



## Evaluation of *in-Vitro* Antioxidant and Antidiarrheal Activities of *Peperomia Pellucida* Methanol Extracts on Albino Mice

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**ABSTRACT:** *Peperomia pellucida* also known as shiny bush is a common plant widely distributed in the tropical regions of the world including Nigeria. In ethnomedicine, it is used to treat hemorrhages, fevers, lower cholesterol levels and serves as a cough suppressant, emollient, and diuretic. This study evaluated the *in vitro* and antidiarrhea activities of *Peperomia pellucida*. For the *in vitro* antioxidant assay the following methods were used: 2-diphenyl-2-picrylhydrazyl (DPPH) scavenging activity, Superoxide scavenging activity, Hydrogen peroxide scavenging activity, Lipid Peroxidation Assay, Total Antioxidant Capacity, 2,2'-Azinobis (3-Ethylbenzothiazoline-6-Sulphonic Acid) (ABTS) radical scavenging activity and Hydroxyl radical scavenging activity. Antidiarrheal activities were studied using Castor Oil-Induced and Intestinal Transit in Mice models. Methanol and aqueous plant extract of *Peperomia pellucida* at various concentration demonstrated impressive *in vitro* antioxidant scavenging activities. The onset of diarrheal (\*\*P<0.01) and the % of charcoal travel (\*P<0.05) in the mice was reduced at 200 mg/kg compared to the control. Conclusively, this study showed that the aqueous and methanol extract of *Peperomia pellucida* has *in vitro* free radical scavenging activities. The plant demonstrated antidiarrheal activities.

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Plants rich in antioxidants and phytochemicals like tannins and alkaloids are reported to relieve cases of diarrhea (Loganga and Foriers, 2010). Due to the increasing side effects and development of resistance to orthodox medicine, new and safe drug sources are being discovered from plant origin. *Peperomia pellucida* belongs to the family Piperaceae. It is an herbaceous plant found mainly in America, Africa and Asia. The species develops during rainy periods and thrives in damp, humid soils and under the shade of trees (Majumder *et al.*, 2011). In ethnomedicine, it is used to treat hemorrhages, fevers, lower cholesterol levels and serves as a cough suppressant, emollient, and diuretic (Zubair *et al.*, 2015). Hence, this study is carried out to evaluate the antidiarrhea and *in vitro* antioxidant activities of *Peperomia pellucida*.

### MATERIALS AND METHODS

**Chemicals:** Methanol, distilled water, 2,2-diphenyl-1-picrylhydrazyl (DPPH), ascorbic acid, ethylenediaminetetraacetic acid (EDTA), sodium cyanide, nitroblue tetrazolium (NBT), phosphate buffer solution (PBS), riboflavin, hydrogen peroxide, deoxyribose, ascorbate, potassium dihydrogen phosphate, potassium hydroxide, ferric chloride,

thiobarbituric acid (TBA), 2,2'-Azinobis (3-Ethylbenzothiazoline-6-Sulphonic Acid) (ABTS), potassium persulfate, ferrous sulfate, acetic acid, trichloroacetic acid (TCA), butanol, conc. tetraoxosulphate (VI) acid, sodium phosphate, ammonium molybdate, sodium chloride, castor oil, activated charcoal and loperamide. All chemicals were of analytical grade and purchased from a reputable chemical store in Benin City.

**Authentication of *Peperomia pellucida*:** Fresh plants of *Peperomia pellucida* were collected from a farmland in Ogbson, Ikpoba Hill, Ikpoba Okha Local Government Area in Benin, Edo State, Nigeria. The plant was identified and authenticated by a taxonomist, Dr. H. A. Akinnibosun in the Department of Plant Biology and Biotechnology, Faculty of Life Sciences, University of Benin.

