



## Distribution of metals in *Labeo coubie* (Ruppel, 1832) from a National Park river in Nigeria

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**ABSTRACT:** This study assesses heavy metals distribution in body parts of *Labeo coubie* (African carp) from River Oli, in Kainji Lake National Park as pollution index of the ecosystem. Fish parts samples (gills, muscles and vertebra bone) were prepared and specifically analyzed for the levels of Pb, Cr, Zn, Cu, Fe and Cd using Atomic absorption spectrophotometry. The concentration of metals in the samples at different concentrations ranged from  $0.001 \pm 0.000$   $\mu\text{g/g}$  for Cd to  $224.87 \pm 4.07$   $\mu\text{g/g}$  of Fe in the fish gill. There is significant ( $p < 0.05$ ) differences in the Pb, Zn, Cu and Fe concentrations across the different fish parts with gills accumulated the highest levels metals while Cr levels significantly ( $p < 0.05$ ) differed in the fish body parts and accumulated more in the muscles ( $10.75 \pm 0.15$   $\mu\text{g/g}$ ). The mean concentrations of metal elements in the fish parts had shown some distinguish connection in its distributions with Pb and Cu; Gills > Vertebra bones, Fe and Zinc; Gills > Vertebra bones > Muscles while Cr was distributed in Muscles > Gill > Vertebra bones. However, it is revealed that *Labeo coubie*, a euryphagus fish probably absorb these metals through ingestion of contaminated food or absorption by the gills and bioaccumulate in different fish parts. It is therefore established that River Oli is contaminated with heavy metals as presence of these metals in fish is an indication of its immediate environment.

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Environmental pollution is generally defined as the contamination of water, soil, or the atmosphere by the discharge of substances that are harmful to living things (Obianime *et al.*, 2017). In this realm, heavy metals are typical pollutants in aquatic environments which are of immediate concern due to their persistence in the environment and toxicity to humans (Alhassan *et al.*, 2016). In different water bodies, heavy metal pollution results from direct atmospheric deposition, geologic weathering or through the discharge of agricultural, municipal, residential or industrial waste products (Dhanakumar *et al.*, 2015; Garcia *et al.*, 2015 and Demirak *et al.*, 2006) and they can have a negative impact on aquatic ecosystems, the food chain and human health (Arantes *et al.*, 2016). Heavy metals can be taken up into fish either from ingestion of contaminated food via the alimentary tract or through the gills and skin (Sfankianakis *et al.*, 2015 and Drevnick *et al.*, 2006). The quantification of potential contaminants in fish tissues can be an important part of water quality assessment programs (Oliveira-Ribeiro *et al.*, 2005) because it can reflect levels found in sediment and water and its distribution in the particular aquatic environment from which they

are sourced (Nhiwatiwa *et al.*, 2011). Heavy metals have been reported to change the genetic, physiological, biochemical and behavioral parameters of aquatic organisms including fish (Mahboob *et al.*, 2016). Fish have been the most popular choice as test organisms for heavy metals because they are presumably the best understood organisms in the aquatic environment and are an important source of protein to man (Murtala *et al.*, 2012). Accumulated heavy metals may lead to morphological alterations in the tissues of fish (Monteiro *et al.*, 2005) and many cause death or sub-lethal pathology of liver, kidneys, reproductive system, respiratory system or nervous system in both invertebrate and vertebrate aquatic animals (Mahboob *et al.*, 2016). However, heavy metals have devastating effects on ecological balance of the recipient environment and a diversity of aquatic organisms (Farombi *et al.*, 2007). Concentrations of these heavy metals in the fish tissues are a critical issue that needs to be addressed as levels of metals in fish usually reflects its presence in water and sediment of the particular water body. There is need for a better understanding of heavy metals composition in different body parts of fish. Therefore, this study

