

NJBMB/020/11

## Effect of *Garcinia kola*, Heckel on bioavailability of two commonly used drugs (Sulphamethazine and Paracetamol)

\*J. O. Ezekwesili – Ofili and C. G. Obeta

Department of Applied Biochemistry, Nnamdi Azikiwe University, Awka, Nigeria

### ABSTRACT

The effects of concurrent administration of *Garcinia kola* Heckel (Guttiferae) seed suspension were investigated on the bioavailability of two commonly used drugs: sulphomethazine and paracetamol in albino rabbits. Two groups of rabbits (n=4) were treated by gavage with a concentration of 0.5g/kg body weight of seed suspension given concurrently with 150mg/kg body weight of sulphonamide and paracetamol respectively. Control groups were given equivalent doses of either drug alone. Blood was withdrawn from the left ear at one hour intervals for five hours. Results showed that *G. kola* seeds decreased significantly ( $p < 0.05$ ) the bioavailability of the two drugs. Relative bioavailability was calculated to be 77.56% for sulphonamide and 75.39% for paracetamol. The time of peak and the peak concentration were also reduced while the concentration at one hour was only significantly different for paracetamol at  $p < 0.05$ . These results suggest that *G. kola* seeds may reduce bioavailability by interfering with drug absorption across the gastrointestinal mucosa.

### INTRODUCTION

*Garcinia kola* Heckel (Guttiferae), is a large forest tree found throughout Western and Central Africa. Widely known in commerce as 'bitter kola, it is used extensively in African traditional medicine for the treatment of various diseases, especially for cough, mouth and respiratory tract infections (Huchkinson and Dalziel, 1954). The seeds are not just grown for pharmaceutical purposes. They are generally harvested and used as adjunct to the true kola nut as social masticatory agent and for esoteric protection against witchcraft and reptiles and scorpion repellency (Esimone et al, 2007).

Studies have reported the use of *G. kola* seeds as aphrodisiac and for the treatment of diarrhoea and dysentery; and extracts of the seeds have been reported to possess a wide range of pharmacological activities which include anti-inflammatory (Adegoke *et al.*, 1998); antihepatotoxic (Iwu, 1985, Iwu *et al.*, 1987; Braide 1991a; Madubunyi, 2010; Akintowa and Essien, 1990); antidiabetic (Iwu *et al.*, 1990); antimicrobial (Madubunyi, 1995; Hussain *et al.*,

1982; Iwu and Igboko, 1982); adaptogenic (Esimone et al, 2007); molluscidal (Okunji and Iwu, 1991); antiasthmatic (Ore and Ekon, 1993) and inhibition of drug metabolism (Braide, 1991b). Most of these pharmacological activities are believed to be related to its antioxidative and immune-enhancing properties (Okunji et al, 1999).

One drawback in the use of *G. kola* seeds is the acclaimed reduction in the effectiveness of drugs in general. This present study is aimed at determining the effect of concurrent ingestion of *G. kola* seeds on the bioavailability of two commonly used drugs, namely, sulphamethazine, a sulphonamide antibiotic, and paracetamol, an analgesic and antipyretic with no anti-inflammatory properties, in albino rabbits.

### MATERIALS AND METHODS

#### Materials:

#### Animals

The animals used for this work were albino rabbits of average weight 1.2 kg obtained from a Rabbitry at Amawbia in Anambra State, Nigeria. The rabbits were randomly assigned to four groups of four animals each, housed in metal cages, and allowed an acclimatization

---

\*Corresponding author:  
ezekejjo@yahoo.com

period lasting 7 days, prior to use for any experiments. The animals had *ad libitum* access to water and standard laboratory animal feed

#### *Plant material*

The *G. kola* seeds were purchased fresh from Eke market, Awka, Anambra State. The seeds were dried, pulverized and stored at 4°C, until use.

#### *Chemicals*

All drugs used were bought from Gauze Pharmaceuticals, Nnamdi Azikiwe University, Awka. All chemicals used were of analytical grade and products of Sigma, South Africa, BDH, Poole, England and Merck, Germany.

### METHODS

#### *Administration of Garcinia kola seed powder and drug*

After an 8 hour overnight fast, the animals in the two test groups, received concurrently by gavage 0.5g/kg body weight of *G. kola* seed powder (suspended in distilled water) and 150 mg/kg body weight each of sulphonamide or paracetamol respectively, while the controls received only equivalent doses of the drugs without *G. kola*. The animals were allowed food and water *ad libitum*.

