Research Article

The Effect of Concurrent Administration of Cadmium and Arsenic through the Food Chain on some Testicular Toxicity Indicators of Rats

Theresa Ezedom[1*], Samuel O. Asagba[2]

[1]Department of Medical Biochemistry, Delta State University, Abraka, Nigeria.
[2]Department of Biochemistry, Delta State University, Abraka, Nigeria.

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ABSTRACT

This study examined the effect of concurrent administration of cadmium and arsenic through the food chain on testicular antioxidant status, protein concentration and expression of mRNA of Bax and Bcl-2 genes in rats. Catfish were exposed to both metals at a concentration of 0.4 mg/metal/100mL for 1 month served as source of protein for the experimental diet which rats were exposed for 1 and 3 months. The metal burden on the feed and testes, activities of antioxidant enzymes, mRNA expression of Bax and Bcl-2 in the testes of rats were then determined using standard procedures. From the results, the activities of the antioxidants enzymes and reduced glutathione (GSH) were significantly ($p<0.05$) decreased after 3 months. Testes total protein and MDA levels were significantly ($p< 0.05$) increased after both periods of exposure. Increase in the level of mRNA expression of Bax gene and a decrease in Bcl-2 gene in the test groups compared to the control after 3 months of exposure were recorded in rats. In conclusion, these results showed that exposure to these metals through the food chain increased oxidative damage leading to alteration in the expression of Bax/Bcl-2 ratio which could have significant consequences in the induction of apoptosis and other associated biochemical cascade.

INTRODUCTION

Several studies on the decline in male reproductive ability as a result of environmental toxicants have been reported (Sukhn *et al*., 2018). Cadmium (Cd) and arsenic (As) are among the major environmental contaminants. Cadmium is gotten primarily from mining, plastics manufacturing, paint pigments, electroplating, alloy preparation, and batteries. Food is the most important source of exposure to Cd for the non-smoking and non-occupationally exposed population (Khan *et al*., 2017). Arsenic exposure could occur through drinking As-contaminated water (Mandal, 2017). Other important sources of exposure to these metals are through food and inhalation. Cadmium and arsenic can exert both carcinogenic and non-carcinogenic effects on various tissues such as the lung, bone, skin, liver, kidney etc. Cadmium has also been found to have adverse effects on the endocrine and

reproductive systems (Pavlova and Atanassova, 2018) and exerts its toxicity through several biochemical mechanisms associated with oxidative stress and cell death (Kumar *et al*., 2017). Cell death could occur through apoptosis, which is characterised by loss of plasma membrane phospholipids asymmetry, enzymatic cleavage of the DNA, condensation of nuclear chromatin and cell segmentation into apoptotic bodies (Naseri *et al*., 2015). The main regulator of this process is the Bcl-2 family proteins. These help in the suppression of cell death. Bax-like subgroups such as Bax, Bak, Bok and Bik have been reported to promote cell death (Leibowitz and Yu, 2010). Although the exact mechanism through which Bcl-2 family proteins regulate the apoptotic pathway has not been fully elucidated, it has been shown to be dependent on protein-protein interactions (Naseri *et al*., 2015). This protein-protein interaction could be induced by mixtures of chemicals in the environment.

According to Sexton and Hattis (2007), a mixture is defined as the combination of two or more environmental agents. The toxicological actions of components in a mixture could be described using the concept of additivity and interaction (which could be synergistic or antagonistic). In order to describe the combined action of the components in the mixture of metals, the most common approach utilized is to carry out experimental studies to compare the effects of the individual components to the effects of the mixture (Anyanwu *et al*., 2018). However, studies on the possible effects of metals and a mixture of metals when provided through the food-chain most especially in Delta State, Nigeria, are scarce in the n literature. Environmentally, humans and animals are exposed to the combinations of various risk elements. The present study thus took account of the effects of long-term exposure to low levels of these elements through the food chain. The importance of the food chain as one of the major sources of human exposure to metals and the increasing rate of male infertility underscore the need for the present study. The aim of the study was to examine the effect of concurrent administration of cadmium and arsenic through the food chain on testicular antioxidant status, protein concentration, and expression of mRNA of Bax and Bcl-2 genes in rats.

