

PRINT ISSN 1119-8362 Electronic ISSN 2659-1499 Full-text Available Online at https://www.ajol.info/index.php/jasem https://www.bioline.org.br/ja

Isolation of Native Fungi from Sawdust Collected at a Sawmill from Adeleke Area, Ogbomosho, Oyo State, Nigeria

¹AKOMOLAFE, AP; ^{1,2}ODELADE, KA; ¹SULAIMON, SO

*¹Department of Science Laboratory Technology, Federal Polytechnic Ayede, Oyo State, Nigeria ²Department of Natural Sciences, Faculty of Pure and Applied Sciences, Precious Cornerstone University, Ibadan, Nigeria

> *Corresponding Author Email: sheriffolaekan186143@gmail.com *ORCID: https://orcid.org/0009-0004-9036-8294 *Tel: +2347025231416

Co-Authors Email: akomolafeap@federalpolyayede.edu.ng; odeladeka@federalpolyayede.edu.ng; kennybramms@gmail.com; sheriffolaekan186143@gmail.com; sulaimonso@federalplolyayede.edu.ng

ABSTRACT: Sawdust is an agricultural waste and could be utilized in several productive and beneficial entities. Therefore, the objective of this paper is to assess the Isolation of Native Fungi from Sawdust collected at sawmill from Adeleke area, Ogbomosho, Oyo state, Nigeria using appropriate standard procedures. Results were obtained at 25° C and 35° C in this study. At 35° C, the result reveals that *Aspergillus carbonarius* cultured with sawdust had the highest enzymatic activity of 3.65 ± 1.414 than with CMC where it had an enzymatic activity of 2.9 ± 0.070 . *Rhizopus nigrican* had an enzymatic activity of 3.5 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 2.7 ± 1.414 . *Aspergillus flavus* had an enzymatic activity of 2.45 ± 0.707 with sawdust than with CMC where it had an enzymatic activity of 1.9 ± 0.707 . *Aspergillus Niger* had an enzymatic activity of 2.4 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 1.6 ± 1.414 . *Trichoderma harzianum* had an enzymatic activity of 2 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 1.6 ± 1.414 . *Trichoderma harzianum* had an enzymatic activity of 2 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 1.6 ± 1.414 . *Trichoderma harzianum* had an enzymatic activity of 2 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 1.3 ± 1.414 . In this study, *Aspergillus niger*, *Aspergillus flavus*, *Aspergillus carbonarius*, *Rhizopus nigrican*, and *Trichoderma harzianum* showed significant cellulolytic ability to degrade sawdust. *Rhizopus nigrican* showed the highest fermentation ability and producing clear zones of 40mm (25°C) and 34mm (35°C) (p<0.001). Hence, the use of *Rhizopus nigrican* in the production of cellulase using sawdust as an agro waste is possible because of its high cellulolytic ability which was observed at both temperatures.

DOI: https://dx.doi.org/10.4314/jasem.v29i3.20

License: CC-BY-4.0

Open Access Policy: All articles published by **JASEM** are open-access and free for anyone to download, copy, redistribute, repost, translate and read.

Copyright Policy: © 2025. Authors retain the copyright and grant **JASEM** the right of first publication. Any part of the article may be reused without permission, provided that the original article is cited.

Cite this Article as: AKOMOLAFE, A. P; ODELADE, K. A; SULAIMON, S. O. (2025) Isolation of Native Fungi from Sawdust Collected at a Sawmill from Adeleke Area, Ogbomosho, Oyo State, Nigeria. *J. Appl. Sci. Environ. Manage.* 29 (3) 841-847

Dates: Received: 20 February 2025; Revised: 13 March 2025; Accepted: 18 March 2025; Published: 31 March 2025

Keywords: Sawdust; cellulose; fermentation; biodegradation; cellulolytic ability

Agricultural wastes are the byproducts from growing crops and/or the initial processing of unprocessed agricultural goods, including fruits, vegetables, dairy, meat, poultry, and other products (El-Ramady *et al.*, 2022a). There are three categories of agro-waste: liquid, solid, and slurry (El-Ramady *et al.*, 2022b). Agro-wastes make up close to 30% of all agricultural products generated worldwide. Several studies have quantified agricultural waste production and

environmental effects, such as the global nutritional and environmental losses caused by food waste (Chen *et al.*, 2020). As a more economical and environmentally friendly alternative to treatment, sustainable management with an emphasis on reducing food waste is recommended (Ogunmoroti *et al.*, 2022). Wood dust is also the byproduct of a few animals, birds, and insects that live in wood, such as the carpenter ants and woodpeckers. The production

