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Clinical outcome of intravitreal gentamicin injection for the treatment of end-stage glaucoma in five rabbits (eight eyes)

Gemma Turner^{1*} , Allyson Groth¹ , Gerry Skinner² , Hayley Volk¹ , Anu O'Reilly¹ 
and Chloe Hardman¹ 

¹Melbourne Eye Vet, Mulgrave, Australia
²The Rabbit Doctors, Collingwood, Australia

Abstract

Background: Glaucoma is a painful and blinding condition that occurs in many species, including rabbits. When medication is no longer effective in maintaining intraocular pressure (IOP), enucleation is the recognized treatment for rabbits with end-stage glaucoma. However, this procedure carries risks relating to the procedure and the anesthesia.

Aim: The aim of this retrospective study was to report the efficacy of intravitreal gentamicin injection in controlling IOP in blind eyes of rabbits with end-stage glaucoma. Ocular and non-ocular complications were retrospectively assessed.

Methods: Medical record review was performed to identify five client-owned rabbits (eight eyes) that were treated by intravitreal injection of 6–20 mg of gentamicin per eye (median 7.18 mg/kg) for chronic, end-stage glaucoma. Treatment was unilateral in two and bilateral in three rabbits. IOP control was assessed via rebound tonometry readings performed approximately 2 weeks, 1 month, 3 months, and 6 months after injection. Total follow-up was between 313 and 1,111 days. Ocular complications were recorded and systemic health was estimated by the owner-answered questionnaire and changes in body weight.

Results: IOP was <25 mmHg in 87.5% of eyes 3 months post-injection. The most common ocular complications were cataracts (62.5%), anterior uveitis (25%), retinal detachment (12.5%), and corneal erosion (12.5%). There were no behavioral or body weight changes suggestive of systemic complications.

Conclusion: 87.5% of rabbit eyes treated with intravitreal gentamicin had controlled IOP 3 months after injection. All eyes were blind at the time of injection. Ocular side effects were common. Investigation of the safety and systemic effects of intravitreal gentamicin injection is required; however, no overt complications were identified in treated rabbits in this study.

Keywords: Chemical ablation, Ciliary body ablation, Gentamicin, Glaucoma, Rabbit.

Introduction

Glaucoma is a painful and blinding condition in animals and people (Bouhenni *et al.*, 2012), and affects three out of 1,000 rabbits (Innes and Williams, 2018), which is higher than their dog and cat counterparts (Innes and Williams, 2018). Despite this, there is less information available about glaucoma in rabbits. This may be because glaucoma is not as easily identifiable in rabbits as pain and distress are difficult to detect and quantify in prey species (Leach *et al.*, 2011). Not only is glaucoma painful, but it can also cause secondary complications including buphthalmia, which can then lead to other painful and globe-threatening conditions such as non-ulcerative and/or ulcerative keratitis (Andrew, 2002). Medical treatment for glaucoma, using topical hypotensive medications, has been shown to be effective in rabbits (Yuschenoff *et al.*, 2020). When these drugs are no longer effective, enucleation is

the recognized treatment for rabbits with end-stage glaucoma (Diehl and McKinnon, 2016; Yuschenoff *et al.*, 2020). Enucleation has its inherent risks relating to both the surgical procedure and the anesthetic. Rabbits have a large orbital vascular sinus in the ventronasal orbit which, if traumatized, can cause life-threatening hemorrhage (Diehl and McKinnon, 2016). Rabbits also have an increased risk of death related to anesthesia of 1.39% to 4.08% compared to 0.05% and 0.11% in dogs and cats, respectively (Brodbeck *et al.*, 2008; Lee *et al.*, 2018).

As an alternative to enucleation for end-stage glaucoma in dogs, intravitreal gentamicin injections have been readily used with reported success rates varying from 65% to 95% (Bingaman *et al.*, 1994; Rankin *et al.*, 2015; Julien *et al.*, 2020). The exact mechanism of how gentamicin affects the ciliary body epithelium is not fully understood. Intravitreal gentamicin injection

*Corresponding Author: Gemma Turner. Melbourne Eye Vet, Mulgrave, Australia.
Email: info@melbournseevet.com.au

procedures are often utilized in dogs due to their lower cost compared with enucleation, as well as the perceived safety advantages afforded by either requiring sedation only or requiring a shorter duration of general anesthesia as compared to enucleation (Julien *et al.*, 2020).