MATERIALS AND METHODS

Preparation of experimental diet

Catfish (first trophic level) were obtained from a local fish pond located in Imoje-Orogun, Delta State, Nigeria. Exposure of the fishes to the metals was done using plastic troughs for a period of 1 month using cadmium chloride ($CdCl_2$) as the source of cadmium and arsenic trioxide (AS_2O_3) as the source of arsenic at a concentration of 0.4 mg of metal/100mL. The fishes were divided into four (4) groups. Group A which served as the control had fish kept in freshwater. In Groups B, C, and D, fishes were exposed to Cd, As, and Cd + As, respectively. The concentration of metal to which the fishes were exposed is in accordance with previous studies (Chaudhari *et al.,* 2015). The water in which the fishes were kept, was changed and re-contaminated every 24 hours. During the 1 month of exposure, fishes were provided with normal commercial feed before being sacrificed, dried, and used as a source of protein in compounding the experimental diet (Table 1). The Cd and As contents of these diets were determined by atomic absorption spectrophotometry**.**

Table 1. Composition of Experimental Diet.

Ingredients	Percentage (%) composition			
	Control	Cd-contaminated feed	As-contaminated feed	(Cd + As) contaminated feed
Control Fish	20	-	-	-
Cd treated fish	-	20	-	-
As teated fish	-	-	20	-
Cd + As treated fish	-	-	-	20
Carbohydrate	55	55	55	55
Fats and oil	10	10	10	10
Fiber (Cellulose)	10	10	10	10
Multivitamin/mineral mix	5	5	5	5

Treatment of animals

The sixty-four (64) adult male albino rats with an average weight of 126.25±3.59 g which were used for the study were gotten from the Animal House of College of Health Sciences, Delta State University, Abraka, Nigeria. The rats were acclimatized for 1 week, given a 12 h light and dark cycle with free access to normal rat chow pellet and water. Thereafter, the animals were categorized into 4 experimental groups with 16 rats per group. Groups A served as the control. Rats in Group B were exposed to the Cd contaminated diet while rats in Groups C and D were exposed to the As and Cd + As compounded diets, respectively. After exposure to the experimental diet for 1 month, half the number of rats in each group were sacrificed and the other half after 3 months of exposure. The animals were sacrificed via cervical dislocation and their testes were excised and used immediately for biochemical analysis. Animal treatments are in accordance with the principles of laboratory animal care of the National Institutes of Health (1985) and have been approved by the Committee on Animal Research and Ethics, Faculty of Basic Medical Sciences, Delta State University, Abraka, Nigeria (Ref: REC/FBMS/DELSU/19/61).

Metal analysis

Weighed samples of the feed and testes of experimental rats of each group were digested separately in beakers with 20 ml of the concentrated acid mixture (98% w/v HNO_3/$HClO_4$; 4:1 v/v) at 100℃. After digestion of samples, the Cd and As concentrations in the tissues and feeds were measured using a Varian AA 1475 spectrophotometer. An International Atomic Energy Agency (IAEA) reference biological sample was used for the evaluation of the accuracy and precision of the analysis.

Biochemical assays

The testes of the rats were weighed, homogenized, centrifuged and the supernatants obtained were used for the determination of the activities of superoxide dismutase (SOD), catalase (CAT), glutathione-s-transferase (GST) based on the methods of Misra and Fridovich (1972), Sinha (1972) and Habig *et al*. (1974), respectively. The levels of reduced glutathione (GSH), total protein, and lipid peroxidation were determined using the methods of Beutler *et al*. (1963), Gornal *et al*. (1949), and Gutteridge and Wilkins (1982), respectively. All chemicals used were of analytical grade.

Preparation of guanidine isothiocyanate (GITC) lysate

Weighed samples of testes from rats in each group and for both durations of the study were macerated aseptically using mortar and pestle under cold conditions and then transferred into a 2 mL Eppendorf tube. GITC solution was activated and the GITC lysate was prepared based on the method of Chomczynski and Sacchi (1987). The lystate was stored at -16°C prior to use.

Isolation and purification of RNA from GITC lysate

Total RNA of the GITC lysate of testes were extracted using Reliaprep RNA Kit, a product of Promega Corporation Madison, Wisconsin, USA. The integrity and purity of RNA obtained were electrophoretically verified by formaldehyde agarose gel stained with ethidium bromide based on the method of Lehrach *et al*. (1977).

cDNA synthesis protocol

In a reverse transcription reaction mixture containing 1x PCR buffer, 0.5 mM deoxy-nucleoside triphosphates (dNTPs), 1 unit of RNase inhibitor, 2.5 μM of oligo d(T)16, and 2.5 units of MuLV reverse transcriptase (Perkin-Elmer, Roche Molecular Systems, Branchburg, New Jersey, USA). One microgram (1μg) of RNA was reverse transcribed into cDNA. This was incubated for 10 mins at room temperature to allow primer annealing, then, the reaction mixture was incubated at 42°C for 15 min, heated to 95°C for 5 min, and chilled at 4°C for 5 min in a GeneAmp thermal cycler (Applied Biosystems, Foster City, CA, USA). Two microliters (2 μL) of the resultant cDNA products were used for PCR amplification.

