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Assessment of Yield, Variability, Heritability and Genetic Advance in Ten Genotypes of Groundnut (*Arachis hypogaea)* **in South-East Nigeria**

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Abstract

Ten genotypes of Groundnut (*A. hypogaea*) were assessed for yield and yield components in a randomized complete block design with three replicates at the Research Farm of the Michael Okpara University of Agriculture, Umudike in 2018 and 2019. In each year, the experiment consisted of 30 plots, each measuring 1m by 1m with a distance of 1m separating the plots and blocks. Seeds were planted at intra and inter-row spacing of 10.5 m to give a population of $40,000$ plants⁻¹. Data were collected on the number of leaves plant⁻¹, number of branches plant⁻¹, length of branches plant⁻¹, days to flowering plant⁻¹, number of pods plant⁻¹, the weight of pods plant⁻¹, number of seeds plant⁻¹, the weight of seeds plant⁻¹, 100 seed weight, pod yield⁻¹ and seed yield⁻¹. Analysis of variance showed that the genotypes were significantly different (P<0.01) in all the characters studied. In both years the number of leaves plant¹, number of seeds plant¹, 100 seed weight, the weight of seeds plant¹ and pod yield⁻¹ were positive and highly significant ($p<0.01$) with seed yield⁻¹. Enormous variability existed among the genotypes of *A. hypogaea* as shown by the genetic component analysis. Genotypes with a high genotypic coefficient of variation, a high broad sense heritability estimates and a high genetic advance for the number of leaves plant⁻¹, the number of seeds plant⁻¹, 100 seed weight, the weight of seedplant⁻¹, pod yield⁻¹ and seed yield⁻¹ could be selected for improvement in seed yield of *A. hypogaea.* High genetic gain (faster progress) could be expected from selection based on the number of leaves plant¹, the number of branches plant¹ and the weight of seeds plant⁻¹ because they are predominantly under the control of addictive genes. *Keywords: Yield, Variability, Heritability, Genetic advance, Groundnut*

Introduction

Groundnut (*Arachis hypogaea* L.) is an annual herb in the Fabaceae family (Batioli *et al*. 2011). It is one of the most important oilseed crops grown in semi-arid and subtropical areas of the world (Ibrahim *et al*. 2022). According to Hamakareem *et al*., (2016), groundnut was taken across the Pacific to the Philippines by the Spaniards before spreading to Asia. The Portuguese imported it into West Africa (Waele and Swanevelders, 2011. The genetic diversity of the genes is classified into four gene pools: primary gene pool consisting of *A. hypogaea* and *A. monticola* secondary consisting of diploid species from section *Arachis* that are crosscompatible with *A. hypogaea*, tertiary consisting of species of section procumbent that are weakly-cross compactable with *A hyprogaea* and the fourth gene pool consisting of the remaining wild species classified into seven other sections (Pasupuleti *et al*. 2013). The wild species are used as forage (Waele and Swanevelder,

2011). A. *hypogaea* is a self-pollinating crop with cleistogamous flowers, natural hybridization can occur to a small extent where bees' activity is high. Flowering begins 17-35 days after seedling emergence depending on the cultivar and environmental conditions. Flowers, simple or compound are borne in the axils of leaves and never at the same node as vegetable branches. One or more flowers may be present at a node (Seabra *et al*. 2019). The stigma becomes receptive to pollen about 24 hours before anthesis and remains so for about 12 hours more, and the dehiscence of anthers takes place 2-3 hours before the opening of the flowers in the morning. Fertilization occurs about 6 hours after pollination (Yusuf *et al*. 2017).

Depending on the prevailing temperatures, the peg or gynophore carrying the ovary and fertilized ovule on its tips appears in 6-10 days and grows to enter the soil (Positive geotropic) where it develops into pods. The tip

orients itself horizontally away from tap roots; (Yadlapalli; 2014), Yol *et al*., (2018) reported that groundnut is derived from the Greek word arachis (legume) and hypogaea meaning "below ground" referring to the formation of pods in the soil. Shalini *et al*., (2016) reported that groundnut is valued as a rich source of energy contributed by oil (48-50%) and protein (25-28%) in the kernels. They provide 564Kcal of energy from 100g of kernels. Groundnut oil is an excellent cooking medium because of its high smoking point (Ibrahim, 2021, Ibrahim, *et al,* 2022). Bodena (2018) reported that groundnut merge fix as much as 190kg of nitrogen per hectare. Groundnut is not produced in commercial quantity in the southeast even with its enormous potential that could favour its production. Identification of genotypes with high seed yield capacities, determination of the relationship between seed yield and yield components and the yield components that influenced seed yield¹ most, calculation of broad sense heritability, genetic advance and genetic gain to be able to predict breeding program for seed yield¹ improvement are the objectives of this research work.