^{*}Corresponding Author Email: sheriffolaekan186143@gmail.com *ORCID: https://orcid.org/0009-0004-9036-8294 *Tel: +2347025231416

of briquettes and sawdust bioremediation reduces air pollution while yielding a useful and valued end product. Sawdust becomes biofertilizers by composting, which binds nitrogen to the soil for 180 days (Asadu et al., 2019). Because of its high lignin and cellulose concentration and low nitrogen level, it is an organic waste that does not degrade in particular, certain microbes. Lignin and cellulose must be completely hydrolyzed by ligninase and cellulase for sawdust to decompose. Both cellulose and lignin are more complex polysaccharides with a high molecular weight; cellulose has a molecular weight of 2 x105 to 2.4 x 106 mole and approximately 2 x103 to 104 glucose monomers (Rusdianto et al., 2021). Numerous Streptomyces species can flourish on potato dextrose agar and with Sabouraud agar different sawdust S. concentrations, including purpureus, S. albidoflavus, S. antibioticus, S. albus, and S. cyaneus (Ejaz et al., 2021). Sawdust biodegradation requires the release of several hydrolyzing enzymes, such as cellulase and ligninase, which hydrolyze cellulose and lignin, respectively (Singhania et al., 2021). Sawdustmicroorganisms decomposing thrive in polysaccharide-rich environments like soil, where efficient hydrolyzing enzymes are continuously generated (Sharma et al., 2019). Hence, the objective of this paper is to assess the Isolation of Native Fungi from sawdust collected at sawmill from Adeleke area, Ogbomosho, Oyo state, Nigeria.

MATERIALS AND METHODS

Sample collection and preparation: Sawdust was aseptically collected from a sawmill, Adeleke area, Ogbomosho, Oyo state, (8°7¹29[°] N 4°13'32[°]E) Nigeria. The sawdust was pretreated by weighing 100g, sun drying, and oven drying at 80°C for 1h and then cooling. The pretreated sawdust was blended with a blender. The blended sample was filtered to small particles sizes with 0.1mm mesh sieve. The filtered sample was then stored in an airtight polythene bag at room temperature until required.

Isolation and Characterization of Fungi: Five grams (5g) of soil sample was collected aseptically into sample bottles from the Precious Cornerstone University disposal site. The soil sample collected was serially diluted and the dilution factor of 10^{-3} and 10^{-5} was used. For five days, the inoculum was incubated at $28\pm2^{\circ}$ C on Potato Dextrose Agar (PDA) supplemented with 0.001mg/l of chloramphenicol. A pure culture was obtained by subculturing the colonies from the culture. Stock cultures were kept on kept on PDA slants and stored at 28° C. The isolates were identified and characterized by phenotypic

characteristics which included colony colour observation and growth pattern studies, as well as microscopic characteristics (Gurung *et al.*, 2020).

Screening for Cellulase Producing Fungi: Spot inoculations of pure cultures' spore suspension were made on plates using a minimal salt medium that contained 1g of carboxymethyl cellulose (CMC) agar, 0.25g of NaNO₃, 0.1g of KH₂PO₄, 0.05g of MgSO₄, 0.06g of CaCl₂, 1.5g of agar agar powder, and 0.001g chloramphenicol in 100ml of H₂O. The plates were then incubated for five days at 28±2°C. Spot inoculations of pure cultures' spore suspension were made on other plates using a minimal salt medium that contained 1g of sawdust, 0.25g of NaNO₃, 0.1g of KH₂PO₄, 0.05g of MgSO₄, 0.06 of CaCl₂, 1.5g of agar agar powder, and 0.001g chloramphenicol in 100ml of H₂O to induce cellulase secretion. The plates were then incubated for five days at a temperature of 28±2 °C. Following incubation, the plates were stained for 15 minutes with 1% Congo red solution and then 10 minutes later with 1M NaCl solution. Each colony's zone of clearance was measured in diameter. To produce cellulase, fungal isolates with the highest zone of clearance were chosen. The fungi isolated are Aspergillus niger, Aspergillus flavus, Aspergillus carbonarius, Rhizopus nigrican and Trichoderma harzianum.

