



Antimosquito Phenylpropanoids from the Stem and Root Barks of *Uvariadendron pycnophyllum* (Diels) R.E.Fr

*¹CHARLES KIHAMPA; ²MAYUNGA H. H. NKUNYA; ²COSAM C. JOSEPH;
³STEPHEN M. MAGESA

¹Department of Environmental Science and Management, Ardhi University, P.O. Box 35176, Dar es Salaam, Tanzania; ²Department of Chemistry, University of Dar es Salaam, P.O. Box 35061, Dar es Salaam, Tanzania; ³National Institute for Medical Research, P.O. Box 81, Amani Research Centre, Muheza, Tanga, Tanzania

ABSTRACT: The phenylpropanoids *O*-methyleugenol, *O*-methylisoeugenol and 2,3-dimethoxycinnamaldehyde, have been isolated as the antimosquitocidal principles of the stem and root bark extracts of *Uvariadendron pycnophyllum* (Diels) R.E. Fr. The extracts and compounds exhibited activity with LC₅₀ values in the range 17-59 ppm against the *Anopheles gambiae* s.s. Giles mosquito larvae, while the constituent phenylpropanoids showed long term mortality effects to adult *An. gambiae* mosquito on impregnated bednets, and mosquito repellency that was stronger than the activity of the standard repellent DEET. @ JASEM

Uvariadendron (Family Annonaceae) is a small genus comprising about 16 species, which are either shrubs or trees, the genus being restricted to tropical Africa and about 6 species are confined to East Africa (Verdcourt, 1971). According to the Flora of Tropical East Africa (Verdcourt, 1971), four *Uvariadendron* species, namely *U. kirkii*, *U. gorgonis*, *U. pycnophyllum* and *U. usambarensis* are reported to occur in Tanzania.

In continuation with investigations of Tanzanian plant species for antimosquito constituents as a contribution to control malaria transmission mosquitoes (Kihampa *et al.*, 2009), we have analyzed the root and stem barks of *Uvariadendron pycnophyllum* (Diels) R.E. Fr. (*Mkene* in Kiswahili) that in preliminary assays exhibited potent antimosquito activity. We now report the antimosquito properties of *O*-methyleugenol (**1**), *O*-methylisoeugenol (**2**), 2,3-dimethoxycinnamaldehyde (**3**) and stigmaterol (**4**) obtained from the stem and root bark extracts. Besides being a source of firewood, building poles, knife and hoe handles, beds, bows and withies, the plant species is not used in any folk medicine.

MATERIALS AND METHODS

Plant Materials: The leaves, stem and root barks of *U. pycnophyllum* were collected from Siggi Valley, 3 km from Kisiwani village along the road to Bombani in the Amani Nature Reserve, East Usambara Mountains in Muheza District, Tanga Region. The plant species was identified on site and its identity was further confirmed at the Herbarium of the Department of Botany, University of Dar es Salaam, where a voucher specimen is deposited.

Extraction and Isolation: The air dried and pulverized root and stem barks were extracted

sequentially with CHCl₃ and MeOH, 2 x 48 h for each solvent. The extracts were stored at -18 °C until further analysed or assayed. The active compounds were isolated following a bioassay-guided isolation from the relevant extracts using the brine shrimp lethality test (BST) (Meyer *et al.*, 1982). Fractionation of the concentrated extracts was carried out by VLC, followed by repeated column chromatography on silica gel and/or Sephadex[®] LH-20 eluting with pet ether and then pet ether containing increasing amounts of EtOAc, and mixtures of MeOH and CHCl₃ (1:1, v/v) respectively. Structural determination was achieved upon analysis of spectral data.

Larvicidal Assay: The assay was carried out according to the WHO protocol (WHO, 1996). Twenty late 3rd or young 4th instar larvae were used per beaker with three beakers per concentration (the water temperature being 25 ± 1°C) and for each test three beakers containing distilled water and test larvae but without sample were used as controls. Larvae mortality and deformities was recorded after every 24 h of continuous exposure and expressed as percent mortality (WHO, 1996). The lethal concentration at which 50% of the test larvae were killed (LC₅₀) was determined using POLO PLUS computer package.

Mosquitocidal Assay: This was performed as described in the literature (Joseph *et al.*, 2004).