Real-time quantitative RT-PCR

A Lightcycler 2.0 system (Roche Applied Systems, USA) was used for the Real-time quantitative RT-PCR to analyze the expression levels of the Bax and Bcl-2 gene relative to glyceraldehyde-3-phosphate dehydrogenase (GAPDH). The real-time quantitative PCR probe design software (Roche Applied Systems, Branchburg, New Jersey, USA) were used to design primer sets (Table 2) for GAPDH, Bax, and Bcl-2. PCR reactions for these primers were first optimized using conventional PCR.

Table 2. Primers Used for the Amplification of Bax and Bcl-2 mRNA in the Real-Time Quantitative Reverse Transcription-Polymerase Chain Reaction (qRT-PCR)

Gene	Primer
GAPDH	F: GGCTCTCTGCTCCTCCCTGTTCTA
	R: TGCCGTTGAACTTGCCGTGG
Bcl-2	F: CTGGTGGACAACATCGCTCTG
	R: GGTCTGCTGACCTCACTTGTG
Bax	F: TTCATCCAGGATCGAGCAGA
	R: GCAAAGTAGAAGGCAACG

For the quantitative Real-Time PCR, 20 μL amplification mixtures (LightCyclerFaststart DNA MasterPLUS SYBR Green Reaction Mix; Roche Applied Science) were prepared as per manufacturer's instructions, containing cDNA (equivalent to 100 ng reverse-transcribed RNA) and 0.5 μM of each primer. The cycling conditions were: 10 min polymerase activation at 95°C and 40 cycles at 95°C for 15 s, 58°C for 15 s, and 72°C for 15 s. After thermal cycling, the gel was run with 3% agarose gel (that is, 3 g of agarose powder in 100 mL of TBE buffer). On cooling, 20 μL EtBr was added. Electrophoresis was run at 100 V for 30 min, watching the controls.

Statistical analysis

All the data are expressed as mean ± standard deviation (SD). Statistical comparisons were performed by one-way analysis of variance (ANOVA) followed by Fisher's least significant difference (LSD) comparison when ANOVA results indicate a statistically significant difference between groups. The SPSS software (version 20) was used in the statistical analysis using multiple comparison tests. A p-value of less than 0.05 ($p < 0.05$) was considered significant.

RESULTS AND DISCUSSION
Results

The results of metal analyses carried out on the feed and testes of rats are shown in Table 3. There were trace amounts of the metals in the control feed and testes most especially for arsenic as the level of cadmium was below the detection limit. The test groups also had slight contamination of these metals as evident by the mole ratio obtained. Though only cadmium was introduced in the feed-in Group B and arsenic only in Group C, metal analysis shows that both metals exist naturally concurrently. The metals accumulated in the testes for the different groups are also indicated.

Table 3. The Concentration of Metals in Feed and Testes of Rats.

Groups	Metal concentration in experimental feed (mg/g)		Mole ratio (Cadmium : Arsenic) x 10^{-7}	Metal concentration in testes (mg/g tissue)	
	Cadmium	Arsenic			
A	Not Detected	0.02±0.56 [a]	-	0.02±0.09 [a]	
B	3.68±0.62 [b]	0.03±0.07 [a]	81.75:1	3.01±1.01 [b]	
C	0.01±0.32 [a]	1.82±0.18 [b]	1:273.03	0.38±0.01 [c]	
D	3.50±0.14 [b]	1.52±0.26 [b]	1.5:1	Cd in group D diet 2.21±0.14 [d]	As in group D diet 0.16±0.01 [c]

Results are expressed as Mean±SD. Values not sharing same superscript in same column differs significantly at (P<0.05). A, Control; B, Cd-contaminated diet; C, As-contaminated diet; D, Cd + As contaminated diet.

Table 4 presents the changes in the body weight and testes/body weight ratio of rats used in the present study for the experimental period of 1 and 3 months. There was a significant (p<0.05) reduction in the body weights of rats in all test groups relative to controls after both periods of study. The weight loss in the 3 months exposure was higher than that of the 1-month exposure and this effect was more severe in rats fed a combination of both metals in the diet. There was a significant (p<0.05) decrease in the testes/body weight ratio of rats fed with Cd-contaminated diet and Cd + As-contaminated diet relative to control after 1-month exposure. Similarly, the testes/body weight ratio of rats exposed to Cd and As singly or in combined form in the diet was significantly decreased relative to control. Again, in both duration of exposure, the effect of the metals on testes/body weight ratio was more severe when combined in the diet.

Table 4. Effect of Food Chain Mediated Exposure to Cd and As on Body Weight Changes and Testes/ Body Weight Ratio of Rats.