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relatively high and ranged from  $180.33 \pm 4.19 \mu\text{g/g}$  in fish muscles to a significant high value of  $224.87 \pm 4.07 \mu\text{g/g}$  in the gills. There are low levels of Cd in the fish samples where concentration in gill is below detectable level while  $0.001 \pm 0.000$  and  $0.001 \pm 0.001$  were recorded for muscles and vertebra bones respectively. Figure 2 depicted the mean concentrations of heavy metals in different body parts of the fish against the maximum tolerable level as recommended by World Health Organisation, 2008. Pb levels in muscles and bones are lower but its concentration of  $3.20 \pm 0.16 \mu\text{g/g}$  in the gills is exceed  $2 \mu\text{g/g}$  maximum tolerable level recommended. Cr levels that ranged from  $0.35 \pm 0.04 \mu\text{g/g}$  and  $10.75 \pm 0.15 \mu\text{g/g}$ , Zn ( $14.75 \pm 0.04 \mu\text{g/g}$  and  $30.60 \pm 2.49 \mu\text{g/g}$ ) and Cd ( $0.001 \mu\text{g/g}$ ) are below tolerable level of  $50 \mu\text{g/g}$  (Cr),  $75 \mu\text{g/g}$  (Zn) and  $<1$  (Cd) recommended for corresponding elements. More also Fe concentrations in all fish parts are considerably above the allowable limit of  $100 \mu\text{g/g}$  while only Cu level of  $53.55 \pm 3.23 \mu\text{g/g}$  in gills exceeded the acceptable level of  $30 \mu\text{g/g}$ . Presence of some these metals at high concentrations are considered as a dangerous source of water pollution, because of its consequential effects on the aquatic resources. These metals are known to induce oxidative stress and/or carcinogenesis by mediating free radicals/reactive oxygen species (Javed *et al.*, 2015). At trace levels, some heavy metals (e.g. Cu, Fe, Zn and Cr) are essential to maintain important biological roles including metabolism of the biological organism such as fish (Abadi *et al.*, 2014) while inadequate supply of these micronutrients could results in a variety of deficiency diseases or syndromes (Kennedy, 2011). However, toxicity could also occur at excessive concentration of these metals (Sivaperumal *et al.*, 2007) leading to a variety of adverse effects and fish diseases. On the other hand, non-essential metals such as Cd and Pb have no proven biological function and their toxicity rises with increasing concentrations (Sfakianakis, 2015) and therefore can lead to poisoning (Binkowski, 2012). From the study, the presence of heavy metals in the different body parts of *Labeo coubie* confirmed the report that heavy metals entering the water bodies can be deposited into aquatic organisms through the effects of bioconcentration, bioaccumulation and the food chain process (Abdel-Baki *et al.*, 2011). It is further evident that heavy metals accumulate in the fish muscles, internal organs and bones as earlier reported in various studies (Kehinde and Adelakun, 2019; Agbon and Omoniyi, 2010; Golonova, 2008 and Dural *et al.*, 2007). Presence of Pb in all fish parts examined in the study agreed with Omwenga, (2003) that Pb accumulates in the bones and soft tissues of fish. Aquatic organisms bio accumulate Pb from water and diet, although there is evidence that Pb

