

Spinal Cord Studies in the African Giant Rat (*Cricetomys gambianus*, Waterhouse)

Olude M.A.^{1,2}, Idowu A.O.¹, *Mustapha O.A.^{1,2}, Olopade J.O.², Akinloye A.K.¹

¹Department of Veterinary Anatomy, College of Veterinary Medicine, Federal University of Agriculture, Abeokuta, Nigeria. ²Department of Veterinary Anatomy, Faculty of Veterinary Medicine, University of Ibadan, Ibadan, Nigeria.

Summary: The African giant rat, AGR, is known for advantageous behavioural patterns among which are cognition and dexterous locomotion. This study investigated the morphological, morphometric and possible functional aspects of the AGR spinal cord (SC) anatomy. Ten adult (5 males and 5 females) AGR were used to determine the gross and histological features of the SC which were typically of rodent features. The mean SC weight and length given as 2.50 ± 0.24 g and 15.87 ± 0.24 cm respectively for the male and 2.32 ± 0.16 g and 15.40 ± 0.61 cm for the female showed no sexual dimorphism ($p < .05$). A positive linear relationship between the tail length and SC weight were found in both sexes ($r = 0.81$ males; $r = 0.95$ females) suggesting significant contribution of the filum terminale to SC weight. Forty-three internal structures including nuclear aggregations and tracts were traced. Eight nuclear aggregations of neurons involved in nociception and limb coordination were observed to be prominent and larger than in laboratory rats. Same was noted for the dorsal, ventral and lateral funicular tracts which control the limbic system. This study provides morphometric baseline research information and delineates the functional aspects of the AGR SC anatomy. The information provided further strengthens the drive proposing the AGR as an indigenous research model for regional anaesthesia and locomotor disease.

Keywords: African giant rat; spinal cord; spinal tract; nuclei; spinal segment; morphometry.

©Physiological Society of Nigeria

*Address for correspondence: drmustyplato@yahoo.co.uk Phone: ±2348035915275

Manuscript Accepted: 2015

INTRODUCTION

Rodents are the largest mammalian order with about fifty percent of the species of mammals being rodents (Sheet, 1989). The African giant rat (AGR) also known as giant pouched rat, is by size one of Africa's largest rodents and is arguably becoming Africa's most intriguing rodent because of its scientific attributes such as the detection of landmines (Verhagen *et al.*, 2003) and also in the medical diagnosis of pulmonary tuberculosis (Weetjens *et al.*, 2009), disease vectors (Durnez *et al.*, 2008), potential pest species status (Peterson *et al.*, 2006) among many others. Earlier investigations on the AGR were centered on reproduction and sustenance in captivity (Oke and Oke, 1999; Akinloye, 2009) while recent studies have focused on interpreting function from morphology (Olude *et al.*, 2009; Ibe *et al.*, 2014). Information on the CNS however, is sparse with a brain bias (Ibe *et al.*, 2010). The SC anatomy has received much less attention (Vera and Meyer-Siegler, 2003). This study therefore, was undertaken to add to the meagre research data on the gross and histological anatomy of

the SC and interpret functional behaviour from the anatomical knowledge of the AGR SC.

MATERIALS AND METHODS

Animals

Ten adult (5 males and 5 females) African giant rats (*C. gambianus*) were obtained from the wild, stabilized in holding cages designed with dark and light compartments to regulate the sleep and wake cycles of the rats. The average body weight of African giant rats was 0.96 ± 0.05 kg for males and 0.91 ± 0.09 kg for females.

Animal Handling

Experimental procedures conformed to the rules and guidelines issued by the University of Ibadan, Ibadan, on health guide for the care and Use of Animals in Experiments.

Animals were anaesthetized by chloroform inhalation. Gross morphometric parameters (body weight, head length, trunk length and tail length) were immediately measured before animals were perfused transcardially

with 4% paraformaldehyde in 0.1M phosphate-buffered saline (PBS), pH 7.4 and post-fixed in 10% formalin for 1 week. The SC was exposed by dissections and laminectomy, harvested and measured using metric instruments. Sections from cervical, thoracic, lumbar, sacral and coccygeal segments were then taken for histology at 5 µm, routinely stained with Thionin stain. Slides were viewed under light microscope (Leica Model DME Microscope, Model: 13595XXX, Leica Microsystems) and images captured with Canon® Power shot S70 camera (PC 1087, No. 033102132). Photomicrographs obtained were traced onto a tracing paper using an HB graded pencil.

Statistical Analysis

All data were analyzed and expressed as mean and standard error of mean using Graph pad prism 5. Statistical significance was determined using t-test and linear regression ($P \leq 0.05$).

RESULTS

Morphometry

The mean SC weight and length were recorded as $2.41 \pm 0.14g$ and $15.63 \pm 0.32cm$ respectively. The mean body measurements and SC measurements were greater in the males than in the females but were all statistically insignificant (Table 1). There was positive relationship between the trunk length and the SC length which was significant for females ($p = 0.0141$) but not for males ($p = 0.1999$). The strength of relationship between the tail length and SC weight were significant in both sexes ($p = 0.037$ males, $p = 0.0044$ females) and were both positively correlated (Figure 1). There was positive relationship between the trunk length [TKL] and the SC length [SCL] which was significant for females ($p = 0.0141$) but not for males ($p = 0.1999$). The strength of relationship between the tail length [TL] and SC weight [SCW] were significant in both sexes ($p = 0.037$ males, $p = 0.0044$ females) and were both positively correlated.

