

THE EFFECT OF MILK SUPPLEMENTATION ON THE DEVELOPMENT AND ACIDIFICATION OF LACTIC BACTERIA

*IGYOR, M.A

Department of Food Science and Technology, University of Agriculture, Makurdi, Nigeria.

* Correspondence author

ABSTRACT

Influence of yeast extract supplementation on the development and acidification of 4 strains of lactic bacteria; *Streptococcus lactis*, *Streptococcus cremoris*, *Lactobacillus casei* and *Lactobacillus acidophilus* were investigated in pasteurised skimmed milk. Three types of yeast extract, M-normal yeast extract, N-yeast supplemented with several vitamins, and O-yeast extract supplemented with group B vitamins, were employed at concentrations of 0.05%, 0.2%, 0.5% and 1.0%. Milk without yeast extract was used as control. Test cultures were inoculated into the various milk products and incubated at 37°C. Total cell counts (cfu/ml) and total titratable acidity (% lactic acid) was estimated at 0,3 and 6 hours intervals. The results showed that both cell counts and acidity increased proportionally with increased levels of yeast extract supplementation. Yeast extract could be used to remedy fermentation problems encountered in milk industry.

Key words: Lactic bacteria, yeast extract, supplementation, milk, and development.

INTRODUCTION:

Lactic bacteria are microorganisms, which in mixed or pure cultures are capable of multiplying in the bossom of milk and curds, to produce at favourable moments, the acidity and aroma sought in the milk industry (Kothari and Nambudripad, 1973). These microorganisms take part largely in the process of the preparation and the maturation of milk products like yoghurts, cheese, butter, etc., and their role lies in the glycolysis.

proteolysis, lipolysis, production of aroma constituents (diacetyl and acetoin) and the effect of inhibiting or stimulating of other microorganisms (Bautista, *et al.*, 1966; Brink ten, *et al.*, 1994; Graciela, *et al.*, 1995). They are the natural contaminants of milk and they produce almost uniquely lactic acid from sugar lactose. Acidity of fermented milk is sufficient to prevent spoilage by proteolytic or other bacteria that are not acid-tolerant (Frazier and Westhoff, 1978). In the milk industry, the lactic bacteria are chosen on the basis of their activity, that is their ability to produce more or less, large quantities of lactic acid in a given time interval and the fact that they are non-pathogens. Their ability to produce lactic acid varies depending on the strain and conditions for growth. The two principal bacterial species, which intervene in the process of lactic fermentation, belong to the group *Streptococcus* and the *Lactobacillus* and are further divided into subgroups. These lactic bacteria are often exigent for their nitrogenous and vitamin nutrients. They are auxotrophic for certain vitamins, amino acids, peptides, and at times purines and pyrimidines (Bracquart and Lorient, 1977; Olmos-Dichara, *et al.*, 1997; Amrane and Prigent, 1998), which limit their multiplication to a small number of favourable media. The presence of these free growth factors in the medium will enhance the multiplication of these bacteria. One of the natural medium in which the lactic bacteria grow is milk. Milk contains a complete range of nutritive elements (glucose, lactose, natural amino acids, minerals and vitamins) that are essential to support growth of lactic bacteria.

The milk industries are often faced with the problem of variation of milk composition. This may arise as a result of adulteration of the milk by practices external to the milk either by accident or design, by the addition of extraneous water and chemical substances (detergents, preservatives, disinfectants, feeds etc) as well as adulterations entering the milk through the cow itself (colostrums, blood, drugs etc) (Early, 1998). Such chemical substances often have bactericidal/bacteriostatic effect on starter cultures. Also, crude milk contains certain quantities of bacteria (with stimulating effects) and several growth factors naturally pre-existing in it. During processing, sterilization or pasteurisation temperatures remove or destroy some of these thermolabile growth factors, thereby depleting the milk of nutritive substances. Milk with high levels of adulterants or low amount of growth factors would sluggishly support the performances of starter cultures (Early, 1998). This constitutes a defect called "Lazy" milk in dairy industry. There is

therefore the need to supplement such milk with more micronutrients to enhance the performances of starter cultures.

Yeast extract is a substance that is complex with high nutritive value. Besides the vitamin B group, it is rich in peptides and amino acids (Smith *et al* 1975). Such a substance could be used to augment nutrients in milk.

It is therefore the objective of this study to determine the effect of supplementation of milk with yeast extracts on the growth and acidification of lactic bacteria.