Fermentation Process for Cellulase Activity of the fungal isolates: The fermentation process was done by comparing the organisms incubated at different temperatures. To determine which temperature had the highest zone of clearance and to assess the fungal isolates' optimal growth, the isolates were incubated at 25° C and 35° C.

enzyme of cellulolytic activity: Screening Microorganisms that produced cellulases were screened on agar plates that were enriched with only CMC as a carbon source. This plate was used as a control. The microorganisms were then screened on another agar plate that included sawdust as the sole carbon source. The plates were kept at 28± 2 °C for five days of incubation. The basis for this qualitative examination is the way congo red interacts with cellulose and its broken-down components, allowing the dve to be retained in the integral biopolymer whereas areas containing cellulose that has been hydrolyzed by enzymes produce clear zones. The enzymatic activity (index) was subsequently calculated by measuring the clear zones using the formula described by Rosa et al. (2020).

$$EA = \frac{Diameter of clear zone}{Diameter of inoculum (colony)}$$
(1)

Statistical Analysis: Graphpad Prism 5 was used for the analysis of the fungal isolates on sawdust and C.M.C at 25°C and 35°C. Two-way ANOVA test was applied to set the degree of significance of the fermentation activity. Also, comparison of the enzyme activity of different isolates was done statistically. In this study, P<0.05; P<0.01 and P<0.001 were used to disapprove the null hypothesis (Oladeji *et al.*, 2020).

RESULTS AND DISCUSSION

The fungi isolated are Aspergillus niger, Aspergillus flavus, Aspergillus carbonarius, Rhizopus nigrican and Trichoderma harzianum Characterization of the fungi isolates:

Aspergillus niger

Colony diameter: 4.5-6.5 cm.

Obverse: Blackish-brown often with yellow mycelium.

Reverse: Creamish-yellow to yellow or orange.

Head: Globose, splitting with age.

Stipe: long, smooth, hyaline to brownish.

Vesicle: Globose, thick-walled, brownish, and fertile on entire surface.

Metulae: present, long, closely packed, brownish.

Phialides: shirt, thick, ampulliform, brownish.

Conidia: Globose to elliptical, rough, dark brown to black.

Mycotoxins: Malformin C, naphthoquinones, nigragillin.

Aspergillus flavus

Colony diameter: 4.0-4.5.

Obverse: Yellowish-green becoming green with age. Reverse: Creamish-yellow

Head: Radiating, becoming loosely columnar with age.

Stipe: long, hyaline.

Vesicle: Dome-shaped, fertile on the entire surface.

Metulae: present, small.

Phialides: small, ampulliform.

Conidia: Globose to subglobose, usually yellowgreen.

Mycotoxins: Aflatoxins B1 and B2, paspalini.

Aspergillus carbonarius

Morphology: Hyphae that form mycelium

Colony: Dark black

Shape: Filamentous with hyphae

Conidiophores: unbranched and may have vesicles at their tips from 3-5 micrometers.

Rhizopus nigrican

Colour: Black or dark gray.

Morphology: Mold-like appearance with a mycelium that branches out and spreads. Reproduction: sexual or asexual.

Trichoderma harzianum

Colour: Greenish hue.

Conidiophores: Elongated and none branching with structures that emerge from the mycelium Conidia: Oval or ellipsoidal Size: 3-5 micrometer

Colony Texture: Colonies often exhibit a fluffy or cotton-like texture.

