NIBMO

Nigerian Journal of Biochemistry and Molecular Biology

The Official Publication of the Nigerian Society of Biochemistry & Molecular Biology (NSBMB). Journal homepage: https://www.nsbmb.org.ng/journals/



Research Article

Evaluation of Nutritional, Phytochemicals and Antioxidant Capacity of *Telfairia Occidentalis* F. and *Vernonia Amygdalina* Delile Leaves

Pass C. Chijindu¹, Onyeka B. Onyeukwu^{2*}, Udomah Ogheneoruese³

OPEN ACCESS

*CORRESPONDENCE

Onyeukwu, O.B. benjamin.onyeukwu@unidel.edu.ng +234-9060500148

ARTICLE HISTORY

Received: 05/03/2024 Reviewed: 26/04/2024 Revised: 11/09/2024 Accepted: 20/09/2024 Published: 30/09/2024

CITATION

Chijindu P.C., Onyeukwu O.B., and Ogheneoruese U. (2022). Evaluation of Nutritional, Phytochemicals and Antioxidant Capacity of *Telfairia Occidentalis* F. and *Vernonia Amygdalina* Delile Leaves. *Nigerian Journal of Biochemistry and Molecular Biology.* 39(3), 170-177, https://doi.org/10.4314/njbmb.v39i3.8

ABSTRACT

This study assessed the nutritional composition, phytochemical constituents, and antioxidant activities of Telfairia occidentalis and Vernonia amygdalina leaves. Standard analytical methods were employed to evaluate the powdered and aqueous extracts of both plants. Findings indicated that T. occidentalis exhibited higher levels of moisture (11.43%), crude fiber (9.11%), protein (8.19%), and crude lipid (3.67%) compared to V. amygdalina. No significant difference was observed in total ash content between the two species. Qualitative phytochemical analysis identified the presence of saponins, cardiac glycosides, terpenoids, triterpenoids, flavonoids, and tannins in both plants, while anthraquinones and steroids were absent. Quantitatively, V. amygdalina contained higher concentrations of phenolics (25.17 \pm 1.03 mg/g), flavonoids (25.86 \pm 0.09 mg/g), saponins (7.61 ± 0.07 mg/g), and alkaloids (6.42 ± 0.07 mg/g). In contrast, T. occidentalis had greater amounts of tannins (12.47 \pm 0.06 mg/g) and cardiac glycosides (2.46 \pm 0.03 mg/g). The antioxidant capacity, measured by DPPH radical scavenging activity, revealed that both plants possess significant antioxidant properties. V. amygdalina demonstrated a higher scavenging ability with an IC₅₀ of $10.37 \pm 1.61 \,\mu g/mL$, compared to *T. occidentalis* at 22.99 \pm 0.61 μ g/mL. However, both were less potent than standard ascorbic acid (0.49 ± 0.001 μg/mL). Total antioxidant capacity (TAC) analysis showed V. amygdalina had a significantly higher TAC (0.941 ± 0.001 mg/g ascorbic acid equivalent) than *T. occidentalis* $(0.830 \pm 0.002 \text{ mg/g})$. These results suggest that *T. occidentalis* and *V. amygdalina* leaves are valuable vegetable sources capable of meeting human nutritional needs and providing defense against oxidative stress-related diseases.

Keywords: Vernonia amygdalina, Telfairia occidentalis, Antioxidant properties, Pumpkin leaf, Bitter leaf, DPPH

INTRODUCTION

Fruits and vegetables are a broad category of plant foods that differ substantially in energy and nutrient content and are generally regarded healthy (Slavin and Lloyd, 2012). Fruits and vegetables are significant providers of nutrients such as vitamins and minerals in meals because they

Copyright © 2024 Chijindu et al. This is an open access article distributed under the Creative Commons Attribution License CC BY 4.0, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

include phytochemicals that fight oxidative stress and act as antioxidants (Basu *et al.*, 2010). Many diseases stem from oxidative stress, which is caused by an imbalance in the relative amounts of vital components of oxidative metabolism (Fatima *et al.*, 2021). Oxidative stress occurs when a system can no longer effectively remove reactive oxygen species and functional metabolites (Goodarzi *et al.*, 2018). An organism's general health is determined by the complex equilibrium between pro- and antioxidants in the cellular environment (Fatima *et al.*, 2021). Antioxidants play a variety of biological roles, even at extremely low

¹Department of Biological Sciences, University of Delta, Agbor, Delta, Nigeria

²Department of Chemical Sciences, University of Delta, Agbor, Delta, Nigeria

³Department of Biology, College of Education, Warri, Delta, Nigeria.

concentrations and they also help to strengthen the immune system and defend the body against diseases caused by the invasion of uncontrolled radicals (Pisoschi and Negulescu, 2011; Sunil, 2014). A number of ageing, autoimmune illnesses, cancer, heart disease, neurological disorders, and rheumatoid arthritis are among the persistent and degenerative ailments that are greatly caused by oxidative stress (Lien *et al.*, 2008; Goodarzi *et al.*, 2018).

