



## EFFECT OF LEAFLITTERS OF NITROGEN FIXING ACACIA TREES ON THE GROWTH OF AFRICAN STAR APPLE (*Chrysophyllum albidum* G.DON): A STEP TOWARDS ENHANCING THE GROWTH OF AN ENDANGERED SPECIES

\*Adelani, D.O., Oni, B.O and Ariyo, O.C.

Federal College of Forestry Mechanization, P.M.B 2273, Afaka, Kaduna, Nigeria

\*Corresponding author's e-mail: [adelani.olusegun@yahoo.com](mailto:adelani.olusegun@yahoo.com); +2347038953146

### ABSTRACT

To meet the population demand for enormous benefits of an endangered species, *Chrysophyllum albidum*, its' slow growth need to be improved through the use of environmentally friendly leaf litters of nitrogen fixing acacia trees. Investigation conducted into effect of leaf litters of nitrogen fixing acacia trees (*Acacia tortilis*, *Acacia seyal*, *Acacia nilotica*, *Acacia senegal*, *Acacia leucophloea* and *Acacia albida*) at the rates of 300 g on the growth of *C. albidum* was laid down using a Completely Randomized Design with five replications. Results showed that leaf litters of nitrogen fixing acacia tree species significantly ( $P < 0.05$ ) enhanced the growth of *C. albidum* seedlings. Tallest plant (25 cm), widest leaf area ( $55.71 \text{ cm}^2$ ) and highest number of leaves (11), significant total fresh weight (48.45 g) and total dry weight (21.55 g) of *C. albidum* were recorded from seedlings planted in soil amended with *A. senegal*. Widest girth (1.92 cm) was recorded from seedlings planted in the soil influenced with *A. leucophloea*. Highest percentage values of 2.22 %, 0.90 % and 1.74 % were recorded for nitrogen, phosphorus and potassium content of *A. senegal*. Soil influenced with *A. senegal* enhanced the growth of *C. albidum* seedlings. Planting of *C. albidum* seedlings in the soil amended with *A. senegal* is recommended for mass production of its seedlings for agro-forestry systems as well as to increase biodiversity conservation.

**Key words:** Leaf litters, Nitrogen fixing tree species, Growth, Population demand, Agro-forestry

### INTRODUCTION

Nigeria is one of the most densely populated countries in Africa, with approximately 200 million people in an area of  $920,000 \text{ km}^2$  ( $360,000 \text{ sq mi}$ ), (Akinyemi and Isiugo-Abanihe, 2014; WHO, 2020) and is also the country with the largest population in Africa (Farrell, 2018; Akanonu, 2019; WHO, 2020) and the seventh largest population in the world (Favre, 2019). The current population of Nigeria is 205,192,124 based on Worldometer elaboration of the latest United Nations data. One of the major worries and concerns of ever dick and harry is how to eradicate poverty and improve food security (Oyebamiji et al., 2019). Mbwambo et al. (2006) stated that agro-forestry is one of the answers that play an important role in increasing food security; improving nutrition and alleviating poverty. However, success of any agro-forestry system relies

heavily on the choice of suitable tree species that should offer diversity of benefits and show compatibility with food crops (Edward et al., 2006) and woody perennials. African star apple, *Chrysophyllum albidum* is one of such tree species.

*Chrysophyllum albidum* is also one of priority and multipurpose tree species of immense benefits in agro-forestry system in Africa. African star apple or White star apple is a climax tree species of tropical rainforest that belongs to the family of Sapotaceae (Wole, 2013; Olaoluwa et al., 2012) which has up to 800 species (Ehiagbonare et al., 2008). The Yoruba name of *C. albidum* is "Osan Agbalumo" (Rahaman, 2012) while in Igbo and Hausa languages, it is called "Udara" or "Udala" and "Agbaluba" or "Agbaluma" respectively (Wole, 2013, Adelani et al., 2016). *C. albidum* has been

noted to be of great economical (Onyekwelu *et al.*, 2011); nutritional and medicinal (Onyekwelu and Stimm, 2011) and industrial (Olaoluwa *et al.*, 2012) values. Ecologically, the tree has an efficient nutrient cycling and the high rate of mineralization of the leaves that improves the quality of the top soil (Aduradola *et al.*, 2005).

The World Agroforestry Centre (ICRAF) has identified *C. albidum* as one of the top five priority species for domestication in the African humid tropics (Tchounjeu *et al.*, 2002). Extensive human activities deplete the population of unregenerated *C. albidum* (Onyekwelu *et al.*, 2011) during irreversible conversion of forest areas to other uses. Despite the importance of *C. albidum*, it has been greatly neglected particularly with respect to their regeneration (Adelani *et al.*, 2014a, Adelani *et al.*, 2016). This calls for deliberate action to save the genetic erosion of this important species. Adelani and Okechalu (2016) stated that *C. albidum* has challenges of slow growth of seedlings and poor germination of seeds. There is need for the use of leaf litters of nitrogen fixing acacia to supply nutrients to enhance its growth for integration with arable crops in agro-forestry systems.

Agroforestry employs the use of fertilizer trees in improving the fertility of soils. Sarvade *et al.* (2014) mentioned that agro-forestry has a great potential of both restoring and maintaining soil fertility and increasing agricultural production. Researchers and planners are recommending agro-forestry systems for increasing agricultural production with improving soil nutrient status (Sarvade *et al.*, 2014). Biomass transfer is an agro-forestry practices that involve cut and carry of litters of agro-forestry tree species from one mother tree to place of restoring soil fertility. Schroth and Sinclair (2003) reported that biomass transfer systems represent an intermediate situation in which the nutrients in the biomass are added to the site where the biomass is applied, but are removed from another site within the same landscape.

