ORIGINAL RESEARCH



Serological survey and risk factors associated with Toxoplasma gondii infection among HIV-infected pregnant women attending Abuja Tertiary Hospital, Nigeria

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Abstract

Background

Toxoplasmosis in pregnancy could induce miscarriage, congenital anomalies in foetuses and encephalitis in HIV-infected people. Hence, there is a need to determine the prevalence of toxoplasmosis in HIV-infected pregnant women to inform clinicians about the significance of maternal toxoplasmosis in antenatal care.

Aim

This study aimed to determine the seroprevalence of Toxoplasma gondii infection, associated CD4+ T-cell profile and sociodemographic risk factors among pregnant women with or without HIV infection attending the University of Abuja Teaching Hospital, Abuja, Nigeria.

Methods

This hospital-based cross-sectional study involved blood samples collected from 160 HIV-infected and 160 HIV-seronegative pregnant women. These samples were analysed for anti-T. gondii (IgG and IgM) and CD4+ T-cell count using ELISA and flow cytometry, respectively. Sociodemographic variables of participants were collected using structured questionnaires.

Results

The overall seroprevalence of anti-T. gondii IgG and IgM was 28.8% and 3.8%, respectively. The seroprevalence of anti-T. gondii IgG and IgM was 29.4% and 4.4%, respectively, among HIV-seropositive pregnant women and 28.1% and 3.1%, respectively, among HIV-seropositive pregnant women and 28.1% and 3.1%, respectively, among HIV-seropegative women. There was no significant association between the seroprevalence of anti-T. gondii-IgG and anti-T. gondii-IgM with age, gestational age, education level, parity or place of residence of HIV-infected pregnant women (P > 0.05). However, there was significant association between the seroprevalence of anti-T. gondii-IgG (P = 0.03) and anti-T. gondii-IgM (P = 0.01) with education level. CD4+ T-cell count varied significantly between HIV-infected and HIV-uninfected pregnant women (P = 0.035).

Conclusion

In this study, the seroprevalence of anti-T. gondii IgG and IgM did not differ in HIV-seropositive or HIV-seronegative pregnant women. However, women with primary T. gondii and HIV coinfection had lower CD4+ T-cell count than those with toxoplasmosis monoinfection.

Key Words

Toxoplasmosis, HIV coinfection, congenital anomaly, cellular immunity, sero-survey

Introduction

Toxoplasma gondii, the aetiological agent of toxoplasmosis, is a zoonotic parasite that has latently affected 33.8% of pregnant women worldwide in the last three decades¹. The ubiquitous obligate intracellular coccidian protozoan infects a wide variety of domesticated animals (such as cats and dogs), birds and humans¹. Clinically, toxoplasmosis is an opportunistic parasitic infection in immunocompromised and immunosuppressed people that has led to serious public health morbidities including physical and/or psychological sequelae for people living with HIV/AIDS². However, in the vast majority of immunocompetent people, *T. gondii* infection is latent, characterised by the persistence of the parasites in the brain, skeletal muscles and heart without causing clinical diseases³.

In chronically infected people who develop cell-mediated immunodeficiency, symptomatic toxoplasmosis is more likely to occur as a result of reactivated infection, especially due to CD4⁺ T lymphopenia⁴ below 100 cells/mm³. Consequently, toxoplasmosis among people living with HIV/AIDS mainly manifests as toxoplasma encephalitis⁵. In pregnant women, toxoplasmosis has been implicated in prenatal and congenital transmission, causing miscarriage or congenitally acquired disorders that primarily affect the central nervous system of neonates⁶. Nigeria has one of the highest HIV prevalences, with 1.4% among adults aged 15-49 years in 2019. HIV prevalence was highest among females aged between 35 and 39 years⁷. According to the recent Nigeria National AIDS Indicator Survey, Abuja was 15th out of 37 states and the capital for highest HIV prevalence. People living in the Abuja suburbs had relatively higher rates of HIV infection compared with those residing in the main Abuja city. Furthermore, higher prevalence was reported among women within the reproductive age group⁷. An overall toxoplasmosis prevalence of 31.5% was reported in Abuja⁸ with similar immunoglobulin M and G (IgM and

IgG) seropositivity, as previously reported in other Nigerian studies^{9,10}. When a pregnant woman contracts *T. gondii* infection in the first trimester which is allowed to be untreated, the risk of miscarriage is significantly high¹¹. However, in the third trimester, untreated *T. gondii* infection increases the risk of toxoplasma-induced congenital anomalies in neonates¹¹. There is a paucity of studies in Nigeria that focus on simultaneous investigation of toxoplasmosis and its impact of CD4⁺ T cellular immunity in pregnant women living with HIV/AIDS in comparison to those who are HIV seronegative. Hence, this study sought to determine the seroprevalence of anti-*T. gondii*, associated CD4⁺ T-cell profile and sociodemographic risk factors among pregnant women with or without HIV infection attending the University of Abuja Teaching Hospital (UATH) in Nigeria.