accumulation in fish, is most probably originated from contaminated water rather than diet (Creti *et al.*, 2010). Although Pb is a naturally occurring substance, its environmental concentrations are significantly increased by anthropogenic sources including lead containing pesticides, through precipitation, fallout of lead dust, road runoff, and community wastewater (Sepe, 2003). Lead (Pb) concentrations recorded in the study (table 1) are comparatively higher to the findings of Okafo *et al.*, (2018), who reported range of  $0.30 \pm 0.15 \mu\text{g/g}$  and  $0.39 \pm 0.18 \mu\text{g/g}$  for *Labeo species* from River Kaduna. Similarly, lower concentrations ranges between  $0.002 \pm 0.001 \mu\text{g/g}$  to  $0.024 \pm 0.004 \mu\text{g/g}$  in gills and  $0.002 \pm 0.000 \mu\text{g/g}$  to  $0.004 \pm 0.001 \mu\text{g/g}$  in muscles of economic important fishes of Aiba reservoir, Nigeria has been documented (Atobatele and Olutona, 2015). However, even at low levels, Pb pollution could cause some adverse effects on fish health and reproduction (Delistry and Stone, 2007). Highest accumulation of Pb found in gills from the study affirmed its deposition in various fish organs including fish gills (Jeziarska and Witeska, 2006), leading to disorders in fish body. The characteristic symptoms of chronic Pb toxicity include changes in the blood parameters with severe damage to erythrocytes and leucocytes and damage in the nervous system (El Badawi, 2005). The concentration of Cr significantly varied in all samples and below the tolerable level of  $50 \mu\text{g/g}$  (WHO, 2008), though higher than the concentrations of  $0.02 \pm 0.01 \mu\text{g/g}$  to  $0.40 \pm 0.13 \mu\text{g/g}$  in the different organs of the six species of fishes from Lake Chad in Doron Buhari, Maiduguri, Borno State, Nigeria (Akan *et al.*, 2009). Chromium is an essential nutrient metal, necessary for metabolism of carbohydrates (Farag *et al.*, 2006). Fish assimilate Cr by ingestion or by the gill uptake tract and accumulation in fish tissues, mainly liver, occurs at higher concentrations than those found in the environment (Pacheco *et al.*, 2013 and Ahmed *et al.*, 2013). The overall toxic impact on organs like gill, kidney and liver may seriously affect the metabolic, physiological activities and could impair the growth and behavior of fish (Vinodhini and Narayanan, 2008). Toxic effects of Cr in fish include: hematological, histological and morphological alterations, inhibition/reduction of growth, production of reactive oxygen species (ROS) and impaired immune function (Reid, 2011 and Vera-Candioti *et al.*, 2011). The mean levels of Zn in this study fall within  $14.56 \pm 0.48 \mu\text{g/g}$  and  $51.82 \pm 2.75 \mu\text{g/g}$  reported for *Chrysichthys nigrodigitatus* in the same study area (Adelakun and Kehinde, 2019), much higher than  $2.19 \pm 0.39 \mu\text{g/g}$  reported for *Labeo species* from River Kaduna (Okafo *et al.*, 2018) but lower than  $65.33 \mu\text{g/g}$  documented for *Tilapia Zilli* from the lower reaches of River Niger (Obodo, 2002). Zn plays a vital role in the

physiological and metabolic process of many organisms (Rajappa *et al.*, 2010). It is essential element in animal diet because it helps in protein synthesis (Amundsen *et al.*, 1997) but may become toxic to fish at concentration above 75 µg/g (WHO, 2008) and could result to mortality, growth retardation, and reproductive impairment (Giardina *et al.*, 2008). However, Zinc does not appear to present a contaminant hazard to *Labeo coubie* within the River Oli catchment of Kainji Lake National Park. For Cu, which can get into aquatic ecosystems from diverse sources for example, from Cu compounds used in agro chemicals, wood preservatives, tie and dye manufacture (Akan *et al.*, 2010). Also, from Cu compounds added in fertilizers and animal feeds as a nutrient to support plant and animal growth. Cu recorded in the study is higher than 7.04±0.03 µg/g in *Chrysichthys nigrodigitatus* and 9.51±0.10 µg/g in *Parachanna obscura* from Ibiekuma stream, Ekpoma, Nigeria (Erhabor *et al.*, 2010) as well as 1.57±0.26 µg/g in *Cynothrissa mento* from Ologe Lagoon, Lagos, Nigeria (Kumolu-Johnson *et al.*, 2010) but is lower than the range of 860-1620 µg/g reported by Anetekhai *et al.* (2007) in *M. vollenhovenii* (a non-fin fish) from the same Ologe Lagoon. This has been associated to greater metal load in Ologe Lagoon because of the presence of Agbara Industrial Estate, which discharges its waste into the lagoon (Kusemiju *et al.*, 2001). However, Cu concentration in gills of the fish in the present study exceeded maximum tolerable level of 30 µg kg<sup>-1</sup> (WHO, 2008) and high doses of Cu may cause anaemia, liver and kidney damage, stomach and intestinal irritation (Tirkey *et al.*, 2012). The high Cu levels in *Labeo coubie* in the study could be attributed to agricultural activities in the catchment especially the use fertilizers and agro chemicals. The River Oli catchment is a high potential area for agricultural practices including crops farming and animal husbandry, therefore Cu compounds added in fertilizers and animal feeds get into river through surface runoff especially during the rainy season. This study show that Iron (Fe) is the most bioaccumulated of all metals in the fish body parts (Table 1), though fall within 126.23 ± 0.06 µg/g and 560.63 ± 0.03 µg/g reported for different body parts of *Chrysichthys nigrodigitatus* and *Parachanna obscura* from Ibiekuma stream, Nigeria (Erhabor *et al.*, 2010). However, the values reported for Fe in the study exceed maximum tolerable level of 100µg/g (WHO,