The leaf litters of nitrogen fixing trees and other agro-forestry tree species are used in biomass transfer method. WAC (2018) stated that nitrogen fixing tree species have the ability to fix atmospheric nitrogen through symbiosis with bacteria or fungi in root nodules. ICRAF (2009)

stated that nitrogen fixing trees enhance soil fertility and improve crop yields through their litters and nitrogen fixing process. The range of N<sub>2</sub>-fixation capacity varies greatly amongst these trees (Giller, 2001). Nygren *et al.* (2012) pointed that annually, N<sub>2</sub>-fixation may add from tens to hundreds of kilograms of N per hectare to an agro-forestry system. The most popular N<sub>2</sub>-fixing trees used in tropical agro-forestry systems include the legumes *Acacia* spp., *Erythrina* spp., *Gliricidia* spp., *Inga* spp., and *Leucaena* spp., which form symbiotic associations with a wide variety of N<sub>2</sub>-fixing bacterial species (Bala *et al.*, 2003).

Palm (1995) noted that nitrogen-fixing trees normally have higher nitrogen concentrations in the biomass than non-fixing species, but this characteristic also varies widely between species. There is dearth of quantified information on the effect of leaf litters of nitrogen fixing trees on the growth of woody perennials. In this light, this investigation was performed to assess the effect of leaf litters of nitrogen fixing acacia trees on the growth of *C. albidum* seedlings.

## MATERIALS AND METHODS

### Experimental Site

The pot experiment of this study was conducted during the wet season (2018) in the screen house of Federal College of Forestry Mechanization, Afaka, Kaduna State, Nigeria. The College is located in the Northern Guinea Savannah ecological zones of Nigeria. It is situated in Chikun Local Government Area of Kaduna State, Nigeria. It lies between Latitude 10° 35' and 10° 34' and Longitude 7° 21' and 7° 20' (Adelani, 2015). The mean annual rainfall is approximately 1000 mm. The vegetation is open woodland with tall broad leaf trees (Otegbeye *et al.*, 2001).

### Collection and Preparation of Experimental Materials

The biomass transfer method which involves the collections of wet leaves was used because some of nitrogen fixing acacia tree species was not located in the same site. The leaves of nitrogen fixing acacia tree species were air dried and pulverized. The sand was collected from the floor of the College dam, passed through 2 mm sieve and soaked in 10 % hydrochloric acid for 24 hours to

remove impurities, organic matter and nutrient residue (Adelani et al., 2014a). The poly pots of 20 x 10 x 10 cm<sup>3</sup> capacity filled with sterilized river sand in the nursery was used.

### Chemical Analysis of Leaf Litters of Nitrogen Fixing Acacia Trees

Each sample of pulverized leaves of nitrogen fixing tree species after air dried was analyzed chemically for nitrogen, phosphorus and potassium (NPK) content at the Federal University of Agriculture Abeokuta, Ogun State, Nigeria laboratory. Determination of total nitrogen was done by Macro Kjeldahi method. Available phosphorus (P) was extracted by Bray-1 method and determined colourimetrically. Extracts from the digestion of the leaves of the agro-forestry tree species were used to determine potassium by flame photometry.

### Experimental Design

The experiment was laid down in a Completely Randomized Design with five replicates. The total sum of 35 seedlings was involved in the experiment. Six treatments and control was used for the study. Leaves of six nitrogen fixing acacia trees (*Acacia tortilis*, *Acacia seyal*, *Acacia nilotica*, *Acacia senegal*, *Acacia leucophloea* and *Acacia albida*) at the rates of 300 g each was mixed with the sterilized river sand and later packed into polythene pot of dimension of 20 x 10 x 10 cm<sup>3</sup>. Seedlings planted in the sterilized river sand without amended with leaf litters of nitrogen fixing trees served as control. Volume of 120 mL of water was applied daily to the seedlings in the polypots. Seedling assessment was carried out after first two

weeks of transplanting to ensure that seedlings were established before the commencement of assessment. Seedling assessment was taken every two weeks for 16 weeks. The mean of every four weeks was tabulated. Parameters assessed include: seedling height (using meter rule), number of leaves and collar girth (using venier calliper). Leaves were counted manually while leaf area was obtained by linear measurement of leaf length and leaf width as described by Ugeese et al. (2008) as stated in the formula below.

$$LA=4.41+1.14LW \quad [1]$$

Where:

LA= leaf area

LW= Production of linear dimension of the length and width at the broadest part of the leaf

### Data Analysis

Data were collected and subjected to analysis of variance (ANOVA) using SAS (2003) software to compare significant means. Significant means at 0.05 was separated using Fishers Least Significant Difference (LSD).

### RESULTS

Tallest plant of 25 cm was recorded from seedlings planted in soil amended with *A. senegal* at 16 WAT. Significant value of 25cm was recorded from seedlings planted in the soil amended with the leaf litters of *A. senegal* as compared to the least value of 3.67cm obtained from seedlings planted in soil not mixed with leaf litters of nitrogen fixing acacia trees (Table 1).

**Table 1: Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on Height (cm) of *C. albidum* Seedlings**

N FAT	Weeks After Transplanting			
	4	8	12	16
<i>A. seyal</i>	16.67 <sup>b</sup>	17.67 <sup>b</sup>	17.67 <sup>b</sup>	21.37 <sup>a</sup>
<i>A. nilotica</i>	17.33 <sup>b</sup>	17.67 <sup>b</sup>	21.00 <sup>ab</sup>	22.67 <sup>a</sup>
<i>A. tortilis</i>	16.67 <sup>b</sup>	17.33 <sup>ab</sup>	19.33 <sup>ab</sup>	20.43 <sup>a</sup>
<i>A. senegal</i>	17.67 <sup>b</sup>	18.67 <sup>b</sup>	23.67 <sup>ab</sup>	25.00 <sup>a</sup>
<i>A. leucophloea</i>	15.00 <sup>c</sup>	16.67 <sup>ab</sup>	18.67 <sup>b</sup>	21.43 <sup>a</sup>
<i>A. albida</i>	16.33 <sup>b</sup>	17.33 <sup>b</sup>	17.67 <sup>b</sup>	22.03 <sup>a</sup>
Control	3.67 <sup>c</sup>	3.67 <sup>c</sup>	4.67 <sup>b</sup>	5.67 <sup>a</sup>
<b>SE±</b>	<b>0.29</b>	<b>0.30</b>	<b>0.34</b>	<b>0.38</b>

\*Means on the same rows having different superscript are significantly different ( $P < 0.05$ )

**Key:** NFAT- Nitrogen Fixing Acacia Trees

### Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Girth of *C. albidum* Seedlings

The widest girth of 1.92 cm was recorded for seedlings planted in the soil amended with *A. leucophloea* at 8 WAT. The narrowest girth of 0.14 cm was recorded from seedlings planted in the soil

not mixed with leaf litters of nitrogen fixing acacia trees. The girth (1.92) of seedlings planted in the soil mixed with *A. leucophloea* at 8 WAT was significantly ( $P < 0.05$ ) different from that of control (0.14cm) (Table 2).