Table 1. Body and Spinal Cord measurements of the AGR (*C. gambianus*), Mean \pm SEM

PARAMETERS	MALE	FEMALE	OVERALL
Body weight (kg)	0.96 ± 0.05	0.91 ± 0.091	0.93 ± 0.05
Trunk length (cm)	24.70 ± 1.30	24.10 ± 1.23	24.40 ± 0.85
Tail length (cm)	33.80 ± 1.56	33.20 ± 0.98	33.5 ± 0.88
Head length (cm)	6.94 ± 0.31	6.78 ± 0.34	6.86 ± 0.22
SC weight (g)	2.50 ± 0.24	2.32 ± 0.16	2.41 ± 0.14
SC length (cm)	15.87 ± 0.24	15.40 ± 0.61	15.63 ± 0.32

$P \leq 0.05$

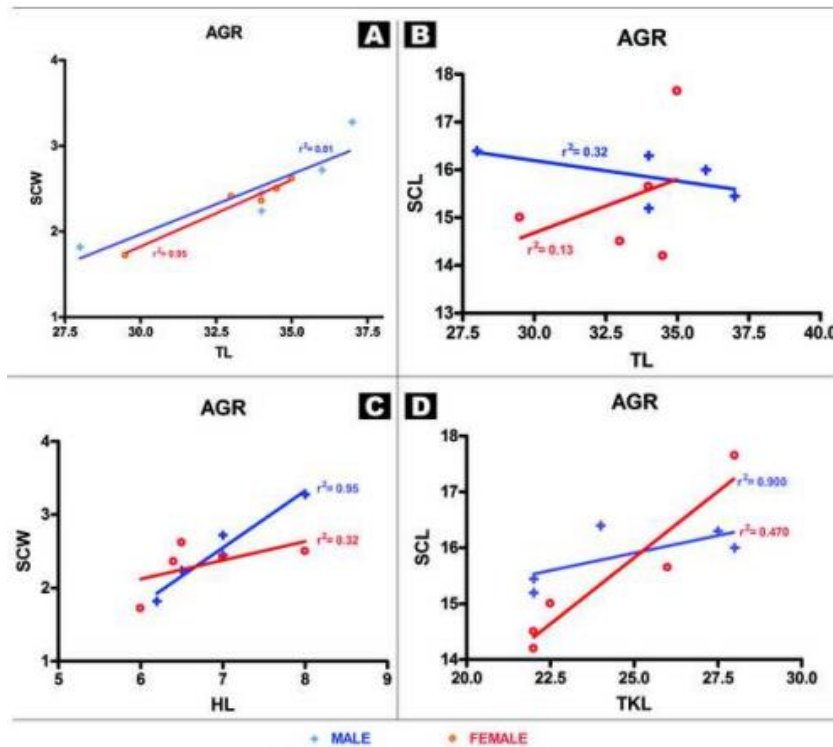


Figure 1: Graphical Representation of the Linear Regression of: (A): Spinal Cord Weight [SCW] versus Tail Length [TL]; (B): Spinal Cord Length [SCL] versus Tail Length [TL]; (C): Spinal Cord Weight [SCW] versus Head Length [HL] and (D): Spinal Cord Length [SCL] versus Trunk Length [TKL]