Group	Bodyweight changes (g)	Testes/body weight ratio
1-month exposure		
Control	(+)3.19±0.96 [a]	1.46±0.13 [a]
Cd-contaminated diet	(-)2.22±0.38 [b]	1.15±0.18 [b]
As-contaminated diet	(-)2.91±0.67 [b]	1.34±0.11 [a]
Cd + As-contaminated diet	(-)3.48±0.64 [c]	0.87±0.06 [b]
3 months exposure		
Control	(+)9.24±1.84 [a]	1.63±0.28 [a]
Cd-contaminated diet	(-)6.52±1.34 [b]	0.70±0.07 [b]
As-contaminated diet	(-)7.81±0.26 [c]	1.03±0.06 [c]
Cd + As-contaminated diet	(-)13.08±0.46 [d]	0.61±0.03 [b]

Results are expressed as Mean±SD. n = 16. Values not sharing the same superscript in the same column differ at (P<0.05).

The effects of the treatment on the total protein and antioxidant status of the testes are shown in Table 5. The levels of total protein and malondialdehyde (MDA) (an index of lipid peroxidation) were significantly (p<0.05) increased when compared to the control for both periods of exposure. After 1 month of exposure, there was a significant (p<0.05) increase in the level of GSH and activities of SOD, CAT, and GST but after 3 months of exposure, these were significantly (p<0.05) decreased.

The changes in the expressions of Bax and Bcl-2 gene mRNA in the testes of rats exposed to Cd and As contaminated diet for both periods of exposure are shown in Figure 1. There was a significant (p<0.05) increase in the level of Bax gene mRNA expression for the Cd and Cd + As a contaminated group when compared to the control after 1 month of exposure. However, there was a significant (p<0.05) increase in the mRNA expression of the Bax gene in the testes for all test groups when compared to the control after 3 months of exposure

The expression of Bcl-2 gene mRNA was found to be downregulated when compared to the control for all groups. In the group exposed to As contaminated diet, this reduction was not significant (p<0.05) after 1-month exposure when compared to the control. However, after 3 months of exposure, there was a significant (p<0.05) decrease in the level of Bcl-2 mRNA for all test groups.
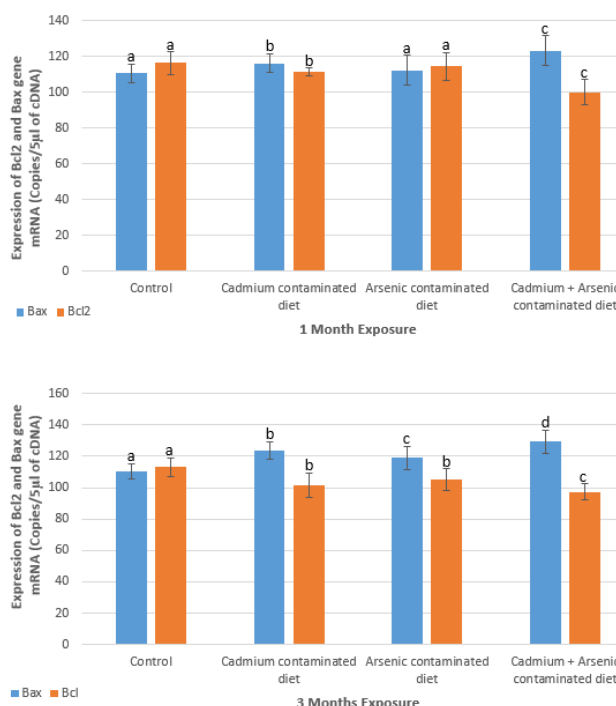
Figure 1. Changes in the Expression of Bcl-2 and Bax Gene in the Tissues of Rats after 1 And 3 Month Exposures to Experimental Diet.

Values not sharing the same superscript within a tissue differ significantly at (P<0.05).

Table 5. Effect of Treatment on Total Protein and Antioxidant Status of Testes.