Fluted pumpkin (Telfairia occidentalis Hook F.) is one of the most widely used and common herbs, cultivated primarily in west and central Africa, particularly in Ghana, Benin, Nigeria, and Cameron (Kayode and Kayode, 2011). The plant is also known by the common names Ugu (Igbo), Egusi-iroko (Yoruba), and Ikong-ubong (Efik and Ibibio) (Kayode and Kayode, 2011). Fluted pumpkin is a tropical vine plant belonging to the cucurbitaceae family (Odiaka and Odiaka, 2011). Nutrients including carbohydrates, proteins, vitamins, lipids, minerals, and fibre are found in fluted pumpkin (Telfairia occidentalis), as well as phytochemicals such phenol, oxalates, glycosides, flavonoids, alkaloids, and resins, some of which are crucial for the manufacturing of pharmaceuticals (Nwite et al., 2013; Imosemi, 2018; Njoku, 2020). Many therapeutic benefits of fluted pumpkin have been linked to the treatment and mitigation of many ailments (Orole et al., 2020).

Vernonia amygdalina is a medium-sized shrub cultivated in the tropics and is known by several common names among the Nigerian populace, including Onugbu (in Igbo) (Achuba, 2018). V. amygdalina grows widespread in Africa and is a member of the Asteraceae family. In several nations, particularly in Africa, the leaves are utilised in complementary and alternative treatment. The chemical components found in V. amygdalina leaves include coumarins, terpenes, flavonoids, polyphenolic chemicals and these compounds have a variety of potential uses, including as anti-inflammatory, anti-cancer, and antioxidant properties (Cho et al., 2020; Fawwaz et al., 2020).

According to reports (Pem and Jeewon, 2015, Rush et al., 2019, Orole et al., 2020 Bokelmann et al., 2022), eating enough green vegetables is crucial for preventing malnutrition and hunger, guaranteeing food security, and helping farmers make money. A major factor in the underutilization of these vegetables by many individuals is a lack of awareness regarding their nutritional and therapeutic benefits. In order to support and validate the use of *V. amygdalina* and *T. occidentalis* as nutraceuticals, this study evaluated the nutritional, phytochemical and antioxidant properties of *T. occidentalis* and *V. amygdalina* leaves.

MATERIALS AND METHODS

Chemicals

Ethanol, chloroform, Mayer's reagent, Dragendorff's reagents, Hager's reagents, Wagner's reagents, sodium hydroxide, sulphuric acid, tannin acid, ascorbic acid,

quercetin, lead acetate, glacial acetic acid, trichloroacetic acid, methanol, phosphate buffer, Anthrone reagent, Folin C & D, Phenol (Sigma-Aldrich), hydrochloric acid, Barium hydroxide, (JHD Shantou) zinc sulphate, potassium ferricyanide, iron chloride (Loba Chemie Mumbia).

Collection and identification of plant samples

Fresh leaves of *Telfairia occidentalis* and *Vernonia amygdalina* were harvested from a Cottage farm in College of Education, Warri (Latitude 5° 3'5.11" N Longitude 5°40'44.11"E, and altitude 13.5-17.5 m.) and the leaves were carefully examined and identified by a Taxonomist at the Department of Botany, University of Lagos and they were deposited at Lagos University Herbarium (LUH) given the voucher numbers LUH 9013 and LUH 9014 respectively.

Preparation and extraction of plant samples

After giving the fresh leaves a thorough rinse and letting them air dry at room temperature, they were pulverized into a fine powder with an electric grinder and quantified. 1000g of the dry powdered plant materials were soaked in 5 liters of double distilled water to produce aqueous extracts, which were then stored at room temperature for 48 hours to ensure a thorough extraction. The extracts were filtered using cotton wool and Whatmann filter paper No. 42 (125mm) after 48 hours and the filtrate was concentrated using a rotary evaporator for 72 hours with a water bath temperature set to 40°C before freeze-drying. After that, the dried residue (raw extract) was kept in storage at 4°C. On each experiment day, aliquot amounts of the crude plant extract residue were weighed and dissolved in distilled water.

Proximate analysis

The proximate composition of the powdered sample was done in order to determine the moisture, ash, crude protein, crude lipids, crude fiber and nitrogen-free extracts (digestible carbohydrate) according to the official method of the Association of Official Analytical Chemists (AOAC, 2002).

Phytochemical Screening

The phytochemical screening of the sample was carried out according to the methods described by Evans & Trease (2002), Harborne (1998) and Sofowora (1993). The sample was screened for saponins, alkaloids reducing sugar, anthraquinones, cardiac glycosides, terpenoids, triterpenoids, steroids, phenolic compounds, Tannins and flavonoids and was quantitatively analyzed.

Evaluation of antioxidant capacity

1, 1-diphenyl -2-picryl hydrazyl (DPPH) assay

With ascorbic acid serving as the standard, the antioxidant capacity was evaluated employing the method of Manzocco *et al.* (1998). To 2 ml of DPPH solution (0.3 mM), 0.2 ml of various aqueous leaf extract concentrations was added.

Following a half-hour dark incubation period, the absorbance at 517 nm was determined. To calculate the % inhibition of DPPH radical scavenging, use the following formula:

% inhibition of DPPH radical = ([$A_0 - A_1$]/ A_0) x 100

Where A_0 is the control absorbance (blank, without extract) and A_1 is the absorbance in the presence of the extract.