MATERIALS AND METHODS

Materials

The Industrial strains of *Streptococcus lactis*; *Streptococcus cremoris*, *Lactobacillus casei*; and *Lactobacillus acidophilus* were provided by Societe Callixte-France; Three yeast extracts N-normal yeast extract, M-yeast extract supplemented with several vitamins and O – yeast extract supplemented with group B vitamins, were provided by Lacto Labo-France. Growth medium was Elliker medium (DIFCO 097401) for cell count and acidity determination. Culture medium for the four strains of lactic bacteria was Eugon broth medium (Biomerieux 51041). Pasteurised skimmed milk was employed in the investigation while milk without yeast extract was used as control.

Microbiological Analysis

The tests were carried out by aseptically inoculating the pasteurised skimmed milk containing the three different yeast extracts at four different concentrations (0.05%, 0.2%, 0.5% and 1%) with 20% test culture organisms (2 ml of culture in Eugon broth) in 10 ml of milk. The test samples were incubated at 37⁰C; cell counts and total titratable acidity were estimated at 0, 3 and 6 hours intervals.

Cell Count

Total viable aerobic plate count (APC) was carried out using pour plate method as described by Harrigan and McCance (1979).

Triplicate 1 ml test samples were aseptically transferred (at appropriate intervals) into 9 ml sterile distilled water as dilute in sterilized test tubes. This was followed by appropriate serial dilutions of the test samples. 1 ml of each dilution was aseptically transferred into sterile petri dishes and was overlaid with molten Elliker medium, which had been cooled down to 45⁰C. The plates were gently swirled and allowed to solidify. The plates were incubated in inverted positions at 37⁰C for a period of 24 hours after which cell counts were determined by multiplying the number of colonies counted by the dilution factors. The results were expressed as colony forming units (cfu/ml) per ml of sample.

Total titratable acidity (TTA)

Total titratable acidity was determined by method described by AOAC (1990).

Triplicate 10ml test samples were titrated against 0.1N NaOH using phenolphthalein as indicator. Percentage total titratable acidity was calculated as % lactic acid.

Statistical Analysis

Analysis of variance (ANOVA) was performed on the total cell count (cfu/mL) and total titratable acidity (TTA) using the methods of Steel and Torrie (1980). The least significant differences (LSD) were calculated using Tukey's test multiple comparisons (Steel and Torrie, 1980).

RESULTS AND DISCUSSIONS

Table 1 shows influence of various yeast extracts on the growth of *S. lact*

Table 1: Influence of Various Yeast Extracts on growth of *S. lactis* in skimmed milk

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | | |
|-------------------------------------|------------|-----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|
| Milk | Time (hrs) | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ |
| Samples | cfu/ml | | | | | | | | | | | | |
| MM | | 1.00x10 ^{4b} ±0.54 | 2.5x10 ^{8b} ±0.06 | 4.0x10 ^{8c} ±0.84 | 1.0x10 ^{4A} ±0.12 | 3.0x10 ^{8C} ±0.81 | 4.5x10 ^{8C} ±0.41 | 1.0x10 ^{4b} ±0.07 | 3.5x10 ^{8c} ±0.61 | 4.5x10 ^{8b} ±0.51 | 1.0x10 ^{4b} ±0.04 | 4.0x10 ^{8b} ±0.07 | 5.0x10 ^{8c} ±0.12 |
| MN | | 1.20x10 ^{4b} ±0.03 | 3.5x10 ^{8b} ±0.02 | 4.5x10 ^{8b} ±0.12 | 1.2x10 ⁴ ±0.01 | 4.0x10 ^{8b} ±0.09 | 5.0x10 ^{8c} ±0.22 | 1.2x10 ^{4b} ±0.00 | 5.0x10 ^{8c} ±0.08 | 5.8x10 ^{8c} ±0.01 | 1.2x10 ^{4b} ±0.11 | 6.0x10 ^{8b} ±0.01 | 1.5x10 ^{9b} ±0.10 |
| MO | | 1.10x10 ^{4b} ±0.01 | 3.5x10 ^{8b} ±0.00 | 4.5x10 ^{8b} ±0.03 | 1.1x10 ^{4b} ±0.03 | 4.5x10 ^{8b} ±0.31 | 6.0x10 ^{8b} ±0.01 | 1.1x10 ^{4b} ±0.01 | 5.0x10 ^{8b} ±0.11 | 5.7x10 ^{8b} ±0.03 | 1.1x10 ^{4b} ±0.00 | 6.0x10 ^{8b} ±0.23 | 8.0x10 ^{8b} ±0.21 |
| M | | 1.0x10 ^{4b} ±0.00 | 6.0x10 ^{4c} ±0.17 | 2.0x10 ^{5b} ±0.01 | | | | | | | | | |
| LSD | | 2.3x10 ⁴ | 1.2x10 ⁴ | 3.9x10 ⁷ | 1x10 ² | 4.2x10 ⁷ | 5.0x10 ⁷ | 2.1x10 ² | 4.7x10 ⁸ | 2.3x10 ⁸ | 1.17x10 ⁸ | 1.85x10 ⁸ | |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN= Milk +Extract N