**Table 2: Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Girth (cm) of *C. albidum* Seedlings**

NFTS	Weeks After Transplanting			
	4	8	12	16
<i>A. seyal</i>	1.46 <sup>a</sup>	1.68 <sup>a</sup>	0.26 <sup>a</sup>	0.21 <sup>a</sup>
<i>A. nilotica</i>	1.60 <sup>a</sup>	1.48 <sup>a</sup>	0.27 <sup>a</sup>	0.22 <sup>a</sup>
<i>A. tortilis</i>	1.36 <sup>a</sup>	1.70 <sup>a</sup>	0.46 <sup>a</sup>	0.41 <sup>a</sup>
<i>A. senegal</i>	1.37 <sup>a</sup>	1.65 <sup>a</sup>	0.31 <sup>a</sup>	0.26 <sup>a</sup>
<i>A. leucophloea</i>	1.29 <sup>a</sup>	1.92 <sup>a</sup>	0.43 <sup>a</sup>	0.38 <sup>a</sup>
<i>A. albida</i>	1.54 <sup>a</sup>	1.48 <sup>a</sup>	0.52 <sup>a</sup>	0.47 <sup>a</sup>
Control	1.00 <sup>a</sup>	1.00 <sup>a</sup>	0.14 <sup>b</sup>	0.14 <sup>b</sup>
<b>SE±</b>	<b>0.25</b>	<b>0.28</b>	<b>0.06</b>	<b>0.06</b>

\*Means on the same rows having different superscript are significantly different ( $P < 0.05$ ).

**Key:** NFAT- Nitrogen Fixing Acacia Trees

### Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Number of Leaves of *C. albidum* Seedlings

Highest number of leaves (11) was recorded from seedlings planted in the soil amended with *A.*

*senegal* at 16 WAT. The number of leaves recorded for seedlings planted in soil amended with *A. senegal* (11) at 16 WAT was significantly ( $P < 0.05$ ) different from that of least value (3.67) recorded for untreated seedlings at 4 WAT (Table 3).

**Table 3: Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Number of Leaves of *C. albidum* Seedlings**

NFAT	Weeks After Transplanting			
	4	8	12	16
<i>A. seyal</i>	7.67 <sup>b</sup>	8.00 <sup>b</sup>	8.67 <sup>ab</sup>	9.67 <sup>a</sup>
<i>A. nilotica</i>	8.00 <sup>b</sup>	8.00 <sup>b</sup>	9.33 <sup>ab</sup>	10.00 <sup>a</sup>
<i>A. tortilis</i>	8.00 <sup>b</sup>	8.33 <sup>b</sup>	8.33 <sup>b</sup>	10.33 <sup>a</sup>
<i>A. senegal</i>	7.33 <sup>c</sup>	8.33 <sup>bc</sup>	9.00 <sup>b</sup>	11.00 <sup>a</sup>
<i>A. leucophloea</i>	7.33 <sup>b</sup>	8.00 <sup>b</sup>	8.67 <sup>b</sup>	10.67 <sup>a</sup>
<i>A. albida</i>	7.67 <sup>b</sup>	8.00 <sup>b</sup>	8.67 <sup>b</sup>	10.67 <sup>a</sup>
Control	3.67 <sup>b</sup>	4.00 <sup>b</sup>	5.00 <sup>a</sup>	5.33 <sup>a</sup>
<b>SE±</b>	<b>0.14</b>	<b>0.14</b>	<b>0.16</b>	<b>0.18</b>

\*Means on the same rows having different superscript are significantly different  $P (< 0.05)$

**Key:** NFAT- Nitrogen Fixing Acacia Trees

### Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Leaf Area of *C. albidum* Seedlings

The widest and significant leaf area of 55.71cm<sup>2</sup> was recorded from seedlings planted in the soil

influenced with *A. senegal* at 4 WAT. The least value of 10.32cm<sup>2</sup> was recorded for leaf area of seedlings planted in unamended soil (control).

**Table 4: Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Leaf Area (cm<sup>2</sup>) of *C. albidum* Seedlings**

NFTS	Weeks After Transplanting			
	4	8	12	16
<i>A. seyal</i>	52.98 <sup>a</sup>	29.22 <sup>b</sup>	26.54 <sup>b</sup>	31.54 <sup>b</sup>
<i>A. nilotica</i>	44.32 <sup>a</sup>	21.95 <sup>b</sup>	13.74 <sup>b</sup>	18.74 <sup>b</sup>
<i>A. tortilis</i>	47.53 <sup>a</sup>	26.95 <sup>b</sup>	13.49 <sup>b</sup>	18.49 <sup>b</sup>
<i>A. senegal</i>	55.71 <sup>a</sup>	28.61 <sup>b</sup>	16.80 <sup>b</sup>	21.80 <sup>b</sup>
<i>A. leucophloea</i>	22.40 <sup>a</sup>	27.60 <sup>a</sup>	16.35 <sup>a</sup>	21.35 <sup>a</sup>
<i>A. albida</i>	34.45 <sup>a</sup>	29.04 <sup>a</sup>	13.27 <sup>b</sup>	18.27 <sup>ab</sup>
Control	10.32 <sup>b</sup>	15.61 <sup>a</sup>	10.34 <sup>b</sup>	15.34 <sup>a</sup>
<b>SE±</b>	<b>2.51</b>	<b>1.60</b>	<b>0.98</b>	<b>1.27</b>

\*Means on the same rows having different superscript are significantly different  $P (<0.05)$

**Key:** NFTS- Nitrogen Fixing Tree Species

#### Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Leaf Area Index of *C. albidum* Seedlings

Significant leaf area index (1.92) was recorded from seedlings planted in the soil amended with *A.*

*senegal* at 4 WAT compared to least value (0.12) recorded in soil not mixed with leaf litters of nitrogen fixing acacia trees at 16 WAT (Table 5).