Systemic absorption of gentamicin, an aminoglycoside antibiotic, is of concern due to its known nephrotoxic effects via tubular necrosis (Kosek *et al.*, 1974). Doses of 4 mg/kg subcutaneously twice daily for 4 weeks have been experimentally shown to cause renal failure and subsequent death in healthy rabbits by 20–27 days post-injection (Ginsburg *et al.*, 1976). In dogs, a study by Rankin *et al.* (2015) has demonstrated that the total plasma concentration of gentamicin post intravitreal injection ranges from 0.21 to 9.71 µg/ml after 25–40 mg of gentamicin per eye with a mean gentamicin cMAX of 2.29 µg/ml at 2.54 hours post-injection. Comparatively, the systemic absorption of intravitreal gentamicin in rabbits is not known, however, it is thought to leave the eye predominantly via the aqueous humor (Kane *et al.*, 1981).

The purpose of this article was to retrospectively investigate the efficacy of intravitreal gentamicin injections in blind eyes of rabbits with end-stage glaucoma.

To the authors' knowledge, this is the first publication investigating the outcomes of intravitreal gentamicin injections for end-stage glaucoma in rabbits.

Materials and Methods

Case selection

Medical records were retrospectively reviewed to identify rabbits with end-stage glaucoma that were treated with intravitreal gentamicin between 2019 and 2021 at an Australian Veterinary Ophthalmology specialist hospital. Records were included when there was documented ophthalmic examination by a board-certified Veterinary Ophthalmologist and a minimum of 6 months follow-up. End-stage glaucoma was defined as a blind eye (absent menace response, absent dazzle reflex, and absent direct and consensual pupillary light reflex) and intraocular pressure (IOP) ≥ 25 mmHg as estimated by rebound tonometry (TonoVet®/TonoVet Plus®) despite the use of topical glaucoma medications. Pre-procedural data gathered included signalment, body weight, eye affected, concurrent ocular disease, ocular and systemic medications, and IOP measurements. Procedural data was collected including anesthetic used, variations in treatment protocol including dosage of gentamicin, dosage, and site of administration of dexamethasone, and post-operative ocular and systemic medication.

Surgical procedure

Written consent was obtained from owners prior to the procedure. Rabbits were sedated using a protocol preferred by the Veterinary Ophthalmologist performing the procedure and included ketamine (10 mg/kg), butorphanol (0.5 mg/kg), and medetomidine (0.1 mg/

kg) intramuscularly for four rabbits and fentanyl (5 mcg/kg) and midazolam (0.2 mg/kg) intravenously for one rabbit.

Intravitreal injection was performed as described in dogs (Rankin *et al.*, 2015). The eye(s) were aseptically prepared with 1% betadine (Povidone-Iodine solution 10%), followed by the application of topical proparacaine hydrochloride 0.5%. Vitreocentesis was performed using a 25-gauge, 5/8-inch hypodermic needle inserted 6–8 mm posterior to the dorsal limbus, and approximately 0.3–0.5 ml of vitreous was aspirated. Gentamicin sulfate (100 mg/ml) was then injected into the posterior segment with a dose ranging from 6 to 20 mg of gentamicin sulfate per eye (maximum of 40 mg per rabbit). Dexamethasone sodium phosphate (3 mg/ml) was injected subconjunctivally in one eye of one rabbit and intravitreally in four eyes of two rabbits. Three eyes of two rabbits were not treated with dexamethasone.

Post-injection treatment varied based on IOP readings immediately after injection and included aqueocentesis and the use of topical anti-glaucoma medications.

Post-operative frequency and type of hypotensive treatment varied but included latanoprost 0.005% (Xalatan; Pfizer Australia Pty Ltd), dorzolamide hydrochloride 2% (Trusopt; Mundipharma Pty Ltd), and brinzolamide 1% (Azopt; Alcon Laboratories Pty Ltd). Other topical medication included diclofenac sodium 0.1% (Voltaren; Novartis Pharmaceuticals Australia Pty Ltd), ketorolac trometamol 0.5% (Acular; Allergan Australia Pty Ltd), and tacrolimus 0.02% (Bova Australia Pty Ltd). All patients received meloxicam oral liquid 1.5 mg/ml (Metacam; Boehringer Ingelheim Pty Ltd) for pain relief.