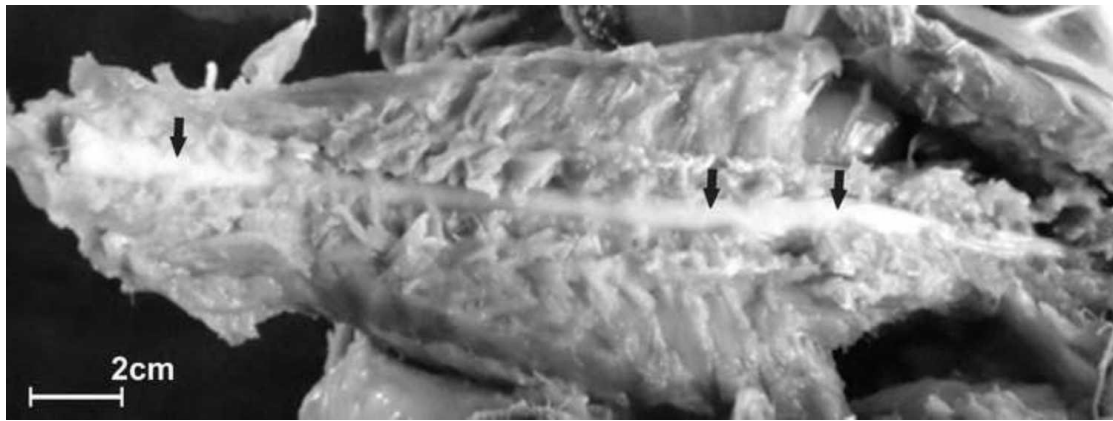


Figure 2: Dorsal view of the SC of the AGR (arrows) after laminectomy

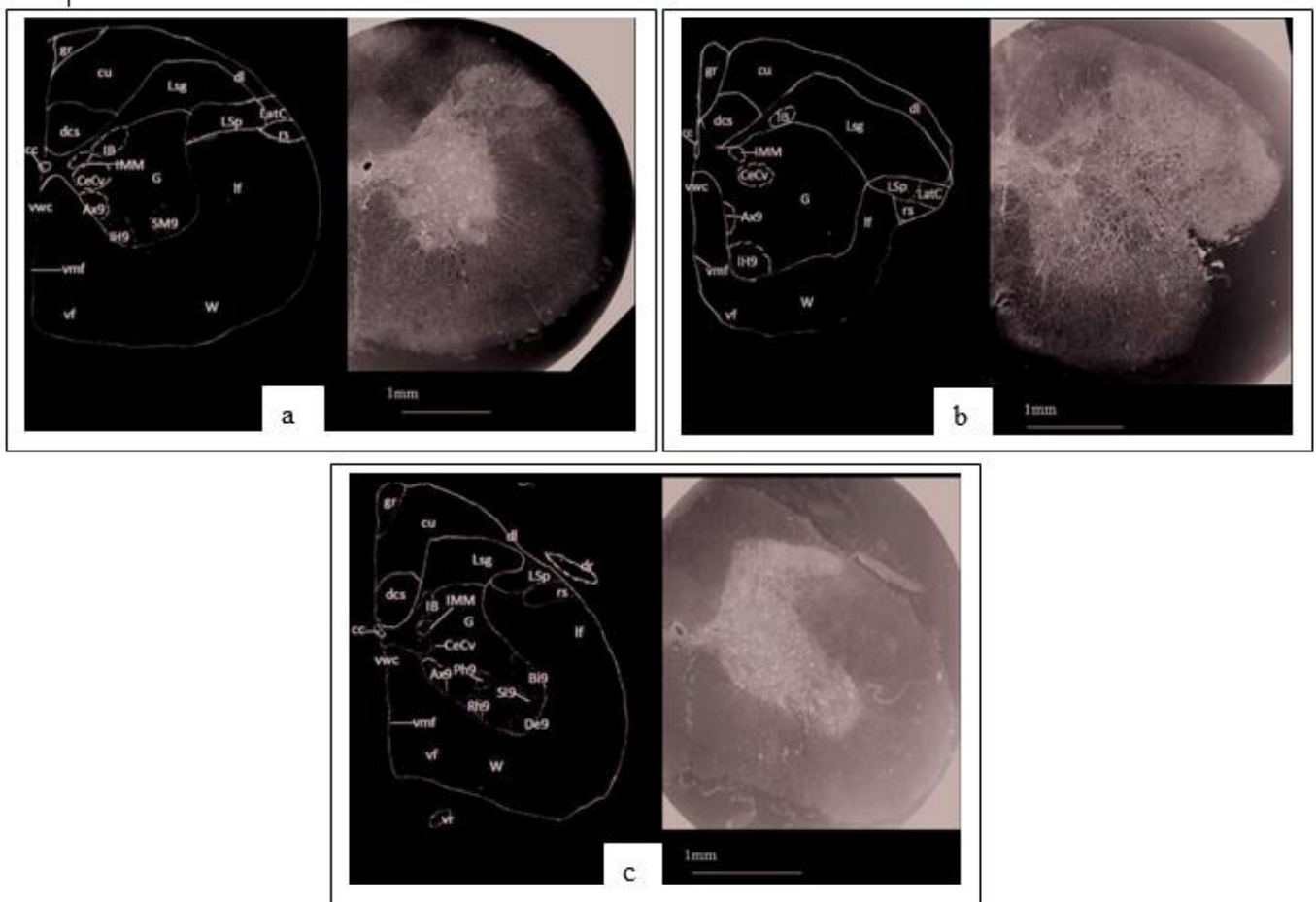


Figure 3. (a) The first cervical spinal segments highlighting the nuclei and tracts. *Left panel: Pencil tracing; Right panel: photomicrograph.* (b) The second cervical spinal segments highlighting the nuclei and tracts. *Left panel: Pencil tracing; Right panel: photomicrograph.* (c) The fifth cervical spinal segments highlighting the nuclei and tracts. *Left panel: Pencil tracing; Right panel: photomicrograph*

Gross morphology

The SC appeared as a whitish cylindrical, tube-like structure situated in the vertebral canal extending from the foramen magnum and continuing as the conus medullaris at vertebra L₄ (n=9) and L₅ (n=1) before terminating as the filum terminale in the coccygeal vertebrae (Figure 2). The SC was covered by the dura matter, which traversed its entire length while the spinal nerves emerged from the SC and exited the vertebral canal through the intervertebral foramina. The cervical enlargement, which contributed to the brachial plexus, spanned from C₄ to T₁ SC segments and was within the nominally corresponding vertebrae

(C₄ to T₁) in all animals. The lumbosacral enlargement, which contributed to the lumbosacral plexus that innervates the hind limb, extended from SC segments L₂ to S₃ and was found about the vertebral levels of T₉ to T₁₂.

Histomorphology

Transverse sections of the SC revealed the typical central canal, gray and white matter. The gray matter, stained deep purple with Thionin, had the typical “H” or “butterfly” shape with a reticular formation complex at the lateral area of the dorsal horn. The ventral horn of the gray matter was larger than the

dorsal horn being widest at the lumbosacral segments followed by the cervical segments and then the thoracic; the coccygeal segments being the narrowest.

Shape variations in the central canal were observed across the SC segments. The central canal of the first cervical segment appeared as a vertical slit (Figure 3a), the second to the sixth cervical segments were horizontally oval (Figures 3b-c) while the seventh and

the eighth cervical segment appeared circular in shape. In the thoracic segments, the first and second segments were vertical slits while the others were vertically oval in shape (Figures 4a-c). The first and second lumbar segments were circular in shape (Figure 5a) while the other lumbar segments, sacral and coccygeal segments were vertically oval in shape (Figures 5b, 6a-b, 7).