Methods Study design

This was a hospital-based cross-sectional study.

Study area

This study was conducted between 2 February 2017 and 28 January 2018 at the Department of Obstetrics and Gynecology and Immunology, UATH, Abuja. The hospital provides healthcare services to the inhabitants of Abuja and neighbouring states including Niger, Kaduna, Kogi and Nasarawa states. The hospital has an average of 3000 deliveries annually. The laboratory investigations were carried out at the Immunology Laboratory of UATH.

Study population

This included consecutively enrolled 160 HIV-seropositive and 160 HIV-seronegative pregnant women who attended antenatal clinics at UATH. All participants were retested for HIV serostatus using the World Health Organization (WHO) algorithm by Uni-Gold Recombigen HIV-1/2 (Trinity Biotech, Wicklow, Ireland) and DetermineTM (Alere, Auckland, New Zealand) proprietary reagents. There were no new HIV-seropositive cases and all the seropositive subjects were on antiretroviral therapy.

Sample size determination

The minimum sample size for this study was determined from the prevalence of 29.1% anti-toxoplasma IgG in pregnant women⁹ as 317. In total, 320 consenting participants were recruited.

Ethical considerations

The ethical approval for this study was obtained from the Research Ethics Committee of the University of Abuja Teaching Hospital (Approval Number: FCT/UATH/HREC/PR/416). Written informed consent was sought from all participants before enrolment into the study. For participants who were not literate and those who may not have understood the medium of communicating the study, translation was done using their local dialects before they consented by nodding their heads, signing the consent form or making a thumb-print on the signature area. All data were treated with utmost confidentiality. The protocols of this study were conducted in accordance with the Helsinki declaration of 1973 (revised in 2000)¹².

Sample collection and processing

The purpose of this work was explained to the subjects before their consent to participate was sought. Interviewer-administered, structured questionnaires were used as a study tool. The questions outlined in the data forms were explained to the subjects and then completed with the required information which included bio-demographic data. Five millilitres of blood was aseptically collected from each

participant via venepuncture.

Three millilitres were dispensed into a plain (no-anticoagulant) container, while the remaining 2 ml was dispensed into an EDTA anticoagulated bottle which was used for CD4⁺ T-cell counts (within 1 h of collection). The blood in the plain container was allowed to clot and retracted. The serum was harvested after separation from the red cells by centrifugation at 10,000 g for 10 min. The serum samples were appropriately labelled and refrigerated (2–8 °C) until ELISA analysis was done within 24 h of collection.

Laboratory analysis Toxoplasma gondii IgG and IgM ELISA

The samples were analysed for the presence of IgG/IgM class antibodies to *T. gondii* by ELISA using Toxo IgG/Toxo IgM ELISA kits (Fortress Diagnostics Limited, Antrim, UK) which are qualitative and quantitative immunoassays for the detection of human antibodies in serum or plasma directed against *T. gondii*. The interpretation of tests and results was carried out based on the manufacturer's instructions.

Flow cytometry of CD4⁺ T lymphocytes

Based on the manufacturer's instructions, the CD4⁺ cell counts in the whole blood were analysed using a PartecTM CyFlow Analyzer (Sysmex, Norderstedt, Germany) Model SL3. This device uses the principle of light scattering (based on dissimilarity in cell size or granularity) and the fluorescence of cells following staining with monoclonal antibodies to markers on the cell surface bound to fluorescent dyes. Flow cytometry data were analysed using FlowJo version 7.6.5 software (Becton Dickinson, Franklin Lakes, NJ, USA) and cell populations of interest were then gated after identification. The generated percentages were multiplied by the total number of lymphocytes in the haemogram to derive absolute values for circulating lymphocytes. Absolute CD4⁺ cell counts were subsequently analysed using a single-platform technique.

Statistical analysis

The data were analysed using Statistical Package for Social Sciences (SPSS) software version 20.0 (IBM, CA, USA). Chisquare was used to determine significant association between the prevalence of anti-*T. gondii* and sociodemographic variables of participants. The *t*-test was used to determine the difference between CD4⁺ T-cell counts between the two groups of participants. One-way ANOVA with Tukey's test was used to determine differences in CD4⁺ T-cell count across the three gestational groups of subjects with HIV/*T. gondii* coinfection and *T. gondii* monoinfection. *P*-values ≤0.05 at confidence interval (CI) of 95% were considered significant.

Results

The overall seroprevalence of anti-*T. gondii* IgG and IgM was 28.8% and 3.8%, respectively. Of the 160 HIV-seropositive and 160 HIV-seronegative pregnant women tested, the seroprevalence of anti-toxoplasma IgG and IgM was 29.4% and 4.4%, respectively, among HIV-seropositive pregnant women and 28.1% and 3.1%, respectively, among HIV-seronegative women (Table 1). The seroprevalence of anti-*T. gondii* IgG and IgM among HIV-seropositive pregnant women was highest among those aged 20–39 years, with 45 (95.7%) and 7 (100.0%) seropositive cases, respectively, of IgG and IgM.