#### **Calculations:**

Serum paracetamol =

$$\frac{\text{Reading of unknown}}{\text{Reading of standard}} \times 323$$

#### ***Determination of serum sulphonamide***

Serum sulphonamide determination was carried out by the method of Bratton and Marshall (1939). The micromodification principle is based on the estimation of free drug in blood. After precipitating the proteins with TCA, the sulphonamide is diazotized and then coupled with N-(1-Naphthyl)-ethylenediamine dihydrochloride after destroying excess nitrate with sulphamic acid. The resulting colour was read at 540nm.

Working standard solution of sulphonamide (2.5mg/L) was obtained by adding 40ml of TCA to 5ml of stock standard solution of sulphonamide (100mg/L) and made up to 299ml with distilled water.

#### *Collection and handling of blood samples*

Blood samples were collected at one hour intervals for five hours and allowed to clot for paracetamol while whole blood was used for sulphonamide estimation. The clotted blood samples were centrifuged at 5,000g for 10 minutes to obtain the sera which were then used for the estimation of serum paracetamol.

#### *Determination of serum paracetamol*

Serum paracetamol was determined by a modified method of Glynn and Kendall (1975). The principle is based on the fact that nitrous acid introduces a nitro group into the paracetamol nucleus. The resulting nitrophenol is coloured deep yellow in alkaline solution.

1 ml of serum was placed in a 25ml centrifuge tube containing 2ml TCA (10%), mixed thoroughly and centrifuged for 2minutes. 1ml of supernatant fluid was transferred to a test tube to which 0.5ml of HCL (6mol/L), followed by 1ml sodium nitrite (100g/L). This was allowed to stand for 2minutes after which 1.0ml of sulphamic acid solution was added. After the mixture had stopped frothing, 1.0ml of sodium hydroxide (100g/L) was added. A standard paracetamol solution (300mg/ml) was likewise treated. Readings were taken at 430nm against a reagent blank

100µl of whole blood was added to 3.0ml of water in a test tube, mixed and 0.9ml of TCA (200mg/l) added. This was allowed to stand for 5 minutes and centrifuged. 2.0ml of the supernatant, standard solution or water were pipetted into test tubes as test, standard or blank respectively to which 0.5ml of HCL (6mol/L), followed by 1ml sodium nitrite (100g/L). This was allowed to stand for 2 minutes after which 2.0ml of ammonium sulphamate (5g/l) was added to each tube. The mixture was left for 2minutes and 1.0ml of coupling agent (500mg/l) added, allowed to stand for 10 minutes and read against the blank at 540nm.

### Calculations

Plasma sulphonamide ( $\mu\text{g/ml}$ ) =

$$\frac{\text{Reading of unknown}}{\text{Reading of standard}} \times 100$$

The mean values (SEM) of the serum/plasma concentrations of the drugs were plotted against time (Figs 1 and 2), and the pharmacokinetic parameters were determined and shown in table 1.

Relative bioavailability for both drugs was calculated by comparing the AUC in the presence and absence of *G. kola* seeds respectively, calculated by using the trapezoid rule. The data were analyzed using the student t test (Graph pad Prism software version 4).  $P < 0.05$  was considered significant.

### Phytochemical tests

Phytochemical analysis was carried out on the crude *G. Kola* seeds, according to established methods (Evans, 1996, Harborne, 1998).

### RESULTS AND DISCUSSION

The effect of *Garcinia kola* seed extract (100 mg/kg) on the bioavailability of two drugs, paracetamol and sulphamethazine was studied. The results (mean  $\pm$  SEM) indicated that concurrent administration of both drugs and *G. kola* seeds significantly ( $P < 0.05$ ) decreased average serum concentration, peak serum concentration and time of peak, which occurred at 2 hours for sulphonamide and at 1 hour for paracetamol, in the groups which received concurrent administration of both drugs and *G. kola* seeds. No peaks were recorded in the groups which received only the drugs (Figs 1 and 2). The initial concentrations at 1 hour were however not affected significantly ( $p < 0.05$ ) for sulphonamide ( $19.10 \pm 0.07 \mu\text{g/ml}$  for test and  $20.00 \pm \mu\text{g/ml}$  for the control). However, for paracetamol, the concentrations at one hour for test and control were  $71.2 \pm \mu\text{g/ml}$  and  $78.00 \pm \mu\text{g/ml}$  respectively, showing a more enhanced interaction. The peak plasma concentration of paracetamol and sulphonamide were  $71.2 \pm 0.00 \mu\text{g/mL}$  at one hour and  $21.4 \pm 0.07 \mu\text{g/mL}$  at two hours, respectively, in the group that received both drug and *G. kola* seeds. The bioavailability of concurrent administration of drug and *G. kola* relative to drug alone within five hours was calculated to be 77.56% for sulphonamide and 75.39% for paracetamol (Table 1).