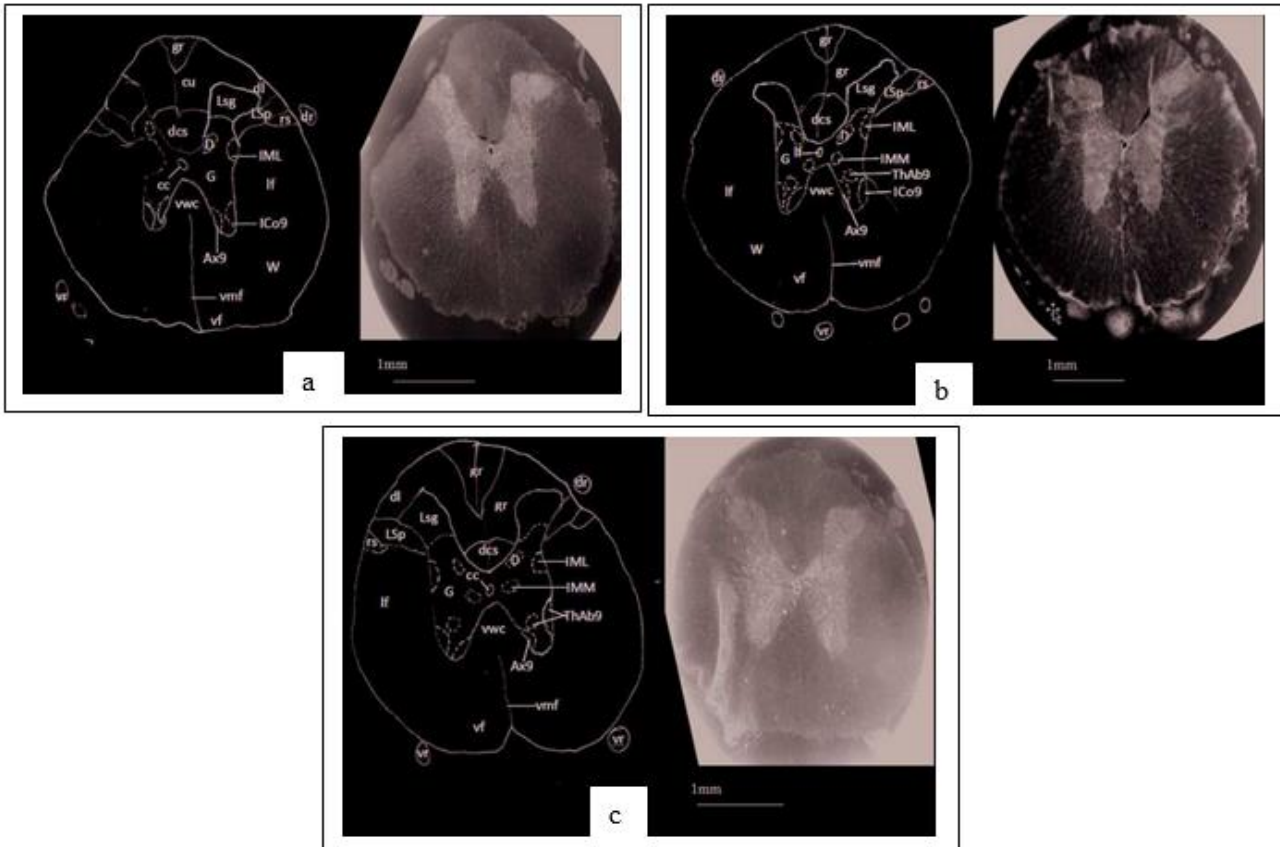


Figure 4. (a) The fourth thoracic spinal segments highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph. (b) The eighth thoracic spinal segments highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph. (c) The twelfth thoracic spinal segments highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph.

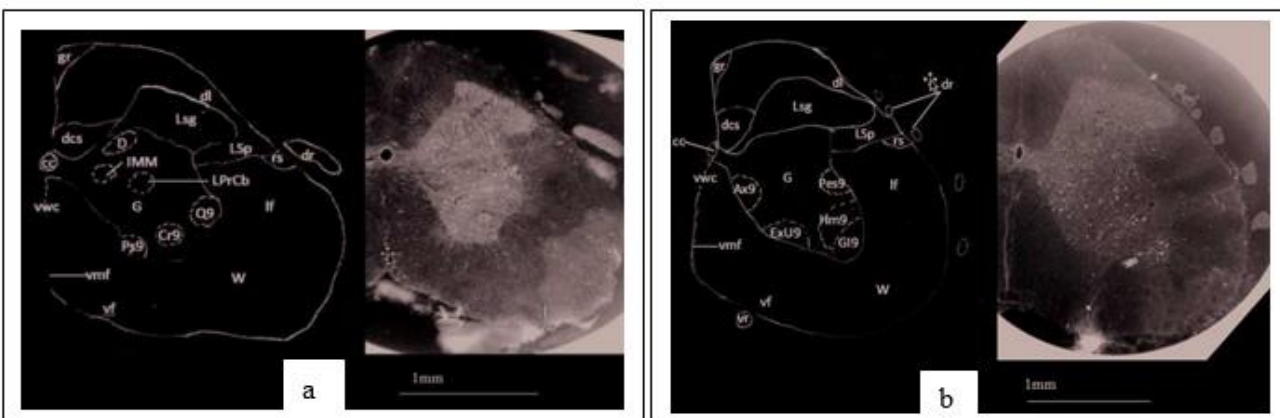


Figure 5. (a) The second lumbar spinal segment highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph. (b) The sixth lumbar spinal segment highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph.