**Aqueous Extraction:** Fresh whole plants of *Peperomia pellucida* were washed with running water to remove dirt. 30 g of the plant was blended with 100 ml of distilled water using an electric blender. The blended mixture was transferred into a large air tight glass container. The mixture was shaken vigorously intermittently for 24 hours. Thereafter the mixture was



reacts with two molecules of thiobarbituric acid (TBA), yielding a pinkish red chromogen with an absorbance maximum at 532 nm. Egg homogenate (250 µl, 10 % in distilled water, v/v) and 50 µl of extract were mixed in a test tube and the volume was made up to 500 µl, by adding distilled water. Finally, 25 µl "FeSO<sub>4</sub>" (0.07 M) was added to the above mixture and incubated for 30 min, to induce lipid peroxidation. Thereafter, 750 µl of 20 % acetic acid (pH 3.5) and 750 µl of 0.8 % TBA (w/v) (prepared in 1.1 % sodium dodecyl sulphate) and 25 µl 20% TCA were added, vortexed, and then heated in a boiling water bath for 60 min. After cooling, 3.0 ml of 1-butanol was added to each tube and centrifuged at 3000 rpm for 10 min. The absorbance of the organic upper layer was measured against 3 ml butanol at 532 nm. For the blank 50 µl of distilled water was used in place of the extract.

**Total antioxidant capacity:** Phosphomolybdate assay system was used to determine the total antioxidant activity of the methanol extract and various fractions (Umamaheswari, 2008). To a reagent solution; sulphuric acid (0.6 M), sodium phosphate (2 mM) and ammonium molybdate (4 mM); 100 µl of each sample was added and incubated at 95 °C in a water bath for 90 min. After cooling to room temperature; absorbance was recorded at 765 nm against reagent blank. Total antioxidant capacity of the ascorbic acid was also estimated for reference. The total antioxidant capacity was determined by using following formula:

Calculated as % scavenging activity (I%) using the following equation:

$$I\% = \frac{A_0 - A_1}{A_0} \times 100$$

Where; A<sub>0</sub>- Absorbance of control; A<sub>1</sub>- Absorbance in the presence of plant extract.

**2,2'- Azinobis (3-Ethylbenzothiazoline-6-Sulphonic Acid) (ABTS) radical scavenging activity:** The ABTS•<sup>+</sup> Stock solution was prepared by reacting ABTS Aqueous solution (7 mM) with 2.45 mM aqueous solution of potassium persulfate in equal amount: the mixture was allowed to stand in the dark at room temperature for 12-16 hours before use. The working solution of ABTS•<sup>+</sup> was obtained by diluting the stock solution in methanol to give an absorbance of 0.70 at a wavelength of 734 nm. Then, 2.0 ml of ABTS•<sup>+</sup> solution was mixed with 1 ml of various concentrations of extract of different solvents of *Peperomia pellucida*. The mixture was then incubated at room temperature for exactly 10 min in the dark. The control was prepared by mixing 2.0 ml of ABTS

•<sup>+</sup> solution with 1 ml of double distilled water. The absorbance was measured against a blank at a wavelength 734 nm using spectrophotometer. Ascorbic acid was used as the standard. The percentage of scavenging activity of each extract on ABTS•<sup>+</sup> was calculated as

$$\% \text{ scavenging activity} = \frac{A_0 - A_1}{A_0} \times 100 \text{ (Jayashree et al., 2015).}$$

Where; A<sub>0</sub>- Absorbance of control; A<sub>1</sub>- Absorbance in the presence of plant extract.

**Hydroxyl radical scavenging activity:** The extent of hydroxyl radical scavenging activity from the fenton reaction was quantified using 2'-deoxyribose oxidative degradation as described by Elizabeth and Rao (1990), modified by (Shinde et al., 2006). Aqueous and methanol extract of various concentration were added into the reaction mixture contained 0.1 ml of deoxyribose, 0.1 ml of FeCl<sub>3</sub>, 0.1 ml of EDTA, 0.1 ml of H<sub>2</sub>O<sub>2</sub>, 0.1 ml of ascorbate, 0.1 ml of KH<sub>2</sub>PO<sub>4</sub>-KOH buffer. The mixture was incubated at 37<sup>o</sup> C for 1 hour. At the end of the incubation period, 1.0 ml of thiobutyric acid (TBA) was added and heated at 95°C for 20 minutes to develop the colour. After cooling, the colour formation was measured spectrophotometrically at 532nm against the blank. The hydroxyl radical scavenging activity was determined as

$$\% \text{ scavenging activity} = \frac{ABc - ABt}{ABc} \times 100$$

Where; ABc = Absorbance of Control; ABt = Absorbance of control

**Antidiarrheal Effect of 70 % Methanol Extract of *Peperomia pellucida* in Mice: Castor Oil-Induced Diarrheal in Mice:** Antidiarrheal activity of methanol plant extract of *Peperomia pellucida* was tested by using castor oil induced method in mice (Uwaya and Idu, 2020). Loperamide was used as standard drug for this study. Nine mice of either sex (20-25 g) were divided into three groups of three mice each. The animals were fasted for 18 hrs prior to the test. Group I mice were treated with distilled water (10 ml/kg), which served as control.