2008). High concentration of Fe in fish gills as evident in the study could cause respiratory distress in fish due to physical clogging of the gills (Dalzell and Macfarlane, 1999) because the precipitated Fe compounds could reduce the gills area available for respiration causing damage to the respiratory epithelium and eventual suffocation of the fish (Abbas *et al.*, 2002). This investigation revealed that Cd values obtained in this study fall within 0.002 ± 0.000µg/g reported for muscles of *Labeo species* in Aiba reservoir, Nigeria (Atobatele and Olutona, 2015). However, small quantities of cadmium could interfere with fish enzymes and cause diseases (Rajappa *et al.*, 2010) including bone defects in animals (Tirkey *et al.*, 2012). Cd was practically less bioaccumulated by the sampled fish species in this study. This could be attributed to the fact that the fishes were conceivably able to excrete the metal at a rate that exceeded the uptake of the metal (Wangboje *et al.*, 2013).

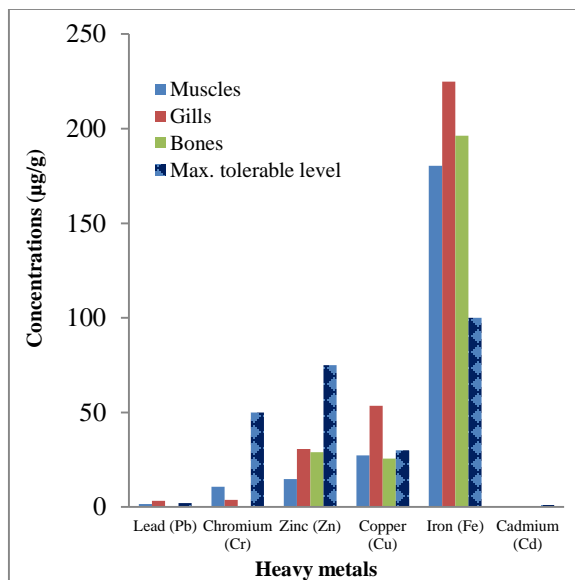
#### *Patterns of heavy metals accumulation in body parts (muscles, gills and vertebra bones) of Labeo coubie:*

The mean concentrations of metal elements in the fish parts had shown some connection in its bioaccumulation patterns. Pb and Cu had similar bioaccumulation forms of Gills > Muscles > Vertebra bones. Fe and Zinc, which are important essential metals exhibited Gills > Vertebra bones > Muscles accumulation pattern while Cr differed with all other metals forms with Muscles > Gill > Vertebra bones order of bioaccumulation in *Labeo coubie* (see table 2). It is apparent from the study that gill samples accumulated highest level of Pb, Cu, Zn and Fe, thus corroborated Storelli *et al.* (2006) and Rashed (2001); that gill is the centre of metal accumulation in fish as a results of its important role in interface with the environment in performing its functions in gas exchange, ion regulation, acid balance and waste excretion (Bajc *et al.*, 2005; Filazi *et al.*, 2003; Shukla *et al.*, 2007). The distribution pattern (gills < muscles < bones) of most metals in the study conformed closely with the work done by (Golnova, 2008) this could be as a result of gills direct metal uptake from water (Storelli *et al.*, 2006) which can be influenced by absorption of metals on to the gill surface (Erdogru and Erbilir, 2007; Dural *et al.*, 2006) while muscles (Uysal *et al.* 2009; Bervoets and Blust, 2003) and bones (Akan *et al.*, 2009) comparably considered to have weak accumulation potential.

