JOPAT Vol 23(2), 1619- 1569, July – December, 2024 Edition. ISSN2636 – 5448 https://dx.doi.org/10.4314/jopat.v23i2.18

Antioxidant activity of *Brachiaria sabia* and *Brachiaria marandu* and Molecular Docking of Constituents targeting Insulin-like Growth Factor 1 Receptor (IGF-1R) for cancer treatment

Ogunlakin Daniel Akingbolabo¹*, Peluola Olujide Ayeni¹*, Adeshina Isaiah Odugbemi¹, Oluwafemi Adeleke Ojo¹, Mojisola Adebimpe Ayomipo¹, Godwin A. Berena¹, Paula Ishola¹, Peter Adeyemi¹, Dorcas Sunday¹, Emmanuel Henry Ezenabor¹, Kevwe Benefit Esievo², Ademola Olabode Ayeleso^{1,3}, Oluyomi S. Adeyemi¹, Gbadebo Emmanuel Adeleke⁴, Oluwatosin A. Adaramoye⁵

¹Biochemistry Programme, Bowen University, Iwo. Osun State, Nigeria.

²Department of Medicinal Plant research and Traditional Medicine, National Institute for Pharmaceutical Research and Development (NIPRD), Abuja, Nigeria.

³ Department of Life and Consumer Sciences, School of Agriculture and Life Sciences, University of South Africa, Roodepoort, South Africa

⁴ Department of Biochemistry, Ladoke Akintola University of Technology, Ogbomosho, Nigeria.

⁵ Department of Biochemistry, University of Ibadan, Ibadan, Nigeria.

*Corresponding authors: gbolaogunlakin@gmail.com; peluola.ayeni@bowen.edu.ng.

Abstract

The methanol extracts of *Brachiaria sabia*, and *Brachiaria marandu* leaves were evaluated for its ability to scavenge free radicals, reduce ferric iron, and function as an iron chelator. The extracts were subjected to HPLC analysis. The two *Brachiaria* species possessed strong antioxidant activity, with *B. marandu* had a higher percentage (%) of phenolic content than *Brachiaria gayana*. *In silico* studies revealed that the two *Brachiaria* grasses tested enhanced insulin-like growth receptor factor-1 receptors (IGF-1R). Quercetin, which is consistent compound in the two grasses, had highest binding affinity to insulin-like growth factor-1 receptors with value of -40.5971. This study therefore reveals that *Brachiaria sabia*, and *Brachiaria marandu* are good antioxidant and Quercetin, a constituent of *Brachiaria* grasses, binds to insulin-like growth factor-1 receptors (IGF-1R) which plays an important role in cancer cell proliferation. *In vivo* and *in vitro* research are also necessary to validate this in silico result.

Keywords: Brachiaria sabia, Brachiaria marandu, Cancer, Antioxidants, Insulin-like growth factor-1 receptors

Introduction

An estimated 11.5 million deaths from cancer are predicted by 2030, making it the second greatest cause of mortality worldwide [1]. Based on research conducted by Bray et al. [2], men are more likely than women to get colorectal, lung, liver, prostate, stomach, and cervical cancers among the 36 distinct forms of cancer. Some conventional and modern techniques used to treat cancer include radiation treatment. chemotherapy, and surgery Numerous disadvantages of these [3]. techniques include toxicity and adverse reactions associated with the use of conventional chemicals in cancer treatment [4]. The discovery of novel, efficient drugs with minimal side effects is necessary for the prevention and treatment of this illness due to inefficiency of conventional the chemotherapeutic approaches [5].

©2024 The authors. This work is licensed under the Creative Attribution 4.0 International license.

The tyrosine kinase receptor known as the insulin-like growth factor-1 receptor (IGF-1R) believed to have an impact on the is development of several malignancies [6]. Through this receptor, the insulin-like polypeptide protein hormone IGF-1 mediates its activities [7] IGF-1 is essential for development and continues to have anabolic effects throughout adulthood that extend to the control of cancer and free radicals. It has been discovered that IGF-IR is markedly overexpressed in a large number of human solid tumours, including sarcomas, hepatocellular pancreatic, carcinoma, ovarian, and gastrointestinal cancers, as well as breast, nonsmall cell lung, and prostate cancer [8-11]. After ligand binding, IGF-1R can initiate the Ras/Raf/MEK/MAPK and PI3K/AKT/mTOR signalling pathways. These pathways then activate several transcription factors, including AP-1, CREB, and ELK-1, to affect angiogenesis, invasion, motility, and cell proliferation [2,13]. Furthermore, accumulating evidence indicates that IGF-1R is essential for tumour growth and plays a role in the critical stages of the metastatic cascade [14,15]. Thus, one of the most sought-after targets for cancer therapy strategies is IGF-1R [16,17].

Antioxidants are abundant in botanicals. Antioxidants are substances that can prevent or lessen the harm that these unstable chemicals do to cells. In addition, antioxidants act as synergists, metal-chelating agents, hydrogen donors, electron donors, peroxide decomposers, enzyme inhibitors, and radical scavengers. Brachiaria sabia and Brachiaria marandu was shown to contain alkaloid [18]. These grasses showed moderate levels of terpenoids and flavonoids. Tannins and saponins, however, were absent. This outcome is consistent with a study conducted by Ogunlakin et al. [19], which found that alkaloids, terpenoids, and flavonoids found in Brachiaria grasses may be advantageous to the growth and production of animals who eat them. Brachiaria marandu had the highest total phenolic content when compared to Brachiaria sabia [18]. The aim of this study is to evaluate the antioxidant and anticancer activity of Brachiaria sabia and Brachiaria marandu via Molecular Docking of Constituents targeting Insulin Growth Factor 1 Receptor (IGF-1R).