Group II (positive control group) received loperamide (5 mg/kg) which served as positive control  
Group III received 200 mg/kg extracts of *Peperomia pellucida*

All doses were administered orally. After 1 hr, all groups received 0.3 ml of castor oil orally. The

animals were placed in cages lined with adsorbent papers and observed for 4 hrs for the presence of diarrhea defined as watery (wet), unformed stool. The time for first excretion of feces and the total number of fecal output by the animals were recorded. The adsorbent paper was weighed before and after the test. The activity of each group was expressed as percentage (%) inhibition of diarrhea. The percentage (%) inhibition of diarrhea was calculated as follows:

$$\% \text{ inhibition of diarrhea} = \frac{A-B}{A} \times 100$$

Where *A* indicates the mean number of diarrhea feces in control group and *B* indicates the mean number of diarrhea feces in test group.

*Castor Oil Induced Intestinal Transit in Mice:* This test was done according to the method of Aye-Than *et al.* (1989) and Uwaya and Idu. (2020). For this test, selected mice were divided into three groups of three mice in each and fasted overnight. 0.1 ml castor oil was given orally in every mouse of each group to produce diarrhea. After 30 mins:

Group I (control group) received distilled water (10ml/kg) orally.

Group II received standard drug (loperamide 5 mg/kg) orally

Groups III received 70 % methanol extract of *P. pellucida* (200 mg/kg) orally

After 1 hr, all animals received 10ml/kg of charcoal meal (10% charcoal suspension in 5% normal saline) orally.

20mins following the charcoal meal administration, all animals were euthanized (sacrificed) via cervical dislocation. The total length and distance covered by the charcoal meal in the intestine, from the pylorus to the caecum was measured and expressed as percentage of the total length of the small intestine from pylorus to the caecum.

$$\text{Intestinal transit \%} = \frac{LTC}{TLSI} \times 100$$

Where; *LTC* – Length travelled by Charcoal; *TLSI* – Total length of small intestine

$$\text{Percentage Inhibition of intestinal transit} = \frac{T_0 - T_1}{T_0} \times 100$$

Where; *T<sub>0</sub>*- intestinal transit of control; *T<sub>1</sub>*- intestinal transit of extract

*Statistical analysis:* Results were represented as mean± standard error of mean (S.E.M.), and ‘n’ represents the numbers of mice per experimental group. Data were subjected to one- way analysis of variance (ANOVA) and differences between samples were determined by Tukey's multiple comparisons test. All data were analyzed using graph pad prism (UK) software version 6. \*P<0.05 indicate significant difference between compared data.

## RESULT AND DISCUSSION

*In vitro* antioxidant activity result: Figure 1 shows that the methanol and aqueous plant extract of *Peperomia pellucida* inhibits DPPH radical activity. The results agree with previous studies (Mutee *et al.*, 2010). But aqueous extract at 0.2 µg/ml gave a better scavenging activity of 83.53 % comparable to that of ascorbic acid which at the same concentration gave a scavenging activity of 83.58 %. DPPH assay is one of the standard and easy colorimeter methods for evaluating antioxidant activities of pure compounds and also plants like cereals, vegetables, herbs and edible seed oils in food and biochemical industries (Cheng *et al.* 2006). Therefore, this assay shows the rich antioxidant activities of *Peperomia pellucida*. Figure 2 shows that the methanol and aqueous plant extract of *Peperomia pellucida* scavenge Superoxide free radicals. The results agree with previous studies (Merlin and Jyoti, 2018). But aqueous extract at the highest concentration scavenged superoxide free radicals better than ascorbic acid with 91.35 % scavenging activity while ascorbic acid had 66.66% scavenging activity. Superoxide play an important role in the formation of other reactive oxygen species such as hydrogen peroxide, hydroxyl radical and singlet oxygen, which induce oxidative damage in lipids, protein and Deoxyribonucleic Acid (DNA) (Venkatachalam and Muthukrishnan, 2012). Also, superoxide has been observed to directly start lipid peroxidation (Ilhami *et al.*, 2010). *Peperomia pellucida* can thus prevent radical chain reaction of superoxide. The methanol and aqueous plant extract of *Peperomia pellucida* scavenge Hydrogen Peroxide free radicals (Figure 3). But at 0.2 µg/ml methanol extract gave better result (92.75 % scavenging activity) than ascorbic acid (74.56 % scavenging activity). The aqueous and methanol plant extract of *Peperomia pellucida* scavenge ABTS free radicals (figure 4). But methanol extract at 0.2 µg/ml had 62.77 % scavenging activity better than that of ascorbic acid having 52.96 %. ABTS radical activity is frequently used for estimating the total antioxidant capacity (TAC) of natural products, including crude extracts. The presence of an antioxidant compound inhibits the formation of ABTS<sup>4+</sup> which is read spectrometrically (Jian *et al.*, 2009). This shows that the crude extract of