Post-procedure monitoring

Data from each revisit included an evaluation of systemic health, as assessed by questions to owners regarding appetite, fecal production, general well-being/activity, and any signs of ocular discomfort. Other data included the rabbit's body weight (measured to 10 g accuracy), IOP results on the treated eye(s), and ocular examination findings, including complications, which were also recorded. The results were collated to time points at approximately 2 weeks, 1 month, 3 months, and 6 months, as well as the last recorded examination.

A retrospective assessment of the rabbits' general health was estimated by a survey answered by the owners with questions based on clinical signs pertaining to pain, as described by the Rabbit Grimace Scale (Keating *et al.*, 2012) and general signs known to be associated with renal injury (Appendices 1 and 2).

The success of the procedure was defined as lowering the IOP to <25 mmHg with or without topical hypotensive medications by 3 months post-procedure.

Ethical approval

The authors confirm that the ethical policies of the journal, as noted on the journal's author guidelines page, have been adhered to. All animals that participated in

this study were client-owned and joined the study after the owner's written consent.

Results

Clinical findings

Eight eyes of five rabbits (three female, two male) were included. The median age of the five rabbits at the time of the procedure was 5.4 years (range of 1.1–7.9 years). Breeds represented included four mini lops and one Blanc de Hotot. The procedure was performed in both eyes of three rabbits, and one eye in two rabbits. Glaucoma was present for a median of 81 days (range of 3–232 days) prior to the procedure. The cause of glaucoma was not specified in all eight eyes.

Procedural outcome

The dose of gentamicin ranged from 6 to 20 mg per eye, with a median of 7.18 mg/kg of gentamicin per rabbit (ranging from 4.2–10.7 mg/kg). The dose of dexamethasone per eye ranged from 0.4 mg to 1 mg per eye, with an average of 0.53 mg/kg of dexamethasone phosphate per rabbit (ranging from 0.18–0.98 mg/kg) (Table 1).

The median duration of follow-up from procedure to final re-examination was 612 days (313–1,111 days). IOP was below 25 mmHg at 2 weeks (12–17 days) in 4/6 eyes (66.7%). Results from two eyes were not available at this time. The IOP was also below 25 mmHg at 1 month (24–35 days) in 6/8 eyes (75%), 3 months (75–84 days) in 7/8 eyes (87.5%), and in 7/7 eyes (168–191 days) post-procedure. One eye was enucleated at 70 days post-procedure due to a persistently high IOP (30 mmHg) and an epithelial erosion and was not included in the 6-month data point.

Median IOP was reduced at 2 weeks (median IOP = 19 mmHg), one (median IOP = 21 mmHg), three (median IOP = 13 mmHg), and 6 months (median = 13 mmHg) compared to the pre-operative IOP results (mean = 44 mmHg) as shown in Figure 1.

The number and frequency of glaucoma drops per eye per day were reduced compared to pre-procedural frequency (median of 3.8 drops) versus at 2 weeks (median 2.4 drops), 1 month (median 2 drops), 3 months (median 0.4 drops), and 6 months (median 0.4 drops).

Ocular complications

Ocular complications included progression of cataracts in 5/8 eyes (62.5%), anterior uveitis in 2/8 eyes (25%), retinal detachment in 1/8 eyes (12.5%), and epithelial erosion in 1/8 eyes (12.5%).

Further surgical intervention was required in 3/8 eyes (37.5%), including repeat gentamicin injection at day 41 in 2/8 eyes (25%) and enucleation at day 70 in 1/8 eyes (12.5%).

Systemic complications

Body weight was measured at each visit using the same scales. Four rabbits had a <1% weight loss and one rabbit gained 4% of its body weight from the time of the procedure to 13 days after the procedure.

Table 1. Individual results for the eight eyes (five rabbits) that received intravitreal gentamicin injection for end-stage glaucoma.