2. Materials and methods

2.1. Plants collection and Extraction

On May 31, 2022, *Brachiaria sabia* and *Brachiaria marandu* were collected from the dairy farm at Bowen University in Iwo, Osun state. The grasses were prepared for extraction by being air dried, pulverized into a powder using an electric grinder, weighed, and kept in dry, clean beakers. 500 g of each powdered sample were extracted with methanol. At 40°C, concentrated filtrates were concentrated in a vacuum using a Stuart-RE300DB rotary evaporator. The extracts were stored in the refrigerator for further use.

2.2. Antioxidant parameters

2.2.1 Iron-chelation antioxidant assay

The ability of the methanolic extracts of *Brachiaria sabia* and *Brachiaria marandu* to chelate ferrous ions was evaluated using a slightly modified procedure of the Ajiboye et al. [20]. This test evaluates the capacity to chelate ferrous ions by stopping Fe^{2+} from binding to ferrozine and forming a vividly colored complex.

2.2.2 OH radical scavenging ability

The ability of the methanolic extracts of *Brachiaria sabia* and *Brachiaria marandu* to scavenge hydroxyl radicals was assessed using the methods reported by Ogunlakin *et al.* [21], with a few minor modifications.

2.2.3 Ferric-reducing antioxidant power (FRAP)

The ferric-reducing antioxidant capability was determined by measuring the capacity of the methanolic extract of *Brachiaria sabia* and *Brachiaria marandu* to change ferric ions (Fe^{2+}) into ferrous ions (Fe^{3+}) . The initial plan for this surgery was changed. It was said to be the only assay that measures antioxidants (or reductants) in a sample directly, in contrast to other methods assessing the suppression of free radicals [21].

2.2.4 DPPH radical scavenging activity

By slightly modifying the procedure outlined by Ogunlakin *et al.* [21], the DPPH radical was used to assess the free radical scavenging

ability of the methanol extract of *Brachiaria* sabia and *Brachiaria marandu*.

High-Performance Liquid Chromatography (HPLC) analysis

Using Ayeni et al.'s [18] technique, High-Performance Liquid Chromatography (HPLC) analysis was performed to determine the phytochemicals present in the methanol extracts of *Brachiaria sabia* and *Brachiaria marandu*. After a few adjustments, the composition gradient looked like this: 5% of methanol (B) for the first two minutes, after which the percentage was adjusted to produce different values between 10 and 60 minutes, spaced out by ten minutes. Water with 2% acetic acid (A) and methanol (B) was the mobile phase.

2.5 In silico studies

2.5.1 Ligand preparation

To prepare ligands, Schrodinger Suite 2020-4's LigPrep module was utilized. The force field known as OPLS_2005 (Optimised Potentials Liquid Simulations) for was utilized to minimize energy. LigPrep converted 2D structures into 3D structures by including hydrogens, accounting for bond lengths and angles. and selecting the conformers' showed the structure that lowest conformational energy. Accurate tautomers, stereochemistry, chirality, and ring conformations are prerequisites for these steps. Using the Epik ionization tool, the ionization state was established at a pH range of 7.0 ± 2.0 [22].

2.5.2 Target structure preparation

In this study, the insulin-like growth factor-1 receptor's crystal structure in the selected Brachiaria species was recovered using the protein data bank (PDB) 7E3H [23]. Using the Maestro v12.6 protein preparation wizard (part of the Schrodinger Suite 2020-4), the protein was made, and the bond order was modified. Furthermore, PROPKA provided the hydrogen atoms at pH 7.0 following the removal of the crystallographic water molecules. Docking was done with the OPLS 2005 force field, constrained minimization with an RMSD of 0.30 being the default, and Epik was also used to construct het states [22].

2.5.3 Receptor grid generation

By enclosing a box around the centroid of the co-crystallized ligands associated with each receptor (the binding pocket), the receptor grid generation module built the grid boxes for each receptor. This technique ensures the docked ligands stay inside the enclosing box by blocking non-specific binding. The co-crystallized ligands of 7E3H were selected to define the grid box to preserve the center of each docked ligand with the same dimensions as the binding pocket [24].

2.5.4 Molecular docking

The prepared and minimized compounds were docked into the receptor's binding region in this work using the XP (Extra-Precision algorithm) docking technique, and the interactions between the ligands and receptor were analyzed. The XP views the receptor as stiff and the ligand sampling as flexible. The OPLS_2005 force field was used in the docking calculations. The 2D interactions of the complexes were investigated using the Schrodinger suite's ligand interaction diagram module [25].

