

Bioactive components in ethanol extract of *Citrullus lanatus* rind using Gas Chromatography- Mass Spectroscopy.

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ABSTRACT

The knowledge of bioactive compounds present in a plant can be helpful in utilizing it for a medicinal purpose in life. The Gas Chromatography-Mass Spectrometry (GCMS) analysis of ethanol extract of Citrullus lanatus rind (CLR) was done to identify bioactive compounds and selectivity or abundance percentage in ethanol extract of CLR. The extract was obtained by fractionating crude ethanol CLR extract in order of increasing polarity, n-Hexane, chloroform, ethylacetate and ethanol. The GC-MS analysis was carried out on GC system comprising Gas Chromatograph interfaced to Mass Spectrometer (GC-MS) instrument. The bioactive compounds obtained were compared with database of spectrum of known components stored in the gas chromatography-mass spectrometry library. The GC-MS analysis of ethanol extract of CLR revealed the presence of androst-4-en-9-thiocyanomethyl-11-ol-3,17-dione a steroid which is a precursor for androgen and estrogen production. As well as cyclooctasiloxane hexadecamethyl and octasiloxane hexadecamethyl in which both of them contain silicon an essential trace element for collagen and bone formation. The presence of these compounds validates the use of ethanol extract of CLR as a good antioxidants, antibacterial agents and stimulant in precursor of male reproductive hormone production. The ethanol extract of CLR can be of great value in ethno-veterinary practice.

Keywords: Bioactive, Ethanol, *Citrullus lanatus* rind, GC-MS

INTRODUCTION

Plants are now occupying significant positions in herbal medicine, homoeopathy, allopathic medicine and aromatherapy¹. Many of these indigenous medicinal plants

are used as spices and food plants they are also sometimes added to foods meant for curing various diseases for medicinal purposes due to the presence of various

bioactive compounds found in their phytochemical composition². Many plants are cheaper and more accessible to most people especially in developing countries than orthodox medicine, and there is lower incidence of adverse effects after use². The awareness of medicinal plants usage is a result of the many years of struggles against illnesses due to which man learned to pursue drugs in barks, seeds, fruit bodies, leaves and other parts of the plants³. The use of medicinal plants had been established to be the most useful in the treatment of diseases and had provided important source of materials to pharmaceuticals industries worldwide through the knowledge of their bioactive components⁴. The phytochemicals found in medicinal plants example of which are steroids, terpenoids, carotenoids, flavanoids, alkaloids, tannins and glycosides are primarily make up of bioactive compounds⁵ These compounds vary widely in chemical structures, functions and are grouped accordingly. Some examples of these bioactive compounds are carotenoids, flavonoids, carnitine, choline, coenzyme Q, carbohydrates, esters, dithiolthiones, phytosterols, phytoestrogens, glucosinolates, polyphenols, taurine also including vitamins and minerals while they stimulate their pharmacological effects.

Most of the bioactive compounds have antioxidant, anti-inflammatory, antimicrobial and anti-carcinogenic properties⁶. Thus, they can exhibit their hepatoprotective, protective effects on cardiovascular diseases and treatment of different diseases. The use of spectrophotometric analysis had been explored for the quantitative analysis of phytochemicals found in plants⁷ while Gas Chromatography-Mass Spectrometer (GC-MS) had been used to confirm the bioactive compounds found in plants⁸. Also, Fourier Transform Infrared (FTIR) spectroscopy analysis was used to analyze the bioactive components of Watermelon rind aqueous extract⁴.

Watermelon, also known as *Citrullus lanatus* is a warm season crop from the cucurbit family⁹. Watermelon biomass can be categorized as three main components which are the flesh, seed, and rind. Watermelon constitutes approximately 68% flesh, the rind 30%, and the seeds 2% of the total weight¹⁰. *Citrullus lanatus* has nutritive values which are beneficial to human health. The plant has pharmacological activities and therapeutic potentials such as antibacterial, antifungal, antimicrobial, antiulcer, antioxidant, anti-inflammatory, gastro-protective, analgesic, laxative, hepato-protective, against prosthetic hyperplasia and atherosclerosis¹¹. The