**Table 5: Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Leaf Area Index of *C. albidum* Seedlings**

NFAT	Weeks After Transplanting			
	4	8	12	16
<i>A. seyal</i>	1.46 <sup>a</sup>	1.68 <sup>a</sup>	0.26 <sup>b</sup>	0.21 <sup>b</sup>
<i>A. nilotica</i>	1.37 <sup>a</sup>	1.65 <sup>a</sup>	0.31 <sup>b</sup>	0.26 <sup>b</sup>
<i>A. tortilis</i>	1.36 <sup>a</sup>	1.70 <sup>a</sup>	0.46 <sup>b</sup>	0.41 <sup>b</sup>
<i>A. senegal</i>	1.92 <sup>a</sup>	1.29 <sup>a</sup>	0.43 <sup>b</sup>	0.38 <sup>b</sup>
<i>A. leucophloea</i>	1.60 <sup>a</sup>	1.48 <sup>a</sup>	0.27 <sup>b</sup>	0.22 <sup>b</sup>
<i>A. albida</i>	1.54 <sup>a</sup>	1.48 <sup>a</sup>	0.52 <sup>b</sup>	0.47 <sup>b</sup>
Control	1.00 <sup>a</sup>	1.00 <sup>a</sup>	0.12 <sup>a</sup>	0.12 <sup>a</sup>
<b>SE±</b>	<b>0.14</b>	<b>0.14</b>	<b>0.03</b>	<b>0.03</b>

\*Means on the same rows having different superscript are significantly different ( $P<0.05$ ).

**Key:** NFAT- Nitrogen Fixing Acacia Trees

#### Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Fresh and Dry Weight of *C. albidum*

A significant total fresh weight of 48.45 g was recorded from seedlings planted in soil amended with *A. senegal*. A significant total dry weight of 21.55 g was recorded from seedling planted in soil mixed with *A. senegal*. Significant fresh leaf

weight (19.05 g) and dry leaf weight (8.30 g) were recorded in seedlings planted in the soil influenced with *A. senegal*. The least values of 3.70 g and 0.45 g were recorded for total fresh weight and total dry weight of seedlings planted in the soil not amended with leaf of nitrogen fixing acacia trees (Table 6).

**Table 6: Effect of Leaf Litters of Nitrogen Fixing Acacia Trees on the Fresh and Dry Weight (g) of *C. albidum***

NFAT	FW			TFW	DW			TDW
	L	S	R		L	S	R	
<i>A.seyal</i>	2.60 <sup>a</sup>	1.35 <sup>a</sup>	2.25 <sup>a</sup>	6.20 <sup>e</sup>	1.10 <sup>a</sup>	0.65 <sup>a</sup>	0.75 <sup>a</sup>	2.50 <sup>f</sup>
<i>A.nilotica</i>	7.50 <sup>a</sup>	6.35 <sup>a</sup>	5.10 <sup>a</sup>	18.95 <sup>d</sup>	3.35 <sup>a</sup>	3.45 <sup>a</sup>	2.00 <sup>b</sup>	8.80 <sup>d</sup>
<i>A.tortilis</i>	9.95 <sup>a</sup>	6.20 <sup>a</sup>	7.45 <sup>a</sup>	23.60 <sup>c</sup>	4.45 <sup>a</sup>	3.50 <sup>b</sup>	3.10 <sup>b</sup>	11.05 <sup>c</sup>
<i>A.senegal</i>	19.05 <sup>a</sup>	12.30 <sup>b</sup>	17.10 <sup>a</sup>	48.45 <sup>a</sup>	8.30 <sup>a</sup>	6.45 <sup>b</sup>	6.40 <sup>b</sup>	21.15 <sup>a</sup>
<i>A.leucophloea</i>	11.40 <sup>a</sup>	7.85 <sup>a</sup>	8.55 <sup>a</sup>	27.80 <sup>b</sup>	4.75 <sup>a</sup>	4.15 <sup>a</sup>	3.65 <sup>b</sup>	12.55 <sup>b</sup>
<i>A.albida</i>	7.05 <sup>a</sup>	3.80 <sup>a</sup>	6.70 <sup>a</sup>	17.55 <sup>d</sup>	3.15 <sup>a</sup>	2.05 <sup>b</sup>	1.95 <sup>b</sup>	7.15 <sup>e</sup>
Control	1.40 <sup>a</sup>	1.00 <sup>a</sup>	1.30 <sup>a</sup>	3.70 <sup>e</sup>	0.18 <sup>a</sup>	0.10 <sup>a</sup>	0.17 <sup>a</sup>	0.45 <sup>f</sup>
<b>SE±</b>	<b>0.67</b>	<b>0.44</b>	<b>0.55</b>	<b>1.66</b>	<b>0.29</b>	<b>0.23</b>	<b>0.21</b>	<b>0.73</b>

\*Means on the same rows having different superscript are significantly different ( $P < 0.05$ ) for Fresh weight and dry weight.

\* Means on the same columns having different superscript are significantly different ( $P < 0.05$ ) for Total fresh weight and Total dry weight.

**Key:** NFAT- Nitrogen Fixing Acacia Trees WAT- Weeks After Transplanting, FW –Fresh Weight, TFW –Total Fresh Weight, DW-Dry Weight -, TDW –Total Dry Weight, L-Leaf, S-Shoot, R –Root.

### Percentage Nutrient Composition of Leaf Litters of Nitrogen Fixing Acacia Trees

Highest percentage values of 2.22 %, 0.90 % and 1.74 % were recorded for nitrogen, phosphorus and

potassium content of *A. senegal*. The least values of 0.04%, 0.23 % and 0.15% were recorded for seedlings planted in soil not influenced with leaf litters of nitrogen fixing acacia trees (Table 7).