Those living in urban areas had more cases of anti-*T. gondii* IgG and IgM antibodies than their rural counterparts (91.5% and 4% versus 100.0% and 0.0%, respectively) (Table 2). The seroprevalence of anti-*T. gondii* IgG and IgM was highest

among HIV-seropositive pregnant women with tertiary education (20 [42.6%] and 5 [71.4%], respectively). Pregnant women at their third trimester (23 [48.9%] and 3 [42.9%]) had the highest prevalence of anti-*T. gondii* IgG and IgM than those at their first and second trimester (19.2% versus 14.2% and 31.9% versus 42.9%, respectively).

Table 1. Seroprevalence of *Toxoplasma gondii* antibodies among pregnant women (n = 320)

HIV status	IgG positive (%)	P-value	IgM positive (%)	P-value
HIV-positive (n = 160)	47 (29.4)	0.5315	7 (4.4)	0.5416
HIV- negative (n = 160)	45 (28.1)		5 (3.1)	
Total	92 (28.8)		12 (3.8)	

HIV-uninfected pregnant women with parity of 1–4 had the highest anti-T. gondii IgG (62.2%) when compared with the nulliparous (22.2%) and those with parity ≥ 5 (15.6%). Similarly, HIV-uninfected pregnant women with parity of 1–4 had the highest anti-T. gondii IgM (60.0%) when compared with the nulliparous (20.0%) and those with parity ≥ 5 (20.0%) (Table 2). There was no significant association between the seroprevalence of anti-T. gondii-IgG and -IgM with age, gestational age, educational level, parity and place of residence of HIV-infected pregnant women (P > 0.005) (Table 2).

The seroprevalence of anti-*T. gondii* IgG and IgM was highest among HIV-seronegative pregnant women between 20 and 39 years (40 [88.9%] and 5 [100.0%], respectively). Those living in urban areas had more cases of anti-*T. gondii* IgG antibodies than their rural counterparts (84.4% versus 80.0%) (Table 3). In contrast, the seroprevalence of anti-*T. gondii* IgM was relatively lower among those residing within urban areas compared with rural residents (15.6% versus 20.0%) (Table 3).

The seroprevalence of anti-*T. gondii* IgG and IgM was highest among HIV-seronegative pregnant women with tertiary education (33 [73.3%] and 5 [100.0%], respectively). Pregnant women at their third trimester had a higher prevalence of anti-*T. gondii* IgG and IgM (23 [51.1%] and 3 [60.0%], respectively) than those at their first and second trimester (13.3% versus 35.6% and 20.0% versus 20.0%, respectively).

Nulliparous HIV-infected pregnant women had the least anti-T. gondii IgG (12.8%) when compared with those with parity of 1–4 (46.8%) and \geq 5 parity (40.4%). However, HIV-infected pregnant women with parity of 1–4 had the highest anti-T. gondii IgM (57.1%) when compared with the nulliparous (14.3%) and those with parity \geq 5 (28.6%) (Table 3). There was significant association between the seroprevalence of anti-T. gondii-IgG and -IgM with education level but only anti-T. gondii-IgM was significantly associated with place of residence (P = 0.01). However, there was no association between age and parity of HIV-seronegative pregnant women (P > 0.05) (Table 3).

HIV-infected pregnant women without anti-T. gondii IgM

Table 2. Seroprevalence of anti-Toxoplasma gondii IgG and IgM by sociodemographic variables of HIV-infected pregnant women

Variable	IgG			IgM		
	Positive (%)	Negative (%)	P-value	Positive (%)	Negative (%)	P-value
Age (years)						
≤19	0 (0.0)	0 (0.0)		5 (3.3)	5 (4.4)	
20–39	45 (95.7)	98 (86.7)	0.778	7 (100.0)	136 (88.9)	0.804
≥40	2 (4.3)	10 (8.9)		0 (0.0)	12 (7.8)	
Place of residence						
Urban	43 (91.5)	84 (74.3)	0.360	7 (100.0)	120 (78.4)	0.790
Rural	4 (8.5)	29 (25.7)		0 (0.0)	33 (21.6)	
Educational status						
No formal education	3 (6.4)	25 (22.1)		0 (0.0)	28 (18.3)	
Primary	14 (29.8)	9 (8.0)	0.061	2 (28.6)	21 (13.7)	0.248
Secondary	10 (21.3)	38 (33.6)		0 (0.0)	48 (31.4)	
Tertiary	20 (42.6)	41 (36.3)		5 (71.4)	56 (36.6)	
Parity						
Nulliparous	6 (12.8)	50 (44.3)		1 (14.3)	55 (35.9)	
Parity 1–4	22 (46.8)	32 (28.3)	0.642	4 (57.1)	50 (32.7)	0.919
Parity ≥5	19 (40.4)	31 (27.4)		2 (28.6)	48 (31.4)	
Gestational age						
1st trimester	9 (19.2)	25 (22.1)		1 (14.2)	33 (21.6)	
2nd trimester	15 (31.9)	38 (33.6)	0.606	3 (42.9)	50 (32.7)	0.119
3rd trimester	23 (48.9)	50 (44.3)		3 (42.9)	70 (45.7)	

^{*}Significant difference determined by chi-square test.