3. Results

3.1 Antioxidant activity and HPLC analysis

Ascorbic acid was utilized as the control in Figure 1, however, Brachiaria sabia and Brachiaria marandu showed increased capacity to scavenge DPPH. Figure 2 shows Brachiaria marandu has a higher ferricreducing antioxidant capacity than Brachiaria sabia. Both plant extracts reduced ferric ions than the control (ascorbic acid). Brachiaria sabia has a greater capacity to scavenge hydroxyl radicals than Brachiaria marandu, however, it still has lower values than the ascorbic acid standard (Figure 3). The concentration-dependent increase in chelating iron was observed in the methanol extracts of Brachiaria sabia and Brachiaria marandu. However, the iron chelating capability of these plants was lower than the benchmark, ascorbic acid (Figure 4). The HPLC phytochemical analyses of B. sabia and B. marandu are shown in Figures 5 and 6, which indicate that B. marandu has more embedded phenolic

compounds than *B. sabia*. Tables 1 and 2 display the percentage (%) content of phenolic compounds for the two Brachiaria grasses. The

results indicate that *B. marandu* had a higher percentage (%) of phenolic content than the other grass.

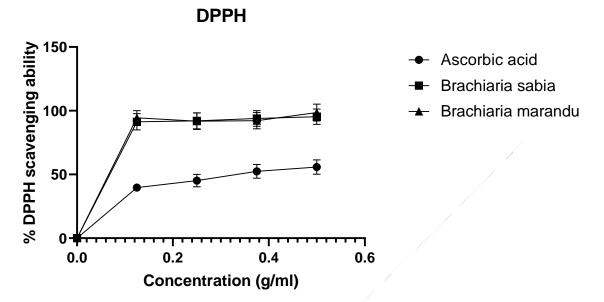


Figure 1: Percentage DPPH scavenging ability of Brachiaria sabia and Brachiaria marandu

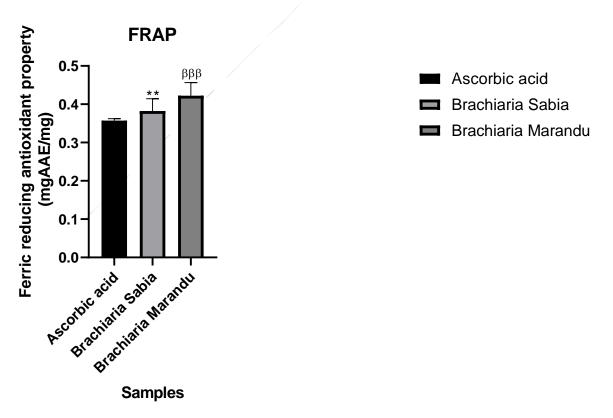


Figure 2: Ferric reducing property of Brachiaria sabia and Brachiaria marandu

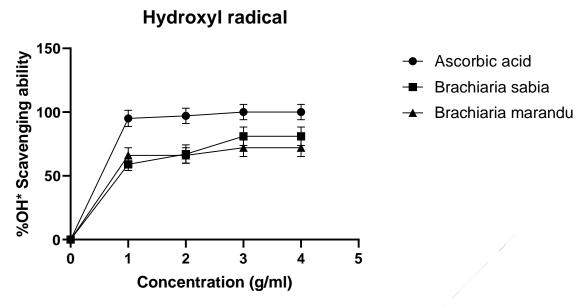


Figure 3: Percentage hydroxyl radical scavenging ability of Brachiaria sabia and Brachiaria marandu.

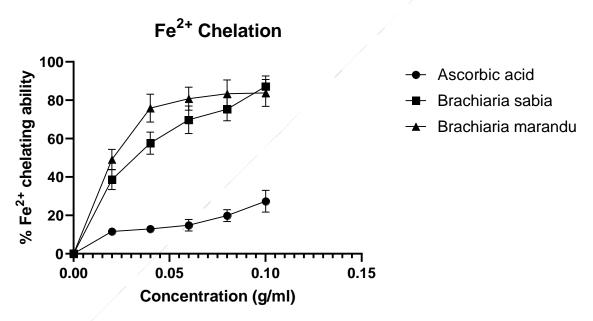


Figure 4: Percentage Iron chelating ability in Brachiaria sabia and Brachiaria marandu.

Ogunlakin et al.

Journal of Phytomedicine and Therapeutics

www.niprdjopat.gov.net; niprdjopat@qmail.com

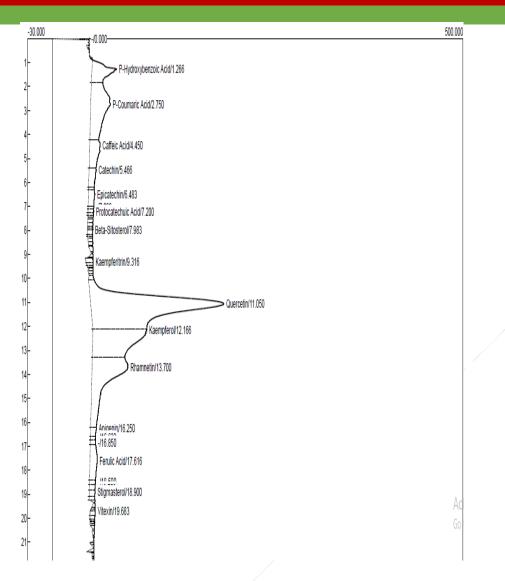


Figure 5: Phytochemical screening of Brachiaria sabia

Phenolic Compounds	% Content	
Para-hydroxy-benzoic acid	4.26	
Para-hydroxy- coumaric acid	10.64	
Caffeic acid	3.39	
Catechin	1.61	
Epicatechin	1.20	
Protocatechuic acid	0.26	
Kaempferitin	0.42	
Quercetin	42.13	
Kaempferol	16.23	
Rhamnetin	14.93	
Apigenin	0.62	
Ferulic acid	3.12	
Stigmastrerol	0.48	
Vitexin	0.33	

Table 1: Phenolic compounds and % content of Brachiaria sabia

Ogunlakin et al.