Citrullus lanatus rind seems to be receiving much attention recently, maybe due to efforts to reduce the environmental toxicants that it may cause since it is regularly discarded when the *Citrullus lanatus* fruit is consumed. It was noted that Kumar *et al.*,⁸ considered the use of GC-MS to study the sugars found in aqueous extract of *Citrullus lanatus* rind. While Bichi *et al.*,⁴ explored the use of GC-MS and FITR to study the bioactive components found in aqueous extract of CLR, but this is in exhaustive because the aqueous extract of CLR cannot be stored or preserved for a long period of time. With the great awareness on the phytochemicals and pharmaceutical importance of CLR recently this will create needs for the CLR stored or preserved for a long period of time by preparing the ethanol extract of CLR. All through extraction, the choice of solvent is preferable to select two solvents with low polarity (n-hexane, chloroform) containing bioactive compounds in highly organic medium, two with medium polarity (dichloromethane, ethanol) containing bioactive compounds in both organic and inorganic media, and one with the highest polarity (water).¹² Meanwhile ethanol had been shown to be used as a preservative because it is effective in knocking out

organisms that could pose danger to consumers¹³.

This study now considered the use of GC-MS to analyze the bioactive components of ethanol extract of CLR and compared with the study on the aqueous extract of CLR that was done by Bichi *et al.*,⁴. This will provide more information on active pharmaceutical ingredients (API) present in CLR when it is stored or preserved in ethanol solvent for long time before consumption by animal or use in livestock since ethanol is a good preservative solvent.

MATERIALS AND METHODS

Plant Specimen

The *Citrullus lanatus* fruit was collected in Abeokuta, Ogun state Nigeria. The rind was carefully removed from the fruit using a sharp knife and cut into small pieces. It was identified and authenticated in Botany department University of Ibadan Oyo state and voucher number UIH 22872 was assigned. The *Citrullus lanatus* rinds were chopped into small pieces and air-dried under room temperature. This was done until it attains a constant dry weight. The dried rind was grinded to powdery form using a grinder (Euro premium blender^R Ultima made in China). The powdered CLR was soaked/macerated in 95% ethanol for 72

hours. The mixture was filtered twice using muslin bag. The obtained filtrate was concentrated using a rotary evaporator (Heidolph laboratory efficient model 517-01002-002 Germany) at 40°C¹⁴.

Fractionation process of crude ethanol extracts of CLR.

The crude ethanol CLR extract was fractionated using the following solvents (n-hexane, chloroform, ethylacetate, ethanol and water) according to their polarity index¹². The fractionation was initiated by pre-absorption process which was done by mixing silica gel to the crude ethanol CLR extract and air-dried overnight. The silica gel was poured into Vacuum liquid chromatograph (VLC) chamber or distillation tube under pressure using pump to closely packed the silica gel very well. A whatman filter paper was placed in between while n-hexane with lowest polarity index was added. This brought out a colored filtrate initially but later became colorless as the volume of the n-hexane increases. It was later stopped after clear n-hexane filtrate which is colorless was obtained. This gave the n-hexane filtrate labelled CLRH, the procedure continued with chloroform based on polarity index¹². This procedure continued with chloroform, ethyl acetate followed by ethanol to obtain ethanol fraction of CLR. These procedures

were carried out at room temperature. Each of the filtrates was concentrated with rotary vacuum evaporator at 40°C¹³. The resulting extracts were weighed and labelled as CLRH, CLRC, CLREA and CLREF for n-hexane, chloroform, ethyl acetate and ethanol fractions respectively.

Identification of bioactive components in ethanol extract of CLR using GC-MS.

The ethanol extract of CLR was sent to Department of Biochemistry and Biotechnology, Kwame Nkrumah University of Science and Technology Kumasi Ghana for gas chromatography-mass spectrometer (GC-MS).

Gas Chromatography Mass Spectrometer Procedure.

The initial temperature was 80°C for 0 minutes, at ramp 10°C/min to 250°C the hold period was minutes and the ramp rate of 5°C/min to 280°C. Later the hold time was 15 minutes injected at 250°C the volume was 0µl split at 10-1 the carrier gas was helium while the solvent delay was 4min and transfer temperature was 280°C the source temperature was 220°C scanned at 50 to 450Da and the column was maintained at 30.0m by 250µm. The spectra of different compounds found in ethanol CLR extract were obtained through GC-MS analysis. The spectra were nomenclated according to their

respective peak spectrum with mass, M; relative mass RM, while selectivity % was calculated as $M/RM \times 100 \%$. The selectivity % indicates the quantity, availability, or amount of a specific

compound found in the ethanol CLR extract (CLRCE). But the classification, molecular formula, molecular weight, and biological uses of each compound were sourced in Pubchem¹⁵.