*Peperomia pellucida* is rich with antioxidants. Figure 5 shows that the methanol and aqueous plant extract of *Peperomia pellucida* reduced Lipid peroxidation activity. But methanol extract had the best Lipid peroxidation reduction activity (66.95 %) at 0.2 µg/ml comparable to that of ascorbic acid having 91.85% lipid peroxidation reduction activity. Lipid peroxidation is a chain of reactions of oxidative degradation of lipids. This process proceeds by a free radical chain reaction mechanism and has been observed in atherosclerosis, heart failure, cancer and other immunological disorders (Satish and Dilipkumar, 2015). Thus, *Peperomia pellucida* can prevent such radical induced illness. Figure 6 shows that the methanol and aqueous plant extract of *Peperomia pellucida* scavenge Hydroxyl free radicals. The results agree with previous studies (Merlin and Jyoti, 2018). But aqueous extract at 0.2 µg/ml had a better scavenging activity of 61.43% comparable to that of ascorbic acid having 76.05%. Among the oxygen radicals, hydroxyl radical is the most reactive and induces severe damage to the adjacent biomolecules such as all proteins, nucleic acid, and almost any biological molecule it touches.

This damage causes aging, cancer and several diseases. Therefore, the removal of hydroxyl radical is probably one of the most effective defenses of a living body against various diseases (Zhu *et al.*, 2006). Thus, *Peperomia pellucida* may help prevent diseases associated with free radicals like cancer. The methanol and aqueous plant extract of *Peperomia pellucida* had the good Total Antioxidant Capacity. But methanol extract had the best Total Antioxidant Capacity 95.63% comparable to that of ascorbic acid having 97.3% at 0.2 µg/ml. (Figure 7).

Reactive oxygen species cause damage to cellular biomolecules such as proteins, nucleic acids, lipids and carbohydrates. Antioxidants interfere with the generation of ROS and also play a crucial role in their inactivation. (Adjimani and Asare, 2015). Under stressful condition, human body produces reactive oxygen species (ROS) which leads to cell damage and health-related problems (Bhadane, Patil and Arts, 2017). like cell aging, cardiovascular diseases, mutagenic changes, and cancerous tumor growth (Ozyurek *et al.*, 2011). Therefore, *Peperomia pellucida* as an excellent source of antioxidants may help to prevent cases of oxidative stress induced illness like cancer, asthma, diabetes, senile dementia, cardiovascular and inflammatory diseases.

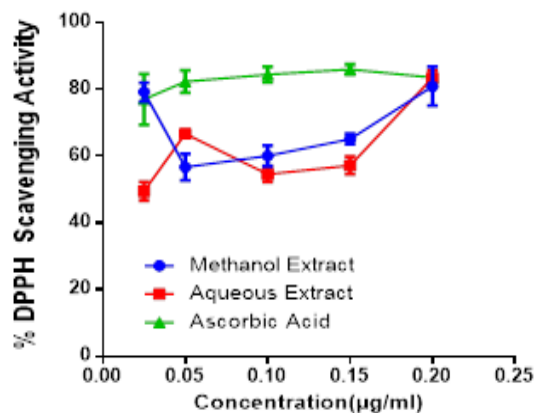


Fig 1: 2, 2-diphenyl-2-picrylhydrazyl hydrate (DPPH) radical scavenging action of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean ± S.E.M., n =3 per group.