Journal of Phytomedicine and Therapeutics

www.niprdjopat.gov.net; niprdjopat@gmail.com

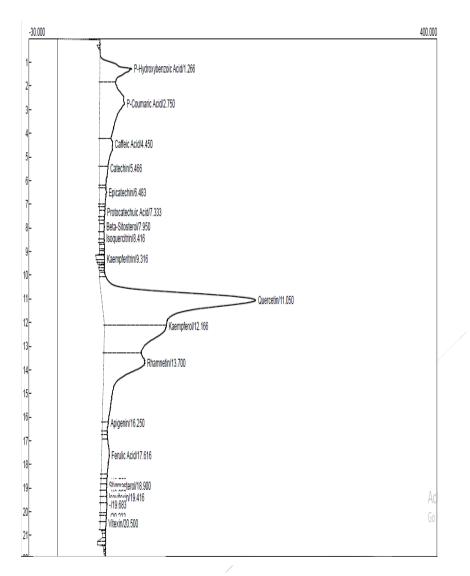


Figure 6: Phytochemical screening of Brachiaria marandu

Table 2: Phenolic compounds and % content of Brachiaria marandu

Phenolic Compounds	% Content
Para-hydroxy-benzoic acid	5.07
Para-hydroxy- coumaric acid	11.56
Caffeic acid	3.56
Catechin	1.62
Epicatechin	1.12
Protocatechuic acid	0.38
Protocatechuic acid	0.36
Kaempferitin	0.41
Quercetin	40.92
Kaempferol	15.17
Rhamnetin	14.18
Apigenin	0.54

www.niprdjopat.gov.net; niprdjopat@gmail.com

Ferulic acid	2.75
Stigmastrerol	0.41
Isovitexin	0.44
Vitexin	0.50

3.2 In silico study

Brachiaria sabia has a greater capacity to scavenge hydroxyl radicals than *Brachiaria marandu*, however, it still has lower values than the ascorbic acid standard (Figure 3). The concentration-dependent increase in chelating iron was observed in the methanol extracts of *Brachiaria sabia* and *Brachiaria marandu*. However, the iron chelating capability of these plants was lower than the benchmark, ascorbic acid (Figure 4). Figure 6 shows the structure of insulin-like growth factor-1 receptors. Table 6 displays the docking scores and MM-GBSA post-grid docking. Quercetin, had the greatest docking score, followed by Kaempferol. It was discovered that in comparison to other bioactive compounds, quercetin, kaempferol, and rhamnetin showed significant binding affinities to insulin-like growth factor-1 receptors (Figure 8).

Table 3: Consistent compounds in B. sabia and B. marandu

Molecule	Name	Smile
Compound 1	Para-hydroxy-benzoic acid	C1=CC(=CC=C1C(=O)O)O
Compound 2	Para-hydroxy-coumaric acid	C1C=C(C=CC1(O)O)/C=C/C(=O)O
Compound 3	Caffeic acid	C1=CC(=C(C=C1/C=C/C(=O)O)O)O
Compound 4	Quercetin	C1=CC(=C(C=C1C2=C(C(=O)C3=C(C=C(C=C3
		02)0)0)0)0)0
Compound 5	Kaempferol	C1=CC(=CC=C1C2=C(C(=O)C3=C(C=C(C=C3O
		2)0)0)0)0
Compound 6	Rhamnetin	COC1=CC(=C2C(=C1)OC(=C(C2=O)O)C3=CC(=
		C(C=C3)O)O)O
Compound 7	Ferulic acid	COC1=C(C=CC(=C1)/C=C/C(=O)O)O

TARGETSWISS-MODEL: Q05688 (IGF1R_BOVIN) Bos taurus (Bovine)

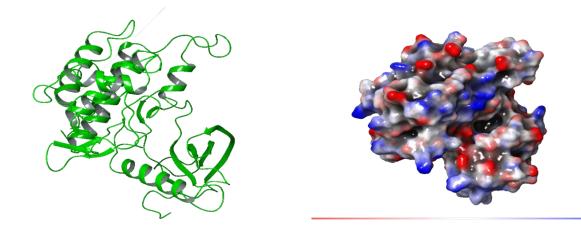
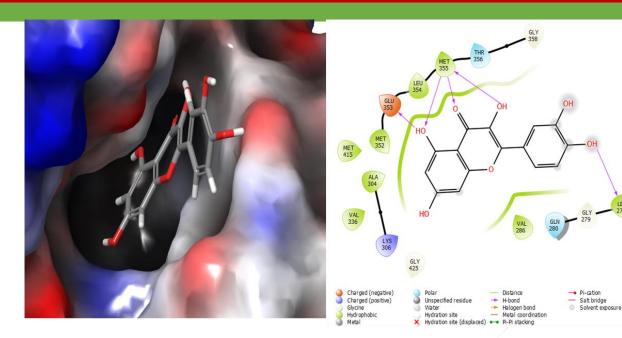