RESULTS AND DISCUSSION

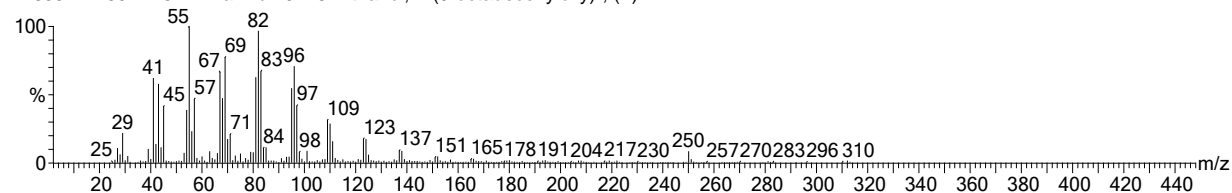
GC-MS analysis on ethanol extract of CLR revealed the following bioactive compounds

with the following different mass spectra showing different peaks.

Number 1

M:635 RM:691 P:5.12 mainlib 19248: Ethanol, 2-(9-octadecenoxy)-, (Z)-

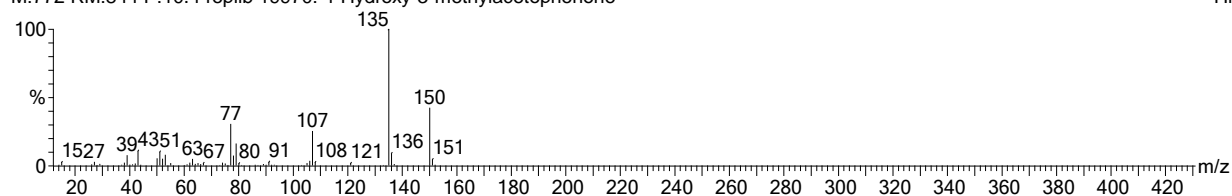
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Number 2

M:772 RM:844 P:10.4 replib 19970: 4-Hydroxy-3-methylacetophenone

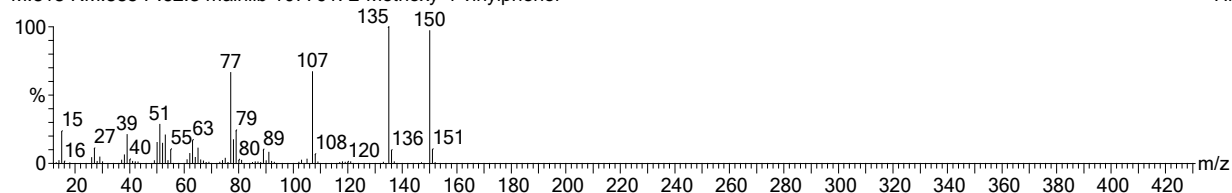
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Number 3

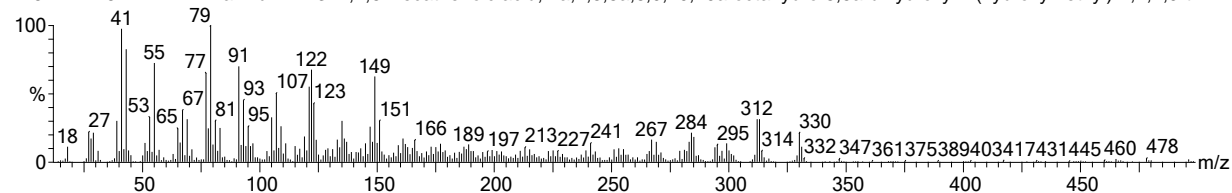
M:818 RM:885 P:62.3 mainlib 107791: 2-Methoxy-4-vinylphenol

Hit: 1



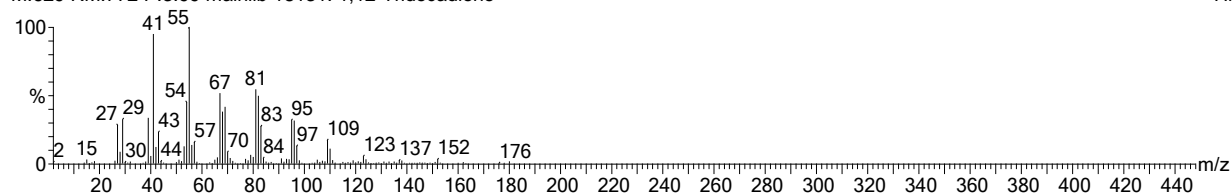
Number 4

M:511 RM:511 P:24.7 mainlib 44123: 2,4,6-Decatrienoic acid, 1a,2,5,5a,6,9,10,10a-octahydro-5,5a-dihydroxy-4-(hydroxymethyl)-1,1,7,9-t... Hit: 1