Table 3. Seroprevalence of anti-Toxoplasma gondii IgG and IgM by sociodemographic variables of HIV-seronegative pregnant women.

Variable	IgG		P-value	IgM		<i>P</i> -value
	Positive (%)	Negative (%)		Positive (%)	Negative (%)	
Age (years)						
≤19	0 (0.0)	1 (0.9)		0 (0.0)	1 (0.6)	
20–39	40 (88.9)	111 (96.5)	0.333	5 (100.0)	146 (94.2)	0.554
≥40	5 (11.1)	3 (2.6)		0 (0.0)	8 (5.2)	
Place of residence						
Urban	38 (84.4)	114 (99.1)		4 (80.0)	148 (95.5)	
Rural	7 (15.6)	1 (0.9)	0.250	1 (20.0)	7 (4.5)	0.010*
Educational status						
No formal educa- tion	0 (0.0)	7 (6.1)		0 (0.0)	7 (4.5)	
Primary	4 (8.9(30 (26.1)	0.033	0 (0.0)	34 (22.0)	0.010
Secondary	8 (17.8)	50 (43.5)		0 (0.0)	58 (37.4)	
Tertiary	33 (73.3)	28 (24.3)		5 (100.0)	56 (36.1)	
Total	45 (100.0)	115 (100.0)		5 (100.0)	155 (100.0)	
Parity						
Nulliparous	10 (22.2)	9 (7.8)		1 (20.0)	18 (11.6)	
Parity 1–4	28 (62.2)	56 (48.7)	0.125	3 (60.0)	81 (52.3)	0.073
Parity ≥5	7 (15.6)	50 (43.5)		1 (20.0)	56 (36.1)	
Gestational age						
1st trimester	6 (13.3)	20 (17.4)		1 (20.0)	25 (16.1)	
2nd trimester	16 (35.6)	44 (38.3)	0.544	1 (20.0)	59 (38.1)	0.999
3rd trimester	23 (51.1)	51 (44.3)		3 (60.0)	71 (45.8)	

^{*}Significant difference determined by chi-square test

Table 4. Toxoplasma gondii serological status and CD4⁺ T-cell counts among HIV-infected pregnant women.

Sero-status	Subjects (n)	Mean±SD CD4⁺	P-value
(a) HIV+ and Toxo IgM+	7	324±23.4	
(b) HIV+ and Toxo IgM-	153	431±55.1	
Total	160	420±52.2	0.035
(a) HIV+ and Toxo IgG+	47	340±31.9	
(b) HIV+ and IgG-	113	372±45.2	
Total	160	330±33.5	0.075

^{*}Significant difference determined by *t*-test

had significantly higher CD4⁺ T-cell count (431±55.1 cells/mm³, mean±SD) than those who were anti-*T. gondii* IgM seropositive (324±23.4 cells/mm³; *P* = 0.035). However, HIV-infected pregnant women without anti-*T. gondii* IgG did not have a significantly different T-cell count (372±45.2 cells/mm³) compared with those who were anti-*T. gondii* IgG seropositive (340±31.9 cells/mm³; *P* = 0.075) (Table 4).Based on *T. gondii* IgG serostatus, it was observed that HIV-infected pregnant women at their third trimester had the lowest mean CD4⁺ T-cell count (284±18.9 cells/mm³), followed by those in their second trimester (323±24.2 cells/mm³), then those in their first trimester (358±28.6 cells/mm³). However,

based on their *T. gondii* IgM serostatus, it was observed that HIV-infected pregnant women at their first trimester had the lowest mean CD4⁺ T-cell count (399 \pm 0 cells/mm³), followed by those in their third trimester (411 \pm 38.5 cells/mm³), then those in their second trimester (523 \pm 41.3 cells/mm³). There was significant difference in the mean CD4⁺ T-cell count of HIV-seropositive pregnant women based on the three gestational groups and *T. gondii* IgM (P = 0.0048) and IgG status (P < 0.001) (Table 5). Based on the *T. gondii* IgG serostatus, it was observed that HIV-seronegative pregnant women at their first trimester had the lowest mean CD4⁺ T-cell count (562 \pm 67.2 cells/mm³), followed by those Https://dx.doi.org/10.4314/mmj.v32i3.9

in their third trimester (582 ± 68.1 cells/mm³), then those in their second trimester (611 ± 72.9 cells/mm³). However, based on their *T. gondii* IgM serostatus, it was observed that HIV-seronegative pregnant women at their first trimester had the lowest mean CD4+ T-cell count (426 ± 0 cells/mm³), followed by those in their second trimester (541 ± 51.7 cells/mm³), then those in their third trimester (598 ± 72.5 cells/mm³). There was no significant difference in the mean CD4+ T-cell count of HIV-seronegative pregnant women based on the three gestation groups and *T. gondii* IgG (P = 0.27) and IgM status (P = 0.32) (Table 6).