Materials and Methods

Ten groundnut genotypes which included, RRB, SUMNUT 10, SUMNUT 20, RMP 91S (Small seed), RMP91 (big seed), JL12, JL24, ICGV 89754 and ICGV 15-87281 were evaluated for growth, pod and seed yieldha¹ in 2018 and 2019 cropping seasons. The experiment was a randomized complete block design with the genotypes as treatment, replicated 3 times and was carried out at the Research Farm of Michael Okpara University of Agriculture, Umudike (Longitude 07° $33^{1}E$, latitude 05^{0} $29^{1}N$ and altitude 122m) with a temperature of 26° C. The seeds were planted at a spacing of 50cm by 50cm with a distance of 1m separating the plots and blocks. Nine stands per plot and ten plots per block, on a land area of 19m by 5m. Weeding was done manually using the hoe and hand-pulling methods as soon as they appeared. Data were collected on the number of leaves plant¹, number of branches plant¹, days to 50% flowering, number of pods plant¹, the weights of pods plant⁻¹, the number of seeds plant⁻¹, the weight of seeds plant¹, 100 seed weight, seed yield h ectare⁻¹ and pod yield hectare⁻¹. Data were analyzed by one-way analysis of variance and significant means separated with the least significant differences (Snedeco and Cochran 1989, Obi, 2001). The gross variability was partitioned into genetic and non-genetic components and phenotypic, genotypic and error variances were estimated using the method of Wrikke and Weber (1986) $\sigma^2 P = \frac{MSG}{I}$ $\frac{SG}{r}$, $\sigma^2 G = \frac{MSG}{r}$ $\frac{SG}{r} - \frac{MSE}{r}$ $rac{\text{SSE}}{\text{r}}$ $\sigma^2 E = \frac{\text{MSE}}{\text{r}}$ $\frac{1}{r}$,

MSG, MSE and r are the mean square *genotypes*, means square error and number of replications while $\sigma^2 P$, $\sigma^2 G$

and σ^2 E are phenotypic, genotypic and error variances respectively. The phenotypic, genotypic and environmental components of variation were estimated according to Singh (2001).

$$
PCV = \frac{\sigma P \times 100}{Mean}, \, GCV = \frac{\sigma G \times 100}{Mean}, \, ECV = \frac{\sigma E \times 100}{Mean}.
$$

PCV, GCV and ECV are phenotypic, genotypic and environmental coefficients of variation respectively. Broad sense heritability was estimated according to Allard (1991). Genetic advance (GA) was calculated according to Johnson *et al*(1995),

$$
G.A = \frac{\sigma^2 G}{\sqrt{\sigma P}} \times K
$$

Where K=2.063(selection differential at 5%), $\sigma^2 G$ = genetic variance.

 \sqrt{QP} = square root of phenotypic variances, Genetic gain (GG) was determined from genetic advance expressed as a percentage of the population mean.

Results and Discussion

The physical and chemical characteristics of the soil experimental site were taken and analyzed before planting. The results are given in Table 1. Soil textural class was sandy loam. Slight. The soil was relatively suitable for the cultivation of *A. hypogaea*, (Baughman *et al*. 2015). The soil was analyzed at the National Root Crop Research Institute Umudike, Soil Science Laboratory. The soil was slightly acidic with pH in $H₂O$ of 5.8 in 2018 and 6.1 in 2019. The rise in organic carbon, total nitrogen and available phosphorus in 2019 was due to the residual effects of the application of organic manure, which released nutrients slowly. Table 2 shows the agro-metrological data of the experimental site in 2018 and 2019. The mean maximum temperatures were 31.75 and 32.22, while the mean rainfall (amount) was 193.56 and 171.76 for 2018 and 2019 respectively. The minimum temperatures in both years are also given. The weather conditions were good enough for the production of *A. hypogaea* in Umudike. The results of the analysis of variance are presented in Tables 3 and 4 which showed that the ten genotypes of *A. hypogaea* differed greatly in their vegetative, the reproductive characteristics as well as pod and seed yield hectare⁻¹. JL 24, ICGV 89754, JL 12, Rmp-91-B and ICGV-15-87281 vegetative characters (number of leaves and number of branches plant¹) performed best in both 2018 and 2019. For reproductive characters (number of seed/pod, weight of seeds/pod and weight of pods) these genotypes recorded superior performance which translated into the height of pod and seed yield $ha⁻¹$ in both years. The genotypes, JL 24, ICGV-89754, JL 12, Rmp-91-B and ICGV-15-87281 could further be reviewed for possible release in Umudike. High variance ratios in Table 5 suggest high variation in the 11 characters of the 10 genotypes of *A. hypogaea*. The correlation matrix showed that the number of leaves plant⁻¹, number of branches plant⁻¹, number of pods plant⁻¹ $\overline{1}$, number of seeds plant¹, weight of seeds plant¹ and pod