**Number 5**

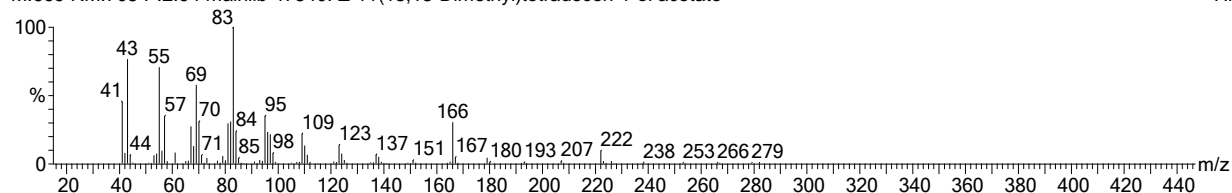
M:629 RM:772 P:3.99 mainlib 18131: 1,12-Tridecadiene

Hit: 3

**Number 6**

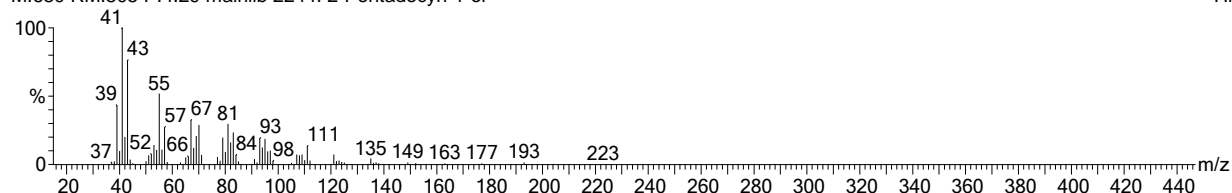
M:669 RM:705 P:2.94 mainlib 47340: Z-11(13,13-Dimethyl)tetradecen-1-ol acetate

Hit: 3

**Number 7**

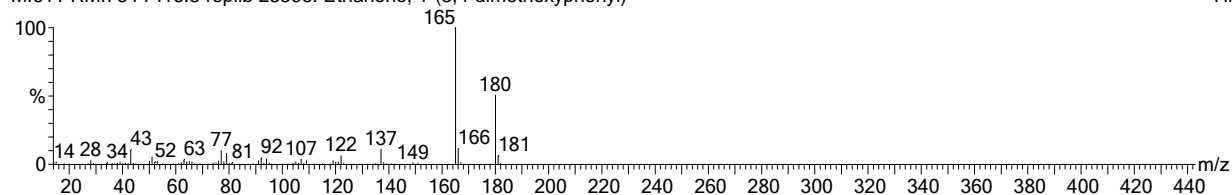
M:680 RM:805 P:4.29 mainlib 2244: 2-Pentadecyn-1-ol

Hit: 1

**Number 8**

M:611 RM:764 P:16.5 replib 23596: Ethanone, 1-(3,4-dimethoxyphenyl)-

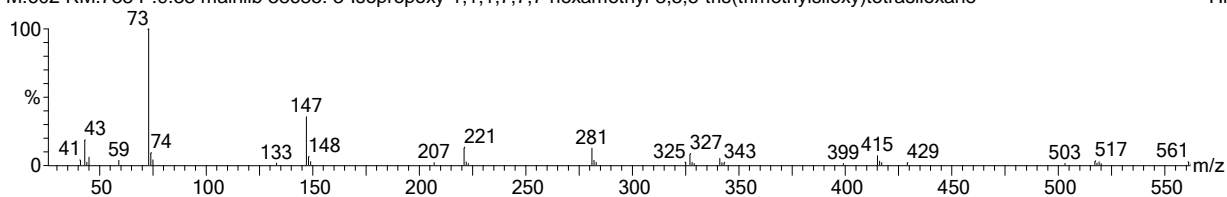
Hit: 1



Number 9

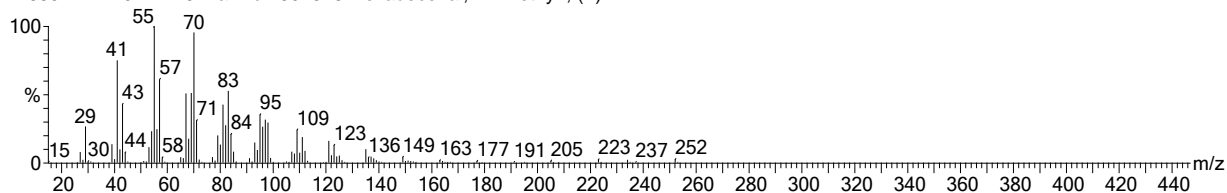
M:602 RM:738 P:9.33 mainlib 38685: 3-Isopropoxy-1,1,1,7,7,7-hexamethyl-3,5,5-tris(trimethylsiloxy)tetrasiloxane