Total anitioxidant capacity

The procedure described by Prieto *et al.* (1999) was employed using ascorbic acid as a reference. In screwcapped tubes, 1.0 mL (0.20–1.00 mg/ml) of the reagent solution (0.6 M sulfuric acid, 28 mM sodium phosphate, and 4 mM ammonium molybdate) was added followed by 0.1 mL of aqueous leaf extract. After being sealed, the tubes spent 90 minutes at 95 °C in a thermal block. After cooling to room temperature, each tube's aqueous solution's absorbance was measured at 695 nm against a blank. Ascorbic acid equivalents (AAE) are used to express the total antioxidant capability.

Statistical analysis

The data are displayed as Mean \pm Standard Error of Mean (SEM) (n = 3). For DPPH, analyses of variance (ANOVA) were conducted using the Least Significance Difference test in order to compare the outcomes while paired sample t-test was used to compare the outcome of the proximate composition, phytochemical analysis and total antioxidant. The threshold for statistical significance was set at p < 0.05. The linear regression plot of the extract's concentration against the percentage of inhibition and the bar charts created with Microsoft Excel were used to determine the IC50.

RESULTS AND DISCUSSION

Results

Results of the proximate composition of T. occidentalis and V. amygdalina is presented in Table 1. T. occidentalis was significantly (p< 0.05) higher in moisture content (11.43%), crude lipid (3.67%), crude fibre (9.11%) and crude protein (8.19%) while V. amygdalina was significantly (p< 0.05) higher in carbohydrates composition (67.20%). There was no significant difference in the total ash composition of T. occidentalis (4.99%) and V. amygdalina (4.79%) at (p> 0.05).

Table 1. Proximate Composition (%) of powdered sample of T. occidentalis (Pumpkin leaf) and V. amygdalina (Bitter leaf)

Samples	Moisture content	Total ash	Crude lipid	Crude fibre	Crude protein	Carbohydrates
Telfairia occidentalis	11.430 ± 0.02^{a}	4.990 ± 0.01^{a}	3.670 ± 0.02^a	9.110 ± 0.02^{a}	8.190 ± 0.02^{a}	62.710 ± 0.01^{a}
Vernonia amygdalina	10.730 ± 0.02^{b}	4.790 ± 0.01^{a}	2.010 ± 0.01^{b}	8.040 ± 0.02^{b}	$7.250 \pm 0.03^{\rm b}$	67.200 ± 0.01^{a}

Results presented as Mean ± SEM. Values with different letters (a-b) have significant differences (p< 0.05).

Tables 2 and 3 show the qualitative and quantitative phytochemicals of T. occidentalis and V. amygdalina, respectively. Alkaloids, saponins, reducing sugars, cardiac glycosides, terpenoids, triterpenoids, phenolics, tannins and flavonoids were detected while anthraquinones and steroids were not detected in both leaves. V. amygdalina contained significantly (p< 0.05) higher saponins (7.61 mg/g), phenolic compounds (25.17 mg/g), and flavonoids (25.86 mg/g) compared to the saponins (3.99 mg/g), phenolic compounds (15.95 mg/g), and the flavonoids (17.95 mg/g) of T. occidentalis. The tannins (12.47 mg/g) and cardiac glycosides (2.46 mg/g) of *T. occidentalis* were significantly (p< 0.05) higher than the tannins (9.29 mg/g) and cardiac glycosides (0.55 mg/g) of *V. amygdalina*. The alkaloids (6.42 mg/g) concentration of V. amygdalina was not significantly different compared to the alkaloids (5.34 mg/g) concentration of T. occidentalis (P> 0.05).

Table 2. Phytochemicals Detected in aqueous extract of *T. occidentalis* (Pumpkin leaf) and *V. amygdalina* (Bitter leaf)

Dhytaahamiaala	Test	Pumpkin	Bitter
Phytochemicals	Test	Leaf	Leaf
Alkaloids	Mayer`s Test	+	++
	Wagner`s Test	+	+
Saponins	Frothing Test	++	+++
Reducing sugar	Fehling`s Test	+	+
Anthraquinones	Borntrager`s	-	-
	Test		
Cardiac glycosides	Keller Killani`s	+	+++
	Test		
Terpenoids	Liebermaan-	++	+++
	Burchard		
Triterpenoids	Liebermaan-	++	+++
	Burchard		
Steroids	Salkowski`s	-	-
	Test		
Phenolic	Lead acetate	+	+++
Compounds	Test		
Tannins	Ferric chloride	++	++
	Test		
Flavonoids	Shinoda`s Test	+	+

+++ = abundant, ++ = moderately present, + = lightly present, - = absent

Table 3. Concentrations (mg/g) of Phytochemicals in aqueous extract of T. occidentalis and V. amygdalina

			Cardiac		Phenolic		
Samples	Saponins	Alkaloids	glycosides	Steroids	Compounds	Flavonoids	Tannins
Telfairia							
occidentalis	3.990 ± 0.02^{a}	5.340 ± 0.02^{a}	2.460 ± 0.03^{a}	0.330 ± 0.02^{a}	15.950 ± 1.02^{a}	17.950 ± 1.07^{a}	12.470 ± 0.06^{a}
Vernonia							
amygdalina	7.610 ± 0.07^{b}	6.420 ± 0.07^{a}	0.550 ± 0.03^{b}	0.740 ± 0.03^{b}	25.170 ± 1.03^{b}	25.860 ± 0.09^{b}	9.290 ± 0.07^{b}

Results presented as Mean ± SEM. Values with different letters (a–b) have significant differences (p< 0.05).