Mosquito Repellency Assay: The assay was conducted as reported in the literature (Innocent *et al.*, 2008) using serial dilutions, the highest concentration being 1% (0.01 g/ml) and the other concentrations were 0.000015, 0.00015, 0.0015, 0.015 and 0.15 mg/cm² corresponding to 10⁻⁵, 10⁻⁴,

10^{-3} , 10^{-2} and 10^{-1} ppm solutions respectively. The screening was done sequentially starting with the lowest dose (0.001%) and ending with the highest one (1%). A test solution (0.5 ml) was dispensed on the right forearm of a volunteer from the wrist to the elbow. The rest of the hand was covered with glove to make it unattractive to the mosquitoes. Acetone (0.5 ml) was dispensed on the left forearm, to act as control. The arms were swapped regularly to eliminate any bias. The control arm was introduced into the cage immediately after releasing the 25 insects and kept there for 3 min. The mosquitoes that had landed on the untreated control arm were recorded. The treated arm was then introduced into the cage and kept there for 3 min. The number of

mosquitoes that landed on the treated arm was also recorded. Each concentration was screened using a fresh batch of mosquitoes. After the bioassay of each concentration, the arms were washed with bar soap, rinsed well with tap water and then allowed to dry for 15-20 min, before application of the next dose of the test sample and the percentage protective efficacy (PE) was calculated as

$$PE = \left(\frac{PCM - PTM}{PCM} \right) \times 100\%$$

where PCM is the percent control mean and PTM is the percent test mean of mosquitoes landing on the control and treated arms respectively.

Table 1 Larvicidal Activity of 1, 2 and 4 Against *An. gambiae* Larvae

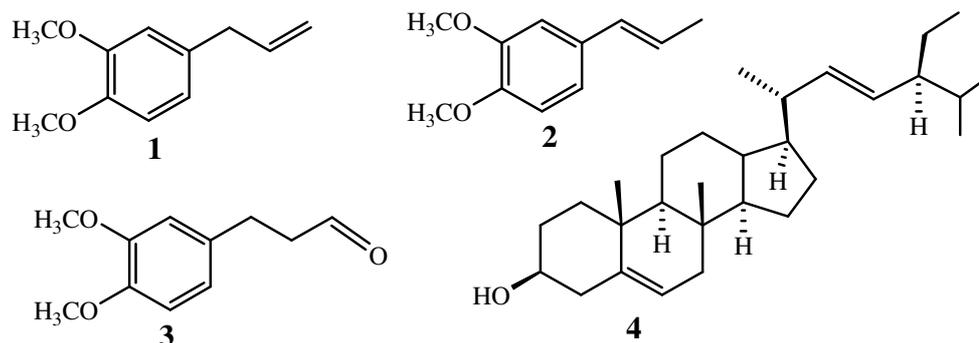
Cp	T (h)	Concentration (ppm)						LC ₅₀ (ppm) 95% CL
		15.62	31.25	62.5	125	250	500	
1	24	23.3 ± 6.7	53.3 ± 8.8	53.3 ± 6.7	76.7 ± 3.3	100 ± 0	100 ± 0	43 (23-69)
	48	80 ± 5.7	80 ± 5.7	100 ± 0	100 ± 0	nd	nd	nd
	72	100 ± 0	100 ± 0	nd	nd	nd	nd	nd
2	24	23.3 ± 3.3	30 ± 5.7	36.7 ± 8.8	70 ± 0	90 ± 0	100 ± 0	59 (34-95)
	48	53.3 ± 6.7	66.7 ± 3.3	93.3 ± 3.3	100 ± 0	100 ± 0	nd	17 (04-26)
	72	100 ± 0	100 ± 0	100 ± 0	nd	nd	nd	nd
4	24	10 ± 5.7	16.7 ± 3.3	66.7 ± 3.3	96.7 ± 3.3	100 ± 0	100 ± 0	46 (31-66)
	48	30 ± 5.7	73.3 ± 13	90 ± 5.7	100 ± 0	nd	nd	22 (11-32)
	72	100 ± 0	100 ± 0	100 ± 0	nd	nd	nd	nd
UPSC*	-	-	-	-	-	-	-	56 (34-87)
UPSM*	-	-	-	-	-	-	-	109 (66-192)
UPRC*	-	-	-	-	-	-	-	56 (34-86)
UPRM*	-	-	-	-	-	-	-	56 (35-86)