MO= Milk +Extract O

M = Milkon

The growth increased with increase in yeast extract concentrations. N-yeast extract showed greater growth stimulation, followed by O-yeast extract.

Table 2. Shows influence of various yeast extracts on the growth of *S.cremoris*.**Table 2: Influence of Various Yeast Extracts on growth of *S.cremoris* in skimmed milk**

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | | |
|-------------------------------------|------------|--------------------------|---------------------------|----------------------------|-------------------------|--------------------------|---------------------------|-------------------------|------------------------|----------------------------|-------------------------|--------------------------|---------------------|
| Milk Samples | Time (hrs) | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ |
| | | cfu/ml | | | | | | | | | | | |
| MM | | 2x10 ^{4a} ±0.0 | 5.5x10 ^{6b} ±0.7 | 2.0x10 ^{7c} ±0.01 | 1.2x10 ^{4a} ±0 | 7.0x10 ^{7b} ±0. | 2.0x10 ^{8c} ±0.0 | 1.2x10 ^{4a} ±0 | 1.0x10 ^{8b} ± | 2.5x10 ^{8c} ±0.01 | 1.2x10 ^{4a} ±0 | 1.5x10 ^{8c} ± | 3.0x10 ⁸ |
| | | 1 | | | .08 | 03 | 1 | 00 | 0.03 | | .01 | 0.10 | 0.01 |
| MN | | 1.1x10 ^{4a} ±0. | 1.5x10 ^{6b} ±0.0 | 2.5x10 ^{6b} ±0.05 | 1.1x10 ^{4a} ±0 | 2.0x10 ^{8a} ±0. | 3.0x10 ^{8a} ±0.0 | 1.1x10 ^{4a} ±0 | 2.5x10 ^{8a} ± | 5.5x10 ^{8a} ±0.05 | 1.1x10 ^{4a} ±0 | 3.0x10 ^{8a} ±0. | 6.0x10 ⁸ |
| | | 1 | 3 | | .01 | 40 | 4 | 04 | 0.10 | | 00 | 1 | 0.7 |
| | | | | | | | | | | | | | 0 |
| MO | | 3x10 ^{4a} ±0.0 | 1.5x10 ^{6a} ±0.0 | 3.0x10 ^{6a} ±0.00 | 1.3x10 ^{4a} ±0 | 1.9x10 ^{8a} ±0. | 2.5x10 ^{8b} ±0.0 | 1.3x10 ^{4a} ±0 | 2.3x10 ^{8a} ± | 5.0x10 ^{8b} ±0.10 | 1.3x10 ^{4a} ±0 | 2.0x10 ^{8b} ±0. | 6.0x10 ⁸ |
| | | 3 | | | .01 | 77 | 5 | 11 | 0.35 | | 04 | 0 | 0.13 |
| M | | 1.5x10 ^{4a} ±0 | 1.5x10 ^{6c} ±0. | 1.2x10 ^{6d} ±0.16 | | | | | | | | | |
| | | .14 | 02 | | | | | | | | | | |
| LSD | | 1.05x10 ⁷ | 1.32x10 ⁶ | 1.17x10 ⁷ | 9x10 ⁷ | 1.25x10 ⁸ | 4.25x10 ⁸ | 1.01x10 ⁷ | 1.93x10 ⁷ | 2.21x10 ⁸ | 9.3x10 ⁷ | 4.1x10 ⁷ | 2.5x10 ⁷ |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN= Milk +Extract N

MO= Milk +Extract O

M = Milk only

Growth of this organism drastically increased with increase in yeast extract levels. O- yeast extract had greater influence on the growth rate of *S. cremoris*, followed by N-yeast extract. M-yeast extract had least effect on the growth of this organism.