Comparative analysis of the fermentative activity of the fungal isolates at 25°C and 35°C: During the fermentation of sawdust at 25°C and 35°C, the cellulolytic organisms isolated showed significant fermentative ability. The fungal isolates grown at 25oC with sawdust were compared statistically with the fungal isolates grown with CMC at 25°C and the fungal isolates grown with sawdust at 35°C were compared with the isolates grown with CMC at 35°C. The graph (Figure 1-2) below shows the significance of each fungal isolate when compared with CMC. Values with (*, **, and ***) are significantly higher than the control at P < 0.05; P < 0.01, and P < 0.001. The statistical analysis of the fermentation activities on sawdust and carboxymethyl cellulose for the cellulolytic activity of the organisms is shown in Figures 1 and 2. In the fermentation process, it was observed that all the fungal isolates showed a significant ability to biodegrade sawdust at P<0.05, P<0.01, P<0.001, although at different rates. Figure 1 at 25°C shows that all the fungal isolates grown with sawdust had the highest fermentation activities than fungal isolates grown with CMC. The fungal isolates showed clear zones ranging from 27mm-40mm. Rhizopus nigrican showed the highest fermentation activity with a clear zone of 40mm at P<0.001 with sawdust than with CMC where it had a clear zone of 32mm. Aspergillus niger had a clear zone of 27mm at P<0.05 with sawdust than with CMC where it had a clear zone of 23mm. Aspergillus flavus had a clear zone of 30mm at P<0.001 with sawdust than with CMC where it had a clear zone of 17mm. Aspergillus carbonarius had a clear zone of 37mm at P<0.01 with sawdust than with CMC where it had a clear zone of 30mm. Trichoderma harzianum had a clear zone of 30mm at P<0.001 on sawdust than with CMC where it had a clear zone of 22mm.

Figure 2 at 35°C shows that fungal isolates grown with sawdust had the highest fermentation activity than the fungal isolate grown with CMC. The fungal isolates showed clear zone ranging from 19mm-34mm. *Rhizopus nigrican* showed the highest fermentation activity with a clear zone of 34mm at P<0.001 than with CMC where it had a clear zone of 28mm. *Aspergillus niger* had a clear zone of 23mm at P<0.001 than with CMC where it had a clear zone of 17mm.



Fig 1: Fermentation activity of cellulolytic organism on sawdust against C.M.C at 25°c



Fig. 2: fermentation activity of cellulolytic organism on sawdust against C.M.C at 35°c

Aspergillus flavus had a clear zone of 25mm at P<0.05 with sawdust than with CMC where it had a clear zone of 20mm. Aspergillus carbonarius had a clear zone of 37mm at P<0.001 with sawdust than with CMC where it had a clear zone of 30mm. Trichoderma harzianum had a clear zone of 19mm at P<0.01 with sawdust than with CMC where it had a clear zone of 12mm.

In Table 1, values of the enzymatic activities of the cellulolytic organisms at 25° C and 35° C are expressed as mean \pm SD for n=2 for each concentration. Values with (*, **, ***) are significantly higher than the standard at P<0.05, P<0.01, P<0.001 respectively.

Comparative analysis on the enzymatic activity of the fungal isolates on sawdust and CMC at 25°C and $35^{\circ}C$: The enzymatic activities of the fungal isolates grown with sawdust at 25°C were compared statistically with the fungal isolates grown with CMC at 25°C. Also, the fungal isolates grown with sawdust at 35°C were compared with the isolates grown with CMC at 35°C (Table 1). This analysis was statistically analyzed at a significance level of P < 0.05, P < 0.01 and P < 0.001. To minimize errors, the mean ±SD for each concentration was determined. At 25°C the enzymatic activity of *Rhizopus nigrican* cultured with sawdust was statistically higher having an enzymatic activity of 3.9 ± 0.414 at P < 0.001 than with CMC where it had an enzymatic activity of 3.1±0.141. Aspergillus flavus had an enzymatic activity of 3.05±0.707 with sawdust than with CMC where it had an enzymatic activity of 1.75 ± 0.070 . Aspergillus niger had an enzymatic activity of 2.8 ± 0.141 with sawdust than with CMC where it had an enzymatic activity of 2.4 ±0.141. Aspergillus carbonarius had an enzymatic activity of 3.65±0.070 with sawdust than with CMC where it had an enzymatic activity of 3.05±0.070. Trichoderma harzianum had an enzymatic activity of 3.1±1.414 with sawdust than with CMC where it had an enzymatic activity of 2.1±0.141.