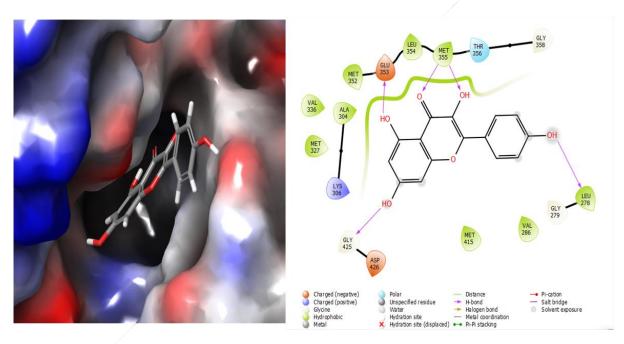
Figure 7: Structure of Insulin-like Growth Factor -1 Receptor

Ogunlakin et al.

<u>www.niprdjopat.gov.net; niprdjopat@gmail.com</u>



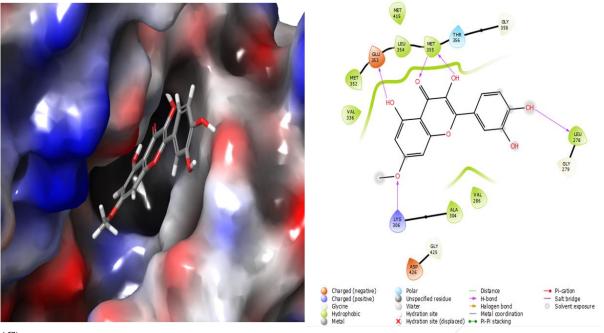
(A)



(B)

Ogunlakin et al.

www.niprdjopat.gov.net; niprdjopat@gmail.com



(C)

Figure 8: Intermolecular interactions of (A) quercetin, (B) kaempferol, and (C) rhamnetin with insulinlike growth factor-1-receptor

Ogunlakin et al.

www.niprdjopat.gov.net; niprdjopat@qmail.com

 Table 4: Docking score and MM-GBSA post rigid docking

Name	Para-hydroxy- benzoic acid (1)	Para-hydroxy-coumaric acid (2)	Caffeic acid (3)	Quercetin (4)	Kaempferol (5)	Rhamnetin (6)	Ferulic acid (7)
Docking Score	-5.142	-4.922	-6.395	-8.425	-8.84	-9.385	-4.774
$MMGBSA_\Delta G_Bind$	-16.3516	-21.5461	-24.6631	-42.811	-39.3028	-40.5971	-29.9584
MMGBSA_AG_Bind Coulomb	80.96294	87.9359	77.43172	-30.0433	-22.2266	-25.3496	75.19584
MMGBSA ΔG Bind Covalent	0.365253	0.707484	1.09186	5.586899	2.900419	6.324156	-3.07835
MMGBSA ΔG Bind HBond	-1.89688	-0.52778	-1.24349	-2.52237	-2.71795	-2.64201	-1.14783
MMGBSA ΔG Bind Lipo	-7.46844	-8.14198	-9.76507	-7.00138	-8.09104	-8.36927	-11.3458
MMGBSA ΔG Bind Solv GB	-70.2267	-73.5759	-69.4857	21.62977	22.12441	22.91623	-66.81
$\underline{MMGBSA} \Delta \mathbf{G} \overline{Bind} \mathrm{vdW}$	-18.0877	-27.9438	-22.6924	-30.4606	-31.2921	-33.4767	-22.7722

4. Discussion

There have been reports that some flavonoids can stop the growth of malignant cells [19,26]. Flavonoids are an important family of natural substances found in fruits, vegetables, and many plants. They are members of a group of secondary plant metabolites that have a polyphenolic structure. Their diverse metabolic and antioxidant characteristics have been associated with an array of medical conditions [27]. Research has demonstrated that Brachiaria grasses are rich in flavonoids [18]. According to Mohammed et al. [28], the concentration of flavonoids and phenolics may vary depending on how polar the extraction solvents are. This result is in line with our previous study [18] which concluded that Brachiaria grasses have an acceptable phenolic content. Studies show that the amount of phenol in a plant directly influences its antioxidant content. Phenolic compounds can decrease, donate hydrogen, and scavenge free radicals [29]. Brachiaria grasses contain substantial levels of phenolics, which may play a major role in their antioxidant properties.

Phytochemical screening was conducted utilising High-Performance the Liquid Chromatography (HPLC) method in order to examine the three grasses that were collected quantitatively. These findings showed that all three grasses have high concentrations of antioxidants such as phenolic acids and flavonoids. Compared to Brachiaria gayana and Brachiaria sabia, Brachiaria marandu has been found to have the greatest amount of bioactive chemicals by HPLC analysis. Brachiaria marandu thus contained a higher concentration of antioxidants. The results of this study are consistent with those of Boudalia et al. [30], who found that Brachiara grasses contain a variety of bioactive compounds that might enhance animal productivity.