Table 3 shows influence of various yeast extracts on the growth of *L. casei*.

Table 3: Influence of Various Yeast Extracts on growth of *L. casei* in skimmed milk

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | | |
|-------------------------------------|-------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|---------------------------|------------------------|
| Milk Samples | Time (hrs) cfu/ml | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ |
| MM | | 1.1x10 ⁶ ±0.03 | 1.0x10 ⁶ ±0.18 | 2.5x10 ⁶ ±0.51 | 1.1x10 ⁶ ±0.22 | 1.5x10 ⁶ ±0.10 | 3.0x10 ⁶ ±0.01 | 1.1x10 ⁶ ±0.94 | 1.6x10 ⁶ ±0.05 | 3.5x10 ⁶ ±0.02 | 1.3x10 ⁶ ±0.16 | 2.0x10 ⁶ ±0.70 | 5.0x10 ⁶ ±0 |
| MN | | 1.0x10 ⁶ ±0.01 | 2.0x10 ⁶ ±0.05 | 3.0x10 ⁶ ±0.06 | 1.0x10 ⁶ ±0.09 | 2.5x10 ⁶ ±0.16 | 4.0x10 ⁶ ±0.00 | 1.0x10 ⁶ ±0.07 | 2.5x10 ⁶ ±0.13 | 4.5x10 ⁶ ±0.01 | 1.0x10 ⁶ ±0.02 | 3.0x10 ⁶ ±0.07 | 5.0x10 ⁶ ±0 |
| MO | | 1.2x10 ⁶ ±0.05 | 1.3x10 ⁶ ±0.02 | 2.5x10 ⁶ ±0.00 | 1.2x10 ⁶ ±0.13 | 3.0x10 ⁶ ±0.04 | 4.0x10 ⁶ ±0.01 | 1.2x10 ⁶ ±0.26 | 2.6x10 ⁶ ±0.05 | 3.0x10 ⁶ ±0.18 | 1.2x10 ⁶ ±0.00 | 2.6x10 ⁶ ±0.01 | 3.5x10 ⁶ ±0 |
| M | | 1.5x10 ⁶ ±0.06 | 2.0x10 ⁶ ±0.11 | 2.5x10 ⁶ ±0.11 | | | | | | | | | |
| LSD | | 8.5x10 ⁷ | 6.5x10 ⁷ | 3x10 ⁸ | 9x10 ⁷ | 5.5x10 ⁷ | 9.5x10 ⁷ | 8.21x10 ⁷ | 8.56x10 ⁷ | 9.91x10 ⁷ | 9.0x10 ⁷ | 2.5x10 ⁷ | 1.9x10 ⁷ |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM = Milk + Extract M

MN = Milk + Extract N

MO = Milk + Extract O

M = Milk only

0.5% of all the three yeast extracts showed no significant difference in the cell counts at 6 hours incubation. At 1% yeast concentration, M and N – yeast extracts had similar values (5.0×10^8 cfu/ml) at 6 hours, while 0-yeast extract had lower cell count – 3.5×10^5 cfu/ml. Nevertheless, increased yeast extract levels resulted in increase in cell count of all test samples.

Table 4 shows influence of various yeast extracts on the growth of *L. acidophilus*.

Table 4: Influence of Various Yeast Extracts on growth of *L.acido* in skimmed milk