	Rabbit 1		Rabbit 2		Rabbit 3		Rabbit 4		Rabbit 5	
	Eye 1	Eye 2	Eye 3	Eye 4	Eye 5	Eye 6	Eye 7	Eye 8	Eye 9	Eye 10
Pre-procedural IOP	53 + TVP	49 + TVP	26 + TV	61 + TV	39 + TV	46 + TV	43 + TVP	31 + TV		
IOP (2 weeks) (mmHg)	35 + TVP	16 + TVP	N/A	N/A	28 + TVP	19 + TVP	3 + TVP	15 + TV		
IOP (1 month) (mmHg)	22 TV	13 TV	22 + TV	16 + TV	29 + TVP ^a	35 + TVP ^a	2 + TVP	25 + TV		
IOP (3 months) (mmHg)	13 TV	17 TV	24 + TV	15 + TV	11 TVP	7 TVP	6 TVP	Removed (day 70)		
IOP (6 months) (mmHg)	16 TVP	19 TVP	19 + TV	11 + TV	9 TVP	9 TVP	6 TVP	N/A		
Dose of gentamicin (mg)	20 mg	20 mg	6 mg	10 mg	10 mg	10 mg	10 mg	10 mg		
Dose and site of dexamethasone injection	1 mg vitreous	1 mg vitreous	None	None	0.5 mg vitreous	0.5 mg vitreous	0.1 mg subconj	None		
Complications	Anterior uveitis, cataract	Anterior uveitis, cataract	None	None	Repeat injection, retinal detachment, cataract	Repeat injection, cataract	None	Removed (day 70), cataract, corneal erosion		

TV: Tonovet Plus (laprine setting), TV: Tonovet (canine setting), IOP: intraocular pressure, Subconj: subconjunctivally; (+): with hypotensive medication.

^a Repeat injection performed day 41, with 10 mg gentamicin and 0.5 mg dexamethasone intravitreally.

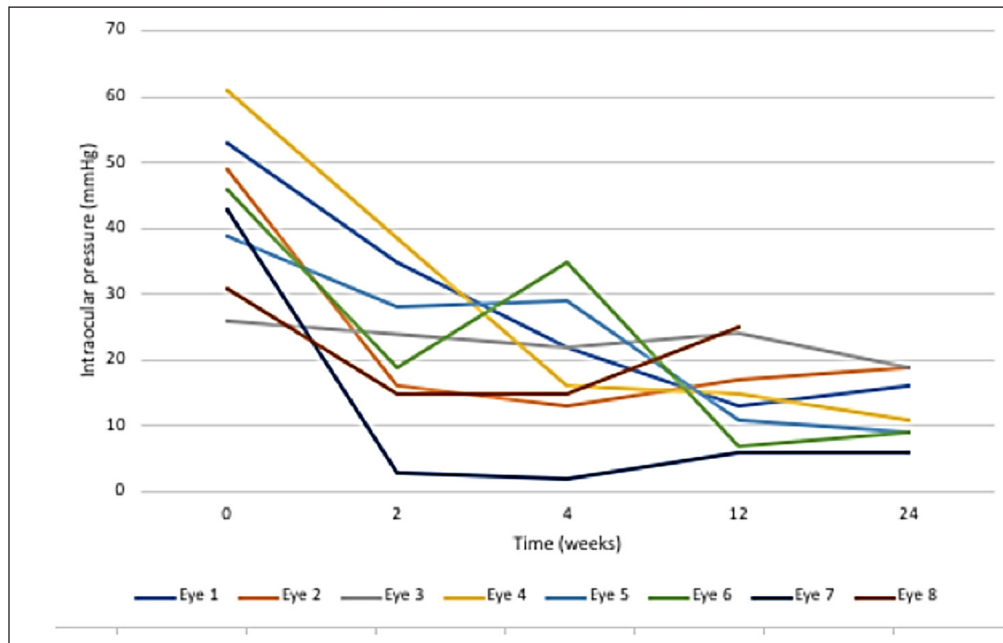


Fig. 1. Intraocular pressures (mmHg) of eight rabbit eyes following intravitreal gentamicin injection.

At the time of article submission, one rabbit (one eye) was euthanized 375 days after the gentamicin injection due to poor mobility secondary to hindlimb arthritis. The remaining four rabbits were alive at the time of writing the article, 313, 307, 956, and 1,111 days post-procedure. Post-procedure hematology and biochemistry were available for one of the five rabbits at 1,058 days after the procedure. The blood tests were performed for reasons unrelated to the study. No abnormalities in parameters relating to renal physiology were noted.

There were no clinical records of owner-perceived discomfort or signs pertaining to an acute kidney injury based on the questions about general appetite, fecal production, demeanor, and signs of ocular discomfort (including squinting and rubbing) posed to the owners at each revisit.