DPPH radical scavenging ability of aqueous extract of T. occidentalis and V. amygdalina leaves are shown in Figure 1. At 50% inhibitory concentration, V. amygdalina aqueous leaf extract was more effective at scavenging free radicals generated by stable DPPH's free radical ($10.37 \pm 1.61 \, \mu g/mL$) than the T. occidentalis aqueous leaf extract ($22.99 \pm 0.61 \, \mu g/mL$) but was significantly (P < 0.05) lower in scavenging DPPH's free radical when compared to the Ascorbic acid standard ($0.49 \pm 0.001 \, \mu g/mL$). The ability of the aqueous extracts of V. amygdalina and T. occidentalis leaves to

scavenge DPPH-radicals was thus demonstrated by the results. Results from the total antioxidants capacity of T. occidentalis and V. amygdalina (Figure 2) revealed a significant difference in the vegetables, with V. amygdalina having a significantly higher value (0.941 \pm 0.001 mg/g AAE) than T. occidentalis (0.830 \pm 0.002 mg/g AAE).

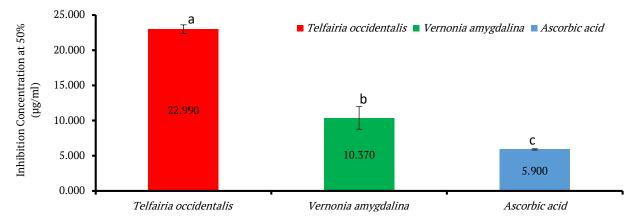


Figure 1. IC50 of *Telfairia occidentalis* and *Vernonia amygdalina* for DPPH radical. Results presented as Mean \pm SEM. Values with different letters (a–b) have significant differences (P < 0.05).

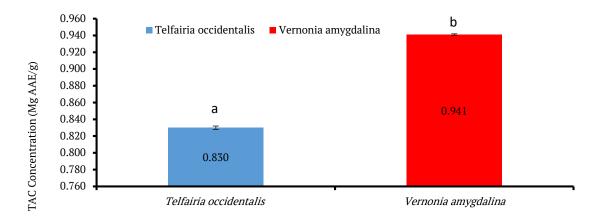


Figure 2. Total Antioxidant Capacity of *Telfairia occidentalis* and *Vernonia amygdalina*. Results presented as Mean \pm SEM. Values with different letters (a–b) have significant differences (P < 0.05).

Discussion

Plants provide nutritional value and can be used medicinally. The proximate analysis revealed that *T. occidentalis* and *V.* amygdalina leaves contained moisture, ash, lipid, fiber, protein, and carbohydrates. The dried leaves of *T. occidentalis* and *V. amygdalina* have fairly low moisture contents, which would prevent the proliferation of spoilage bacteria and extend the leaves' shelf life. The moisture content reported in this study is lower than the moisture content of leaves reported by Auwal et al. (2023) and slightly higher than the moisture content of leaves reported by Omimakinde et al. (2018). Elevated levels of moisture have been shown to support the preservation of the protoplasmic content of cells (Gbadamosi et al. 2011). It also promotes the activity of hydrophilic enzymes and coenzymes that are necessary for leafy green vegetable metabolism (Ihenacho and Udebuani, In contrast, the results of this study indicate that *T.* occidentalis leaves have more moisture content, are more quickly digested after consumption and are more prone to bacterial degradation during storage than V. amygdalina leaves. The amount of ash on leaves is an indicator of the mineral content. The ash content reported in this study is lower than the 8.3% and 8.6% reported by Adeyeye and Omolayo, (2011) and Auwal et al. (2023) respectively. The low amount of crude fat reported in this study is consistent with the findings of Banerjee et al. (2012), who stated that vegetables have minimal fat content. It is well recognised that the fatty acids found in vegetables promote membrane fluidity and facilitate gaseous exchange between intracellular and extracellular osmosis (Lovejoy, 2002). The crude fibre value in this study agrees with the result of Iheanacho (2024) who reported 10.63% crude fibre of *T. occidentalis*. Olaoye et al. (2023) reported (2.32 % and 2.15 %) of crude fibre of some leaves. Eating foods high in dietary fibre has been linked to lower blood cholesterol levels and a lower risk of heart disease, hypertension, diabetes, colon cancer, and breast cancer, among other illnesses (Ebana et al., 2019, China et al., 2021). The result of this study is consistent with the 8.72% of crude protein reported by Spranghers et al. (2017). Proteindense plant foods are those that have more than 12% of their calories from protein (Ali, 2020). This provides evidence of the low protein content of leafy greens. The leaves in this study have a high amount of carbohydrates. Previous authors have reported high content of carbohydrates in leaves (Omimakinde et al., 2018, Ali, 2020, Auwal et al., 2023, Olaoye et al., 2023). The high concentrations of carbohydrates in the leaves suggest that they provide a significant contribution to the food total energy content. Carbohydrates are primarily used by the human body as a fuel and energy source to support daily activities and physical exercises. Phytochemicals are evidence that these leaves contain bioactive compounds with potential medical use. The phytochemicals reported in this study support previous researches carried out by various authors (Elejere et al., 2019, Okoye and Orakwue, 2019, Ali, 2020, Udosen and Osu, 2022) who have reported the presence