Numerous investigations conducted over the past 20 years have indicated that this receptor plays a part in cell transformation, cancer cell growth, and metastatic processes [31,32]. Numerous studies have demonstrated a connection between this signalling system and the risk of getting cancer, even though no

recurring cancer-specific mutations of the IGF-1R or its ligands have been identified to date [33,34]. The most frequent observations linked to IGF-1R overexpression or the formation of autocrine or paracrine signalling loops are related to dysregulated IGF signalling. Carcinogenic signalling loops are more prevalent and have been linked to a wide range of human cancers, whereas elevated expression levels of IGF-1R have been linked to colorectal and breast cancers. Paracrine signaling has mainly been described for breast cancer, where stromal cells have been shown to produce IGF-1 and IGF-2. Population studies have further highlighted the importance of IGF signaling in some of the most common cancers [35-38]. Elevated IGF-1 levels have been linked to an increased risk of cancer diagnosis, according to published evidence from epidemiological research [39,40]. Systematic evaluations of this data led to the inference that circulating IGF-1 levels are, in fact, associated with a risk of various common malignancies, even though the population studies did not always reach the same conclusions [41-43]. Prostate cancer, premenopausal breast cancer, and colorectal cancer were identified as having the strongest correlations between elevated levels of IGF-1 and the likelihood of receiving a cancer diagnosis [44-46]. Noteworthy, however, is that a major, comprehensive investigation identified no significant overall links between common germline variation in IGF1 and other genes implicated in IGF-1 metabolism and breast cancer [47-49].

IGF signalling and the IGF-1R have been implicated in human malignancies, according to a considerable body of research from mechanistic and epidemiological investigations. New therapeutic techniques that can be used with existing traditional therapy regimens have shown promise in targeting the IGF 1R. The findings indicate that seven different chemicals were consistently found in all three of the examined grasses: parahydroxy-benzoic acid, para-hydroxy-coumaric acid, caffeic acid, quercetin, kaempferol, and ferulic acid. The molecule that is most prevalent and constant in the two grasses, quercetin (-40.5971), had the highest binding affinity to the insulin-like growth factor-1 receptor. This is

seen by their relative extent of negative values, which indicate that, in comparison to other substances, they both require the least amount of energy to bind insulin-like growth factor-1 receptors. This work is consistent with that of Chen et al. [50] and Nagini *et al.* [51], which reported that flavonoids and phenolic acids exhibited a hydrophobic interaction with the intracellular structure of IGF-1 receptors.

5. Conclusion

Brachiaria marandu and sabia are excellent antioxidants. The existence of phenolic compounds has led to these results, which demonstrate strong antioxidant action. The most prevalent and reliable component in both grasses, quercetin, exhibited the highest binding affinity to insulin-like growth factor-1 receptors. Accordingly, this study shows that Quercetin, a constituent of Brachiaria grasses, binds to insulin-like growth factor-1 receptors (IGF-1R) which plays an important role in cancer cell proliferation. *In vivo* and *In vitro* research are also necessary to validate this in silico result.

Funding: Not applicable.

Conflict of Interest: Not applicable.

Ethical approval: Not applicable.

Informed consent: Not applicable.

References

- [1] Stukalin I, Ahmed NS, Fundytus AM, Qian AS, Coward S, Kaplan GG, Hilsden RJ, Burak KW, Lee JK, Singh S, Ma C. (2022). Trends and projections in national United States health care spending for gastrointestinal malignancies (1996–2030). Gastroenterology. 162(4):1098-110.
- [2] Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide

for 36 cancers in 185 countries. CA: a cancer journal for clinicians. 68(6):394-424.

- [3] Ndlovu NL, Mdlalose WB, Ntsendwana B, Moyo T. (2024).
 Evaluation of Advanced Nanomaterials for Cancer Diagnosis and Treatment. Pharmaceutics. 16(4):473.
- [4] Dutta Gupta Y, Mackeyev Y, Krishnan S, Bhandary S. (2024). Mesoporous silica nanotechnology: promising advances in augmenting cancer theranostics. Cancer Nanotechnology. 15(1):9.
- [5] Dehelean CA, Marcovici I, Soica C, Mioc M, Coricovac D, Iurciuc S, Cretu OM, Pinzaru I. (2021). Plantderived anticancer compounds as new perspectives in drug discovery and alternative therapy. Molecules. 26(4):1109.
- [6] Peng Y, Tan J. (2023). The Relationship between IGF Pathway and Acquired Resistance to Tyrosine Kinase Inhibitors in Cancer Therapy. Frontiers in Bioscience-Landmark. 28(8):163.
- [7] Al-Samerria S, Radovick S. (2021) The role of insulin-like growth factor-1 (IGF-1) in the control of neuroendocrine regulation of growth. Cells. 10(10):2664
- [8] Lv QL, Huang YT, Wang GH, Liu YL, Huang J, Qu Q, Sun B, Hu L, Cheng L, Chen SH, Zhou HH. (2016). Overexpression of RACK1 promotes metastasis by enhancing epithelial-mesenchymal transition and predicts poor prognosis in human glioma. International journal of environmental research and public health. 13(10):1021.
- [9] Holly JM, Biernacka K, Perks CM. (2019). The neglected insulin: IGF-II, a metabolic regulator with

www.niprdjopat.gov.net; niprdjopat@gmail.com

implications for diabetes, obesity, and cancer. Cells. 8(10):1207.