Hit: 3

**Number 10**

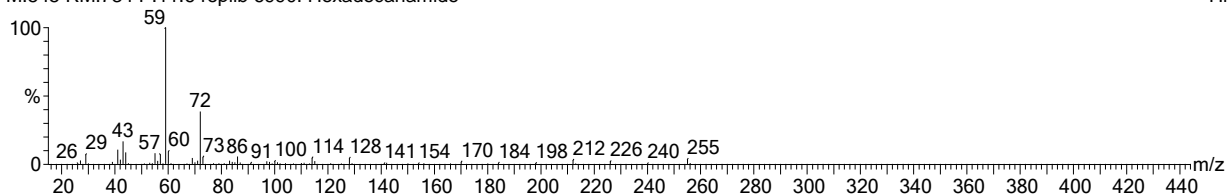
M:680 RM:740 P:4.29 mainlib 19015: 8-Hexadecenal, 14-methyl-, (Z)-

Hit: 2

**Number 11**

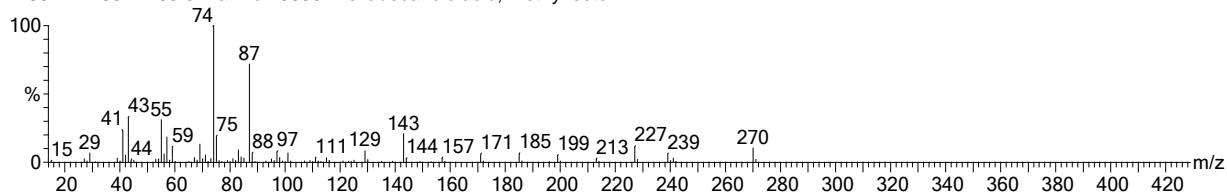
M:545 RM:734 P:11.6 replib 6990: Hexadecanamide

Hit: 3

**Number 12**

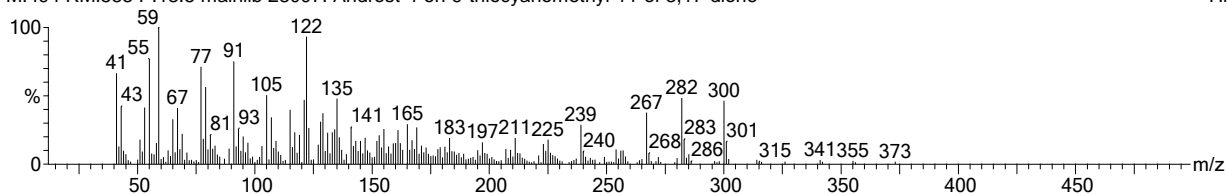
M:807 RM:852 P:63.9 mainlib 40690: Hexadecanoic acid, methyl ester

Hit: 1

**Number 13**

M:494 RM:555 P:13.5 mainlib 28007: Androst-4-en-9-thiocyanomethyl-11-ol-3,17-dione

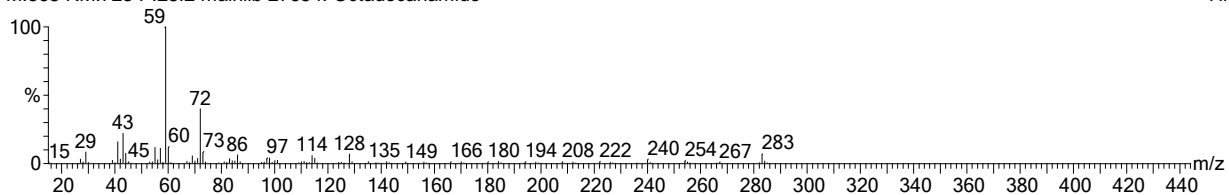
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Number 14