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | | |
|-------------------------------------|-----------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|--|
| Milk Sampler | Time (hrs) of/incl | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | |
| MM | | $1.0 \times 10^4 \pm 0.92$ | $1.5 \times 10^5 \pm 0.50$ | $3.0 \times 10^6 \pm 0.08$ | $1.0 \times 10^4 \pm 0.00$ | $2.0 \times 10^6 \pm 0.01$ | $3.5 \times 10^8 \pm 0.09$ | $1.0 \times 10^4 \pm 0.02$ | $2.3 \times 10^6 \pm 0.01$ | $4.0 \times 10^8 \pm 0.00$ | $1.0 \times 10^4 \pm 0.01$ | $2.5 \times 10^6 \pm 0.19$ | |
| MN | | $1.6 \times 10^4 \pm 1.61$ | $1.0 \times 10^5 \pm 0.02$ | $3.0 \times 10^6 \pm 0.11$ | $1.0 \times 10^4 \pm 0.06$ | $1.9 \times 10^6 \pm 0.09$ | $3.2 \times 10^8 \pm 0.06$ | $1.0 \times 10^4 \pm 0.01$ | $2.5 \times 10^6 \pm 0.43$ | $4.8 \times 10^8 \pm 0.82$ | $1.0 \times 10^4 \pm 0.01$ | $1.5 \times 10^6 \pm 0.02$ | |
| MO | | $1.3 \times 10^4 \pm 0.06$ | $2.0 \times 10^5 \pm 0.10$ | $3.0 \times 10^6 \pm 0.02$ | $1.3 \times 10^4 \pm 0.17$ | $3.0 \times 10^5 \pm 0.74$ | $4.5 \times 10^6 \pm 0.01$ | $1.8 \times 10^4 \pm 0.07$ | $3.0 \times 10^5 \pm 0.14$ | $5.0 \times 10^6 \pm 0.01$ | $1.5 \times 10^4 \pm 0.10$ | $3.7 \times 10^5 \pm 0.74$ | |
| M | | $1.5 \times 10^4 \pm 0.12$ | $2.5 \times 10^4 \pm 0.01$ | $2.7 \times 10^6 \pm 0.14$ | | | | | | | | | |
| LSD | | 1.15×10^2 | 4.5×10^7 | 2.5×10^8 | 9.1×10^2 | 8.57×10^8 | 6.5×10^7 | 7.5×10^3 | 4.3×10^7 | 1.9×10^7 | 6.2×10^2 | 9.7×10^7 | |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN= Milk +Extract N

MO= Milk +Extract O

M = Milk only

The cell counts increased with increase in yeast extract levels for all the different yeast extracts. O – yeast extract had greater effect on *L. acidophilus* followed by M-yeast extract. However at 0.05% yeast extract concentration, there was no significant difference in the cell count among the three yeast extracts at 6 hours incubation, though all values were superior to the value of the control (2.7×10^4 cfu/ml).

Table 5 shows the influence of various yeast extracts on the acidification of *S. lactis*.

Table 5: Influence of Various Yeast Extracts on acidification of *S. lactis* in skimmed milk

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | |
|-------------------------------------|-----------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|
| Milk Samples | Time (hrs) % lactic acid | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ |
| MM | | 10.00 ^a ±0.06 | 12.50 ^b ±0.04 | 21.25 ^b ±0.01 | 10.10 ^a ±0.00 | 13.75 ^b ±0.01 | 22.50 ^b ±0.01 | 10.21 ^a ±0.01 | 15.00 ^b ±0.03 | 24.50 ^b ±0.0 | 10.25 ^a ±0.03 | 16.25 ^b ±0.00 |
| MN | | 10.00 ^a ±0.00 | 12.50 ^b ±0.09 | 21.75 ^b ±0.05 | 10.12 ^a ±0.00 | 12.25 ^b ±0.04 | 24.25 ^b ±0.01 | 10.20 ^a ±0.07 | 13.00 ^b ±0.12 | 25.00 ^b ±0.0 | 10.25 ^a ±0.01 | 13.75 ^b ±0.02 |
| MO | | 10.05 ^a ±0.01 | 14.03 ^b ±0.05 | 22.50 ^b ±0.30 | 10.12 ^a ±0.06 | 11.50 ^b ±0.01 | 22.50 ^b ±0.16 | 10.10 ^a ±0.03 | 15.25 ^b ±0.00 | 22.50 ^b ±0.01 | 10.15 ^a ±0.01 | 16.00 ^b ±0.21 |
| M | | 10.00 ^a ±0.02 | 11.50 ^a ±0.00 | 13.75 ^a ±0.23 | | | | | | | | |
| LSD | | 0.12 | 0.50 | 0.32 | 0.09 | 0.63 | 1.13 | 0.12 | 1.94 | 0.33 | 0.95 | 1.71 |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN= Milk +Extract N

MO= Milk +Extract O

M = Milk only

The total titratable acidity produced by this lactic bacterium increased with increased levels of the different yeast extracts. N – yeast extract showed greater effect, followed by M-yeast extract. However at 1% yeast extract concentration there was no significance differences in the acidity between M and O yeast extracts at 6 hours incubation.

Table 6 shows influence of various yeast extracts on the acidification of *S. cremoris*.