and concentrations of phytochemicals such as alkaloids, flavonoids, tannins, saponins, phenols, terpenoids, steroids, cardiac glycosides etc. Researches have shown that flavonoids may enhance the body's defense against several disorders such as injury, cancer, ageing, atherosclerosis, inflammation, and neurodegeneration by increasing the immune system's antioxidant levels (Pal et al., 2012, Dilipkumar and Preeti, 2013). According to reports, phenols have antioxidant properties that guard against oxidative damage to cells by scavenging free radicals (Ugwu et al., 2013). Because phenolics can scavenge or neutralise free radicals, they have anti-inflammatory properties and reduce the risk of heart disease (Omale and Okafor, 2008). There have been reports that digestive disorders can be treated with leaves that contain tannins (Akindahunsi and Salawu, 2005). The metabolism and growth of living things are significantly influenced by alkaloids (Edeoga et al., 2006). It is a good chemical for plants since it keeps parasites and predators away. The antibacterial action of the alkaloid has been attributed to the presence of recognized antimicrobial agents (Usunobun and Okolie, 2016). The potential of natural compounds to function as antioxidants can usually not be determined by one asssay (Rahman et al., 2015). Hence, we evaluated the antioxidant potency of aqueous leaf extract of V. amygdalina and T. occidentalis using two techniques in this study. In both technique used for the evaluation of antioxidant capacity, V. amygdalina was significantly higher than T. occidentalis although both vegetables recorded antioxidant ability, capable of scavenging free radicals. Enhancing the quality of life by preventing or delaying the development of chronic illnesses and potentially saving a significant amount of money on medical services are two major benefits of free radical scavenging and antioxidant capability (Alam et al., 2013; Anigboro et al., 2019). A number of researchers have investigated the antioxidant capacity of leaves and vegetables (Iyamah et al., 2014; Agambi et al., 2017; Anigboro et al., 2019; Chijindu et al., 2022; Hasibuan et al., 2023; Onyeukwu et al., 2023), indicating their potential as pharmacological and therapeutic ability to combat oxidative stress. The bioactive components of the plant's leaves may be responsible for the extract samples' demonstrated antioxidant properties in this investigation. Fruits and vegetables contain the highest concentrations of naturally occurring antioxidants, such as carotenoids, tocopherols, flavonoids, phenolic acids, vitamin C, and certain minerals (Abirami et al., 2015; Kulczyński et al., 2020). Compared to manufactured antioxidants, natural antioxidants have no negative health effects on humans (Azeem et al, 2021). A large class of phytochemical substances known as flavonoids and phenols are synthesized by plants during metabolic processes as secondary metabolites connected to physiological processes such as decreasing or eliminating oxidative radicals and regulating plant growth. The process underlying this antioxidant action is that reactive oxygen species produced

are neutralized by phenols and flavonoids' functional hydroxyl groups (Kumar and Pandey, 2013.

CONCLUSION

This study revealed that *T. occidentalis* (pumpkin leaf) and *V. amygdalina* (bitter leaf) are rich in antioxidants and can be used to combat oxidative stress related diseases in which the participation of reactive oxygen species has been implicated. This research has revealed the importance of pumpkin and bitter leaf for human consumption, as traditional herbs and also as possible source for the formulation of drugs as both plants are good source of antioxidants.

AUTHORS' CONTRIBUTIONS

Conceptualization and supervision: PCC; Experimentation: UO and PCC; Data analysis: OBO; Writing-original draft preparation: UO and PCC; Writing-review and editing: OBO and PCC; Resources: OBO, PCC and UO. All authors approved the final version of the manuscript.

FUNDING STATEMENT

This research did not receive any funding.

CONFLICT OF INTEREST

No conflict of interest declared.

ACKNOWLEDGEMENT

We thank the staff of Faculty of Pharmacy Laboratories, University of Lagos for providing their facility and for their support during the analysis.