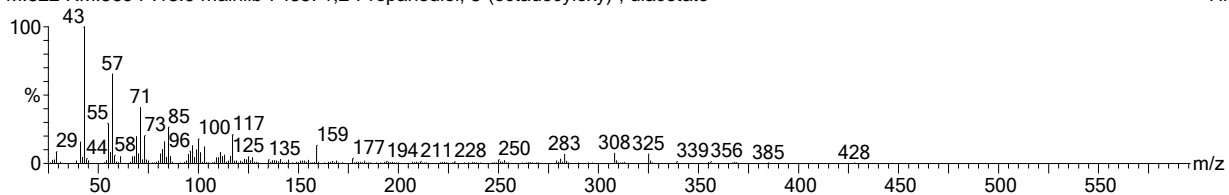
M:568 RM:723 P:28.2 mainlib 27654: Octadecanamide

Hit: 1

**Number 15**

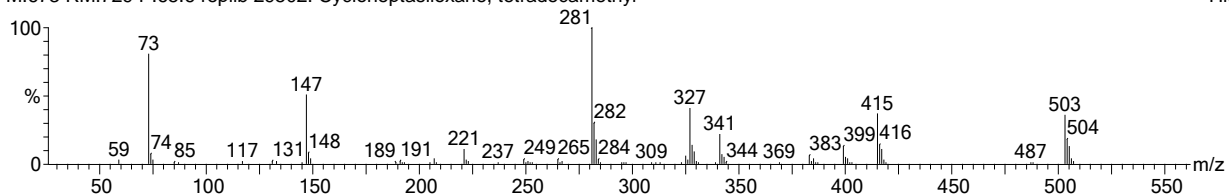
M:522 RM:569 P:13.9 mainlib 7435: 1,2-Propanediol, 3-(octadecyloxy)-, diacetate

Hit: 1

**Number 16**

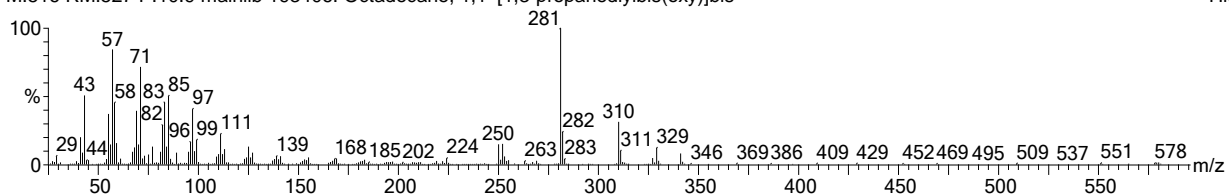
M:673 RM:729 P:65.6 replib 29502: Cycloheptasiloxane, tetradecamethyl-

Hit: 1

**Number 17**

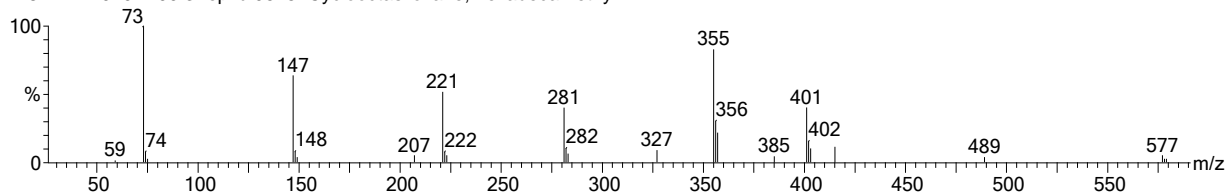
M:516 RM:527 P:10.9 mainlib 193406: Octadecane, 1,1'-[1,3-propanediylbis(oxy)]bis-

Hit: 3

**Number 18**

M:811 RM:919 P:90.3 replib 9645: Cyclooctasiloxane, hexadecamethyl-

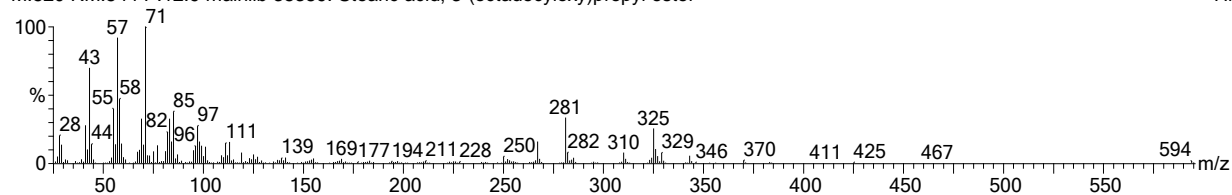
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Number 19