The retrospective questionnaire involved more in-depth questions regarding pain, formulated based on the Rabbit Grimace Scale, and in-depth questions pertaining to possible acute kidney injuries, within the immediate post-procedural period. Questions regarding changes in appetite, hiding, changes to socialization, squinting, lethargy, teeth grinding, vocalization, overt aggression, changes in fecal production, weight loss, changes in drinking, and urination habits and any overt signs of abdominal pain did not highlight any abnormalities. All five rabbit owners stated that they would recommend the injection to friends and family as a treatment option for end-stage glaucoma in rabbits.

Discussion

The success rate of intravitreal gentamicin injection for end-stage glaucoma in rabbits was 87.5% at 3 months. This was similar to the outcome as described by Rankin *et al.* (2015) in dogs at 1-month post-procedure, however, differed from other studies which indicated a success rate as low as 65%, in a retrospective evaluation between 1985 and 1993 (Bingaman *et al.*, 1994), and as high as 95% at 3 months in a retrospective analysis of 108 canine eyes between 2013 and 2019 by Julien *et al.* (2020).

The decision to describe a normal IOP as <25 mmHg was based on the literature on normal IOPs in rabbits (Williams, 2021). Alternative values of normal IOP in rabbits (Pereira *et al.*, 2011) indicate that the values obtained, following the intravitreal gentamicin injections in this study, are still outside of the normal ranges for the values set by Pereira *et al.* (2011). Further investigation is required to determine the true normal IOP in rabbits.

The dose of gentamicin ranged from 6 to 20 mg per eye. The median dose per body weight of intravitreal gentamicin was 7.18 mg/kg and ranged from 4.2 to 10.7 mg/kg. The dose variation between rabbits was based on ophthalmologist preference. Comparatively, the study conducted in dogs by Rankin *et al.* (2015) had a median dose of gentamicin per body weight of 2.57 mg/kg and ranged from 0.61 to 7.50 mg/kg which correlated to a plasma concentration of 0.2–9.7 ul/ml. Comparisons between dogs and rabbits are difficult to interpret as drug metabolism and distribution

differ, however, the authors believe that the total concentration of intravitreal gentamicin should be less than the known systemic dose of gentamicin for rabbits of 8 mg/kg once daily (Plumb, 2020) to reduce risk of nephrotoxicity. Caution should be exercised when considering intravitreal gentamicin in rabbits with known renal insufficiency or those that are receiving other nephrotoxic drugs.

There were two eyes in one rabbit (25%) that required a second intravitreal injection, with the time from the first injection being 41 days from the first injection. In dogs, repeat injection was only required in 12 eyes (11%) of eyes with a mean time from the first injection of 158 days (Julien *et al.*, 2020). The authors recognize that the difference in the percentage of rabbits needing repeat injections compared to dogs could be due to a number of factors including a smaller number of patients, varied gentamicin protocols, and limitations on the total dose of gentamicin per eye that could be safely administered to rabbits, due to rabbits generally being smaller in size compared to dogs.

The most common ocular complication was the progression or development of cataracts in 62.5% of eyes as compared to the study in dogs where the most common ocular complication was phthisis bulbi in 59.2% of eyes (Julien *et al.*, 2020). In this study, obvious signs of phthisis bulbi were not noted in any examination postoperatively. The authors do acknowledge, however, that any reduction in globe size could not be accurately assessed as globe diameters were not measured pre- and post-procedure.

The second most common ocular complication was anterior uveitis in two out of eight eyes (25%), followed by one eye with a corneal epithelial erosion and one eye with a retinal detachment (12.5% incidence each). The authors believe that whilst not insignificant, these complications may be effectively managed through the addition of topical medication. This was shown in the two eyes with anterior uveitis which was managed with the use of topical 0.1% diclofenac twice daily and oral 1.5 mg/ml meloxicam once daily. Resolution of the uveitis was noted on days 17 and 55 post-procedure, right and left eye, respectively.

Systemic complications were difficult to ascertain and were based solely on the lack of a downward trend in body weight and the lack of overt clinical signs of illness at the post-operative revisits and via a retrospective questionnaire at the time of article submission. There was a <1% reduction in body weight in any rabbit, and this was thought to be insignificant. This is a crude measure of overall health; however, it is a common clinical sign seen with many illnesses in rabbits, including kidney disease (Harcourt-Brown, 2013).