Fungal Isolates	CMC at 25°C	Sawdust at 25°C	CMC at 35°C	Sawdust at 35°C
Rhizopus nigrican	3.1±0.141	3.9±0.414***	2.7±1.414	3.5±1.414***
Aspergillus flavus	1.75 ± 0.070	3.05±0.707***	1.95±0.707	2.45±0.707*
Aspergillus niger	2.4±0.141	2.8±0.141*	1.6 ± 1.414	2.4±1.414***
Aspergillus carbonarius	3.05±0.070	3.65±0.070**	2.9±0.707	3.65±1.414***
Trichoderma harzianum	2.1±0.141	3.1±1.414***	1.3±1.414	2±1.414**

Table 1: comparative analysis of the enzymatic activity of the fungal isolates at 25°c and 35°c

At 35° C Aspergillus carbonarius cultured with sawdust had the highest enzymatic activity of 3.65 ± 1.414 than with CMC where it had an enzymatic activity of 2.9 ± 0.070 . *Rhizopus nigrican*

had an enzymatic activity of 3.5 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 2.7 ± 1.414 . *Aspergillus flavus* had an enzymatic activity of 2.45 ± 0.707 with sawdust than with CMC

where it had an enzymatic activity of 1.95 ± 0.707 . *Aspergillus niger* had an enzymatic activity of 2.4 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 1.6 ± 1.414 . *Trichoderma harzianum* had an enzymatic activity of 2 ± 1.414 with sawdust than with CMC where it had an enzymatic activity of 1.3 ± 1.414 .



Fig. 3: comparison of the cellulolytic activity of the fungal isolates at 25° c and 35° c on sawdust.

The statistical analysis of the cellulolytic activity of the fungal isolates on sawdust was done by comparing their cellulolytic activity at 25°C and 35°C. Figure 3 shows that *Trichoderma hazianum* was statistically higher at 25°C at p<0.001 than at 35°C. While *Aspergillus carbonarius* had no significant value at p>0.05. *Aspergillus flavus* had a significant value of p<0.001 at 25°C than at 35°C *Aspergillus niger* had a significant value of p<0.05 at 25°C than at 35° and *Rhizopus nigrican* had a significant value of p<0.05 at 25°C than at 35°C.

In this study, different fungal isolates were used in the biodegradation of sawdust to produce cellulase because of its high cellulolytic compound in conjunction with (Christopher *et al.*, 2023). The fungal isolates identified and characterized from the sawdust decomposing soil were *Aspergillus niger*, *Aspergillus flavus*, *Rhizopus nigrican*, *Trichoderma harzianum*, *Aspergillus carbonarius* in contrast with Oshoma *et al.*, (2020) that used fungal isolates from contaminated soils. As adaptable cellulase producers in the bioprocessing business, these fungi have been reported by several authors (Tomico-Cuenca *et al.*, 2021). In contrast to (Fierro *et al.*, 2022) findings, which suggested that *P. chrysogenum* is a suitable choice for cellulase high productivity yield among the Penicillium species, this study found that Rhizopus nigrican is a good microorganism to produce cellulase. From the screened fungal isolates result all isolates had varied zones of clearance for the cellulase production profile (Carbonero-Pacheco *et al.*, 2023). According to the results of the screening for cellulolytic activity, Rhizopus nigrican had the highest zone of clearance (40 mm) and is considered to be a highly productive and efficient species of cellulolytic fungus. The enzymatic activity of the fungal isolate was done following (Coronado-Ruiz *et al.*, 2018)

The statistical analysis of the fermentation activities on sawdust and carboxymethyl cellulose to the cellulolytic activity was carried out between the organisms. In the fermentation process, it was observed that all the fungal isolates showed a significant ability to biodegrade sawdust at P<0.05, P<0.01, and P<0.001 although at different rates (Oshoma *et al.*, 2020).

According to this study, the temperature at 25°C shows that all the fungal isolates grown with sawdust had the highest fermentation activities than fungal isolates grown with CMC. The fungal isolates showed clear zone ranging from 27mm-40mm. Rhizopus nigrican showed the highest fermentation activity with a clear zone of 40mm at P<0.001. Aspergillus niger had a clear zone of 27mm at P<0.05, Aspergillus flavus had a clear zone of 30mm at P<0.001 Aspergillus carbonarius had a clear zone of 37mm at P<0.01 and Trichoderma harzianum had a clear zone of 30mm at P<0.001. At 25°C fungal isolates grown with CMC showed low fermentation activity compared to those grown with sawdust (Jumare et al., 2022). The fungal isolates showed clear zones ranging from 18 mm to 32 mm at P<0.05, P<0.01, P<0.001. Rhizopus nigrican showed the highest fermentation activity with a clear zone of 32mm. Aspergillus niger had a clear zone of 23mm, Aspergillus flavus had a clear zone of 18mm, Aspergillus carbonarius had a clear zone of 31mm and Trichoderma harzianum had a clear zone of 22mm.