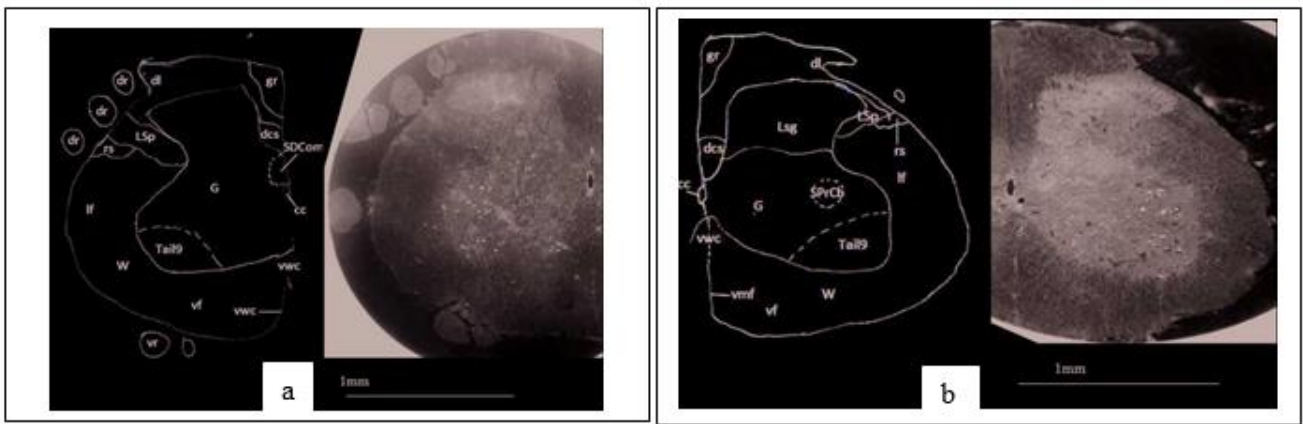


Figure 6. (a) The second sacral spinal segment highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph. (b) The fourth sacral spinal segment highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph.

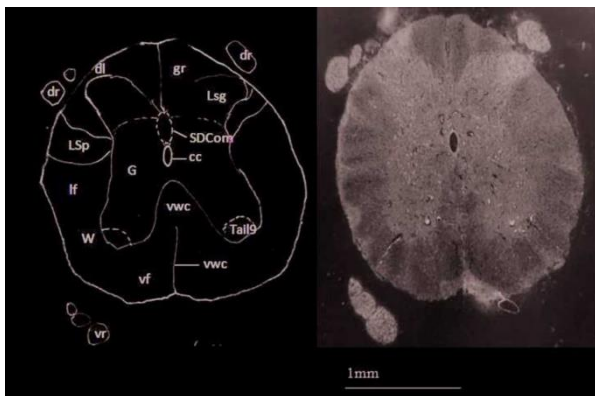


Figure 7. The second coccygeal spinal segment highlighting the nuclei and tracts. *Left panel:* Pencil tracing; *Right panel:* photomicrograph.

LPrCb	Lumbar paracerebellar nucleus
LSp	Lateral spinal nucleus
Pes9	Pesmotor neuron of lamina 9
Ph9	Phrenic motor neuron of lamina 9
Ps9	Psoas motor neuron of lamina 9
Q9	Quadriceps motor neuron of lamina 9
Rh9	Rhomboid muscle motor neuron of lamina 9
Rs	Rubrospinal tract
SDCom	Sacral dorsal commissural nucleus
SI9	Supraspinatus and infraspinatus motor neurons of lamina 9
SM9	Sternomastoid motor neurons of lamina 9
SPPrCb	Sacral precerebellar nucleus
Tail9	Tail muscle motor neuron of lamina 9
ThAb9	Thoracoabdominal wall muscle motor neuron of lamina 9
Vf	Ventral funiculus
Vmf	Ventral median fissure
Vr	Ventral root
Vwc	Ventral white commissure
W	White matter

Table 2. Abbreviations and definitions of structures of the SC traced

Abbreviation	Definition
Ax9	Axial muscle motor neuron of lamina 9
Bi9	Bicep motor neuron of lamina 9
Cc	Central canal
CeCv	Central cervical nucleus
Cr9	Cremaster motor neuron of lamina 9
Cu	Cuneate fasciculus
D	Dorsal nucleus (Clarke)
Dcs	Dorsal corticospinal tract
De9	Deltoid motoneuron of lamina 9
DI	Dorsolateral fasciculus
Dr	Dorsal root
ExU9	External urethral sphincter motor neuron of lamina 9
G	Gray matter
GL9	Gluteal motor neuron of lamina 9
Gr	Gracile fasciculus
Hm9	Hamstring motor neuron of lamina 9
IB	Internal basilar nucleus
ICo9	Intercostals muscle motor neuron of lamina 9
IH9	Infraspinatus muscle motor neuron of lamina 9
IML	Intermediolateral column
IMM	Intermediomedial column
Lsg	Laminae of spinal gray matter
LatC	Lateral cervical nucleus
Lf	Lateral funiculus