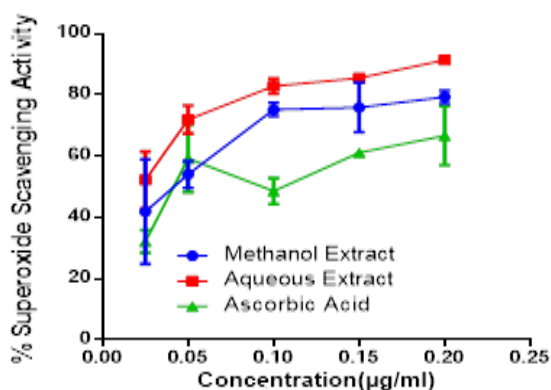


Fig 2: Superoxide radical scavenging action of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean ± S.E.M., n =3 per group.

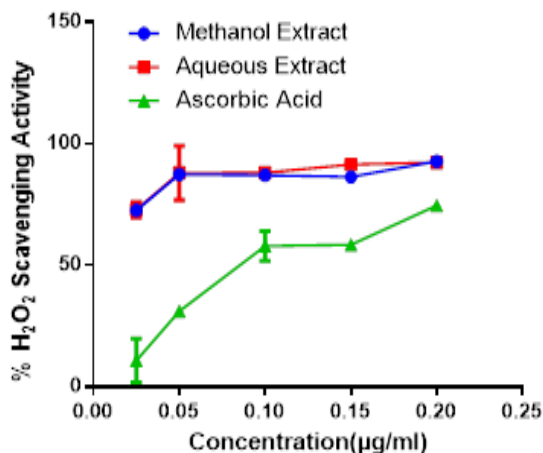
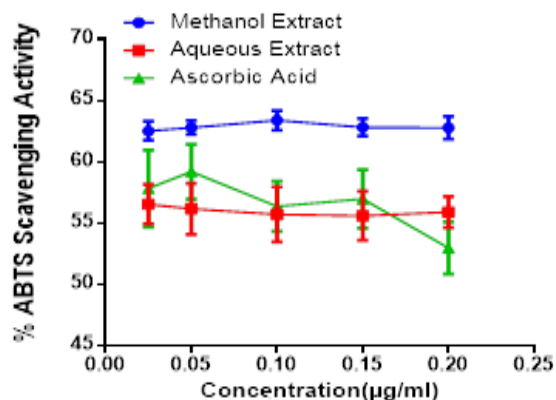
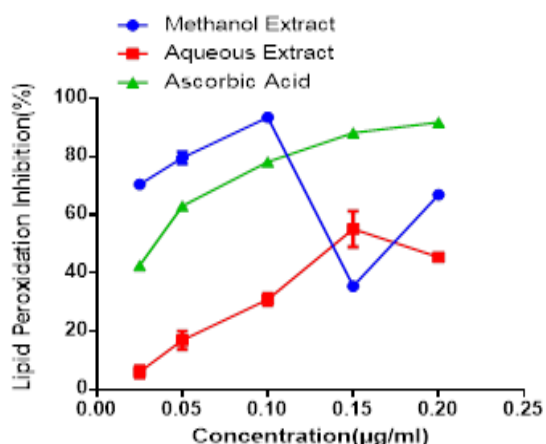


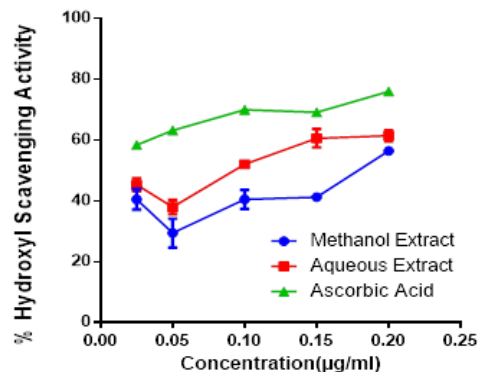
Fig 3: Hydrogen Peroxide radical scavenging action of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean ± S.E.M., n =3 per group.



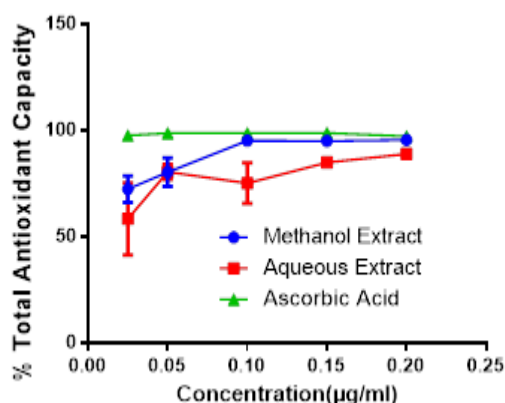
**Fig 4** 2,2- azino bis-3-ethyl benzthiazoline-6 sulphonic acid (ABTS) radical scavenging action of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean  $\pm$  S.E.M., n=3 per group.