REFERENCES

- Abirami, A., Nagarani, G. and Siddhuraju, P. (2015). Hepatoprotective effect of leaf extracts from Citrus hystrix and C. maxima against paracetamol induced liver injury in rats. *Food Science and Human Wellness*, 4:35–41. Doi:10.1016/j. fshw.2015.02.002
- Achuba, F.I. (2018). Role of bitter leaf (Vernonia amygdalina) extract in prevention of renal toxicity induced by crude petroleum contaminated diets in rats. *International Journal of Veterinary Science and Medicine*, 6: 172-177
- Adeyeye, E. I. and Omolayo, F. O. (2011). Chemical composition and functional properties of leaf protein concentrate of *Amaranthus* hybridal and *Telfairia* occidentalis. *Agriculture and Biology Journal of North America*,2:499-
 - 511.https://doi.org/10.5251/abjna.2011.2.3.499.511.
- Aganbi, E., Onyeukwu, O. B., Avwioroko, J. O. and Tonukari, N. J. (2017). Effect of fermentation on sensory, nutritional and antioxidant properties of mixtures of aqueous extracts of Hibiscus sabdariffa (zobo) and *Raphia hookeri* (raffia) wine. *Nigerian Journal of Science and Environment*, 15(1): 66 74.
- Akindahunsi, A. A., and Salawu, S. O. (2005). Phytochemical screening of nutrient and antinutrient composition of selected tropical green leafy vegetables. *African Journal of Biotechnology*, 4: 497-501.

- Alam, M.N., Bristi, N.J. and Rafiquzzaman, M. (2013). Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharmaceutical Journal*, 21(2):143–152.
- Ali, M. (2020). Determination of proximate, phytochemicals and minerals composition of *Vernonia amygdalina* (Bitter Leaf). *Nutraceutical Research*, 1(1): 1-8
- Anigboro, A.A., Avwioroko, O.J., Ohwokevwo, O.A. and Pessu, B. (2019). Bioactive components of *Ficus exasperata, Moringa* oleifera and Jatropha tanjorensis leaf extracts and evaluation of their antioxidant properties. *Eurasian Journal of Biosciences*, 13: 1763–1769.
- Association of Official Analytical Chemists. (2002). Official method of analysis. 15th edition, *Association of Officials Analytical Chemists, Washington DC*, pp. 910-928.
- Auwal, Y., Bardea, M.I. Imam, N., Murtalaa, A. (2023). Proximate analysis of *Telfairia occidentalis* (Fluted Pumpkin) and *Telfairia* pedata (Oyster Nut) leaves consumed in katsina metropolis: A comparative study. *Recent Advances in Natural Sciences*, 1(8): 1-4.
- Azeem, A.M.A., Mounir, A.M. and El-shahat, A.N. (2021). Studying the anti-diabetic effect of gamma- irradiated pumpkin seeds. *Pakistan Journal of Zoology*, 54(2):851-857.
- Banerjee, A., Datta, J. K. and Mondal, N. K. (2012). Biochemical changes in leaves of mustard under the influence of different fertilizers and cycocel. *Journal of Agricultural Technology*, 8(4): 1397–1411.
- Basu, A., Rhone, M. and Lyons, T. (2010). Berries: emerging impact on cardiovascular health. *Nutrition Review*, 68: 168-77.
- Bokelmann, W., Huyskens-Keil, S., Ferenczi, Z., Stöber, S. (2022). The role of indigenous vegetables to improve food and nutrition security: Experiences from the project hortinlea in Kenya (2014–2018). *Frontiers in Sustainable Food Systems*, 6:1-19. Doi: 10.3389/fsufs.2022.806420
- Chijindu, P.C., Biadoyo, A.O. and Atubi, O. (2022). Mineral composition and antioxidant analysis of the leaves of monkey's potato (Solenostemon monostachyus (P. Beauv) Briq). *Nigerian Journal of Natural Products and Medicine*, 26(1): 28-32.
- China, M. A., Precious, O. N. and Owuno, F. (2021). Utilisation of fluted pumpkin (*Telfairia occidentalis*) seed milk for the production of textured vegetable protein. *European Journal of Agriculture and Food Sciences*, 3: 4-10. https://doi.org/10.24018/ejfood.2021.3.4.81.
- Cho, S.Y., Kim, H.W., Lee, M.K., Kim, H.J., Kim, J.B., Choe, J.S. and Jang, H.H. (2020). Antioxidant and anti-inflammatory activities in relation to the flavonoids composition of pepper (Capsicum annuum L.). *Antioxidants*, 9(10):1-10 https://doi.org/10.3390/antiox9100986.
- Dilipkumar, P., and Preeti, V. (2013). Flavonoids: A powerful and abundant source of antioxidants. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3): 95-98
- Ebana, R. U., Edet, U. O., Anosike, K. I., Etok, C. A. and Kanu, T. O. (2019). Nutritional analysis and wine production potentials of Telfairia occidentalis (fluted pumpkin) leaves and Cucumis sativus L.(cucumber) using baker's