**Table 7: Percentage Nutrient Composition of Leaf Litters of Nitrogen Fixing Acacia Trees**

NFAT	Nitrogen (%)	Phosphorus (%)	Potassium (%)
<i>A. seyal</i>	1.89	0.79	1.67
<i>A. nilotica</i>	2.08	0.81	1.73
<i>A. tortilis</i>	1.91	0.80	1.71
<i>A. senegal</i>	2.22	0.90	1.74
<i>A. leucophloea</i>	2.16	0.79	1.72
<i>A. albida</i>	1.98	0.84	1.70
<b>Control</b>	<b>0.04</b>	<b>0.23</b>	<b>0.15</b>

**Key:** NFAT - NFAT- Nitrogen Fixing Acacia Trees

### DISCUSSION

Tallest plant was recorded from seedlings planted in soil amended with *A. senegal*. Similar observation has been reported by Adelani (2019) who recorded highest number of growth parameters for *Citrus tangelo* planted in the soil amended with *Pentaclethra microphylla*. It can be inferred that *A. senegal* contained and released higher number of nitrogen for plant growth compared to other nitrogen fixing tree species investigated. This result is in consonance with the reports of Ahmed *et al.* (2006) and Silvia and Victor (2008). The widest leaf area was recorded from seedlings planted in the soil influenced with *A. senegal*. This result is in line with the reports of Odeyemi *et al.* (2015) and Adelani (2019).

Highest leaf area index was recorded in seedlings planted in the soil amended with *A. senegal*. It can be deduced that *A. senegal* was able to supply nutrients for highest leaf area to cover land area. Similar observation has been reported by Adelani *et al.* (2017a) and Adelani (2019). Highest percentage of nitrogen, phosphorus and potassium were recorded for *A. senegal*. Similar observation has been reported by Adelani *et al.* (2014b) and Adelani *et al.* (2017a).

The excellent performance of *A. senegal* in term of enhancing growth parameters is adduced to functions of its component nitrogen. Nitrogen is most critical element require for plant growth. Nitrogen is the mineral element that plant requires

in larger amounts and is a constituent of many plant cell components, including amino and nucleic acid (Nathan, 2009, Hu and Schmid halter, 2005). It has ability to improve water absorption and protein synthesis and equally promotes cell division as well as elongation of seeds and seedlings. Similar observation has been reported by Adelani *et al* (2017a).

Nitrogen is part of various enzymatic proteins that catalyses and regulates plant-growth process (Sinfield *et al.*, 2010). It is a component of chlorophyll (Anderson, 2015). Nitrogen has been called the growth element because it is a vital part of protoplasm. Protoplasm is the seat of cell division (Abod and Siddiqui, 2002). Improved growth recorded for seedlings planted in the soil amended with *A. senegal* is traceable to its ability to utilize and mobilize metabolites better to growing points.

It can be concluded from this investigation that *A. senegal* was the best source of nitrogen. Various investigations such as Camberato (2001) (Turfgrass), Ohlund and Nasholm (2001) (*Picea abies*), Warren and Adams (2002) (*Pinus pinaster*) and Khamis *et al.*(2013) (*Populus euphratica*) have reported the efficacy of sources of nitrogen in enhancing the growth of plant.

Phosphorus content of *A. senegal* enhanced the growth of *C. albidum* seedlings. Similar observation has been reported by Adelani *et al.* (2017b). Phosphorus is an important component of *Gliricidia sepium* (Adelani *et al.*, 2014b) which help in the germination of seeds (Mengel and Kirkby 2001; Smith, 2014) and seedling growth (Adelani *et al.*, 2014a). Phosphorus is considered a primary nutrient for plant growth (Hinsinger, 2001) and is needed to sustain optimum plant production and quality (Zepata and Zaharah, 2002). The element is essential for cell division, reproduction, and plant metabolism; moreover, its role is related to the acquisition, storage, and use of energy (Epstein and Bloom, 2004).

In addition, phosphorus plays an important role in lateral root morphology (Williamson *et al.*, 2001) and root branching (Lopez-Bucio *et al.*, 2003) and influences not only root development, but also the

availability of nutrients (Jin *et al.*, 2005). Therefore, plants have developed various strategies for obtaining optimum phosphorus from soils, including increases in root surface area, specific root length (SRL), and root-shoot ratio (Tang *et al.*, 2009; Xu *et al.*, 2012). The growth-promoting role of phosphorus application has been reported previously (Williamson *et al.*, 2001, Waraich *et al.*, 2005; Pandey *et al.*, 2006). This is in consonance with the reports of Hudai *et al.* (2007) and Cicek *et al.* (2010).

The present study also confirmed the results of Jin *et al.* (2005) who reported that phosphorus application increased total root length and average root diameter. In most species, phosphorus deficiency results in decreased average root diameter (Hill *et al.*, 2006) however, some species, such as *Arabidopsis thaliana*, developed larger roots in phosphorus deficient conditions (Ma *et al.*, 2001).

Based on the findings in this investigation, *A. senegal* reported highest potassium content which influenced the growth parameters of *C. albidum* seedlings. The essential characteristics of its potassium content accounted for highest growth parameters of *C. albidum* planted in it. Potassium is an essential element that functions in activation of enzymes., (Mengel, 2001; Marschner, 2012), the translocation of photosynthates and the synthesis of cellulose, a building block of every plant cell wall (Anghinoni and Bissani, 2004; Havlin *et al.*, 2005; Sardans and Penuelas, 2005; Wang *et al.*, 2013), transplant of sugars from leaves to fruits, and production and accumulation of oils (Romheld and Kirky, 2010). Some studies have shown that potassium fertilization resulted in a marked improvement in water use efficiency (Ashraf *et al.*, 2001).

Adequate potassium supply is essential to enhancing drought resistance by increasing root elongation and maintaining all membrane stability (Datnoff *et al.*, 2007; Wang *et al.*, 2013). An efficient potassium status may facilitate osmotic adjustment which maintains higher turgor pressure, relative water content and lower osmotic potential, thus improving the ability of plants to tolerate drought stress (Kant and Kafkafi,2002; Egilla *et*

*al.*, 2005). Potassium plays a crucial role in turgor regulation within the guard cell during stomatal movement (Wang *et al.*, 2013). This element improves winter hardiness, the rigidity of young stems and increases diseases resistance. The least growth parameters recorded in seedlings planted in soil not influenced with leaf litters of nitrogen fixing acacia trees could be traced to lack sufficient nutrients as nitrogen, phosphorus and potassium. This result is in consonance with the reports of Adelani *et al.* (2014b).