Systemic complications associated with the injection were assessed via questions based on owner-perceived assessments of their rabbit. The history taken at the

time of each review was less comprehensive than the retrospective questionnaire. The retrospective nature of assessing subtle behavioral and clinical signs can mean that some of these signs may have been overlooked if the owners were not actively monitoring for these specific signs before and after the procedure. Utilizing the Rabbit Grimace Scale at the time of the procedure, by presenting the image scale to the owner, maybe more useful, as owners may be more aware of the subtle signs of discomfort that may have been inadvertently overlooked.

The main concern associated with the intravitreal gentamicin procedure was whether there was significant systemic absorption of gentamicin which has the potential to cause renal damage. The systemic absorption of intravitreal gentamicin is not known for rabbits with normal, healthy eyes. Furthermore, the absorption of gentamicin from an inflamed eye may be greater than that of a non-inflamed eye. Therefore, without further diagnostics, such as documenting plasma concentrations of gentamicin as performed in dogs by Rankin *et al.* (2015) in dogs, it is difficult to quantify and understand the potential for renal damage. A future prospective study could investigate the total plasma concentration of gentamicin after intravitreal injection in rabbits. The vitreal half-life of gentamicin in rabbits is known to be between 10 and 12 hours in normal rabbit eyes (Kane *et al.*, 1981) and samples can be obtained around this time to determine the peak plasma concentration. Further studies could also look at the role of intravitreal dexamethasone and its systemic absorption and complications. The authors note that, although the role of intravitreal dexamethasone and its systemic absorption and complications were not investigated in this study, no outward side effects were observed.

Limitations of the study were associated with its retrospective nature. These included variations in intravitreal injection protocols, sedation protocols, post-operative medication, and the timing of post-procedural examinations. Variations include, but are not limited to, variation in the volume of aspirated vitreous, dose of gentamicin, type, dose, and injection site used for concurrent glucocorticoids. Data such as histopathology on the enucleated eye in this study may have assisted in understanding the effects of gentamicin in the rabbit, including the identification of intraocular tumors, that have been documented in 39.5% and 62.5% of dogs and cats post gentamicin injection respectively (Duke *et al.*, 2013a, 2013b).

Other limitations of this study include the overall small sample size of eyes, conflicting data available on normal rabbit IOPs, and the difficulty in assessing the systemic effects of gentamicin.

Overall, enucleation is the preferred treatment for a blind, glaucomatous eye refractory to medication as it negates the requirement for any post-procedural

topical medication—whether that is a topical non-steroidal anti-inflammatory or topical glaucoma medication. Intravitreal gentamicin, with or without dexamethasone sodium phosphate injections, can be an appropriate alternative for rabbits as they have both a higher anesthesia risk and a higher risk of substantial blood loss during surgery. Intravitreal gentamicin may also be considered for those owners who have cost-prohibitive reasons to not consider eye removal.

Acknowledgments

None.

Conflicts of interest

The authors declare no conflicts of interest.

Author contributions

Gemma Turner collated data and wrote the manuscript. The procedure was performed by Chloe Hardman, Anu O'Reilly, and Hayley Volk. The manuscript was reviewed and edited by Chloe Hardman, Anu O'Reilly, Allyson Groth, Gerry Skinner, and Hayley Volk.

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Appendix 1.

Questionnaire for rabbit owners.


1. Did you find any of the following **PRIOR** to the gentamicin injection procedure?
 - a. Reduced appetite Y/N/Unsure
 - b. Feces production normal/reduced/none/Unsure
 - c. Hiding Y/N/Unsure