Group	MDA (units/g testis)	Total Protein (mg/g)	SOD (units/g testis)	CAT (μmole H_2O_2/min/mg protein)	GST (μmol CDNB – GSH complex formed/min/mg protein)	GSH (mg/g testis)
1 Month Exposure						
A	56.86±1.56[a]	6.33±0.70[a]	34.47±0.20a	67.45±1.48[a]	7.75±1.14 [a]	31.00±1.41[a]
B	72.53±1.25[b] (27.56%)	8.75±1.09[b] (38.23%)	44.27±0.24[b] (28.43%)	88.30±1.83[a] (30.91%)	10.48±1.58[b] (35.23%)	39.33±2.31[b] (26.87%)
C	69.38±0.70[c] (22.02%)	7.50±0.21[b] (18.48%)	40.29±0.28[b] (16.88%))	71.85±4.77[b] (6.52%)	8.10±0.10 [a] (4.52%)	33.04±2.82[a] (6.58%)
D	89.51±5.30[d] (57.42%)	8.25±0.26[b] (30.33%)	57.49±1.25[c] (66.78%)	88.00±5.65[c] (30.46%)	13.76±3.83[c] (77.54%)	49.00±4.24[d] (58.06%)
3 Months Exposure						
A	59.10±1.75[a]	8.00±0.08 [a]	30.73±0.24[a]	145.40±1.30[a]	8.15±0.20 [a]	32.34±3.82[a]
B	86.44±1.30[b] (46.26%)	17.00±0.17[b] (112.50%)	18.74±0.91[b] (-39.02%)	92.30±1.97[b] (-36.52%)	3.23±0.13[b] (-60.37%)	22.02±2.82[b] (-31.91%)
C	81.04±0.62[b] (37.12%)	15.00±0.14[c] (87.5%)	24.13±0.66[c] (21.48%)	102.89±4.14[c] (-29.24%)	7.17±0.06 [a] (-12.02%)	29.21±4.24[c] (-9.68%)
D	97.02±3.70[c] (64.16%)	19.33±0.11[d] (141.62%)	14.46±0.59[b] (-52.45%)	78.15±3.18[d] (-46.25%)	2.38±0.12[b] (-70.80%)	20.03±4.24[b] (-38.06%)

Results are expressed as Mean±SD. Values not sharing the same superscript in the same column differ significantly at (P<0.05). A, Control; B, Cd-contaminated diet; C, As-contaminated diet; D, Cd + As contaminated diet. Percentage changes are presented in parenthesis

Discussion

This study examined the effect of concurrent administration of cadmium and arsenic through the food chain on testicular toxicity in rats. Environmentally, humans are exposed to cadmium (Cd) and arsenic (As) at low concentrations either voluntarily through supplementation or involuntarily through intake of contaminated food and water or contact with contaminated soil, dust, or air (Giaccio *et al*., 2012). These metals could have adverse impacts on male reproductive health.

Metal analysis on the compounded experimental feed showed trace contamination of the control and the test groups. The mole ratio clearly indicated the trace presence of As in the Cd-contaminated diet and the trace presence of Cd in the As-contaminated diet. It was observed in the present study, that the control testes also had some very minimal level of contamination of the metals. This could be attributed to the pervasiveness of Cd and As in the general environment (Horiguchi *et al*., 1996).

The present study revealed a reduction in the body weight and testes/body weight ratio of rats exposed to the experimental diets for both periods of exposure. The severe effect on body weight changes and testes/body weight ratio of rats observed after 3 months exposure could be attributed to toxicity of the metals due to their increased accumulation

relative to 1 month exposure. Again, the findings also indicated that the combination of the metals in Group D had a more pronounced negative effect on the weight of rats. This is in accordance with the report of Mahaffey *et al*. (1981) who showed that both As and Cd decreased body weight and food utilization with more pronounced effects for the mixture.

It has been shown that testicular germ cells are susceptible to oxidative damage by free radicals due to the presence of polyunsaturated fatty acids in the plasma membrane (Moazamian *et al*., 2015). This leads to the excessive generation of ROS which could outweigh the cell's antioxidant defense system (Nna *et al*., 2019). Thus, markers of oxidative stress and antioxidant enzymes were analyzed in the testes of experimental rats. After 1 month of exposure, the activities of the antioxidant enzymes (SOD and CAT), and GST, a phase 1 drug-metabolizing enzyme were all increased in the testes by the Cd and As (singly and in the mixture) contaminated diet. This is apparently traceable to the low level of an oral dose of the metals in the diet as well as the short duration of exposure. Ezedom *et al*. (2020) also reported an increase in the activities of SOD and CAT upon exposure to cadmium and arsenic. An increase in the activities of the antioxidant enzymes could also be attributed to the ability of the testes to tolerate the metal stress (Zhang *et al*., 2007). Ezedom and Asagba (2016) have also reported

an increase in oxidative enzymes in testes of rats after 1-month exposure of rats to Cd and As contaminated diets. The reduction in the activities of these enzymes after 3 months of exposure may imply enzyme inactivation caused by excess reactive oxygen species (ROS) formation, displacement of an essential cofactor such as Zn or Cu, or binding to thiol groups of the enzyme (Casalino *et al*., 2002). The current study also showed that exposure to Cd and As caused a significant ($p < 0.05$) increase in membrane lipid peroxidation (LPO) in the testes of exposed rats for both durations of study which is an indication of oxidative stress and could lead to intra and inter-molecular cross-links in proteins and nucleic acids occasioned by LPO products such as MDA (Dooley *et al*., 2001).