Discussion

In this study, the overall seroprevalence of anti-*T. gondii* (both IgG and IgM) among HIV-positive and HIV-negative pregnant women was found to be 33.8% and 31.2%, respectively. There was a slight increase in seroprevalence of both IgG and IgM antibodies (29.4% and 4.4%) among the pregnant HIV-positive population over their HIV-negative counterparts (28.1% and 3.1%). This difference could be due to the immunosuppression consequences in HIV-positive individuals which primarily manifests as a drop in their CD4⁺ T-cell counts and function¹³.

Table 5. Comparison of CD4* T-cell counts by gestation of HIV-infected pregnant women with serological evidence of toxoplasmosis.

\CD4+ T-cell count	(cells/mm³) mean±SD		
HIV-seropositive	Anti-T. gondii IgG positive (n = 47)	HIV-seropositive	Anti-T. gondii IgM positive (n = 7)
1st trimester (n = 9)	358±28.6	1st trimester (n = 1)	399±0
2nd trimester (n = 15)	323±24.2	2nd trimester (n = 3)	523±41.3
3rd trimester (n = 23)	284±18.9	3rd trimester (n = 3)	411±38.5
F	37.7689	F	26.8822
P-value	<0.0001*	P-value	0.0048*

^{*}Significant mean difference determined by one-way ANOVA

Tukey HSD post-hoc test for HIV-seropositive subjects with T. gondii IgG

1st trimester vs 2nd trimester: Diff. = -35.0000; 95% confidence interval (CI) = -58.1781 to -11.8219; P = 0.0019.

1st trimester vs 3rd trimester: Diff. = -74.0000; 95% CI = -95.6137 to -52.3863; P = 0.0000.

2nd trimester vs 3rd trimester: Diff. = -39.0000; 95% CI = -57.2440 to -20.7560; P = 0.0000.

Tukey HSD post-hoc test for HIV-seropositive subjects with T. gondii IgM

1st trimester vs 2nd trimester: Diff. = 124.0000; 95% CI = -40.2969 to 288.2969; P = 0.11.

1st trimester vs 3rd trimester: Diff. = -115.0000; 95% CI = -279.2969 to 49.2969; P = 0.14.

2nd trimester vs 3rd trimester: Diff. = -239.0000; 95% CI = -355.1755 to -122.8245; P = 0.003.

Table 6. Comparison of CD4⁺ T-cell counts by gestation of HIV-seronegative pregnant women with serological evidence of toxoplasmosis.

CD4 ⁺ T-cell count (cells/mm ³) mean±SD					
HIV- seronegative	Anti- T . gondii IgG positive ($n = 45$)	HIV- seronegative	Anti- <i>T. gondii</i> IgM positive (<i>n</i> = 5)		
1st trimester $(n = 6)$	562±67.2	1st trimester (n = 1)	426±0		
2nd trimester $(n = 16)$	611±72.9	2nd trimester ($n = 1$)	541±51.7		
3rd trimester $(n = 23)$	582±68.1	3rd trimester ($n = 3$)	598±72.5		
F	1.3594	F	2.1255		
<i>P</i> -value	0.2679	P-value	0.3199		

^{*}Significant mean difference determined by one-way ANOVA.

Tukey HSD post-hoc test for HIV-seronegative subjects with T. gondii IgG

1st trimester vs 2nd trimester: Diff. = 49.0000; 95% confidence interval (CI) = -32.1196 to 130.1196; P = 0.32.

1st trimester vs 3rd trimester: Diff. = 20.0000; 95% CI = -57.6800 to 97.6800; P = 0.81.

2nd trimester vs 3rd trimester: Diff. = -29.0000; 95% CI = -84.1643 to 26.1643; P = 0.42.

Tukey HSD post-hoc test for HIV-seronegative subjects with T. gondii IgM

1st trimester vs 2nd trimester: Diff. = 115.0000; 95% CI = -488.9721 to 718.9721; P = 0.60.

1st trimester vs 3rd trimester: Diff. = 172.0000; 95% CI = -321.1412 to 665.1412; P = 0.30.

2nd trimester vs 3rd trimester: Diff. = 57.0000; 95% CI = -436.1412 to 550.1412; P = 0.80.