## REFERENCES

- Abod, S.A., and Siddiqui, M. T. (2002). Growth response of teak to (*Tectona grandis* L.f.) seedlings to nitrogen, phosphorus and potassium fertilizers. *Pertanika Journal of Tropical Agricultural Science* 25 (2):107-113.
- Anderson, P. (2015). N.P.K and Fe, what are they good for? *Tree Service* 1(1):1-4. Accessed on 1/16/2015. [www.Limbwalker](http://www.Limbwalker)
- Anghinoni, I and Bissani, C.A. (2004). Fosforo e adubos fosfatados. In: Bissani, C. A (Ed). *Fertilidade dos solos e manejo de adubacao de culturas* V.I.Porto Alegre:Genesis pp117-137.
- Adelani, D. O., Suleiman, R. A., Akesode, H.A and Akande, M.T. (2014a). Effect of sources and rates of organic fertilizer on the growth of *Chrysophyllum albidum* seedlings. *Organic Agriculture Research: A Catalyst for Sustainable National Agricultural Transformation Agenda*. In: Olabiyi, T.I and Bolarinwa, I.F (Eds). *Proceedings of the 10<sup>th</sup> National Conference on Organic Agriculture* pp 65-73.
- Adelani, D. O., Suleiman, R.A. Aduradola, M. A and Akesode, H.A. (2014b). Assessment of leaf litters of some tree species on growth of *Zea mays* (L) in Northern Guinea Savanna Ecology. *Journal of Organic Agriculture and Environment* 2:117-130.
- Adelani, D. O. (2015). Effect of hydro-priming and potassium nitrate priming on the germination of *Balanites aegyptiaca*. *Applied Tropical Agriculture*, 20 (2): 17-23.
- Adelani, D. O., Aduradola, M. A and Maisamari, I. J. (2016). Storability and pre-sowing treatments of *Chrysophyllum albidum* seeds: A step towards biodiversity conservation. In: Borokini, I.T and Babalola, F.D. (Eds); *MDG<sub>5</sub> to SDG<sub>s</sub>: Towards Sustainable Biodiversity Conservation in Nigeria. Proceedings of Joint Biodiversity Conservation Conference of Nigeria Tropical Biology Association (NTBA) and Nigeria Chapter of Society for Conservation Biology (NSCB) Conference*. Pp 80-86.
- Adelani, D. O and Okechalu, S. O. (2016). Effect of organic seed pelletings and storage periods on the early seedling growth of *Chrysophyllum albidum* seedlings. *Biological and Environmental Sciences Journal for the Tropics* 13 (3): 99 – 109.
- Adelani, D. O., Amos, O.S., Ogunsanwo, J.A. and Peter, S. (2017a). Effect of botanical Pelletings and storage periods on the germination of sweet orange (*Citrus sinensis*) seeds. *Biological and Environmental Sciences Journal for the Tropics*, 14(3): 139-145.
- Adelani, D.O., Amos, O.S and Maikano, S. (2017b). Effect of botanical pelletings and storage periods on the germination of African star apple (*Chrysophyllum albidum* G. Don). *Journal of Agricultural Science and Environment* 17(2): 37-49.
- Adelani, D.O. (2019). Effect of leaf litters of nitrogen fixing trees on the growth of *Citrus tangelo* seedlings. *Sustainable Development Goals through Appropriate Forest Management Strategies*. In: V. A. J. Adekunle, O.Y. Ogunsanwo, N. A. Adewole and P.I. Oni (eds). *Proceedings of 41<sup>st</sup> Annual Conference of Forestry Association of Nigeria held in Abuja, FCT between 7<sup>th</sup>-11<sup>th</sup> October, 2019*. pp1131-1140.
- Aduradola, A.M., Adeola, B. F and Adedire, M.O. (2005). Enhancing germination in seeds of