Table 6: Influence of Various Yeast Extracts on acidification of *S. cremoris* in skimmed milk

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | | |
|-------------------------------------|-----------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|
| Milk Samples | Time (hrs) % lactic acid | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ |
| MM | | 10.00 ^a ±0.13 | 10.50 ^b ±0.03 | 10.60 ^c ±0.22 | 10.12 ^a ±0.01 | 10.60 ^a ±0.27 | 11.25 ^b ±0.00 | 10.15 ^a ±0.02 | 10.60 ^a ±0.07 | 11.25 ^b ±0.04 | 10.25 ^a ±0.00 | 11.25 ^a ±0.01 | 13.75 ^b ±0.16 |
| MN | | 10.00 ^a ±0.05 | 10.25 ^b ±0.01 | 11.50 ^b ±0.01 | 10.11 ^a ±0.11 | 10.75 ^a ±0.16 | 11.25 ^a ±0.02 | 10.12 ^a ±0.01 | 10.80 ^b ±0.06 | 13.25 ^a ±0.51 | 10.15 ^a ±0.19 | 11.50 ^a ±0.11 | 16.25 ^a ±0.00 |
| MO | | 10.10 ^a ±0.00 | 10.75 ^b ±0.05 | 11.25 ^b ±0.00 | 10.12 ^a ±0.42 | 10.75 ^a ±0.42 | 11.25 ^a ±0.06 | 10.12 ^a ±0.05 | 10.30 ^a ±0.65 | 11.25 ^b ±0.08 | 10.16 ^a ±0.17 | 10.60 ^b ±0.23 | 13.25 ^a ±0.10 |
| M | | 10.00 ^a ±0.03 | 11.50 ^a ±0.18 | 12.60 ^a ±0.13 | | | | | | | | | |
| LSD | | 0.10 | 0.62 | 0.43 | 1.14 | 0.95 | 1.01 | 0.17 | 0.65 | 1.70 | 0.45 | 1.03 | 2.14 |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN= Milk +Extract N

MO= Milk +Extract O

M = Milk only

The trend of increase in acid production with increased levels of yeast extract supplementation was observed. At 1% level of yeast extracts, N-yeast extract exhibited greater acid stimulation, while there were no significant differences between the acidity values from M and O yeast extracts (13.75% lactic acid and 13.25% lactic acid respectively).

Table 7 shows influence of various yeast extracts on the acidification of *L. casei*.

Table 7: Influence of Various Yeast Extracts on acidification of *L. casei* in skimmed milk

| Concentration of yeast extract. (%) | | | | | | | | | | | | |
|-------------------------------------|-----------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|----------------|
| | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | |
| Milk Samples | Time (hrs) % lactic acid | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ |
| MM | | 10.00 ^a ±0.18 | 11.75 ^b ±0.28 | 15.00 ^c ±0.15 | 10.10 ^a ±0.06 | 12.25 ^b ±0.10 | 16.25 ^c ±0.12 | 10.16 ^a ±0.01 | 13.50 ^b ±0.00 | 17.50 ^c ±0.01 | 10.15 ^a ±0.29 | 13.2 |
| MN | | 10.10 ^a ±0.07 | 11.75 ^b ±0.16 | 16.25 ^c ±0.03 | 10.13 ^a ±0.00 | 11.50 ^b ±0.20 | 14.86 ^c ±0.62 | 10.14 ^a ±0.11 | 12.25 ^b ±0.11 | 17.86 ^c ±0.15 | 10.15 ^a ±0.06 | 13.75 |
| MO | | 10.15 ^a ±0.01 | 10.50 ^b ±0.23 | 14.67 ^c ±0.01 | 10.15 ^a ±0.07 | 10.50 ^b ±0.00 | 12.00 ^c ±0.04 | 10.16 ^a ±0.08 | 13.00 ^b ±0.05 | 16.25 ^c ±0.02 | 10.15 ^a ±0.10 | 13.50 |
| M | | 10.00 ^a ±0.00 | 12.50 ^b ±0.04 | 15.00 ^c ±0.29 | | | | | | | | |
| LSD | | 0.10 | 0.56 | 1.10 | 0.94 | 0.70 | 1.31 | 0.16 | 1.10 | 1.54 | 0.13 | 0.42 |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN= Milk +Extract N

MO= Milk +Extract O

M = Milk only

The results revealed that acid production increased with increase levels of all the three yeast extracts. At 0.5% yeast extract concentration, there was no significant difference in the acidity between M and N- yeast extracts at 6 hours incubation.