- [10] Fabrizio P, Alcolei A, Solari
 F (2024). Considering
 Caenorhabditis elegans Aging on a
 Temporal and Tissue Scale: The
 Case of Insulin/IGF-1 Signaling.
 Cells. 13(3):288.
- [11] Wang SS. (2024). Advancing biomarker development for diagnostics and therapeutics using solid tumour cancer stem cell models. Tumori Journal. 110(1):10-24.
- [12] Tian X, Wang J, Jiang L, Jiang Y, Xu J, Feng X. (2022). Chemokine/GPCR signalingmediated EMT in cancer metastasis. Journal of Oncology. 2022(1): 2208176..
- [13] Aung TH, Shivamallu C, Prasad S. (2023). In silico screening and molecular docking of spiroindimicin AH targeting Insulin Growth Factor 1 Receptor (IGF-1R) for cancer treatment. Drug Discovery. 17:e20dd1923.
- [14] Liu G, Zhu M, Zhang M, Pan F. (2023). Emerging role of IGF-1 in prostate cancer: a promising biomarker and therapeutic target. Cancers. 15(4):1287.
- [15] Werner H. (2023). The IGF1 signaling pathway: from basic concepts to therapeutic opportunities. International journal of molecular sciences. 24(19):14882.
- [16] Patel M, Eckburg A, Gantiwala S, Hart Z, Dein J, Lam K, Puri N. (2021). Resistance to molecularly targeted therapies in melanoma. Cancers. 13(5):1115.
- [17] Romano S, Nele V, Campani V, De Rosa G, Cinti S.

(2024). A comprehensive guide to extract information from extracellular vesicles: a tutorial review towards novel analytical developments. Analytica Chimica Acta. 2024:342473.

- [18] Ayeni PO, Obafemi BA, Adeleke GE, Ogunlakin AD, Odugbemi AI, Ayeleso AO, Ojo OA. (2023). Phytochemical analysis of Brachiaria grasses from Bowen University dairy farm and molecular docking of constituents with insulin-like growth factor binding protein. Informatics in Medicine Unlocked. 43:101386.
- Ogunlakin AD, Ojo OA, [19] Gyebi GA, Akinwumi IA. Adebodun GO. Avokunle DI. Ambali OA, Ayeni PO, Awosola OE, Babatunde DE, Akintunde EA. (2023).Elemental evaluation. nutritional analysis, GC-MS analysis and ameliorative effects of Artocarpus communis JR Forst. & G. Forst. seeds' phytoconstituents on metabolic syndrome via in silico approach. Journal of Biomolecular Structure and Dynamics. 2023:1-21.
- [20] Ajiboye BO, Ojo OA, Oyinloye BE, Akuboh O, Okesola MA, Idowu O, Kappo AP. (2020). In vitro antioxidant and inhibitory of polyphenolic-rich activities extracts of Syzygium cumini (Linn) Skeels leaf on two important enzymes relevant to type II diabetes mellitus. Pak Journal of Pharmaceutical Sciences. 33(2):523-9
- [21] Ogunlakin AD, Ojo OA, Osagie PO, Ubogu O, Adegoke AA, Ogunlakin OA. (2024). Effect of Pentaclethra macrophyla Benth. Leaf on expression of P53, IRS,

HsD17β2, FTO, and CYP11a genes in letrozole-induced polycystic ovarian syndrome rats. Comparative Clinical Pathology. 2024:1-2.

- [22] Du J, Sun H, Xi L, Li J, Yang Y, Liu H, Yao X. (2011). Molecular modeling study of checkpoint kinase 1 inhibitors by multiple docking strategies and prime/MM–GBSA calculation. Journal of computational chemistry. 32(13):2800-9.
- [23] Prakash P, Gayathiri E, Periyasami Rahaman Μ, G, Pandiaraj S, Pratheep T, Selvam K, Chaudhari SY, Thirumalaivasan N, Thomas J, Hatami M. (2023.Exploring the potential of targeting insulin-like growth factor-1 through network pharmacology, molecular docking, molecular dynamics, and experimental validation of antioxidant and anti-inflammatory activities. South African Journal of Botany. 162:707-18.
- [24] Prabhu S, Vijayakumar S, Manogar P, Maniam GP, Govindan N. (2017). Homology modeling and molecular docking studies on type II diabetes complications reduced PPARγ receptor with various ligand molecules. Biomedicine & Pharmacotherapy. 92:528-35.
- Reddy SV, Reddy KT, [25] Kumari VV, Basha SH. (2015). Molecular docking and dynamic simulation studies evidenced plausible immunotherapeutic anticancer property by Withaferin A targeting indoleamine 2. 3dioxygenase. Journal of Biomolecular Structure and Dynamics. 33(12):2695-709.
- [26] Forni C, Rossi M, Borromeo I, Feriotto G, Platamone G, Tabolacci C, Mischiati C, Beninati