In the present study, a significant ($p < 0.05$) increase in GSH levels in the testes was observed after 1-month exposure but it was significantly decreased at the end of 3 months. The increase in tissue GSH content after a 1-month of exposure might be attributed to the system trying to mop up free radicals generated by the metal accumulation in these organs. A high level of GSH as observed after 1 month of exposure could protect against cell death. The decreased GSH level observed after 3 months exposure could be due to the oxidation of GSH by free radicals or as a result of depletion of the sulfhydryl group of cysteine moiety in GSH due to its high affinity for Cd and As forming Cd/As-GSH complex or its electron donor ability (Akinboro *et al*., 2022). A reduction in the level of GSH might initiate the onset of cytotoxicity.

In the present study, a non-significant ($p < 0.05$) increase in the level of Bax gene mRNA in the testes of rats was recorded but down-regulated for Bcl-2 after 1 month of exposure. After 3 months of exposure, Bcl-2 was significantly ($p < 0.05$) downregulated in all test groups while Bax was significantly ($p < 0.05$) up-regulated in all test groups. Interactions between Bcl-2 and Bax regulate cytochrome c release from mitochondria and establish baseline sensitivity to apoptotic stimuli (Hata *et al*., 2015). Induction of apoptosis has been utilized for the treatment of cancer, but cancer cells have developed different strategies to resist death by apoptosis (Nachmias *et al*., 2004). One such strategy is an increase in the expression of anti-apoptotic Bcl-2 family proteins. Upon binding to Bax, Bcl-2 can prevent pore formation and cytochrome c release. On the other hand, an increase in the expression of Bax can induce cell death and is utilized in eliminating tumor cells. Based on different reports (Naseri *et al*. 2015) showing a reduction in the expression of Bax and a corresponding increase in the expression of Bcl-2 in different drug-resistant tumor cells, it could be hypothesized that induction of mitochondrial apoptosis pathway by Cd and As in the testes of rats through

the food chain might be mediated through the Bcl-2 and Bax proteins.

Christian *et al*. (2016) reported that the disruption of the blood-testis barrier (BTB) could be responsible for the sensitivity of the testes to Cd and As toxicity. Rodent testes under normal conditions have higher levels of metallothionein (MT) than other organs (liver and kidney excluded), therefore, the high susceptibility of the testes to Cd toxicity could possibly be related to genetic background. MT is a metal-binding protein that protects cells from the toxicity of metals by binding to them (Rana, 2018).

Very little information is available on the interactive effects of As and Cd upon ingestion. Schmolke *et al*. (1992) and Mahaffey *et al*. (1981) have shown using sub-chronic dietary studies in rats that neither metal significantly affected the accumulation of the other in kidney, liver, or brain tissue. Mahaffey *et al*. (1981) found that both As and Cd increased RBC count and decreased hematocrit; responses to the mixture were less than additive. One of the most striking findings from this study was the pronounced effect of the diet containing a combination of cadmium and arsenic as compared to diets containing the individual metals on most of the parameters studied which is evident in the percentage changes from the controls. These findings may have arisen from the additive effect and/or synergism of the two metals on the parameters studied. Thus, the result of the present study is in consonance with the observation of Choudhury and Mudipalli (2008) that showed that As (as arsenite) and Cd were more lethal to rats when administered as a mixture than when injected alone. In the studies of Choudhury and Mudipalli (2008) each metal lowered the LD_{50} for the other, and the two metals at single fixed doses caused greater than additive lethality as compared to either alone.

CONCLUSION

In conclusion, this study has provided evidence that consumption of food contaminated with Cd and As through the food chain could be a contributory factor to reproductive disorders in male rats. The results of the present study showed that Cd and As accumulated through the food chain can potentially affect negatively the weight and testes/body weight ratio of rats most especially as the duration of exposure increases with the mixture of metals having a greater effect than either of the metals alone. Exposure to the metals also altered the activities of antioxidant enzymes leading to an increased level of LPO by ROS, induced histological changes, and altered the normal ratio of the mRNA expression of Bax and Bcl-2genes in the testes of exposed rats and these effects could lead to loss of reproductive functions. The up-regulation of the mRNA expression of Bax and downregulation of Bcl-2gene

expression after both periods of exposure could be hypothesized as mediating induction of apoptotic pathway leading to cell death and consequently, loss of cell function. The findings of the study also revealed that the combined metals had more pronounced biochemical and histological effects on the testes of the exposed rats than the individual metals, especially in the three months exposed rats.