Cp = compound, nd = not determined, CL = class limits, UPSC = *U. pycnophyllum* stem bark chloroform extract, UPSM = *U. pycnophyllum* stem bark methano extract, UPRC = *U. pycnophyllum* root bark chloroform extract, UPRM = *U. pycnophyllum* root bark methanol extract, *Kihampa et al 2009

RESULTS AND DISCUSSION

Bio-assayed guided fractionation of the antimosquito chloroform extract of the stem bark, and chloroform and methanol extracts of the root barks yielded *O*-methyleugenol (1) and *O*-methylisoeugenol (2) as the major constituents of the root bark, as well as 2,3-dimethoxycinnamaldehyde (3) and stigmasterol (4). The stem bark methanol extract was less active than the other three extracts, hence it was not analysed further for constituent compounds. Structures of the isolated compounds were identified based on analysis of spectroscopic data (Mohammed *et al.*, 1985; Mussa, 2000).

The phenylpropenoids 1 and 2, and compound 4 displayed larvicidal activity at different levels (Table 1). There was no significant difference in the observed activity between the tested compounds after 24 h of larvae exposure. Furthermore, no significant difference in the activity was observed between the three compounds 1, 2 and 4, and that of the crude chloroform extracts of the stem and root barks, and that of the root bark methanol extract (Table 1). Compound 3 was not assayed for larvicidal activity due to paucity of the obtained amount. Long term exposure of the larvae to the samples up to 72 h showed significant increase in larvicidal activity as deduced from the LC₅₀ value after 48 and 72 h larvae exposure for 1, 2 and 4 (Table 1).



Compounds 1 and 2 were also assayed for activity against adult mosquitoes in the tarsal contact and repellency bioassays. Table 2 shows that after 3 min exposure compounds 1 and 2, and the standard insecticide Fendona® were inactive against *An. gambiae* mosquitoes in all the tested concentrations. After 1 h holding time compounds 1 and 2 exhibited a knock down effect of 30 and 10 % respectively at the highest concentration tested (200 mg/m²). At the same time interval (1 h) the standard insecticide

showed strong activity even at the lowest concentration tested (100 mg/m²), the knock down effect being 80 %. The mortality caused by the standard insecticide tended to decrease with extension of holding time but the activity for compounds 1 and 2 showed an increasing trend. These results suggest that compounds 1 and 2 could be interesting candidates for the development of bednet impregnated mosquitocides.

Table 2 Insecticidal Activity of 1, 2 and Fendona® Against *An. gambiae* Mosquito

Compound	Time	% Mortality/Concentration (mg/m ²)				
		100	125	150	175	200
1	3 min	0±0	0±0	0±0	0±0	20±0
	1 h	0±0	0±0	0±0	0±0	30±3.3
	24 h	0±0	10±3.3	20±0	30±0	40±5.8
	48 h	0±0	10±0	20±5.8	20±5.8	30±0
2	3 min	0±0	0±0	0±0	0±0	0±0
	1 h	0±0	0±0	0±0	0±0	10±0
	24 h	0±0	10±0	20±0	20±0	30±5.8
	48 h	10±0	40±0	40±0	40±0	50±0
Fendona®	3 min	0±0	0±0	0±0	0±0	0±0
	1 h	80±0	80±11.5	80±11.5	100±0	100±0
	24 h	46.7±6.7	80±11.5	80±0	80±0	100±0
	48 h	30±0	66.7±6.7	80±0	80±0	80±0

In the mosquito repellency assays, the activity of compounds 1 and 2 were compared to those of the standard repellent DEET (Table 3 and Figure 1). The graph in Figure 1 shows that the activity trend of the two compounds was almost linear. The repellency and 4 were not tested in the tarsal contact and repellency activity due to paucity of the amount isolated.

activities of the two compounds 1 and 2 were significantly higher than the standard repellent DEET, compound 2 being more effective than 1. Compounds 3.