- and palm wine yeast strains. World News of Natural Sciences, 22: 12-30.
- Edeoga, H. O., Omobuna, G. and Uche, L. C. (2006). Chemical composition of *Hyotissuaveoleus* and *Ocimum gratissium* hybrids from Nigeria. *African Journal of Biotechnology*, 5(910):892-895.
- Elejere, C. P., Okpara, C. E. and Ukwujiagu, C. U. (2019). The phytochemical contents of Telferia occidentalis (fluted pumpkin) and the effect of its aqueous extract on blood glucose and enzymatic antioxidant of stress induced rats. *Journal of Dietitians Association of Nigeria* (JDAN), 10: 52-57.
- Fatima, N., Baqri, S.S.R., Alsulimani, A., Fagoonee, S., Slama, P., Kesari, K.K., Roychoudhury, S. and Haque, S. (2021). Phytochemicals from Indian ethnomedicines: promising prospects for the management of oxidative stress and cancer. *Antioxidants*, 10:1606-1609.
- Fawwaz, M., Pratama, M., Hasrawati, A., Widiastuti, H. & Abidin, Z. (2020). Total carotenoids, antioxidant and anticancer effect of penaeus monodon shells extract. *Biointerface Research in Applied Chemistry*, 11(4):11293–11302. Doi:10.33263/Briac114.1129311302
- Gbadamosi, I. T., Moody, J. O. and Lawal, A. M. (2011). Phytochemical screening and proximate analysis of eight ethnobotanicals used as antimalaria remedies in Ibadan, Nigeria. *Journal of Applied Bioscience*, 44: 2967 2971. https://www.m.elewa.org/JABS/2011/44/1.pdf.
- Goodarzi, S., Rafiei, S., Javadi, M., Haghighian, H. k. and Noroozi, S. A. (2018). Review on antioxidants and their health effects. *Journal of Nutrition and Food Security*, 3 (2): 106-112.
- Harborne, J. B. (1998). Phytochemical methods: A guide to modern techniques of plant analysis. 3rd ed. *Chapman and Hall Int*. (Ed).NY, pp. 49–188.
- Hasibuan, P.A.Z., Keliat, J.M. and Lubis, M.F. (2023). Microwave-assisted extraction impacts on pharmacological activity of Vernonia amygdalina Delile leaf extracts and ft-ir application for phytochemicals analysis. *RASĀYAN Journal of Chemistry*, 16(3):1677-1685. Doi:10.31788/RJC.2023.1638330.
- Iheanacho, I. (2024). Comparative analysis of proximate and mineral compositions of Jatropha tanjorensis L.and Telfairia occidentalis Hook F. leaves cultivated in Zaria, *Advance Journal Chemistry* B, 6: 17-30.
- Ihenacho, K.M.E. and Udebuani, A.C (2009). Nutritional composition of some leafy vegetables consumed in Imo State, Nigeria. *Journal of Applied Science and Environmental Management*, 13(3):35-38. https://doi.org/10.4314/jasem.v13i3.55349.
- Imosemi, I.O. (2018). Review of the toxicity, medicinal benefits, pharmacological actions and morphological effects of Telfairia occidentalis hook. F. *European Journal of Pharmaceutical and Medical Research*, 5(7): 22-32.
- Iyamah, P., Fasola, T. and Idu, M. (2014). DPPH free radical scavenging activity of some medicinal plants used in the treatment of malaria in South-Western, Nigeria. *Scopmed Spatula D.D.* 4(4): 213-222

- Kayode, A.A. and Kayode, O.T. (2011). Some Medicinal Values of Telfairia occidentalis: A review. *American Journal of Biochemistry and Molecular Biology*, 1(1):30–38.
- Kulczyński, B., Gramza-Michałowska, A. and Królczyk, J.B. (2020). Optimization of extraction conditions for the antioxidant potential of different pumpkin varieties (cucurbita maxima). *Sustainability*, 12(4):1-21. Doi:10.3390/su12041305.
- Kumar, S. and Pandey, A.K. (2013). Chemistry and biological activities of flavonoids: an overview. *Scientific World Journal*, 2013:1-16. doi: 10.1155/2013/162750
- Lien, A. P., Hua, H. and Chuong, P. (2008). Free radicals, antioxidants in disease and health; review. *International Journal of Biomedical Science*, 4(2):89-96.
- Lovejoy, J. C. (2002). The influence of dietary fats in insulin resistance. *Current Diabetes Reports*. 2 (5):430-440.
- Manzocco, L., Anese, M. and Nicoli, M.C. (1998). Antioxidant properties of tea extracts as affected by processing. *Lebens-mittel-Wissenschaft Und-Technologie*, 31 (7–8), 694–698.
- Njoku, T.R.F., Udofia, H.S, Nwoko, C.O., Ihejirika, C.E., Mgbahuruike, L.U., Ebe, T.E., Egbauawa, I.O. and Ezike, M. N. (2020). Heavy metal concentration level in fluted pumpkin (*Telfairia occidentalis*) grown around Obio/Akpor, Rivers State, Nigeria: Its health implications. *Journal of Environmental Science and Public Health*, 4:16-31.
- Nwite, J.C., Keke, C.I., Obalum, S.E., Essien, J.B., Anaele, M.U. and Igwe, C.A. (2013). Organo-mineral amendment options for enhancing soil fertility and nutrient composition and yield of fluted pumpkin. *International Journal of Vegetable Science*, 19:188 199.
- Odiaka, N. I. and Odiaka, E. C. (2011). Gold mine in indigenous vegetable: The case of fluated pumpkin (*Telfairia Occidentalis Hook F.*) for economic growth. *Acta Horticulturae*, 911: 279-284.
- Okoye, E. I. and Orakwue, F. C. (2019). The chemical evaluation and anti-microbial screening of extracts from seeds and leaves of *Telfairia Occidentalis* (Fluted Pumpkin). *Chemistry Research Journal*, 4(3):98-104
- Olaoye, A., Jelili, A. and Olatunji, F. K. (2023). Heavy Metal Contents and Proximate Analysis of *Telfairia occidentalis* (fluted pumpkin) Brought from Dagbolu along Ikirun, Olorunda Local Government and Irepodun Village Osogbo Local Government, Osun State. *Journal of Chemistry and Nutritional Biochemistry*, 4(1): 21–26. DOI: https://doi.org/10.48185/jcnb.v4i1.794
- Omale, J. and Okafor, P. (2008). Comparative antioxidant capacity, membrane stabilization, polyphenol composition and cytotoxicity of the leaf and stem of Cissusmultistriata. *African Journal of Biotechnology*, 7(17):3129-3133.
- Omimakinde, A. J., Oguntimehin, I., Omimakinde, E. A. and Olaniran, O. (2018). Comparison of the proximate and some selected phytochemicals composition of Fluted Pumpkin (*Telfairia occidentalis*) leaves and pods. *International Biological and Biomedical Journal*, 4(4):206-212.