Nuclear tracing

Forty-three (43) anatomical structures including nuclear aggregations and tracts were traced (Table 2). It describes the abbreviations and definitions of structures of the SC traced out, which represents the list of structures in the histological pictures. Tracts of the dorsal, ventral and lateral funiculi were observed in the white matter. The dorsal funiculus consisted of the gracilis fasciculus, cuneate fasciculus and also a descending tract - the dorsal corticospinal tract. The gracilis fasciculus extended all through the SC segments, the cuneate fasciculus were indistinct at the caudal spinal segments (lumbar to the coccygeal segments) while the dorsal corticospinal tracts ran through the cervical to the sacral spinal segments and became indistinct at the coccygeal spinal segments (Figures 3 - 7). The lateral funiculus also consisted of a descending tract - rubrospinal tract (Figures 3b, 3c). Eight nuclei namely central cervical, dorsal (Clarke's), internal basilar, lateral cervical, lumbar paracerebellar,

lateral spinal, sacral dorsal commissural and sacral precerebellar nuclei were identified and traced (Figures 3a-b, 4a-c, 5a-b, 6a-b, 7).

The SC appeared as a whitish cylindrical, tube-like structure situated in the vertebral canal extending from the foramen magnum and continuing as the conus medullaris at vertebra L₄ (n=9) and L₅ (n=1) before terminating as the filum terminale in the coccygeal vertebrae (Figure 2). The SC was covered by the dura matter, which traversed its entire length while the spinal nerves emerged from the SC and exited the vertebral canal through the intervertebral foramina. The cervical enlargement, which contributed to the brachial plexus, spanned from C₄ to T₁ SC segments and was within the nominally corresponding vertebrae (C₄ to T₁) in all animals. The lumbosacral enlargement, which contributed to the lumbosacral plexus that innervates the hind limb, extended from SC segments L₂ to S₃ and was found about the vertebral levels of T₉ to T₁₂.

DISCUSSION

The basic features of the AGR SC were typical of rodents (Hebel and Stromberg, 1976; Bjugn *et al.*, 1989). The SC accounts for about 0.26% of body weight of the AGR. It is smaller than the rabbit which weighs 5-7g (about 0.5% body weight) (Farang *et al.*, 2012) but greater than that of the horse which weighed 250-300g (about 0.06% body weight) (Nickel *et al.*, 2004).

Bjugn *et al.* (1989) reported the SC length of mice as 4.4cm (55.7% body length) which is smaller than that of the AGR 15.63cm (64.2% body length). SC length has also been described in several animals amongst which are: 34.7 cm in Wistar rats (70.4% body length) (Hebel and Stromberg, 1976; Aguh *et al.*, 2013); 34.7 cm in rabbits (99.1% body length) (Farang *et al.*, 2012); 53.8 cm in goats (61.6% body length) (Kahvecioglu *et al.*, 1995); 167.2 cm in horses (68.6% body length) (Sadullah *et al.*, 2013); 106.8 cm in donkeys (53.4% body length) (Ocal and Haziroglu, 1988); 61.5 cm in brockets (64.7% body length) (Lima *et al.*, 2010).

The positive linear relationship ($p \leq 0.05$) between the tail length and SC weight of males and females established that the variability observed in the SC weight may be explained by the tail length in both sexes, indicating that the bulk of the filum terminale contributed significantly to the weight of the SC. This was further substantiated by a relatively higher tail: body length ratio of 1.07 in the AGR compared to 1.00 in mice, (Brian and William, 2000) 0.47 in fox squirrel, 0.87 in red squirrel, 0.50 in California ground squirrel (Virginia, 2008) and 0.43 in greater cane rats (Fitzinger, 1995).

The anatomical location of the cervical and lumbosacral enlargements - between C₄ and T₁ and between L₂ and S₃ respectively - were typical with laboratory rats (Bjugn *et al.*, 1989). These anatomical

positions seem characteristic of rodents as several authors have reported slightly different positions in most domestic animals. The cervical enlargement is found between C₇ and C₈ in pigs (Dellmann and McClure, 1975), C₅ and T₁ in the rabbits (Farang *et al.*, 2012), C₆ and T₁ in the dogs (Miller *et al.*, 1964), C₆ and T₂ in buffalo and camels (Abu-zaid, 1982; Mansour, 1983) and C₅ - T₂ in Indian sheep and donkeys (Mansour, 1980; Rao, 1990). While the lumbosacral enlargement lies between L₂ and S₃ spinal segments in AGR, the following positions have been documented in domestic animals: between L₂ and S₁ in the donkeys (Mansour, 1980), L₆ and S₁ in camels (Mansour, 1983), L₄ and S₃ in rabbits (Farang *et al.*, 2012), the last three lumbar and first two sacral in buffalo (Abu-zaid, 1982), L₄ and S₁ in sheep (Rao, 1990), L₆ and L₇ in the pigs and L₄ and S₁ in dogs (Dellmann and McClure, 1975).

The enlargements at the cervical and lumbosacral segments provide innervations to the fore and hind limbs respectively; contributing to the brachial and lumbosacral plexuses (Bjugn *et al.*, 1989; Rahmanifar *et al.*, 2008). Worthy of note is that the extents of the enlargements, which began earlier in spinal segments, are more extensive in rodents and may characteristically add to limb efficiency than other mammals. Thus; regional anaesthesia for surgical maneuvers in the AGR can be readily achieved based on the knowledge of the extent and anatomical locations of these enlargements (Jonathan and Gerbrand, 2005).