The prevalence rates from this study are higher than the prevalence of 9.8% and 12.8% among an HIV-positive subgroup and HIV-negative pregnant women, respectively, reported in South Africa¹⁴. The overall seroprevalence in this study is in conformity with a previous seroprevalence of 31.5% recorded in the central city of Abuja⁸. The prevalence in this study was higher than the 29.9% recorded in Zaria but lower than others carried out in Lagos and Maiduguri where 40.2% and 48.9% were recorded, respectively^{10,15}. The variations of findings from these studies could be due to differences in laboratory methods adopted by the studies, sample size, HIV and antiretroviral therapy statuses of study participants and endemicity of toxoplasmosis.

Lagos is a riverine area with a high prevalence of outdoor activities, whereas Maiduguri dwellers have been ravaged by national insecurity which grossly affected their standard of living and the socioeconomic status of residents, including access to potable water and hygienic meat and vegetables. Studies reported within and outside Africa have recorded seroprevalence rates among pregnant women of 83.6% in Ethiopia¹⁶, 92.5% in Ghana¹⁷, 48.3% in India¹⁸ and 56.6% in Brazil¹⁹. It is possible that geographical and hot climatic conditions in these countries that favour sporulation could account for these wide margins in prevalence^{20,21}. The climatic condition in our study area may be relatively less favourable for the survival of oocysts compared with studies that reported a very high seroprevalence of toxoplasmosis. Prevalence rates as low as 10% have been reported in the United States and the UK^{22,23}. The lower rates found in developed countries are not surprising as these countries have higher standards of living, good potable water and proper waste disposal systems that might reduce the risk of contracting T. gondii infection.

The highest prevalence of *T. gondii* IgG and IgM antibodies was observed among women in the age group of 20–39 years. This was similar in both subgroups. Seroprevalence is said to increase with age among both immunocompromised and immunocompetent pregnant women⁸. This may be an indicator of susceptibility to *T. gondii* infection due to occupational hazards such as farming and contact with poultry in older populations, compared with younger populations who are less involved in these activities²⁴. These findings corroborate the reports from Zaria in Nigeria²⁵ and Saudi Arabia²⁶.

Toxoplasmosis is a food-borne infection. IgG and IgM seropositivity were found to be higher among the urban dwellers than their rural counterparts in both the HIV-positive and HIV-negative pregnant women. This may be because urban dwellers are more likely to patronise fast-food vendors and restaurants with poor sanitary conditions. This action may predispose pregnant women, especially the 'working class', to consume poorly prepared vegetables and undercooked meat products from unprofessional vendors. A similar finding was reported by Nasir et al.¹⁵

There was a statistically significant relationship between attaining tertiary level of education among the HIV-negative pregnant women and a higher prevalence of IgG and IgM antibodies.

Although it is generally accepted that highly educated people are expected to know about infection prevention measures, we observed that this category of pregnant women might have contracted food-borne toxoplasmosis because their occupation might have affected their feeding habits, as previously explained. To further buttress this, urban residents had a higher prevalence of anti-*T. gondii* IgG than rural residents. However, this opposes other findings in studies that showed no significant association between different levels of education^{26,27}.

The highest seroprevalence of anti-*T. gondii* IgG antibodies for both HIV-positive and HIV-negative pregnant women was found in the third trimester (51.1% versus 48.9%) in this study. This could be due to the immunosuppressive effects of pregnancy at a late stage (third trimester) tending to predispose these women to higher chances of contracting *T. gondii* infection. It has been demonstrated that congenital transmission also increases with gestational age, with the highest rates in the third trimester¹¹. On the other hand, disease severity has been reported to decrease with gestational age, with first-trimester infection resulting in miscarriage more often²⁸.

The mean CD4⁺ T-cell counts were found to be significantly lower in HIV-positive pregnant women with anti-*T. gondii* IgM antibodies compared with those with IgM seronegative results. Similar findings were also reported by Nazari et al.¹³ This might be due primarily to the synergistic effects of HIV infection and toxoplasmosis²⁹.

In HIV infection, due to immunosuppression, opportunistic infection occurs with *T. gondii* because of depletion of CD4

T cells, impaired production of IL-12 and interferon gamma (IFN-γ) and impaired cytotoxic T-lymphocyte activity²⁹. There is decreased in vitro production of IL-12 and IFN-y, and decreased expression of CD154 in response to T. gondit²⁹. It could be inferred that the women with HIV infection should be considered at high risk for developing toxoplasmosis, especially when CD4⁺ T-cell count is very low. CD4⁺ T-cell counts of less than 100 cells/mm³ have been implicated in reactivation of latent infection which could lead to clinical neurologic disease. There are several toxoplasmosis crosssectional studies across different subpopulations. However, very few have focused on simultaneous investigation of toxoplasmosis and its impact on CD4+ T cellular immunity in pregnant women living with HIV/AIDS and those who were HIV seronegative. In this study, it was shown that primary toxoplasmosis in HIV infection significantly altered CD4+ T-cell count of pregnant women. Hence, these findings will assist obstetricians and gynaecologists in the early diagnosis and management of T. gondii infection in pregnant women, especially HIV-coinfected individuals, to prevent intrauterine complications.