- d. Reduced socialization with people and/or other animals Y/N/Unsure
 - e. Squinting Y/N/Unsure
 - f. Sleeping more/reluctance to move Y/N/Unsure
 - g. Teeth grinding Y/N/Unsure
 - h. Cheek flattening Y/N/Unsure
 - i. Change in nostril shape Y/N/Unsure
 - j. Whiskers point outwards Y/N/Unsure
 - k. Ears held backward Y/N/Unsure
 - l. Vocalization Y/N/Unsure
 - m. Overt aggression Y/N/Unsure
2. Did you find any of the following develop within 24–72 hours **AFTER** the gentamicin injection procedure, if so please comment on the details:
- a. Appetite normal, reduced, none, unsure, lasted for:
 - b. Feces production normal, reduced, none, unsure, lasted for:
 - c. Hiding Y/N/Unsure, stopped:
 - d. Reduced socialization with people and/or other animals Y/N/Unsure, stopped:
 - e. Squinting/closing eyes Y/N/Unsure, stopped:
 - f. Sleeping more/reluctance to move Y/N/Unsure, stopped:
 - g. Teeth grinding Y/N/Unsure, stopped:
 - h. Cheek flattening Y/N/Unsure, stopped:
 - i. Change in nostril shape Y/N/Unsure, stopped:
 - j. Whiskers point outwards Y/N/Unsure, stopped:
 - k. Ears held backwards Y/N/Unsure, stopped:
 - l. Vocalization Y/N/Unsure, stopped:
 - m. Overt aggression Y/N/Unsure, stopped:
3. To your knowledge, did your rabbit develop any of the following signs post gentamicin injection procedure:
- a. Weight loss Y/N/Unsure
 - b. Increased drinking Y/N/Unsure
 - c. Increased urination Y/N/Unsure
 - d. Abdominal pain (i.e. pain on touching the abdomen) Y/N/Unsure
 - e. Weakness Y/N/Unsure
4. To your knowledge, was your rabbit diagnosed with kidney disease prior to the gentamicin procedure?
- a. Y/N, if yes when was the diagnosis?
5. To your knowledge was your rabbit diagnosed with kidney disease after the gentamicin procedure?
- a. Y/N—if yes, when?
6. Based on your experience, would you recommend the procedure to a friend's rabbit or recommend a different treatment (i.e. eye removal)?
- a. Yes, I would recommend the injection.
 - b. No, I would recommend another treatment method.

Appendix 2. Rabbit Grimace Scale (15).



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The Rabbit Grimace Scale

Research has demonstrated that changes in facial expression provide a means of assessing pain in rabbits.

The specific facial action units shown below comprise the Rabbit Grimace Scale. These action units increase in intensity in response to post-procedural pain and can form part of a clinical assessment alongside other validated indices of pain.

The action units should only be used in awake animals. Each animal should be observed for a short period of time to avoid scoring brief changes in facial expression that are unrelated to the animal's welfare.

	Action units		
	Not present "0"	Moderately present "1"	Obviously present "2"
Orbital tightening <ul style="list-style-type: none"> • Closing of the eyelid (narrowing of orbital area) • A wrinkle may be visible around the eye 			
Cheek flattening <ul style="list-style-type: none"> • Flattening of the cheeks. When 'obviously present', cheeks have a sunken look. • The face becomes more angular and less rounded 			
Nostril shape <ul style="list-style-type: none"> • Nostrils (nares) are drawn vertically forming a 'V' rather than 'U' shape • Nose tip is moved down towards the chin 			
Whisker shape and position <ul style="list-style-type: none"> • Whiskers are pushed away from the face to 'stand on end' • Whiskers stiffen and lose their natural, downward curve • Whiskers increasingly point in the same direction. When 'obviously present', whiskers move downwards 			
Ear shape and position <ul style="list-style-type: none"> • Ears become more tightly folded / curled (more cylindrical) in shape • Ears rotate from facing towards the source of sound to facing towards the hindquarters • Ears may be held closer to the back or sides of the body 			

Read the original paper: Keeling DCJ, Thomas AA, Redmill PA, Leach MC (2012) Evaluation of BMLA cream for preventing pain during tail docking of rabbits: Changes in physiological, behavioural and facial expression responses. PLOS ONE 7(8): e44437. doi:10.1371/journal.pone.0044437

For guidance on using the Rabbit Grimace Scale, additional images of each action unit, research papers that underpin the techniques, and for grimace scales in other species, visit: www.nc3rs.org.uk/grimacescales

To request copies of this poster, please email: enquiries@nc3rs.org.uk
 The NC3Rs provides a range of 3Rs resources at www.nc3rs.org.uk/resources
 Images kindly provided by Dr Matthew Leach, Newcastle University
 The Rabbit Grimace Scale would not have been developed without the continuing work of the Pain and Animal Welfare Sciences Group (PAWS) at Newcastle University