Bioavailability, which is the fraction of an administered drug that reaches the systemic circulation, is dependent on the extent of drug absorption, and is thus crucial to its therapeutic value. Whereas for intravenously administered drugs the bioavailability is 100% or 1.0, drugs administered by other routes have bioavailability values relative to the IV route. Orally administered drugs reach the systemic circulation amidst various adverse factors such as solubility, pH of the gastrointestinal tract; first pass metabolism, etc., which interfere with absorption within the gastrointestinal tract. The test drugs, sulphonamide and paracetamol are both well absorbed in the gastrointestinal tract. Interactions may however occur between the drugs and food or other stomach contents, which would reduce their absorption and hence their bioavailability. Concurrent administration of *G. kola* seeds with the two drugs reduced their bioavailability most likely by interfering with absorption across the gastrointestinal mucosa. Esimone et al (2002) reported similar in vivo interaction between aqueous seed extracts of *G. kola* and ciprofloxacin, a broad spectrum antibiotic.

The notion that *G. kola* seeds can be used to neutralize poisons and to reduce the efficacy of drugs has been of long time ethnobotanical knowledge in the South – Eastern part of Nigeria. Patients undergoing ethnotherapy other than those requiring *G. kola* are normally advised to avoid chewing the seeds during the duration of therapy. On the other hand, people who have accidentally taken poison are advised to chew the seeds as initial first aid. Phytochemical analyses of *G. kola* seeds in this work revealed mainly the presence of flavonoids (Table 2), similar to reports by previous workers, consisting mainly of bioflavonoids, xanthenes and triterpenes which have been implicated for most of the properties (Iwu, 1985; Monago and Akhidue, 2002). The freshly cut seeds were however observed to release resinous exudates and a suspension of the dried seeds was colloidal. It is not apparent at this stage which of these components, singly or collectively, might be responsible for these observed effects. However, it is suggested that the colloidal suspension may have formed an entrapment for

the drug molecules, thus reducing their absorption across the gastrointestinal tract.

*Garcinia kola* seeds extract was reported to have protective effect against paracetamol induced hepatotoxicity (Akintowa and Essien, 1990), and CCL4 – induced erythrocyte damage (Farombi, 2002). This was attributed to its antioxidant properties (Olatunde et al, 2002). It is also probable that part of its effectiveness might be due to its interference with drug absorption as observed in this work.

Chronic ingestion of *Garcinia kola* seeds has however caused untoward pharmacological effects in rats (Braide, 1990; Braide and Grill, 1990; Braide, 1991b, Atawodi et al, 1995,). Reports have also shown considerable levels of oxalates, tannins and saponins in the seeds which may be potentially toxic when ingested in high amounts for long periods (Monago and Akhidue, 2002).

This work has validated the claim that *G. kola* seeds reduce the bioavailability of drugs when taken concurrently and as such by the same mechanism may also be exploited in reducing the absorption of chemicals and poisons..

The seeds of *G. kola* continue to be significant in traditional medical practices, for both cultural and therapeutic purposes. In view of its numerous benefits, the use plant however should not preclude its use with orthodox healthcare. However, timing is of essence. It is suggested that *G. kola* seeds should be eaten at least five hours after taking the last dose of a drug to allow enough time for sufficient/adequate absorption of drug to occur.

## References

Adaramoye, O. A. and Akinloye, O. (2000). Possible protective effect of Kolaviron on CCl<sub>4</sub>- induced erythrocyte damage in rats. *Biol. Sci. Reports*. 20 (4): 259-264.

Adegoke GO, Kumar MV, Sambaiah K, Lokesh BR (1998). Inhibitory effect of *Garcinia kola* on lipid peroxidation in rat liver homogenate. *Indian J Exp. Biol.* 36(9): 907-910.

Akintonwa, A. and Essien, A. R. (1990). Protective effects of *Garcinia kola* seed extract against paracetamol-induced hepatotoxicity in rats. *J. Ethnopharmacol.* 29(2): 207-211.