Conclusion

In this study, the seroprevalence of anti-*T. gondii* IgG and IgM did not differ in the HIV-seropositive or HIV-seronegative pregnant population. However, women with primary *T. gondii* and HIV coinfection had lower CD4⁺T-cell count than those with toxoplasmosis monoinfection.

Place of residence and level of education among HIV-negative pregnant women have significant association with toxoplasmosis. There is a need to educate pregnant women about preventive strategies and risk factors that could predispose them to *T. gondii* infection and to prevent consequential congenital infections.

Authors' contributions

Conceptualisation and design of paper: MMZ, AYI. Collection of data: MMZ, AYI, RO, TY, INA. Implementation of research: MMZ, AYI, RO, TY, INA. Data analysis and interpretation: MMZ, AYI, INA. Preparation of manuscript: MMZ, AYI, RO, TY, INA. All authors checked and approved the final manuscript before submission.

Competing interests

None to declare.

References

- 1. Rostami A, Riahi SM, Gamble HR, Fakhri Y, Nourollahpour Shiadeh M, Danesh M, et al. Global prevalence of latent toxoplasmosis in pregnant women: a systematic review and meta-analysis. Clin Microbiol Infect. 2020;26(6):673-83. doi: 10.1016/j.cmi.2020.01.008.
- 2. Ayi I, Sowah AO, Blay EA, Suzuki T, Ohta N, Ayeh-Kumi PF. *Toxoplasma gondii* infections among pregnant women, children and HIV seropositive persons in Accra, Ghana. Trop Med Health. 2016;44:17. doi: https://doi.org/10.1186/s41182-016-0018-5.
- 3. Mendez OA, Koshy AA. *Toxoplasma gondii:* entry, association, and physiological influence on the central nervous system. PLoS Pathog. 2017;13(7):e1006351. doi: https://doi.org/10.1371/journal.ppat.1006351.
- 4. Rezanezhad H, Sayadi F, Shadmand E, Nasab SDM, Yazdi HR, Solhjoo K, et al. Seroprevalence of *Toxoplasma gondii* among HIV Patients in Jahrom, Southern Iran. Korean J Parasitol. 2017;55:99-103. doi: https://doi.org/10.3347/kjp.2017.55.1.99.
- 5. Machala L, Malý M, Hrdá S, Rozsypal H, Stanková M, Kodym P. Antibody response of HIV-infected patients to latent, cerebral and

- recently acquired toxoplasmosis. Eur J Clin Microbiol Infect Dis. 2009;28:179-82. doi: https://doi.org/10.1007/s10096-008-0600-9.
- 6. Katawa G, Kolou M, Nadjir LK, Ataba E, Bomboma G, Karou SD. CD4 T-lymphocytes count in HIV-*Toxoplasma gondii* co-infected pregnant women undergoing a prevention of mother-to-child transmission program. J Biosci Med. 2018;6:76-84. doi: https://doi.org/10.4236/jbm.2018.64006.
- 7. Nigerian National AIDS Indicator and Impact Survey. Press Release. 2019 [cited 2020 Apr 21]. Available from: https://www.unaids.org/en/resources/presscentre/pressreleaseandstatementarchive/2019/march/20190314_nigeria.
- 8. Uttah EC, Ajang R, Ogbeche J, Etta H, Etim L. Comparative seroprevalence and risk factors of toxoplasmosis among three subgroups in Nigeria. J. Nat Sci Res. 2013;3(8):23-9. Available from: https://www.iiste.org/Journals/index.php/JNSR/article/view/6891/6997.
- 9. Ishaku BS, Ajogi I, Umoh UJ, Lawal I, Randawa AJ. Seroprevalence and risk factors for *Toxoplasma gondii* infection among antenatal women in Zaria, Nigeria. Res J Med Sci 2009;4(2):483-8. doi: https://doi.org/10.9734/AJMAH/2017/31528.
- 10. Deji-Agboola AM, Busari OS, Osinupebi OA, Amoo AOJ. Seroprevalence of *Toxoplasma gondii* antibodies among pregnant women attending antenatal clinic of Federal Medical Centre Lagos, Nigeria. Int J Biol Med Res. 2011;2(4):1135-9.
- 11. Thiebaut R, Leproust S, Chene G, Gilbert R. Effectiveness of prenatal treatment for congenital toxoplasmosis: a meta-analysis of individual patient's data. Lancet. 2007;369(9556):115-22. doi: https://doi.org/10.1016/S0140-6736(07)60072-5.
- 12. Declaration of Helsinki 2000. World Medical Association [cited 2020 Apr 29]. Available from: https://www.wma.net/what-we-do/medical-ethics/declaration-of-helsinki/doh-oct2000/.
- 13. Nazari N, Bozorgomid A, Janbakhsh A, Bashiri F. *Toxoplasma gondii* and human immunodeficiency virus co-infection in western Iran: a cross sectional study. Asian Pac J Trop Med. 2018;11:58-62. doi: https://doi.org/10.4103/1995-7645.223562.
- 14. Kistiah K, Barragan A, Winiecka-Krusnell J, Karstaedt A, Frean J. Seroprevalence of *Toxoplasma gondii* infection in HIV positive and HIV negative subjects in Gauteng, South Africa. South Afr J Epidemiol Infect. 2011;26(4):225-8. doi: https://doi.org/10.1080/10158782.2011. 11441457.
- 15. Nasir IA, Aderinsayo AH, Mele HU, Aliyu MM. Prevalence and associated risk factors of *Toxoplasma gondii* antibodies among pregnant women attending Maiduguri Teaching Hospital, Nigeria. J Med Sci. 2015;15(3):147-54. doi: https://doi.org/jms.2015.147.154.
- 16. Zemene E, Yewhalaw D, Abera S, Belay T, Samuel A, Zeynudin A. Seroprevalence of *Toxoplasma gondii* and associated risk factors among pregnant women in Jimma town, Southwestern Ethiopia. BMC Infect Dis. 2012;12:337. doi: https://doi.org/10.1186/1471-2334-12-337.
- 17. Ayi I, Edu A, Apea-Kubi K, Boamah D, Bosompem K, Edoh D. Sero-epidemiology of toxoplasmosis amongst pregnant women in the Greater Accra Region of Ghana. Ghana Med J. 2009;43:107-14. doi: https://doi.org/10.4314/gmj.v43i3.55325.
- 18. Singh S, Pandit AJ. Incidence and prevalence of toxoplasmosis in Indian pregnant women: a prospective study. Am J Reprod Immunol. 2004;52:276-83. doi: https://doi.org/10.1111/j.1600-0897.2004.00222.x.
- 19. Vaz RS, Thomaz-Soccol V, Sumikawa E, Guimaraes ATB. Seroprevalence of *Toxoplasma gondii* antibodies in pregnant women in southern Brazil. Parasitol Res. 2010;106:661-5. doi: https://doi.org/10.1007/s00436-009-1716-2.
- 20. Nijem KI, Al-Amleh S. Seroprevalence and associated risk factors of toxoplasmosis in pregnant women in Hebron district, Palestine. East Mediterr Health J. 2009;15:1278-84. Available from: http://applications.emro.who.int/emhj/1505/15_5_2009_1278_1284.pdf.