- Onyeukwu, O.B., Dibie, D.C. and Njideaka, O.T. (2023). *Hibiscus sabdariffa* - uses, nutritional and therapeutic benefits - A review. *Journal of Bioscience and Biotechnology Discovery*, 8(2): 18-23. Doi:10.31248/IBBD2023.178.
- Orole, T.R., Orole, O.O., Aisoni, E.J., Ishyaku, J. and Muhammed, Y.S. (2020). Comparative study of the physicochemical properties of male and female fluted pumpkin (*Telfairia occidentalis*). *The Journal of Medical Research*, 6(2), 55-61.
- Pal, D., Saha, S., and Singh, S. (2012). Importance of pyrazolemolety in the field of cancer. *International Journal of Pharmacy and Pharmaceutical Science*, 4(2): 98-104.
- Pem, D. and Jeewon, R. (2015). Fruit and Vegetable Intake: Benefits and Progress of Nutrition Education Interventions- Narrative Review Article. *Iranian journal* of Public Health, 44(10): 1309–1321.
- Pisoschi, A.M., and Negulescu, G.P. (2011). Methods for total antioxidant activity determination: A review. *Biochemistry and Analytical Biochemistry*, 1(1):106-118.
- Prieto, P., Pineda, M. and Aguilar, M. (1999). Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: Specific application to the determination of vitamin E. *Analytical Biochemistry*, 269(2):337-341. Doi: 10.1006/abio.1999.4019.
- Rahman, M., Badrul, I., Mohitosh, B. and Khurshid, A.H.M. (2015) In vitro antioxidant and free radical scavenging activity of different parts of *Tabebuia pallida* growing in Bangladesh. *BMC Research Notes*, 8:1-9.
- Rush, E., Savila. F., Jalili-Moghaddam, S. and Amoah, I. (2019). Vegetables: New Zealand children are not eating enough. *Frontiers in Nutrition*, 5:1-5.

- Doi:10.3389/fnut.2018.00134
- Slavin, J.L. and Lloyd, B. (2012). Health benefits of fruits and vegetables. *Advances in Nutrition*, 3: 506-516.
- Sofowora, A. (1993). Medicinal plants and traditional medicine in Africa; *John Wiley and Sons*, Ltd, Ife, Nigeria, pp. 55-201.
- Spranghers, T., Ottoboni, M., Klootwijk, C., Ovyn, A., Deboosere, S., De Meulenaer, B., Michiels, J., Eeckhout, M., De Clercq, P. and De Smet, S. (2017) Nutritional composition of black soldier fly (*Hermetia illucens*) prepupae reared on different organic waste substrates, *Journal of the Science of Food and Agriculture*, 7(8):2594-2600. https://doi.org/10.1002/jsfa.8081
- Sunil, K. (2014). The importance of antioxidant and their role in pharmaceutical science A review. *Asian Journal of Reserch in Chemical and Pharmaceutical Sciences*, 1: 27-44
- Trease, G.E. and Evans, W.C. (2002). Phytochemicals. In: *Pharmacognosy*. 15th ed. Saunders Publishers, London, pp. 42-44, 221- 229, 246- 249, 304-306,331-332, 391-393.
- Udosen, I.R and Osu, S.R. (2022). Phytochemistry, and effects of Telfairia occidentalis leaf extracts on the growth and haematological properties of Wistar Albino Rats. *Journal of Applied Science and Environmental Management*, 26(2): 317-322.
- Ugwu, O.P.C., Nwodo, O.F.C., Joshua, P., Aburbakar, B., Ossai, E.C. and Odo, C.E. (2013). Phytochemical and acute toxicity studies of *Moringa oleifera* ethanol leaf extract. *International Journal of Life Sciences, Biotechnology and Pharma Research*, 2(2):66-71.
- Usunobun, U. and Okolie, P. N. (2016). Phytochemical analysis and proximate composition of *Vernonia amygdalina*. *International Journal of Scientific World*. 4(1):11-14.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher. The publisher remains neutral with regard to jurisdictional claims.

Submit your next manuscript to NJBMB at https://www.nsbmb.org.ng/journals