The ventral horn of gray matter coordinates the motor neuron; this explains its relative bigger size compared to the dorsal horn (Gruener and Biller, 2008). The ventral horn appears wider at the cervical region and the lumbosacral region than other segments. This corresponds to the cervical and lumbosacral enlargements (Gruener and Biller, 2008). The AGR uses its tail to dig, defend itself and has been reported to stand on it (personal observation). It is also known to burrow more with the forelimbs and shows high locomotor dexterity (Ajayi, 1977). This probably accounts for the ventral horns of cervical and lumbosacral regions being more developed than other segments.

The dorsal (Clarke's) and central cervical nuclei are particularly essential for the coordination of movement and balance (Gruener and Biller, 2008). These nuclei appeared well developed in the AGR and might explain the dexterous limb movements and balance shown by the AGR. The AGR also has been documented as a fast running, burrowing and shovelling rodent (Olude *et al.*, 2010). The lateral cervical, lateral spinal, sacral dorsal commissural nuclei are responsible for nociception (Rea, 2009) and are therefore important for their defence. The internal basilar nuclei are responsible for voluntary motor control and procedural learning relating to routine behaviours and habits (Weyhenmeyer and Gallman,

2007). The AGR has been shown to adapt well to training to detect landmines and diagnose Tuberculosis (Weetjens, 2010). The sacral pre-cerebellar nucleus also relays unconscious proprioception motor (lower extremities and trunk) feedback to the cerebellum.

Shape variation of central canal and tracts observed in the white matter. The central canal of each segment and the tracts observed in the white matter of the SC of the AGR were similar to that of rats (Watson *et al.*, 2008).

Conclusion

This study documents baseline data on the morphometric and morphologic features of the SC of AGR, thus contributing to the knowledge of anatomy of the AGR and providing useful information on its regional anaesthesia. It also strengthens the drive in adopting the AGR as a convenient indigenous research model and could assist further researches especially in the study of SC diseases/injuries within the African context.

Acknowledgement

Special thanks to Mr. Anise for helping with the preparation of the histological slides.

REFERENCES

- Abu-zaid, S. (1982). Some gross anatomical studies on the prenatal and postnatal morphological features on the spinal cord of the water buffalo (*Bos Bubalis L.*), Ph.D Thesis, (Anatomy), Faculty of Veterinary Medicine, Cairo University.
- Aguh, B. Yahaya, A. Saidu, L. Ayeku, P. Agba A. (2013). Correlation of body weight and other morphometric measurements in Albino rats (*Rattus norvegicus*). Sci J of Biol. Sci. 2(3): 39-44.
- Ajayi, S. (1977). Field observation on the African Giant Rat (*Cricetomys gambianus*) in southern Nigeria. East Afr. Wild J. 15(3): 191-198.
- Akinloye, A. (2009). Structural and hormonal studies of the female giant rats (*Cricetomys gambianus*, Waterhouse) at different stages of oestrous cycle, PhD Thesis, Faculty of Veterinary Medicine, University of Ibadan, Nigeria.
- Bjugn, R., Boe, R., Haugland, H. (1989). A stereological study of the ependyma of the mouse spinal cord with a comparative note on the choroid plexus ependyma. J Anat. 166: 171-178.
- Brian, K. and William R. (2000). Body Weight and Tail Length Divergence in Mice Selected for Rate of Development. J Exp Zool (Mol Dev Evol). 288: 151-164.
- Dellmann, H. and McCLURE, R. (1975). Central nervous system of domestic animals in "Sisson-Grossman" The Anatomy of Domestic Animals. 5th ed. Vol. 2 reviewed by R. Getty. WB Saunders-Philadelphia, London, Toronto.
- Durnez L., Eddyani M., Mgode G., Katakweba A., Katholi C. Machang'u R., Kazwala R., Portaels F. and Leirs H. (2008). First detection of Mycobacteria in African rodents and insectivores, using stratified pool screening. Appl Environ Microbiol. 74: 768-773.
- Farag F., Elayat M., Wally Y., Elkarmoty A. (2012). Morphometric Studies on the Spinal cord Segments of the Domestic Rabbit (*Oryctolagus cuniculus*). J Vet Anat. 5(2): 33-47.
- Fitzinger J. (1995). Cane Rats. In R Nowak, ed. Walker's Mammals of the World Johns Hopkins Med J., 1650-1651.
- Gruener G., Biller J. (2008). Spinal cord anatomy, Localization, and Overview of Spinal cord syndromes. Am. Acad. Neurol. 14(3): 11-35.
- Hebel R., Stromberg M. (1976). The Nervous System. In: Anatomy of the laboratory rat. The Wilkins and Wilkins company, Baltimore, Md. U.S.A. Library of congress cataloging in publication data. 119-121
- Ibe C., Onyeanusi B., Hambolu J. (2014). Functional morphology of the brain of the African giant pouched rat (*Cricetomys gambianus* Waterhouse, 1840), Onderstepoort J Vet Res. 81(1): 644 -650.
- Ibe C., Onyeanusi B., Hambolu J., Ayo J. (2010). Sexual dimorphism in the whole brain and brain stem morphometry in the AGR (*Cricetomys gambianus*, Waterhouse, 1840). Folia morphol. 69(2): 69-74.
- Jonathan, R. and Gerbrand J. (2005). Applied Epidural Anatomy. Contin. Educ. Anaesth. Crit. Care Pain, 5(3): 98-100.
- Kahvecioglu K., Ozcan S., Cakir M. (1995). Anatomic studies on the medulla spinalis of the Angora goat (Excluding the coccygeal segments) Yuzuncu Yil Universitesi Veteriner Fakultesi Dergisi. 6(1-2), 76-80.
- Lima F., Santos A., Lima B., Vieira L., Hirano L. (2010). Topographic anatomy of the spinal cord and vertebromedullary relationships in *Mazamagou azoubira* Fisher, 1814 (Artiodactyla; Cervidae) Acta Scientiarum Bio Sci. 32(2), 189-194.
- Mansour A. (1980). Some morphological features of the Spinal cord in Donkey. MVSc Thesis (Anatomy). Faculty of Veterinary Medicine, Assuit University.
- Mansour A. (1983). Some anatomical features of the system anervosum centrale of *Camelus dromedarius*. Ph.D. Thesis (Anatomy). Faculty of Veterinary Medicine, Assuit University.
- Miller M., Christensen G., Evans H. (1964). Anatomy of the Dog, 1st. Ed. WB Saunders Comp. Philadelphia, London, Toronto.
- Nickel R., Schummer A., Seiferle E. (2004). Lehrbuch der Anatomie der Haustiere: Nerven system, Sinnesorgane, endocrine Drusen. Stuttgart, Germany: Parey.
- Ocal M., Hazirolu R. (1988). Comparative morphological studies on the spinal cord of the donkey: I. The cross sectional areas of the Spinal