AUTHORS' CONTRIBUTIONS

Author TE and SOA contributed to conception, design and execution of the study, acquisition of materials required for the study, analysis and interpretation of data, drafted the manuscript, critically revised the manuscript, agreed on the journal to which the article was submitted, reviewed and agreed on all versions of the article before submission, during revision, the final version accepted for publication and agreed to be accountable for all aspects of work ensuring integrity and accuracy.

FUNDING STATEMENT

CONFLICT OF INTEREST

The authors declare that they have no competing interests

REFERENCES

Anyanwu, B. O., Ezejiofor, A. N., Igweze, Z. N. and Orisakwe, O. E. (2018). Heavy metal mixture exposure and effects in developing nations: An update. *Toxics*, 6(4), 65, doi:10.3390/toxics6040065

Akinboro, A., Adedosu, O. T., Badmus, J. A., Adegbola, P. I., Abubakar, M. Y., James, R.O. and Aborisade, A. B. (2022). *Aspalathus Linearis* extract ameliorate haematological disorder, dyslipidaemia and tissue toxicity associated with arsenic exposure in rats. *Phytomedicine Plus*, 2(1), 100171.

Beutler, E., Duron, O. and Kelly, B. M. (1963). Improved method for determination of blood glutathione. *Journal of Laboratory and Clinical Medicine*, 61, 882-888.

Casalino, E., Calzaretti, G., Sblano, C. and Landriscina, C. (2002). Molecular inhibitory mechanisms of antioxidant enzymes in rat liver and kidney by cadmium. *Toxicology,* 179(1-2), 37–50.

Chaudhari, R. T., Harshad, R. K. and Thorat, S. R. (2015). Effect of cadmium chloride on the biochemical charecteristics of fresh water fish, *Cyprinus carpio*. *International Journal of Management and Social Science,* 3(3), 85-89.

Chomczynski, P. and Sacchi, N. (1987). Single-step method of RNA isolation by acid guanidiniumthiocyanate–phenol–chloroform extraction. *Analytical Biochemistry,* 162(1), 156-159.

Choudhury, H. and Mudipalli, A. (2008). Potential considerations and concerns in the risk characterization for the interaction profiles of metals. *Indian Journal of Medical Research,* 128(4), 462-483.

Christian, E. I., Olatoye, T. R., Bamidele, F. S., Ojo, O. E. and Ademoye, K. A. (2016). Cadmium-induced testicular toxicity, oxidative stress and histopathology in Wistar rats: sustained effects of polyphenol-rich extract of *Vernonia amygdalina* leaf. *Journal of Interdisciplinary Histopathology*, 4(3), 54-62.

Dooley, P. A., Tsarouhtsis, D., Korbel, G. A., Nechev, L. V., Shearer, J., Zegar, I. S., Harris, C. M., Stone, M. P. and Harris, T. M. (2001). Structural studies of an oligodeoxynucleotide containing a trimethylene interstrand cross-link in a 5'-(CpG) motif: model of a malondialdehyde cross-link. *Journal of the American Chemical Society*, 123(8), 1730-1739.

Ezedom, T., Asagba, S. and Tonukari, N. J. (2020). Toxicological effects of the concurrent administration of cadmium and arsenic through the food chain on the liver and kidney of rats. *Journal of Basic and Applied Zoology,* 81(1), 1-12.

Ezedom, T. and Asagba, S.O. (2016). Effect of a controlled food-chain mediated exposure to cadmium and arsenic on oxidative enzymes in the tissues of rats. *Toxicology Reports*, 3, 708–715.

Giaccio, L., Cicchella, D. De Vivo, B., Lombardi, G. and De Rosa, M. (2012). Does heavy metals pollution affects semen quality in men? A case of study in the metropolitan area of Naples (Italy). *Journal of Geochemical Exploration*, 112, 218–225.

Gornall, A., Bardsmill, C. T. and David, M. M. (1949). Determination of serum protein by means of biuret reaction. *Journal of Biological Chemistry*, 177(2), 751-766.

Gutteridge, J. M. C. and Wilkins, C. (1982). Copper dependent hydroxyl radical damage to ascorbic acid. Formation of a thiobarbituric acid-reactive product. *FEBS Letters*, 137(2), 327-340.

Habig, W. H., Pabst, M.J. and Jakoby, W. B. (1974). Glutathione S-transferases. The first enzymatic step in mercapturic acid formation. *Journal of Biological Chemistry*, 249 (22), 7130-7139.

Hata, A. N., Engelman, J.A. and Faber, A. C. (2015). The BCL2 Family: Key mediators of the apoptotic response to targeted anticancer therapeutics. *Cancer Discovery,* 5(5), 475–87.