**Table 1.** Mean (±) concentration of heavy metals in muscles, gills and vertebra bones of *Labeo coubie* from the study area (µg/g dry weight).

Fish parts	The mean concentration of heavy metals in fish flesh (µg/g)					
	Pb	Cr	Zn	Cu	Fe	Cd
Muscles	1.50±0.02 <sup>b</sup>	10.75±0.15 <sup>c</sup>	14.75±0.04 <sup>a</sup>	27.35±0.23 <sup>a</sup>	180.33±4.19 <sup>a</sup>	0.001±0.000
Gills	3.20±0.16 <sup>c</sup>	3.80±0.22 <sup>b</sup>	30.60±2.49 <sup>b</sup>	53.55±3.23 <sup>b</sup>	224.87±4.07 <sup>b</sup>	BDL
Bones	0.45±0.02 <sup>a</sup>	0.35±0.04 <sup>a</sup>	29±10.87 <sup>b</sup>	25.65±5.54 <sup>a</sup>	196.30±21.00 <sup>a</sup>	0.001±0.001

Note: Values are mean values ± standard error. Mean with different superscript within the same column are significantly different (p<0.05)



**Fig 2.** Mean concentrations of heavy metals in the muscles, gills and vertebra bones of *Labeo coubie* from the study area against WHO (2008) maximum tolerable level.

**Table 2.** Bioaccumulation patterns of heavy metals in body parts (muscles, gills and vertebra bones) of *Labeo coubie*

Metal elements	Pattern of the bioaccumulation of heavy metal fish parts
Pb	Gills > Muscles > Vertebra bones
Cr	Muscles > Gill > Vertebra bones
Zn	Gills > Vertebra bones > Muscles
Cu	Gills > Muscles > Vertebra bones
Fe	Gills > Vertebra bones > Muscles

**Conclusion:** There is the possibility of the heavy metals originating from the chemical fertilizers and all forms of pesticides in the surrounding communities of the Park possibly being washed down into the river through runoff. Hence, *Labeo coubie*, a euryphagus fish probably absorb these metals through ingestion of contaminated food or by absorption by the gills and bioaccumulate in different body parts. It is then pertinent to facilitates appropriate action towards protection and sustainable environment for wildlife conservation in the Park and its surrounding communities.

**REFERENCES**

Abadi, DRV; Dobaradaran, S; Nabipour, I; Lamani, X; Ravanipour, M; (2014). Comparative investigation of heavy metal, trace, and macro element contents in commercially valuable fish species harvested off from the Persian Gulf. *Environ Sci Pollut Res.*

Abbas, HH; Zaghloul, KH; Mousa, MA (2002). Effect of some heavy metal pollutants on some biochemical and histopathological changes in

Blue tilapia, *Oreochromis aureus*. *Egypt. J. Agric. Res.*, 80: 1395-1411.

Adelakun, KM; Kehinde, AS (2019). Heavy metals bioaccumulations in *Chrysichthys nigrodigitatus* (Silver catfish) from River Oli, Kainji Lake National Park, Nigeria. *Egyptian Journal of Aquatic Biology & Fisheries* 23(5): 253 – 259

Agbon, AO; Omoniyi, IT (2010). Heavy metals in some Fish species of Oyan Lake, Southwest Nigeria. *Nig. J. Fisheries*. Vol.7 No1 and 2: 91 - 99.