- 21. Tenter AM, Heckeroth AR, Weiss LM. *Toxoplasma gondii*: from animals to humans. Int J Parasitol. 2000;30(12-13):1217-58. doi: https://doi.org/10.1016/s0020-7519(00)00124-7.
- 22. Jones JL, Kruszon-Moran D, Sanders-Lewis K, Wilson M. *Toxoplasma gondii* in the United States, 1999-2004, decline from the prior decade. Am J Trop Med Hyg. 2007;77(3):405-10.
- 23. Nash JQ, Chissel S, Jones J, Warburton F, Verlander NQ. Risk factors for toxoplasmosis in pregnant women in Kent, United Kingdom. Epidemiol Infect. 2005;133(3):475-83. doi: https://doi.org/10.1017/s0950268804003620.
- 24. Aqeely H, El-Gayar EK, Khan DP, Najmi A, Alvi A. Seroepidemiology of *Toxoplasma gondii* amongst pregnant women in Jaza province, Saudi Arabia. J Trop Med. 2014;2014:913950. doi: https://doi.org/10.1155/2014/913950.
- 25. Ogoina D, Onyemelukwe GC, Musa BO, Obiako RO. Seroprevalence of IgM and IgG antibodies to Toxoplasma infection in

- healthy and HIV positive adults from northern Nigeria. J Infect Dev Countr. 2013;7(5):398-403. doi: https://doi.org/10.3855/jidc.2797.
- 26. Al- Harthi AS, Menal H, Ghazi HO. Seroprevalence of *Toxoplasma gondii* among pregnant women in Makkah, Saudi Arabia. Um Al Qura Univ J Sci Med Eng. 2006;8(2):217-27.
- 27. Gelaye W, Kebede T, Hailu A. High prevalence of anti-Toxoplasma antibodies and absence of *Toxoplasma gondii* infection risk factors among pregnant women attending routine antenatal care in two hospitals of Addis Ababa, Ethiopia. Int J Infect Dis. 2015;34:41-5. doi: https://doi.org/10.1016/j.ijid.2015.03.005.
- 28. Ghasemi FS, Rasti S, Piroozmand A, Bandehpour M, Kazemi B, Mousavi SG, et al. Toxoplasmosis-associated abortion and still birth in Tehran, Iran. J Matern Fetal Neonatal Med. 2016;(2):248-51. doi: https://doi.org/10.3109/14767058.2014.996127.
- 29. Basavaraju A. Toxoplasmosis in HIV infection: an overview. Trop Parasitol. 2016;6(2):129-35.