Atawodi S, Mende P., Pfundstein, B., Preusmann, R., and Spiegelhalter, B.(1995). Nitrosamines and nitrosamide formation in natural stimulants: *Cola acuminata*, *Cola nitida* and *Garcinia kola*. *Food Chem Toxicol.* 33: 625-630.

Braide VB (1990). Pharmacological effects of chronic ingestion of *Garcinia kola* seeds in rats. *Phytother. Res.* 4: 39-41.

Braide V. B. (1991). Antihepatotoxic and biochemical effects of kolaviron, a biflavonoid of *G. kola* seeds. *Phytother. Res.* 5:35-37.

Braide, V. B. (1990). Pharmacological effects of chronic ingestion of *Garcinia kola* seeds in rats. *Phytother. Res.* 4: 39-41.

Braide, V. B. and Grill, V. (1990). Histological alterations by a diet containing seeds of *Garcinia kola*: Effect on liver, kidney and intestine in the rat. *Gegenbaurs Morphol. Jahrb.* 136(1): 95-101.

Braide, V. B. (1991a). Antihepatotoxic biochemical effects of kolaviron, a biflavonoid of *Garcinia kola*. *Phytother. Res.* 5: 35-37.

Braide, V. B. (1991b). Inhibition of drug metabolism by flavonoid extract (Kolaviron) of *Garcinia kola* seeds in rats. *Phytother. Res.* 5:38-40.

Bratton A.C. and Marshal E.K. (1939). A new coupling component for sulfanilamide determination. *J. Biol. Chem.* 128: 537 - 550

Esimone, C. O.; Nwafor, S. V.; Okoli, C. O.; Chah, K. F.; Uzuegbu, D. B.<sup>1</sup>; Chibundu, C.; Eche, M. A.; Adikwu, M. U. (2002) In Vivo Evaluation of Interaction Between Aqueous Seed Extract of *Garcinia kola* Heckel and Ciprofloxacin Hydrochloride American Journal of Therapeutics: Volume 9 ( 4): 275-280

Esimone C.O., Adikwu M.U., Nworu C.S., Okoye F.B.C. and Odimegwu D.C. (2007). Adaptogenic potentials of *Camellia sinensis*, *Garcinia kola* and *Kola nitida* seeds. *Scientific Res and Essay* 2(7): 232-237.

Evans WC (1996). *Trease and Evans Pharmacognosy*. WB Saunders Company Ltd, London–Philadelphia–Toronto –Sydney–Tokyo., p.612

Farombi, O. E. (2000). Mechanisms for the hepatoprotective action of kolaviron: Studies on hepatic enzymes, microsomal lipids and lipid peroxidation in carbon tetrachloride treated rats. *Pharmacol. Res.* 42(10): 75-80.

Glynn J.P. and Kendall S.E. (1975) Paracetamol measurement *Lancet* 1:1147

Harborne J.B. (1998). *Phytochemical Methods: a guide to modern techniques in plant analysis*. 3<sup>rd</sup> Edition. Chapman and Hall. London, New York.

Hussain RA, Owegby AG, Waterman PG (1982) Kolanone, a novel polyisoprenylated benzophenone

with antimicrobial properties from the fruit of *Garcinia kola*. *Planta Medica* 44: 78-81.

Hutchinson J, Dalziel JM (1954). *Flora of West Tropical Africa*, 2nd edn., Vol1, HMSO, London, p. 295.

Iwu, M. (1985). Antihepatotoxic constituents of *Garcinia kola* seeds. *Experientia* 41: 699 – 700.

Iwu M.M. and Igboko O. (1982). Flavonoids of *Garcinia kola* seeds. *J. Nat Prod.* 45: 650-651.

Iwu MM, Igboko OA, Onwuchekwa UA, Olaniyi CO (1987). Evaluation of the hepatotoxic activity of the biflavonoids of *G. kola* seed. *J. ethnopharmacol.* 21(2): 127-138.

Iwu, M.M. Igboko, O. A. and Tempesta, M. S. (1990). Antidiabetic and aldose reductase activities of biflavonones of *Garcinia kola*. *J. Pharm. Pharmacol.* 42(4): 290-292.

Madubunyi I. I. (1995). Antimicrobial activities of the constituents of *Garcinia kola*. *Int J. Pharmacog* 33 (3): 232-235.

Madubunyi I.I. (2010) Antiohepatotoxic principles of *Garcinia kola* seeds. *Comp Clin Pathol.* 20(5): 481-485.