- African Star Apple, *Chrysophyllum albidum* (G. Don). *Journal of Food, Agriculture and Environment* 3 (2): 292-294.
- Ahmed, A.S., Mohammed, M., Bashir, A., Navar, B and Khan, H. (2006). Effect of various levels of nitrogen, phosphorus and potash on the yield of garlic. *Sarhad Journal of Agriculture* 22 (1): 341-349.
- Akanonu, P. (2019). How big is Nigeria's power demand? Accessed on 28/04/2020. <https://www.energyforgrowth.org/memo/how-big-is-nigerias-power-demand>. Accessed
- Akinyemi, A., and Isiugo-Abanihe, U. C. (2014). Demographic dynamics and development in Nigeria. *African Population Studies*, 27(2): 239–248.
- Ashraf, M., Ahmad, A and Neilly, T.M. (2001). Growth and photosynthetic characteristics in pearl millet under water stress and different potassium supply. *Photosynthetica*, 3: 389-394.
- Bala, A., Murphy, P. J, Osunde, A.O and Giller, K.E. (2003). Nodulation of tree legumes and ecology of their native rhizobial populations in tropical soils. *Applied Soil Ecology*, 22: 211–223.
- Camberato, J.J. (2001). Nitrogen in soil and fertilizers. *Turfgrass Foundation News* 8(1):6-10.
- Cicek, E., Yilmaz, F and Yilmaz, M. (2010). Effect of N and NPK fertilizers on early field performance of narrow-leaved ash, *Fraxinus angustifolia*. *Journal of Environmental Biology*. 31(1–2):109–114.
- Datnoff, L.E., Elmer W and Huber, D.M. (2007). Mineral nutrition and plant disease. *The American Phytopathological Society. St Paul Minnesota, U.S.A.* 278PP. ISBN 978-0-89054-346-7.
- Edward, E., Chamshama, S. A. O and Mugasha, A. G. (2006). Growth performance of lesser-*Leucaena* species/provenances at Gairo inlang plateau, Morogoro, Tanzania. *Southern African Forestry Journal*, 208: 53-62.
- Egilla, J.N., Davies, J.F.T and Boutton, T.W. (2005). Drought stress influences leaf water content photosynthesis and water use efficiency of *Hibiscus Rosa-sinensis* at three potassium concentration. *Photosynthetica*, 43:135-140.
- Ehiagbonare, J. E., Onyibe, H. I. and Okoegwale, E. E. (2008). Studies on the isolation of normal and abnormal seedlings of *Chrysophyllum albidum*. A Step Towards Sustainable Management of the taxon in the 21<sup>st</sup> Century. *Scientific Research and Essays*, 3(12):567-570.
- Epstein, E and Bloom, A. J. (2004). *Mineral nutrition of plants: Principles and perspectives* (Second Edition). Sunderland, M. A: Sinauer Associates, Inc. 402p.
- Farrell, K. (2018). An Inquiry into the Nature and causes of Nigeria's Rapid Urban Forum 29,277-298. <https://doi.org/10.1007/s12132-018-9335-6>
- Favre, L. (2019). 10 Interesting Facts About Nigeria. Africa's largest economy is home of hundreds of languages and a rich film industry. <https://www.usnews.com/news/best-countries/articles/2019-07-02/10-interesting-facts-about-nigeria> Accessed on 28/04/2020.
- Giller, K.E. (2001). *Nitrogen Fixation in Tropical Cropping Systems*, CAB International, Wallingford, UK, 2<sup>nd</sup> edition, 423P
- Havlin, J. L., Tisdale, S. L., Beaton, J.D and Nelson, W.L. (2005). *Soil Fertility and Fertilizers: An Introduction to Nutrient Management*. Pearson Education, Inc, Upper Saddle River, New Jersey 07458. pp 244-254.
- Hill, J.O., Simpson, R. J, Moore, A. D and Chapman, D.F. (2006). Morphology and response of roots of pasture species to phosphorus and nitrogen nutrition. *Plant and Soil*, 286(1–2): 7–19.
- Hinsinger, P. (2001). Bioavailability of soil inorganic P in the rhizosphere as affected by root-induced chemical changes: a review. *Plant and Soil*, 237(2):173–195.
- Hudai, S. M , Sujauddin, M., Shafinat, S and Uddin, M. S. (2007). Effects of phosphorus and potassium addition on growth and nodulation of *Dalbergia sissoo* in the nursery. *Journal of Forestry Research*, 18(4):279–282
- Hu, Y and Schmidhalter, L. E. (2005). Drought and salinity: A comparison of their effect on mineral nutrition of plants. *Journal of Plant Nutrition and Soil Science* 168:541-549.
- ICRAF. (2009). Creating an evergreen Agriculture in Africa for food security and environmental

- resilience, World Agroforestry Centre, Nairobi, Kenya, pp 24, 114pp.
- Jin, J., Wang, G.H., Liu, X., Pan, X and Herbert, S. J. (2005). Phosphorus application affects the soybean root response to water deficit at the initial flowering and full pod stages. *Soil Science and Plant Nutrition*, 51(7):953–960.
- Kant, S and Kafkafi, U. (2002). Potassium and abiotic stresses in plants. In: Pasricha, N.S, Bansal, S.K (eds) Potassium for sustainable crop production. *Potash Institute of India, Gurgaon*, pp 233–251
- Khamis, M.H., Atia, M.G and Ali, H.M. (2013). Impact of nitrogen and phosphorus sources on the growth efficiency of *Melia azedarach* and *Populus euphratica* in Wadi El Natrun, Egypt. *Journal of Forest Products and Industries* 2 (5):13-19.
- López-Bucio, J., Cruz-Ramírez, A and Herrera-Estella, L. (2003). The role of nutrient availability in regulating root architecture. *Current Opinion in Plant Biology*, 6(3):280–287. pmid:12753979
- Ma, Z., Bielenberg, D. G., Brown, K. M and Lynch, J. P. (2001). Regulation of root hair density by phosphorus availability in *Arabidopsis thaliana*. *Plant, Cell and Environment*, 24(4):459–467.
- Marschner, P. (2012). Marschner's Mineral Nutrition of Higher plants, 3<sup>rd</sup> edition, Academic Press: London, UK, PP 178-189.
- Mbwambo, L., Mndolwa, M. A., Gillah, P. R., Balama, C. and Kitojo, D. H. (2006). Some wood properties of five tree species grown under Agroforestry system in Shinyanga and Coast Regions. In: Chamshama, SAO *et al.* (Ed). *Proceedings of the Second National Agroforestry and Environment Workshop: Partnerships and Linkages for Greater Impact in Agroforestry and Environmental Awareness*. Pp 164-170.
- Mengel, K., Kirkby, E. A., Kosegarten, H and Appel, T. (2001). Principles of plant nutrition, vol 5. Springer Netherlands, pp1–849.
- Nathan, M.V. (2009). Soils, Plant Nutrition and Nutrient Management. Master Gardener Core Manual pp 20.
- Nygren, P., Fernández, M. P., Harmand J. M and Leblanc, H. A. (2012). Symbiotic dinitrogen fixation by trees: an underestimated resource in agroforestry systems? *Nutrient Cycling in Agroecosystems*, 94(2/3): 123-160. doi:10.1007/s10705-012-9542-9
- Odeyemi, O.M., Olubode, O.O and Makinde, E. A. (2015). Effect of organic and conventional production systems on nutrient uptake and residual soil nutrient status of pawpaw (*Carica papaya*) orchard. *Journal of Organic Agriculture and Environment* 3: 46-55.
- Ohlund, J and Nasholm, T. (2001). Growth of conifer seedling on organic and inorganic nitrogen sources. *Tree Physiology*, 21:1319-1326.
- Olaoluwa, T. A., Muhammad, N.O and Oladiji, A.T. (2012). Biochemical assessment of the mineral and some antinutritional constituents of *Aspergillus niger* fermented *Chrysophyllum albidum* seed meal. *African Journal of Food Science* 6 (1): 20-28.
- Onyekwelu, J.C. and Stimm, B. (2011). *Chrysophyllum albidum* In: A, Roloff; H, Weisgerber; U, Lang; B, Stimm (Eds): *Enzyklopadia der Holzgewachse*, Wiley-VCH, Weinheim, 59.Erg. Lfg. 10/11, 12PP.
- Onyekwelu, J.C., Stimm, B., Mosandi, R and Olusola, J.A. (2011). Domestication of *Chrysophyllum albidum* from rainforest and derived savannah ecosystem – Phenotype variation and selection of elite trees. *Conference on International Research on Food Security, Natural Resource Management and Rural Development*. Tropentage, pp. 7.
- Otegbeye, G. O., Owonubi, J. J and Oviasuyi, P. K. (2001). Interspecific variation growth Eucalyptus growing in northern Nigeria. In : Popoola. L. Abu. J. E and Oni, P. I (eds). *Proceedings of 27<sup>th</sup> Annual Conference of the Forestry Association of Nigeria* Pp 12-16.
- Oyebamiji, N.A., Jamala, G.Y and Adelani, D.O. (2019). Effects of agroforestry tree biomass and urea on maize tasselling and silking production. *World Scientific News* 120 (2): 250-258.
- Palm, C. A. (1995). Contribution of agroforestry trees to nutrient requirements of intercropped plants. In: Sinclair, F. L. (ed.) *Agroforestry: Science, Policy and Practice*. Kluwer