S. (2021). Flavonoids: A myth or a reality for cancer therapy?. Molecules. 26(12):3583.

- [27] Kumar S, Chhabra V, Shenoy S, Daksh R, Ravichandiran V, Swamy RS, Kumar N. (2024). Role of flavonoids in modulation of mitochondria dynamics during oxidative stress. Mini Reviews in Medicinal Chemistry. 24(9):908-19.
- [28] Mohammed EA, Abdalla IG, Alfawaz MA, Mohammed MA, Al Maiman SA, Osman MA, Yagoub AE, Hassan AB. (2022). Effects of extraction solvents on the total phenolic content, total flavonoid content, and antioxidant activity in the aerial part of root vegetables. Agriculture. 12(11):1820.
- [29] Lang Y, Gao N, Zang Z, Meng X, Lin Y, Yang S, Yang Y, Jin Z, Li B. (2024). Classification and antioxidant assays of polyphenols: A review. Journal of Future Foods. 4(3):193-204.
- [30] Boudalia S, Smeti S, Dawit M, Senbeta EK, Gueroui Y, Dotas V, Bousbia A, Symeon GK. (2024). Alternative Approaches to Feeding Small Ruminants and Their Potential Benefits. Animals. 14(6):904.
- [31] Su C, Mo J, Dong S, Liao Z, Zhang B, Zhu P. (2024). Integrinβdisorders and cancers: 1 in molecular mechanisms and therapeutic Cell targets. Communication Signaling. and 22(1):71.
- [32] Liu H, Tang L, Li Y, Xie W, Zhang L, Tang H, Xiao T, Yang H, Gu W, Wang H, Chen P. (2024). Nasopharyngeal carcinoma: current views on the tumor microenvironment's impact on drug

resistance and clinical outcomes. Molecular Cancer. 23(1):20.

- [33] Solarek W, Czarnecka AM, Escudier B, Bielecka ZF, Lian F, Szczylik C. (2015). Insulin and IGFs in renal cancer risk and progression. Endocr Relat Cancer. 22(5): R253-64.
- [34] Szablewski L. (2024). Insulin Resistance: The Increased Risk of Cancers. Current Oncology. 31(2):998-1027.
- [35] Gallagher EJ, LeRoith D. (2011). Minireview: IGF, insulin, and cancer. Endocrinology. 152(7):2546-51.
- [36] Malaguarnera R, Belfiore A. (2022). The emerging role of insulin and insulin-like growth factor signaling in cancer stem cells. Frontiers in endocrinology. 5:79919.
- [37] Nwabo Kamdje AH, Seke Etet PF, Kipanyula MJ, Vecchio L, Tagne Simo R, Njamnshi AK, Lukong KE, Mimche PN. (2022). Insulin-like growth factor-1 in signaling the tumor microenvironment: Carcinogenesis, cancer drug resistance, and therapeutic potential. Frontiers in Endocrinology. 13:927390.
- [38] Smulders L, Deelen J. (2024). Genetics of human longevity: From variants to genes to pathways. Journal of Internal Medicine. 295(4):416-35.
- [39] Xie Z, Yang F. (2021). The effects of lycopene supplementation on serum insulin-like growth factor 1 (IGF-1) levels and cardiovascular disease: A dose-response metaanalysis of clinical trials. Complementary Therapies in Medicine. 56:102632.
- [40] Szydlowska-Gladysz J, Gorecka AE, Stepien J, Rysz I, Ben-

Skowronek I. (2024). IGF-1 and IGF-2 as Molecules Linked to Causes and Consequences of Obesity from Fetal Life to Adulthood: A Systematic Review. International Journal of Molecular Sciences. 25(7):3966.

- [41] Giuliani C, Garagnani P, Franceschi C. (2018). Genetics of human longevity within an ecoevolutionary nature-nurture framework. Circulation Research. 123(7):745-72.
- [42] Werner H, Laron Z. (2023). Insulin-like growth factors and aging: Lessons from Laron syndrome. Frontiers in Endocrinology. 14:1291812.
- [43] Athanasouli F, Georgiopoulos G, Asonitis N, Petychaki F, Savelli A, Panou E, Angelousi A. (2021).
 Nonfunctional adrenal adenomas and impaired glucose metabolism: a systematic review and metaanalysis. Endocrine. 74(1):50-60.
- [44] Loh NY, Wang W, Noordam R, Christodoulides C. (2022). Obesity, fat distribution and risk of cancer in women and men: a Mendelian randomisation study. Nutrients. 14(24):5259.
- [45] Pati S, Irfan W, Jameel A, Ahmed S, Shahid RK. (2023). Obesity and cancer: A current overview of epidemiology, pathogenesis, outcomes, and management. Cancers. 15(2):485.
- [46] Garczorz W, Kosowska A, Francuz T. (2024). Antidiabetic Drugs in Breast Cancer Patients. Cancers. 16(2):299.
- [47] Xu GP, Chen WX, Zhao Q, Zhou H, Chen SZ, Wu LF. (2019). Association between the insulinlike growth factor 1 gene rs2195239 and rs2162679 polymorphisms and

www.niprdjopat.gov.net; niprdjopat@gmail.com

cancer risk: a meta-analysis. BMC medical genetics. 20:1-10.

- [48] Werner H. (2022). BRCA1: an endocrine and metabolic regulator. Frontiers in endocrinology. 13:844575.
- [49] Neagu AN, Bruno P, Johnson KR, Ballestas G, Darie CC.
 (2024). Biological Basis of Breast Cancer-Related Disparities in Precision Oncology Era. International Journal of Molecular Sciences. 25(7):4113.
- [50] Chen X, Ma R, Wu W, Gao R, Shu Y, Dong M, Guo M, Tang D, Li D, Ji S. (2024). Wighteone, a prenylated flavonoid from licorice, inhibits growth of SW480 colorectal cancer cells by allosteric inhibition of Akt. Journal of Ethnopharmacology. 118195.
- [51] Nagini S, Palrasu M, Bishayee A. (2024). Limonoids from neem (Azadirachta indica A. Juss.) are potential anticancer drug candidates. Medicinal Research Reviews. 44(2):457-96.