Igyor M. A (2005) Nig. J. Biotech. 16 (1) 40 – 54.

However at 1% extract concentration, M – yeast extract had superior values of (21.25% lactic acid) followed by N – yeast extract (19.25% lactic acid) while O–yeast extract had the least value 17.50% lactic acid.

Table 8 shows influence of various yeast extracts on the acidification of *L. acidophilus*.

Table 8: Influence of Various Yeast Extracts on acidification of *L. acidophilus* in skimmed milk

| Concentration of yeast extract. (%) | | 0.05 | | | 0.2 | | | 0.5 | | | 1.0 | |
|-------------------------------------|-----------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|--------------------------|----------------|
| Milk Samples | Time (hrs) % lactic acid | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ | t ₂ | t ₀ | t ₁ |
| MM | | 10.00 ^a ±0.01 | 11.25 ^b ±0.55 | 15.00 ^c ±0.23 | 10.15 ^a ±0.19 | 11.25 ^b ±0.30 | 15.50 ^b ±0.07 | 10.15 ^a ±0.17 | 10.75 ^a ±0.01 | 17.00 ^c ±0.95 | 10.15 ^a ±0.01 | 14.50 |
| MN | | 10.00 ^a ±0.11 | 10.25 ^b ±0.00 | 15.00 ^c ±0.04 | 10.15 ^a ±0.01 | 11.75 ^b ±0.43 | 16.25 ^b ±0.17 | 10.16 ^a ±0.02 | 10.25 ^b ±0.00 | 17.50 ^c ±0.06 | 10.16 ^a ±0.01 | 14.25 |
| MO | | 10.10 ^a ±0.37 | 10.00 ^b ±0.46 | 11.10 ^c ±0.02 | 10.15 ^a ±0.01 | 11.10 ^a ±0.07 | 14.50 ^c ±0.01 | 10.15 ^a ±0.12 | 11.50 ^b ±0.00 | 15.00 ^b ±0.11 | 10.20 ^a ±0.04 | 12.50 |
| M | | 10.00 ^a ±0.11 | 11.50 ^b ±0.06 | 13.75 ^b ±0.06 | | | | | | | | |
| LSD | | 0.11 | 0.63 | 0.12 | 0.91 | 0.73 | 0.66 | 0.17 | 1.10 | 1.97 | 0.15 | 1.53 |

Means not followed by the same superscript in the same column are significantly different from each other at 5% probability level.

± = Standard deviation of triplicate analysis

LSD = Least Significant difference

MM= Milk +Extract M

MN = Milk +Extract N

MO = Milk +Extract O

M = Milk only

Acidification increased with increased concentration of yeast extracts. While there was no significant difference in acidification between M and N yeast extracts at 0.5% concentration at 6 hours incubation, yeast extract had greater acidity than M-yeast extract at 6 hours incubation at 1% yeast extract level. N-yeast extract had higher acid values 16.25% lactic acid than M - yeast (15.50% lactic acid) at 0.2% concentration while O-yeast extract had the least value 14.50% lactic acid.

The results above demonstrated that different yeast extracts with their various concentrations exerted various influences on the growth and acid production of these lactic bacteria. Growth of the organisms and the acidification increased proportionally with levels of yeast extract supplementation. These results agreed with earlier works of Braequart *et al* (1978); Olmos – Diechara, *et al*, (1997); Amrane and Prigent, (1998) who showed the exigency of lactic bacteria for certain amino acid and vitamins. These results also agreed with Oliveira, *et al* (2001) who showed that yeast extract supplementation improved the performance of lactic bacteria in milk products.

The *Streptococci* and *Lactobacilli* are often used as mixed starter cultures in dairy industry. Apart from their health and nutritional benefits (Gilliland, 1990), they also produce various compounds such as organic acids, diacetyl, hydrogen peroxide and bacteriocin or bactericidal proteins during lactic fermentations (Lindgård and Dobrogosz, 1990). The antimicrobial properties of lactobacilli are of specific interest in developing strongly competitive starter cultures for food fermentation. Lactobacilli exert strong antagonistic activity against microorganisms including food spoilage organisms and pathogens by production of other inhibitory substances such as bacteriocins (Brink *et al* 1994). Ogunbanwo *et al* (2003) demonstrated that yeast extract supplementation produced large quantities of bacteriocin from lactobacilli without impairing the qualities of the products. Yeast extract could therefore be employed to solve fermentation problems in the milk industry without adversely affecting the quality of the product.