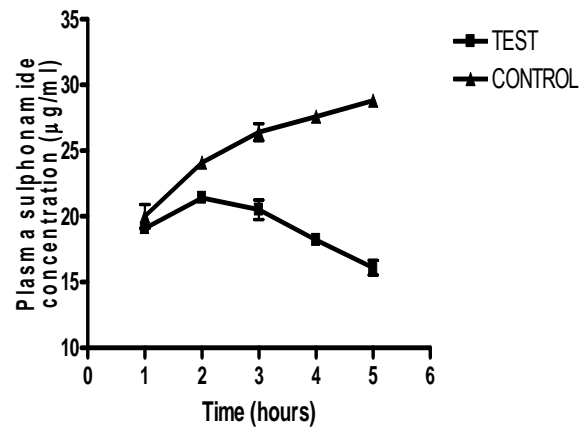
Monago C.C, and Akhidue V. (2002). Estimation of tannin, saponin, oxalate, cyanogenic and cardiac glycosides in *Garcinia kola*. *J. Applied Sci Environ Manage.* 6(1): 22-25.

Okunji CO, Iwu, MM (1991). Molluscicidal activity of *Garcinia kola* biflavanones. *Fitoterapia.* 62: 74 – 76.

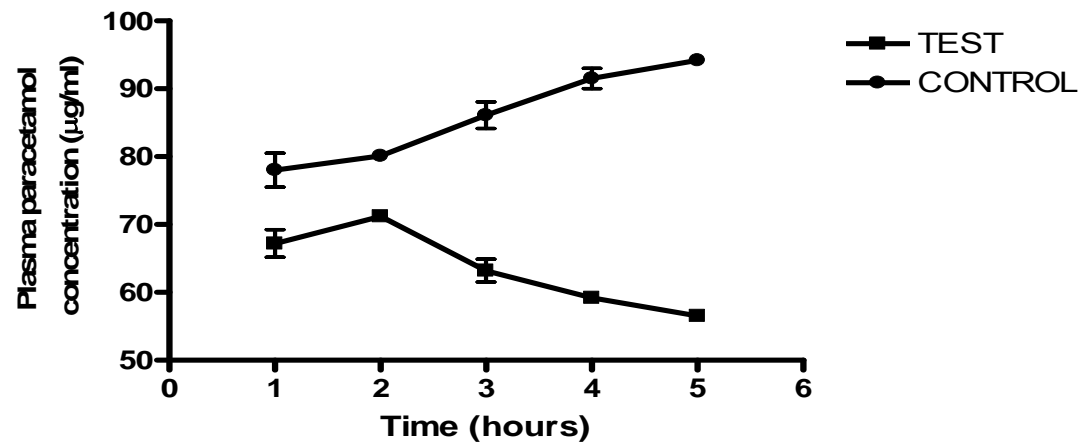
Okunji C. O., Schanchy, D. J. and Iwu, M. M. (1999). Challenges and issues involved in the standardization of *G. Kola* formulations. Abstract Int'l Conference on ethnomed. and Drug Discovery; Silver Spring, Maryland, USA, Nov 3-5, p. 29

Orie N.N. and Ekon, E. U (1993) Bronchio-dilatory effect of *Garcinia kola*. *East Afr Med J.* 70: 143-145.

**Fig 1: Plasma sulphonamide concentrations with and without G. kola administration**



**Fig 2: Plasma paracetamol concentrations with and without G. kola administration**



**Table 1: Some Pharmacokinetic parameters**

<b>Drug</b>	<b>Relative Bioavailability</b>	<b>Concentration at 1hr (<math>\mu\text{g/ml}</math>)</b>	<b>Peak Conc. (<math>\mu\text{g/ml}</math>)</b>	<b>Time of peak</b>
Sulphonamide	77.56%	Test- 19.10 $\pm$ 0.00 Control- 20.00 $\pm$ 2.15	21.4 $\pm$ 0.07	2 Hours
Paracetamol	75.39%	Test - 71.20 $\pm$ 5.46 Control - 78.00 $\pm$ 7.20	71.2 $\pm$ 0.00	1 Hour

**Table 2: Phytochemical Constituents**

<b>Phytochemicals</b>	<b>Results</b>
Alkaloids	-
Saponins	+
Tannins	+
Flavonoids	++
Proteins	+
<b>Glycosides:</b>	
Cyanogenic	-
Anthracene	+
Steroid	+
<b>Carbohydrate:</b>	
Reducing sugar	+
starch	+
sterols/Terpenoids	+