- cord segments. Ankara Universitesi Veteriner Fakultesi Dergisi. 35(1): 55-68.
- Oke O., Oke B. (1999). Vaginal cytological changes during the oestrous cycle of the adult female African giant rats (*Cricetomys gambianus*, Waterhouse). Trop. Vet. 17: 169-180.
- Olude M., Olopade J., Adebayo A., Mustapha O. (2010). Macro-anatomical investigations of the skeletons of the African giant rat (*Cricetomys gambianus* Waterhouse 1840) II: Fore limb. Eur J Anat. 14(1): 19-23.
- Olude M., Olopade J., Fatola I., Onwuka S. (2009). Some aspects of the neurocraniometry of the African giant rat (*Cricetomys gambianus*, Waterhouse). Folia Morphol. 68: 224-227.
- Peterson T., Papes M., Reynolds M., Perry N., Hanson B., Regnery R., Hutson C., Muizniek B., Damon I., Carroll D. (2006). Native range ecology and invasive potential of *Cricetomys* in North America. J Mammal. 8: 427-432.
- Rao G. (1990). Anatomical studies on the ovine spinal cord. Ann Anat. 171: 261-264.
- Rahmanifar F., Mansouri S., Ghazi S. (2008). Histomorphometric study of the Spinal cord segments in the chicks and adult male ostrich (*Struthio camelus*). Iran J Vet Res. 4(9): 1-7.
- Rea P. (2009). The role of the lateral spinal nucleus in nociception. A thesis presented for the degree of Doctor of Philosophy in Neuroscience and Molecular Pharmacology, Faculty of Biomedical and Life Sciences, University of Glasgow.
- Sadullah B., Durmus B., Muhammet L.S. (2013). The segmental morphometric properties of the horse cervical spinal cord: a study of cadaver. Sci World J. <http://dx.doi.org/10.1155/2013/734923>.
- Sheets B. (1989). Cranial anatomy of *Jaculus orientalis* (rodentia, dipoidea): new evidence for close relationship of diploid and muroid rodents. Submitted to the committee on undergraduate honors of Baruch College of the City University of New York in partial fulfillment of the requirements for the degree of Bachelor of art in biology with honors.
- Vera P., Meyer-siegler, K. (2003). Anatomical location of macrophage migration inhibitory factor in urogenital tissues, peripheral ganglia and lumbosacral spinal cord of the rat. BMC Neurosci. 4: 17-26.
- Verhagen, R., Cox C., Machang'u R., Weetjens B., Billet M. (2003). Preliminary results on the use of *Cricetomys* rats as indicators of buried explosives in field conditions. In: Mine detection dogs: training operations and odour detection. Geneva: Geneva International Centre for Humanitarian Demining: 175-193.
- Virginia H. (2008). Patterns of body and tail length and body mass in sciuridae. J mammal. 89(4): 852-873
- Watson C., Paxinos G., Kayalioglu G. (2008). The Spinal cord: A Christopher and Dana Reeve foundation text and atlas. Elsevier Ltd.
- Weetjens, B. (2010). How I taught rats to sniff out land mines, TED Talks, Rotterdam, 2010, (accessed 2011-09-16).
- Weetjens B., Mgode G., Machang'u R., Kazwala R., Mfinanga G., Willa F., Cox C., Jubitana M., Kanyagha H., Mtandu R., Kahwa A., Mwessongo J., Makingi G., Mfaume S., Steenberge J., Beyene N., Billet M., Verhagen R. (2009). African pouched rats for the detection of pulmonary tuberculosis. Int J Tuber Lung Dis. 13: 737-43.
- Weyhenmeyer J., Gallman E. (2007). Rapid review of neuroscience. Mosby Elsevier: 120