Table 3 Repellency Activity of the Isolated Compounds and DEET

Compound	Percentage Repellency/Concentration (mg/ml)					RC ₅₀ (mg/cm ²)
	10 ⁻⁵	10 ⁻⁴	10 ⁻³	10 ⁻²	10 ⁻¹	
1	45±1.7	71±1.2	91±0	100±0	100±0	5.84 x 10 ⁻⁶
2	25±1.2	46±0	74±0.6	85±0.6	92±0	1.58 x 10 ⁻⁴
DEET	0±0	41±5.8	46±0	98±5.8	100±0	6.12 x 10 ⁻⁴

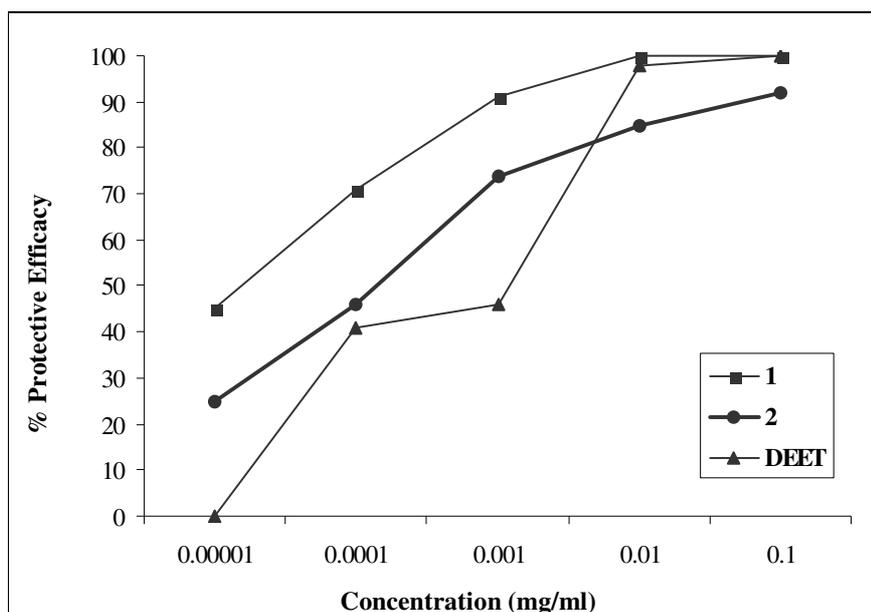


Fig.1 .. Percentage Variation of Mosquito Repellency Efficacy of Compounds 1, 2 and DEET Against *An. gambiae* Mosquitoes

Acknowledgement: We thank the Germany Academic Exchange Services (DAAD) for a study grant for the research work that was also undertaken at Amani Medical Research Center in Muheza, Tanga, Tanzania. Financial support through Sida/SAREC support to the Faculty of Science at the University of Dar es Salaam is gratefully acknowledged. We thank Mr. Frank Mbago of the Herbarium, Department of Botany at the University of Dar es Salaam for location and identification of the investigated plant species.

REFERENCES

- Innocent, E; Gikonyo, NK; Nkunya, MHH (2008). Repellency property of long chain aliphatic methyl ketones against *Anopheles gambiae* s.s. Tanzania. *J Health Res* 10: 50–54.
- Joseph, CC; Ndoile, MM; Malima, RC; Nkunya, MHH (2004). Larvicidal and mosquitocidal extracts, a coumarin, isoflavonoids and pterocarpanes from *Neorautanenia mitis*. *Trans. Royal Soc Trop Med Hyg* 98: 451-455.
- Kihampa, C; Joseph, CC; Nkunya, MHH; Magesa, SM; Hassanali, A; Heydenreich, M; Kleinpeter, E (2009). Larvicidal and IGR activity of extract of Tanzanian plants against malaria vector mosquitoes. *J Vector Borne Dis* 46: 145-152.
- Mohammed, I; Waterman, PG (1985). Chemistry in the Annonaceae, XVII. Phenylpropenes from *Uvariadendron connives* seeds. *J Nat Prod* 48: 328-329.
- Mussa, LM (2000). Flavonoids, eugenol derivatives and terpenoids from three coastal plants of Annonaceae. *MSc Thesis*, University of Dar es Salaam.
- Meyer, BN; Ferrigni, NR; Putnam, JE; Jacobsen, LB; Nichols, DE; McLaughlin, JL (1982). Brine Shrimp: A convenient general bioassay for active plant constituents. *Planta Med* 45: 31–34
- Verdcourt, B (1971). *Flora of Tropical East Africa: Annonaceae*. Crown Agents, London.
- WHO (1996). Report of the WHO informal consultation on the evaluation and testing of insecticides. WHO, Geneva, pp 32-36 and 50-52.