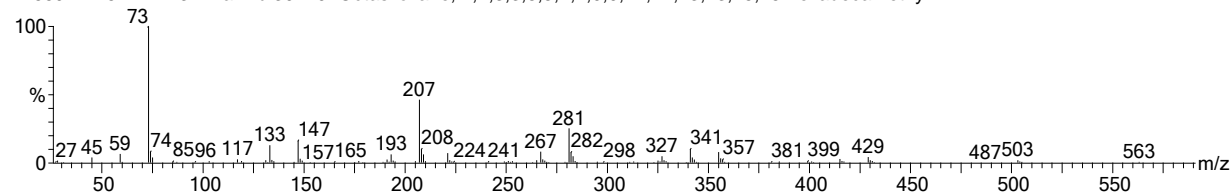
M:520 RM:544 P:12.9 mainlib 35389: Stearic acid, 3-(octadecyloxy)propyl ester

Hit: 2

**Number 20**

M:669 RM:674 P:2.31 mainlib 39429: Octasiloxane, 1,1,3,3,5,5,7,7,9,9,11,11,13,13,15,15-hexadecamethyl-

Hit: 3



The serial number in the table below follows same trend as seen in the numbering of spectra shown above.

Table 1: Different bioactive compounds found in ethanol *Citrullus lanatus* rind extract using Gas Chromatography-Mass Spectrometry analysis showing their chemical classifications and some of their uses.

S/	M	R	Selec	Compound name	Structur	Molec	Classifica	Natural Uses
N		M	tivity		e	ular	tion	
O			%			weight		
1	6	6	91.44	Ethanol,-2-(9,12-	$C_{20}H_{42}O_2$	314	Alcohol	Useful in medicine as cough syrups, tonic as a good solvent, color additives, enhanced flavour, cosmetics and beauty lotions.
	3	9	92.8	Octadecadienyloxy)-				
	1	0						
2	7	8	91.82	4-Hdroxy-2-	$C_9H_{10}O_2$	150.2	Ketone	Antimicrobial

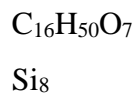
	8	5	243	methyacetophenone.				and	
	6	6						antioxidant	
								effects	
3	8	8	92.42	2-Methoxy-4-vinylphenol	$C_9H_{10}O_2$	150.2	Carbohydr	Calorific	or
	1	8	938				ate	Energy	
	8	5							
4				Decatrienoic			Carboxyli	Antioxidants	
				acid,1a,2,5,5a,6,9,10,10a			c acids		
	5	5		Octahydro-5,5a-	$C_{10}H_{14}O_2$				
	1	1		dihydroxy-4-					
	1	1	100	(hydroxymethyl 1,1,7,9.		166.2			
5	6	7	81.47	1,12-Tridecadiene	$CH_3(CH_2$	184.4	Alkene	Useful	in
	2	7	668		$)_{11}CH_3$			research	
	9	2						laboratory	as
								distillation	
								chaser, it can	
								cause	skin
								irritation.	
6	6	7		11(13,13-			Ketone		
	6	0	94.89	Dimethyltetradecen-1-ol	$C_{12}H_{24}O_2$				
	9	5	362	acetate		200.3			
7	6	8	84.47	2-Pentadecyn-1-ol	$C_{15}H_{32}O$	228.4	Ketone	Antibacterial	
	8	0	205					effects against	
	0	5						E.Coli/Antidia	
								rrheal against	
								Clostridium	
								butyricum.	
8	6	7	79.97	Ethanone,-1-(3,4-	$C_{16}H_{26}O$	234.4	Ketone	Odor	
	1	6	382	dimethoxyphenyl				agent/Unmask	
	1	4						unpleasant	

									odor
9	6	7	81.57	3-Isopropoxy-1,1,1,7,7,7-	C ₁₈ H ₅₂ O ₇	577.2	Organosili	Collagen	
	0	3	182	hexamethyl-3,5,5-	Si ₇		con	formation.	
	2	8		tris(trimethylsiloxy(tetrasi				Pubcem ¹⁵ .	
				loxane					
10	6	7	91.89	8-Hexadecanal, 4-methyl	C ₁₆ H ₃₂ O	240.4	Aldehyde	Flavouring	
	8	4	189					agent of Food	
	0	0						additives.	
								Action on	
								cellular	
								location at	
								endoplasmic	
								reticulum,extra	
								cellular and	
								membrane	
11	5	7	74.25	Hexadecanamide	C ₁₆ H ₃₃ N	255.4	Fatty	Found in aging	
	4	3	068		O		amide	mouse brain.	
	5	4						Also use in	
								skin	
								conditioning ¹⁵	
12	7	8	87.88	Hexadecanoic	acid	C ₁₆ H ₃₂ O ₂	256.4	Palmitic	
	4	5	235	,methyl ester.				acid/Orga	
	7	0						nic acid	
								Therapeutic	
								effect on	
								multiple	
								meloma cells,	
								no effects on	
								normal	
								peripheral	
								blood	
								mononuclear	
								cells (PBMN).	