**Fig 5** % Lipid peroxidation inhibition of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean  $\pm$  S.E.M., n=3 per group



**Fig 6** Hydroxyl radical scavenging action of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean  $\pm$  S.E.M., n=3 per group.



**Fig 7** Total antioxidant capacity of methanol and aqueous plant extract of *Peperomia pellucida*. Values are represented as mean  $\pm$  S.E.M., n=3 per group

**Table 1:** Effect of 70 % Methanol Extract *Peperomia pellucida* on Castor oil- induced diarrhea in Mice

Groups	Treatment	Weight of Diarrhea Faeces (g)	Onset of Diarrhea (mins)	Total No of Diarrhea Faeces	% Inhibition of Diarrhea
CONTROL	Castor oil + Distilled water (2ml/kg p.o)	0.35 $\pm$ 0.02	4.44 $\pm$ 1.79	7.33 $\pm$ 0.33	-
STANDARD	Castor oil+ Loperamide (5mg/kg i.p)	0.22 $\pm$ 0.12	171.60 $\pm$ 25.17***	1.67 $\pm$ 0.88**	77.27%
200 mg/kg	Castor oil + Extract of plant	0.30 $\pm$ 0.01	27.85 $\pm$ 0.59**	6.67 $\pm$ 0.88	9.08%

Values were expressed as mean S.E.M. (n= 3). Statistically significant difference from control \*\*P< 0.01, \*\*\*P<0.001

**Castor oil- induced diarrhea result:** The 70 % methanol plant extract of *Peperomia pellucida* at 200 mg/kg reduced (\*\*P<0.01) the onset of diarrheal in the mice compared to the control as seen in Table 1. There was no significant reduction in the total no of diarrheal feces and the weight.

**Intestinal transition result:** The methanol plant extract of *Peperomia pellucida* at 200 mg/kg reduced

(\*P<0.05) the % of charcoal travel in the mice compared to the control as seen in Table 2. The 70 % methanol plant extract of *Peperomia pellucida* at 200 mg/kg was seen to suppress GIT motility with 28.16 %. Research has shown the presence of reducing sugar, tannin, steroid and alkaloid in *Peperomia pellucida* (Pulak *et al.*, 2011). Earlier studies showed that anti-dysenteric and antidiarrheal activities of medicinal plants were due to tannins, alkaloids,

saponins, flavonoids, sterols, tannins and phenolics present in plant extracts (Loganga and Foriers, 2010; Galvez *et al.*, 2013). They are reported to inhibit release of autacoids and prostaglandins, thereby inhibit motility (Loganga and Foriers, 2010; Galvez *et al.*, 2013). The antidiarrheal activity of *Peperomia*

*pellucida* leaves may be because of its phytochemical constituents. The plant demonstrated a better effect of antidiarrheal activity via suppression in GIT motility. Thus, *Peperomia pellucida* can help in managing a diarrheal condition.

**Table 2:** Effect of 70 % Methanol Extract *Peperomia pellucida* on Intestinal transition in Mice

Groups	Treatment	Total Length of Intestine (cm)	Distance Traveled By Charcoal (cm)	% of Charcoal Travel	% Suppression on GIT Motility
CONTROL	Castor oil + Distilled water (2ml/kg p.o)	35.59 ± 0.02	32.92±4.57	92.50 ±3.90	–
STANDARD	Castor oil+ Loperamide (5mg/kg i.p)	39.43± 4.68	27.83±4.57	70.58± 5.3*	29.42%
200 mg/kg	Castor oil + Extract of plant	41.72±3.91	29.97±0.95	71.83 ±2.74*	28.16 %

Values were expressed as mean S.E.M. ± n= 3. Significant difference from control \*P< 0.05.

**Conclusion:** This study showed that the aqueous and methanol extract of *Peperomia pellucida* demonstrated impressive *in vitro* antioxidant activities. The plant also demonstrated an antidiarrheal activity via suppression on GIT motility.

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