protein levels increased in broilers treated with 1 g or 2 g phyto-genic additives/kg diet, including oregano, compared with control broilers. Levkut *et al.* (2011) observed that supplementing the diet with OEO resulted in a higher concentration of plasma proteins only at 29 days compared with the control group, while no significant effect was observed in the total period (42 days). In contrast, Traesel *et al.* (2010) did not observe a significant difference in plasma protein concentrations between broilers treated with a mixture of essential oils (oregano, sage, rosemary, and pepper) at 100 mg/kg and control groups.

The leukogram results are presented in Table 5. A significant increase in total and differential leukometry was observed in the OEO600 and OEO900 groups compared with the control and OEO300 groups, indicating that supplementing OEO at 300 mg/kg in feed can be used without significantly changing leukogram values. The differential leukometry indicated that lymphocytes were affected most by the treatment, with a lower value being recorded for the positive control and a higher relative value for the OEO treatments compared with the negative control. The 600 and 900 mg OEO/kg levels apparently triggered a response above the normal values reported by Jain (1993), Cardoso & Tessari (2003), and Campbell (2007), being an expressive non-specific stimulus of immune-mediated response. In contrast, a significant decrease was observed in the total and differential leukometry of the antibiotic-administered group, suggesting either a compromised cell-mediated response owing to depression of leukopoiesis or a decrease in the number of peripheral leukocytes (Schmidt *et al.*, 2007). According to Hong *et al.* (2012), oregano supplementation increased antibody levels significantly in broiler chickens as a potential stimulator of the immune system. Silva-Vázquez *et al.* (2018) also observed an increase in leukocyte and lymphocyte numbers when treating broilers with two types of OEO.

Table 5 Leukogram values for broilers supplemented with different levels of oregano essential oil or antibiotic

Leukogram measures	Treatments ¹					CV (%)	P-value
	NC	PC	OEO (mg/kg)				
			300	600	900		
Leukocytes ($\mu\text{L} \times 10^3$)	31.33 ^a	25.83 ^b	33.58 ^a	45.83 ^c	46.92 ^c	16.15	0.000
Lymphocytes ($\mu\text{L} \times 10^3$)	20.50 ^a	14.42 ^b	22.12 ^c	24.16 ^d	26.36 ^e	15.50	0.000
Heterophils ($\mu\text{L} \times 10^3$)	8.73 ^a	8.82 ^a	9.27 ^a	16.86 ^b	13.76 ^c	28.32	0.000
Monocytes ($\mu\text{L} \times 10^3$)	2.21 ^a	2.53 ^a	2.08 ^a	4.75 ^b	6.80 ^c	30.94	0.000
Heterophil /lymphocyte ratio	0.43 ^a	0.62 ^b	0.42 ^a	0.61 ^b	0.51 ^b	24.06	0.000

Means with different letters in the same row differ statistically at $P < 0.05$ by the Student-Newman-Keuls (SNK) test

¹NC: negative control, PC: positive control (colistin sulfate), OEO: oregano essential oil

Treatment with OEO300 resulted in a heterophil/lymphocyte ratio (H/L) within the normal reference range (negative control). Long-term stress such as nutritional deficiency can affect the H/L ratio, and the magnitude of the changes in leukocyte proportions depends on stress intensity and persistence (Cirule *et al.*, 2012).

The broilers from the positive control group displayed increased activities of the hepatic enzymes ($P < 0.05$) AST, GGT, and ALP compared with the other treatments (Table 6). The increases in AST and GGT enzyme activities in the positive control group suggest a significant change in the liver. Some studies have reported that this increase may be because of hepatocyte damage resulting from necrosis or alterations in cell membrane permeability and can be attributed to recent liver dysfunction (Luedde *et al.*, 2014; Yang *et al.*, 2015). Levkut *et al.* (2011) also observed increased ALP activity in broilers from the control group compared with those treated with OEO in the diet. Increased levels of the enzyme ALP can also represent hepatic involvement, although this enzyme is not sensitive to a diagnosis because of its low activity in hepatobiliary tissue (Campbell, 2007). Reis *et al.* (2018) found no differences in the AST or ALT activities of broilers supplemented with a phyto-genic additive composed of carvacrol, thymol, and cinamaldehyde. Carvacrol, the major constituent of oregano essential oil, exhibits hepatoprotective activity (Alagawany *et al.*, 2015).

The increase in serum urea observed with all the treatments, and that for uric acid in the OEO900 and positive control groups, may be due to the higher coefficient of apparent nitrogen metabolism obtained in the broilers from this group (Dias *et al.*, 2015). Nonetheless, the possibility of impaired renal function and metabolic disorders in broilers treated with the highest OEO levels cannot be ruled out. Increases in the

serum levels of these metabolites occur when 30% or less of the kidney is functional, or because of pre- and post-renal causes (Fudge & Joseph, 2000; Kaneko, 2008).

Table 6 Serum biochemistry values for broilers supplemented with different levels of oregano essential oil or antibiotic

Enzymes/metabolites	Treatments ¹					CV (%)	P-value
	NC	PC	OEO (mg/kg)				
			300	600	900		
Aspartate aminotransferase (IU/L)	276.43 ^a	487.91 ^b	276.98 ^a	327.65 ^a	329.61 ^a	32.16	0.000
Alanine aminotransferase (IU/L)	28.81 ^a	36.28 ^b	28.67 ^a	32.59 ^c	32.59 ^c	20.82	0.038
Gamma glutamyltransferase (IU/L)	10.25 ^a	19.09 ^b	7.61 ^a	11.67 ^a	11.67 ^a	35.32	0.000
alkaline phosphatase (IU/L)	1733 ^a	2278 ^b	1781 ^a	1815 ^a	1815 ^a	20.63	0.007
Urea (mg/dL)	0.47 ^a	0.57 ^b	0.45 ^a	0.55 ^b	0.67 ^b	21.22	0.000
Creatinine (mg/dL)	5.50 ^{ab}	4.33 ^b	4.58 ^b	4.33 ^b	6.75 ^a	30.97	0.001
Uric acid (mg/dL)	15.13 ^a	22.33	15.14 ^a	17.90 ^a	22.80 ^b	28.91	0.000

Means with different letters in the same row differ statistically ($P < 0.05$) by the Student-Newman-Keuls (SNK) test

¹NC: negative control, PC: positive control (colistin sulfate), OEO: oregano essential oil

Conclusions

Broilers treated with OEO displayed a stimulated immune system, while broilers treated with 300 mg OEO/kg of diet presented values within the reference range. The use of OEO in the diets did not harm the performance and carcass characteristics of broilers. At a concentration of 300 mg/kg, OEO can be used in broiler diets as a phyto-genic additive, especially in diets without antimicrobial performance enhancers.

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Authors' Contributions

GE, AVCG and CARL designed the study, collected and analysed the data, drafted and revised the manuscript. LSLA, BST and FD collected the data. GE, AVCG, FD, CARL and VSC analysed the data and revised the manuscript.

Conflict of Interest Declaration

The authors declare that there was no conflict of interest.

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