Horiguchi, H., Sato, M., Konno, N. and Fukushima, M. (1996). Long-term cadmium exposure induces anemia in rats through hypoinduction of erythropoietin in the kidneys. *Archives of Toxicology*, 71, 11–9.

Khan, M.A., Khan, S., Khan, A. and Alam, M. (2017). Soil contamination with cadmium, consequences and remediation using organic amendments. *The Science of the Total Environment,* 601–602: 1591–1605.

Kumar, K. M. K., Kumar, N. M., Nagesh, R., Patil, R. H., Babu, R. L., Sharma, S.C. and Manjunath, K. (2017). Arsenic induces oxidative stress and apoptosis in human lung cancer cells. *International Journal of Life Sciences*, 5 (4): 551-560.

Lehrach, H., Diamond, D., Wozney, J. M. and Boedtker, H. (1977). RNA molecular weight determinations by gel electrophoresis under denaturing conditions, a critical re-examination. *Biochemistry*, 16(21), 4743–51.

Leibowitz, B. and Yu J. (2010). Mitochondrial signaling in cell death via the Bcl-2 family. *Cancer Biology and Therapy*, 9(6):417–22.

Mahaffey, K. R., Capar, S. G., Gladen, B. C. and Fowler, B. A. (1981). Concurrent exposure to lead, cadmium, and arsenic. Effects on toxicity and tissue metal concentrations in the rat. *Journal of Laboratory and Clinical Medicine*, 98(4), 463-81.

Mandal, P. (2017). An insight of environmental contamination of arsenic on animal health. *Emerging Contaminants,* 3(10), 17-22.

Misra, H. P., and Fridovich, I. (1972). The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *The Journal of Biological Chemistry*, 247(10), 3170–3175.

Moazamian, R., Polhemus, A., Connaughton, H., Fraser, B., Whiting, S., Gharagozloo, P. and Aitken, R. J. (2015). Oxidative stress and human spermatozoa: diagnostic and functional significance of aldehydes generated as a result of lipid peroxidation. *Molecular Human Reproduction,* 21(6), 502–515.

Nachmias, B., Ashhab, Y. and Ben-Yehuda, D. (2004). The inhibitor of apoptosis protein family (IAPs): an emerging therapeutic target in cancer. *Seminars in Cancer Biology*, 14(4), 231–43.

Naseri, M. H., Mahdavi, M., Davoodi, J., Tackallou, S. H., Goudarzvand, M., & Neishabouri, S. H. (2015). Up regulation of Bax and down regulation of Bcl2 during 3-NC mediated apoptosis in human cancer cells. *Cancer Cell International*, 15(1).doi:10.1186/s12935-015-0204-2.

Nna, V. U., Bakar, A. B. A., Ahmad, A., Eleazu, C. O. and Mohamed, M. (2019). Oxidative stress, NF-κB-mediated inflammation and apoptosis in the testes of streptozotocin–induced diabetic rats: combined protective effects of malaysian propolis and metformin. *Antioxidants,* 8(10), 465.

Pavlova, E. and Atanassova, N. (2018). Impact of cadmium on male fertility. *Acta Morphologica et Anthropologica*, 25(1-2), 108-116.

Rana, S. V. S. (2018). Protection of metal toxicity by melatonin-recent advances. *EC Pharmacology and Toxicology*, 6.9, 851-864.

Schmolke, G., Elsenhans, B., Ehtechami, C. and Forth, W. (1992). Arsenic-copper interaction in the kidney of the rat. *EC Pharmacology and Toxicology*, 11(5), 315-21.

Sexton, K. and Hattis, D. (2007). Assessing cumulative health risks from exposure to environmental mixtures - three fundamental questions. *Environmental health perspectives*, 115(5), 825–832. https://doi.org/10.1289/ehp.9333

Sinha, A. K. (1972). Colorimetric assay of catalase. *Analytical Biochemistry*, 47(2), 389-394.

Sukhn, C., Awwad, J., Ghantous, A. and Zaatari, G. (2018). Associations of semen quality with non-essential heavy metals in blood and seminal fluid: data from the environment and male infertility (EMI) study in Lebanon. *Journal of Assisted Reproduction and Genetics,* 35:1691–1701.

Zhang, F. Q., Wang, Y. S., Lou, Z. P. and Dong, J. D. (2007). Effect of heavy metal stress on antioxidative enzymes and lipid peroxidation in leaves and roots of two mangrove plant seedlings (*Kandelia candel* and *Bruguiera gymnorrhiza*). *Chemosphere*, 67(1), 44-50.

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