yield hectare¹ associated positively and highly significantly ($P<0.01$) with seed yield hectare⁻¹ in 2018 and 2019, (Table 5). This suggests that improvement in the performance of these characters will lead to an increase in seed yield hectare^{-1}. From Table 6, the weight of seed plant⁻¹ (0.641 in 2018, 0.599 in 2019) was the single reproductive factor that directly contributed most to seed yield ha⁻¹. The weight of pods plant⁻¹ (0.406 in 2018, 0.375 in 2019) was the second individual reproductive factor that directly influenced seed yield $ha⁻¹$. Pod yield $ha⁻¹$ was the third most important single factor that directly influenced seed yield. This implies that Agronomic practices and other factors that could improve the performance of the weight of seeds plant ha $\frac{1}{1}$, weight of pod plant¹ and pod yield ha⁻¹ will lead to an increase in seed yield $ha⁻¹$ of these genotypes. The effectiveness of selection in any crop depends on the extent and nature of phenotypic and genotypic variability present in different agronomic traits of the population (Yeshiwas, *et al*. 2017). Generally, genetic parameters including genotypic coefficient of variation, heritability and genetic advance are prerequisites for genetic improvement of crops. A high genotypic coefficient of variation indicates the availability of high variation. The very low value of variation indicates that selection will not be effective for such traits because of narrow genetic variability (Dikshit, *et al*. 2015). From Table 7, very high variance ratios were observed among most of the characters studied.

Table 8 shows the means, estimates of phenotypic and genotypic variances, genetic advance and genetic gain in 2018 and 2019. Estimates of Phenotypic and Genotypic coefficients of variation were high in length of branches plant¹, number of leaves plant¹, number of branches plants¹, number of pods plants¹, weight of pods plants⁻¹, weight of seeds plants⁻¹, pod yield⁻¹and seed yield¹ High genotypic coefficient of variation indicates greater variability from these characters, thereby suggesting abundant scope from improvement through selection among these variations. Since phenotypic coefficients of variation were slightly higher than the genotypic coefficient of variation for all the characters studied, this indicates the presence of slight environmental influence to some degree in the phenotypic expression of the characters. Pandy *et al*. (2010) and Mulugeta, *et al*. (2012) also observed similar findings. The same trends were reported by Chinatu and Ukpaka (2016) in *Piper guineense*, Eze *et al*. (2016) In Taro, Chinatu, *et al*., (2016) in cucumber wide range of variation was observed from most of the characters under study. Shoba, *et al*. (2012) and Meta and Monpara (2010) reported high phenotypic and genotypic coefficient of variation for pod yield. Pradham and Patra (2011) reported low heritability estimates for days to 50% flowering and pod yield – *A. hypogaea*. Johnson et al. (1995) reported that heritability estimates together with genetic advances are more important than heritability alone to predict the effect of selecting the best individual genotypes. Genetic advances and genetic gains indicated moderate to high variability for length of branches, number of leaves, number of seeds/plants, weight of pods/plant, pod yield/ha and

seed yield/ha in both years.

Conclusion

This study revealed that the genotypes JL 24, ICGV-89754, JL12, Rmp-91-B and ICGV-15-87281 performed well in Umudike. Further studies could be carried out for it to be released to farmers to encourage groundnut production in the Southeast. The high phenotypic and genotypic coefficients of variation with very low environmental coefficients of variation observed among most of the seed yield/ha and yield components studied revealed enormous and exploitable variability which is mostly heritable. Moderate progress in the improvement of seed yield of *A. hypogaea* could be achieved by selecting genotypes with the higher number of leaves plant¹, longer and more branches plant', higher seed weight plant', higher pod weights $plan{t}^{-1}$, higher pod and seed yield hectare⁻¹ since they have high broad sense heritability genotypic coefficient of variation and genetic advance. High genetic gain (faster progress) could be expected from selection based on the number of leaves plant¹, number of branches $plant⁻¹$ and weight of seeds plant^{-1} because they are predominantly under the control of addictive genes.

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Physical Characteristics	2018	2019	Method of Analysis
Sand $(\%)$	75.6	74.2	Hydrometer (Jackson, 1962)
Silt $(\%)$	10.3	11.8	Hydrometer (Jackson, 1962)
Clay $(\%)$	14.0	13.9	Hydrometer (Jackson, 1962)
Textural class	Sandy-loam	Sandy-loam	
Chemical properties			
PH (H ₂ 0)	5.8	6.1	pH meter
Organic Carbon $(\%)$	1.12	1.3	Flame Photometric (Kjedahl, 1983)
Total Nitrogen (%)	0.097	0.15	Kjedahl method (Kjedahl, 1983)
Available $P(mgKg^{-1})$	33.4	22.7	Flame photometric (Kjeldahl, 1983)
Exchangeable $K(CmolKg^{-1})$	0.221	0.3	Oxidation (Kjeldahl, 1983)
Exchangeable $Ca(CmolKg^{-1})$	3.20	4.0	A. A. S. (Kjeldahl, 1983)
Exchangeable $Mg(CmolKg^{-1})$	0.96	1.1	A. A. S. (Kjeldahl, 1983)
Cation exchange capacity (CmolKg ⁻¹)	5.72	7.0	
Base Saturation	83.21	84.9	

Table 1: Physicochemical properties of the experimental site in 2018 and 2019

Source: National Root Crops Researches Institute Umudike, Abia State, Nigeria

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ean, r ange, ph enotypic, Genotypic and environ mental varian ces, Phenotypic, genotypic riation, Heritability, Genetic advance and Genetic Gain

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