At the temperature 35° C it also shows that fungal isolates grown with sawdust had the highest fermentation activity than the plates containing fungal isolates grown with CMC. The fungal isolates showed clear zone ranging from 19mm-34mm. *Rhizopus nigrican* showed the highest fermentation activity with a clear zone of 34mm at P<0.001. *Aspergillus niger* had a clear zone of 23mm at

P<0.001, Aspergillus flavus had a clear zone of 25mm at P<0.05 Aspergillus carbonarius had a clear zone of 37mm at P<0.001 and Trichoderma harzianum had a clear zone of 19mm at P<0.01. At 35°C fungal isolates grown with CMC showed low fermentation activity compared to those grown with sawdust 2025/3/13. The fungal isolates showed clear zones ranging from 12mm-30mm at P<0.05, P<0.01, P<0.001. Rhizopus nigrican showed the highest fermentation activity with a clear zone of 28mm Aspergillus niger had a clear zone of 20mm, Aspergillus carbonarius had a clear zone of 30mm and Trichoderma harzianum had a clear zone of 12mm.

Conclusion: This study demonstrated that sawdust, which is an agricultural waste, can promote the growth of fungal isolates by facilitating the breakdown of cellulose, the main component required for cellulase production, under various temperature conditions. This finding suggests that utilizing sawdust for cellulase production is a promising approach that can effectively address environmental pollution caused by improper waste disposal.

Acknowledgments: The authors extend their appreciation to the Tertiary Education Trust Fund (TETFUND) for funding this research through a grant awarded under the Institution-Based Research (IBR) intervention, 2023, to the Federal Polytechnic Ayede, Oyo State, Nigeria.

Declaration of Conflict of Interest: The authors declare no conflicts of interest.

Data Availability Statement: Data are available upon request from the second author.

REFERENCES

- Asadu, CO; Egbuna, SO; Chime, TO; Eze, CN; Kevin, D; Mbah, G O; Ezema, AC (2019). Survey on solid wastes management by composting: Optimization of key process parameters for biofertilizer synthesis from agro wastes using response surface methodology (RSM). Artif. Intell. Agric. 3, 52–61. https://doi.org/10.1016/j.aiia.2019.12.002
- Carbonero-Pacheco, J; Aguilar, J; Raya, MC; Trapero, A; Gaju-Ricart, M; Agustí-Brisach, C (2023). Diversity of Cellulolytic Microorganisms Associated with the Subterranean Termite Reticulitermes grassei. J. Fungi. 9(3), 294. https://doi.org/10.3390/jof9030294

Chen, C; Chaudhary, A; Mathys, A (2020). Nutritional and environmental losses embedded in global food waste. Resour. Conserv. Recycl. *160*, 104912.