- Academic Publishers, Dordrecht, pp. 105–124.
- Pandey, S.T., Singh, P and Pandey, P. (2006). Site specific nutrient management for *Withania somnifera* at subtropical belt of Uttaranchal. *International Journal of Agricultural Sciences*, 2:626–628
- Rahaman, O. (2012). A review of medicinal value of *Chrysophyllum albidum* (African star apple). *African Traditional Medicine*. 1(1):1-3 (<http://searchwarp.com/rahaman>).
- Romheld, V and Kirkby, E.A. (2010). Research on potassium in agriculture: needs and prospects. *Plant Soil*, 335:155-180.
- Sardans, J and Penuelas, J. (2005). Drought decreases soil enzymes activity in Mediterranean holm oak forest. *Soil Biology and Biochemistry*, 37: 455-461.
- Sarvade, S., Singh, R., Prasad, H and Prasad, D. (2014). Agroforestry practices for improving soil nutrient status. *Popular Kheti* 2(1): 60-64.
- SAS (2003). Statistical Analysis System. SAS release 9.1 for windows, SAS Institute Inc. Cary, NC, USA.
- Schroth, G and Sinclair, F. L. (2003). *Trees, Crops and Soil Fertility Concepts and Research Methods*. CABI Publishing. UK, USA. Pp. 423.
- Silvia, G and Victor, M. L. (2008). Effect of nitrogen fertilization on yield and colour of red garlic (*Allium sativum* L) cultivars. Instituto Nacional de *Technogra. Experimental Agropecuaria. Meridoza, Argentinian Ciencia Investigation Agraria* 35 (1): 57-64.
- Sinfield, J.V., Fagerman, D and Colic, O. (2010). Evaluation of sensing technologies for on-the-go detection of macro-nutrients in cultivated soil. *Computers and Electronic in Agriculture*, 70:1-18.
- Smith, R.A. (2014). *Plant Nutrients*. 1(1): 1-4. [http://www/plant\\_nutrient.htm](http://www/plant_nutrient.htm). Accessed on 10/03/2015
- Tang, C., Han, X. Z., Qiao, Y. F and Zheng, S. J. (2009). Phosphorus deficiency does not enhance proton release by roots of soybean [*Glycine max* (L.) Murr.]. *Environmental and Experimental Botany*, 67(1): 228–234.
- Tchounjeu, Z., Kengue, J and Leakey, R.R.B. (2002). Domestication of *Dacryodes edulis*: state-of –the-art. *Forests, Trees and livelihoods*, 12:3-13.
- Ugese, F. D., Baiyeri, K. P and Mbah, B.N. (2008). Leaf area determination of Shea butter tree (*Vitellaria paradoxa* C.F Gaertn). *International Agrophysics*, 22:167-170.
- Wang, M., Zheng, Q., Shen, Q and Guo, S. (2013). The critical role of potassium in plant stress response. *International Journal of Molecular Science*, 14:7370-7390.
- Waraich, E. A., Ahmad, Z., Ahmad, R., Saifullah, M and Ashraf, M.Y. (2015). Foliar applied phosphorous enhanced growth, chlorophyll contents, gas exchange attributes and PUE in wheat (*Triticum aestivum* L.). *Journal of Plant Nutrition*, 38(12):1929–1943.
- Warren, C.R and Adams, M.A. (2002). Phosphorus effects, growth and partitioning of nitrogen to Rubisco in *Pinus pinaster*. *Tree Physiology* 22:11-19.
- Williamson, L., Ribrioux, S., Fitter, A and Leyser, O. (2001). Phosphate availability regulates root system architecture in Arabidopsis. *Plant Physiology*, 126 (2):875–882.
- World Agroforestry Centre. (2018). Interaction between trees and crops. Agroforestry extension manual for Kenya <http://www.worldagroforestry.org/Units/Library/Books/>. Accessed on 24/07/2018. Pp6.
- World Health Organization (2020). Maternal Health in Nigeria: Generating Information for action. [www.who.int/reproductivehealth/maternal-health-nigeria/accessed\\_on\\_28/04/2020](http://www.who.int/reproductivehealth/maternal-health-nigeria/accessed_on_28/04/2020). pp4.
- Wole, O. (2013). Unlimited nutritional benefits of African star apple. *Natural Health* 1(1):1-4.
- Xu, B.C., Niu, F.R., Duan, D. P, Xu, W. Z and Huang, J. (2012) Root morphological characteristics of *Lespedeza davorica* (L.) intercropped with *Bothriochloais chaemum* (L.) Keng under water stress and P application conditions. *Pakistan Journal of Botany*, 44(6):1857–1864.
- Zapata, F and Zaharah, A. R. (2002). Phosphate availability from phosphate rock and sewage sludge as influenced by addition of water soluble phosphate fertilizers. *Nutrient Cycling in Agroecosystems*, (1) 63:43–48.