13								Steroid	Metabolite in
	4	5		Androst-4-en-9-		$C_{21}H_{27}N$			androgen and
	9	5	89.00	thiocyanomethyl-11-		O_3S			estrogen
	4	5	901	ol3,17-dione.			373.5		production ¹⁵
14	5	6	88.26	9-Octadecenamide		$C_{18}H_{35}N$	281.5	Amide or	Useful in food
	4	2	367			O		Stearylami	additives,
	9	2						de	Hypnotics and
									sedatives ¹⁵
15	5	5	91.73	1,2-Propanediol	,3-	$C_{25}H_{48}O_5$	428.6	Ester	Therapeutic
	2	6	989	(Octadecyloxy)-dio-					methods and
	2	9		acetate					pharmaceutical
									compositions for
									treating warts
									with tellurium
									compounds ¹⁵
16	6	7	92.31	Cycloheptasiloxane		$C_{14}H_{42}O$	519.6	Alkane	
	7	2	824	Tetradecamethyl		7Si_7			
	3	9							
17								Lipid	Nucleic acid
	5	5				$C_{39}H_{80}O_2$			transfer carrier,
	1	2	97.91	Octadecane1,1,-(1,3-					compound for
	6	7	271	propanediybis(oxy)bis.			581		nucleic acid
18								Organosil	Collagen
	8	9				$C_{16}H_{48}O_8$		oxane and	formation
	1	1	88.24	Cyclooctasiloxane,hexade		Si_8		Macrocycl	e.
	1	9	81	camethyl			593.2		
19	5	5		Steric	acid,3-			Ester	
	2	4	95.58	(Octadecyloxy)propyl		$C_{39}H_{78}O_3$			
	0	4	824	ester			595		

20

Heterocyclic
Organic
Compound
Natural active
pharmaceutical
ingredients for
oxidative
activities



6	6		Octasiloxane,1,1,3,3,5,5,7		
6	7	99.25	,7,9,9,11,13,13,15-		
9	4	816	hexadecamethyl	607.3	

Discussion

This study revealed that Octahydro-5,5a-dihydroxy-4-(hydroxymethyl) decatrienoic acid has the highest selectivity % of 100% which is a carboxylic acid with molecular formula $C_{10}H_{14}O_2$ and molecular mass of 166 followed by hexadecamethyloctasiloxane has the next selectivity % of 99.25% which is a heterocyclic acid with the molecular formula $C_{18}H_{54}O_7Si_8$ molecular mass of 607. It was noted that these two bioactive compounds are good natural active pharmaceutical ingredients as antioxidants. This suggests that ethanol CLR will possibly be a good antioxidant agent. The least bioactive compounds found in the ethanol extract of CLR were 1-(3,4-dimethoxyphenylethanone a ketone with a selectivity % of 79.9% while Hexadecanamide a fatty amide with selectivity % of 74.3%, both have anticancer effects. This indicates that ethanol CLR has a slight effect against cancer growth and diseases.

It was noted in this study that the following bioactive compounds and groups ethanol-2-(9,12-octadecadienyloxy)-; ethanone-1-(3,4-dimethoxyphenyl); 8-hexadecanal 4-methyl; methyl ester hexadecanoic acid ;

hexadecamethyl octasiloxane in alcohol, carbonyl, aldehyde, palmitic acid, heterocyclic organic acid respectively were seen in this study and earlier study⁴ This shows that ethanol extract of CLR possess good antioxidant and antimicrobial potentials. In addition to all other bioactive compounds found in ethanol CLR seen in Table 1 in this study and in aqueous CLR⁴. The ethanol CLR extract it is also found to contain androst-4-en-9-thiocyanomethyl-11-ol-3,17-dione which is a precursor for androgen and estrogen production male animals. As well as cyclooctasiloxane hexadecamethyl and Octasiloxane,1,1,3,3,5,5,7,7,9,9,11,13,13,15-hexadecamethyl in which both contain silicon an essential trace element for collagen and bone formation¹⁵.

CONCLUSION

Conclusively, in this study it can be established that the ethanol extract of CLR will better preserve CLR for livestock consumption as well as improving the bioactive compounds to make it suitable for male reproductive hormone production, collagen and bone

formation. It can also be useful in ethnoveterinary practice.

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