CONCLUSION

The stimulating effect of yeast extract on the growth and acid production of lactic bacteria could be exploited to overcome problems of “Lazy” milk, due to inhibition and proliferation of undesirable bacteria encountered frequently in the milk industry.

REFERENCES

- Amrane, A. and Prigent, Y (1998). Influence of yeast extract in the growth of *Lactobacillus helveticus*. World J. of Microbiology and Biotechnology. **14**(4): 529-534.
- AOAC (1990) Official Methods of Analysis. 15th edn. Association of Official Analytical Chemists, Washington D.C.
- Bautista, E.S.; Dahiya, R.S.; and Speck, M.L. (1966). Identification of compounds causing symbiotic growth of *Streptococcus thermophilus* and *Lactobacillus bulgaricus* in milk. J. Dairy Res. **33**: 299-307.
- Bracquart, P. and Lorient, D. (1977). Effet des acides amines sur la croissance de *Streptococcus thermophilus*, Michwissenschaft. **32**: 221-224.
- Bracquart P., Lorient, D and Alais, C (1978) Effet des acides amines sur la croissance de *Streptococcus thermophilus*, Michwissenschaft. **33**: 341-344.
- Brink ten, B; Minekns, M; Vander Vossen, J.M.B.M; Leer, R.J and Huisin't Veld,J.H (1994).Antimicrobial activity of lactobacilli. J.Appl. Bacteriol. **77**: 140-148
- Early,R. (1998) Liquid Milk and Cream. In: The technology of dairy products. ed. R. Early. Blackie Academic & Professionals. Chapman & Hall. London. Pp.1-49.
- Frazier, W.C. and Westhof, D.C. (1998). Contamination, Preservation and Storage of Milk and Milk Products. In: Food Microbiology. 3rd ed. Edited by W.C. Frazier and D.C.Westhof. Tata McGraw – Hill Publishing Company Limited, New Delhi. Pp. 278 – 305.
- Gilliland, S.E. (1990). Health and Nutritional benefits from lactic acid bacteria. FEMS Microbial. Rev. **87**:175 – 178.
- Graciela,M; Vignolo, M; de Kairuz, M.N; Aida de Ruiz, H.A.A.P; and Oliver, G. (1995). Influence of growth conditions on the production of lactocin 705, a bacteriocin produced by *Lactobacillus casei* CRL 705. J. Appl. Bacteriol. **78**: 5-10.
- Harrigan, W.F. and McCance, M.E (1979). Laboratory Methods in Food and Dairy Microbiology. Academic Press Incorporated London. Pp. 157-265.
- Kothari, S.L.; Nambudripad, V.K.N. (1973). Isolation and identification of stimulatory growth substance of cheese cultures. J. Dairy Science **56**: 423-428.

Igyor M. A (2005). Nig. J. Biotech. 16 (1) 40 – 54.

Lindgren, S.W. and Dobrogosz, W.J. (1990). Antagonistic Activities of lactic acid bacteria in Food and Fermentation. FEMS Microbiol. Rev. **59**: 1416-1424.

Ogunbanwo, S.T., Sanni, A.I and Onilude, A.A. (2003). Influence of Cultural conditions on the production of bacteriocin by *Lactobacillus brevis* OGI. African Journal of Biotechnology. 179-184.

Olmos-Dichara, A., Ampe, F., Uribelarrea, J.L; Parcilleux, A; and Goma, G. (1997). Growth and Lactic acid production by *Lactobacillus casei ssp. rhamnosus* in batch and membrane bioreactors. Influence of yeast extract and tryptone enrichment. Biotechnology Letters. **19**: 709-714

Oliveira, M.N., Sodini, I. Remeuf, F. and Corrieu, G. (2001). Effect of Milk Supplementation and culture composition on acidification, textural properties and microbiological stability of fermented milks containing probiotic bacteria. International Dairy Journal **11**:9: 946

Smith, J.S; Hiller, A.J. and Lees, G.S. (1975). The Nature of the Stimulation of the growth of *Streptococcus lactis* by yeast extract. J.Dairy Res. **42**: 123-138.

Steel, R.G.D and Torrie, J.H. (1980). Principles and Procedures of Statistics. McGraw- Hill. New York. 168 – 179.