https://doi.org/10.1016/j.resconrec.2020.104912

- Christopher, M; Sreeja-Raju, A; Kooloth-Valappil, P; Gokhale, D. V; Sukumaran, RK (2023). Cellulase Hyper-Producing Fungus Penicillium janthinellum NCIM 1366 Elaborates a Wider Array of Proteins Involved in Transport and Secretion, Potentially Enabling a Diverse Substrate Range. *BioEnerg. Res. 16*(1), 61–73. https://doi.org/10.1007/s12155-022-10407-3
- Coronado-Ruiz, C; Avendaño, R; Escudero-Leyva, E; Conejo-Barboza, G; Chaverri, P; Chavarría, M (2018). Two new cellulolytic fungal species isolated from a 19th-century art collection. *Sci. Rep.* 8(1), 7492. https://doi.org/10.1038/s41598-018-24934-7
- Ejaz, U; Sohail, M; Ghanemi, A (2021). Cellulases: From Bioactivity to a Variety of Industrial Applications. *Biomimetics*. *6*(3), 44. https://doi.org/10.3390/biomimetics6030044
- El-Ramady, H; Brevik, EC; Bayoumi, Y; Shalaby, TA; El-Mahrouk, ME; Taha, N; Elbasiouny, H; Elbehiry, F; Amer, M; Abdalla, N; Prokisch, J; Solberg, S. Ø; Ling, W (2022a). An Overview of Agro-Waste Management in Light of the Water-Energy-Waste Nexus. *Sustainability*. 14(23), 15717. https://doi.org/10.3390/su142315717
- El-Ramady, H; Brevik, EC; Bayoumi, Y; Shalaby, TA; El-Mahrouk, ME; Taha, N; Elbasiouny, H; Elbehiry, F; Amer, M; Abdalla, N; Prokisch, J; Solberg, S. Ø; Ling, W (2022b). An Overview of Agro-Waste Management in Light of the Water-Energy-Waste Nexus. *Sustainability*. 14(23), 15717. https://doi.org/10.3390/su142315717
- Fierro, F; Vaca, I; Castillo, NI; García-Rico, RO; Chávez, R (2022). Penicillium chrysogenum, a Vintage Model with a Cutting-Edge Profile in Biotechnology. *Microorganisms*, 10(3), 573. https://doi.org/10.3390/microorganisms10030573
- Gurung, K; Dasila, K; Pandey, A; Bag, N (2020). Curvularia eragrostidis, a new threat to large cardamom (Amomum subulatum Roxb.) causing leaf blight in Sikkim. J. Bios, 45(1), 113. https://doi.org/10.1007/s12038-020-00086-7

- Helal, GA; Khalil, RR; Galal, YG; Soliman, SM; Abd Elkader, RS (2022). Studies on cellulases of some cellulose-degrading soil fungi. Arch. Microbiol. 204(1), 65. https://doi.org/10.1007/s00203-021-02705-9
- Jumare, FI; Ahmad, MI; Abdulkadir, N; Bello, AY; Gani, M; Usman, MH (2022). Characterization of microorganisms associated with the degradation of sawdust and woodchips. Bayero J. Pure Appl. Sci. 15(1). 69-73. https://doi.org/10.4314/bajopas.v15i1.9
- Ogunmoroti, A; Liu, M; Li, M; Liu, W. (2022). Unraveling the environmental impact of current and future food waste and its management in Chinese provinces. Resou. Environ. Sustain. 9.100064.

https://doi.org/10.1016/j.resenv.2022.100064

- Oladeji, OS; Odelade, KA; Oloke, JK (2020). Phytochemical screening and antimicrobial investigation of Moringa oleifera leaf extracts. Afr. J. Sci. Technol. Innov. Dev. 12(1), 79-84. https://doi.org/10.1080/20421338.2019.1589082
- Oshoma, CE; Emeka-Ifebi, AE; Ahmed El-Imam, A (2020). Cellulase Production using Fungi Grown on Sawdust. RJEES. 5((1)), 56-65.
- Rosa, E; Ekowati, C N; Handayani, T.T; Ikhsanudin, Apriliani, F; Arifiyanto, A (2020). A: Characterization of entomopathogenic fungi as a control of natural biological American cockroaches (Periplaneta americana). Biodiversitas, 21 (11).

- Rusdianto, AS; Amilia, W; Sinta, VJD (2021). The Optimization Of Cellulose Content In Tobacco Stems (Nicotiana tabaccum L.) With Acid Extraction Method And Alkaline Extraction Method. Int. J. Food Agric. Nat. Resour. 2(2), 13-19. https://doi.org/10.46676/ij-fanres.v2i2.28
- Sharma, HK; Xu, C; Qin, W (2019). Biological Pretreatment of Lignocellulosic Biomass for Biofuels and Bioproducts: An Overview. Waste and Biomass Valorization, 10(2), 235-251. https://doi.org/10.1007/s12649-017-0059-y
- Singhania, RR; Ruiz, HA; Awasthi, MK; Dong, CD; Chen, CW; Patel, AK (2021). Challenges in cellulase bioprocess for biofuel applications. Sustain. Energy Rev. 151, 111622. Renew. https://doi.org/10.1016/j.rser.2021.111622
- Tomico-Cuenca, I; Mach, RL; Mach-Aigner, AR; Derntl, C (2021). An overview on current molecular tools for heterologous gene expression in Trichoderma. Fungal Biol. Biotechnol. 8(1), 11. https://doi.org/10.1186/s40694-021-00119-2