Ahmed, MK; Kundu, GK; Al-Mamun, MH; Sarkar, SK; Akter, MS; Khan, MS (2013). Chromium (VI) induced acute toxicity and genotoxicity in freshwater stinging catfish, *Heteropneustes fossilis*. *Ecotoxicol. Environ. Saf.* 92: 64-70.

Akan, JC; Abdulrahman, FI; Sodipo, OA; Akandu, PI (2009). Bioaccumulation of Some Heavy Metals of Six Fresh Water Fishes Caught from Lake Chad in Doron Buhari, Maiduguri, Borno State, Nigeria. *J. Appl. Sci. Environ. Sanit.* 4 (2): 103-114.

Akan, JC; Abdulrahman, FI; Sodipo, OA; Chiroma, YA (2010). Distribution of Heavy Metals in the Liver, Kidney and Meat of Beef, Mutton, Caprine and Chicken from Kasuwan Shanu Market in Maiduguri Metropolis, Borno State, *Res. J. Appl. Sci. Eng. Tech.* 2(8): 743-748.

Alhassan, AB; Balarabe, ML; Gadzama, IMK (2016). Assessment of some heavy metals in macrobenthic invertebrate and water samples collected from Kubanni reservoir, Zaria, Nigeria. *Federal University Wukari Trends in Sci. Tech. J.* 1(1):55-60.

Amundsen, PA; Staldvilk, FJ; Lukin, A; Kashulin, N; Popova, O; Restetnikov, Y (1997). Heavy metals contaminations in fresh water fish from the border region between Norway and Russia. *Sci. Total Environ.* 201:211-4.

Anderson, RA (2005). Essentiality of Chromium in Humans. *Sci. Total Environ.* 86; 75-81

Anetekhai, MA; Akin-Oriola, GA; Aderinola, OJ; Akintola, SL (2007). Trace metal concentration in *Macrobrachium vollenhovenii* from Ologe Lagoon, Lagos, Nigeria. *J. Afrotrop. Zool.*, Special Issue: 25-29.

- Arantes, FP; Savassi, LA; Santos, HB; Gomes, MVT; Bazzoli, N (2016). Bioaccumulation of mercury, cadmium, zinc, chromium, and lead in muscle, liver and spleen tissues of a large commercially valuable catfish species from Brazil. *Annals of the Brazilian Academy of Sciences*, doi.org/10.1590/0001-3765201620140434
- Atobatele, OE; Olutona, GO; (2015). Distribution of three non-essential trace metals (Cadmium, Mercury and Lead) in the organs of fish from Aiba Reservoir, Iwo, Nigeria. *Toxicology Reports 2* (2015) 896–903.
- Ayeni, JSO (2007). Participatory management plan in Kainji Lake National Park. *ENVIRON-CONSULT*: Lagos. 156p.
- Bajc, Z; Gacnik, KS; Jenci, V; Doganoc, DZ (2005). The Content of Cu, Zn and Mn in Slovenian freshwater fish. *Slov. Vet. Res.*, 42, 15-21.
- Bervoets, L; Blust, R (2003). Metal concentrations in water sediment and gudgeon (*Gobiogobio*) from a pollution gradient: relationship with fish condition factor. *Environ. Pollut.* 126: 9-19.
- Binkowski, MN (2012). Preliminary results of cadmium and lead concentration in pectoral muscles of mallards and coots shot in 2006 in southern Poland, *J. Microbiol. Biotech. Food Sci.* 1: 120-1128.
- Cretì, P; Trinchella, F; Scudiero, R (2010). Heavy metal bioaccumulation and metallothionein content in tissues of the sea bream *Sparus aurata* from three different fish farming systems. *Environ. Monit. Assess.* 165: 321-329.
- Dalzell, DJB; Macfarlane, NAA (1999). The toxicity of iron to brown trout and effects on the gills: a comparison of two grades of iron sulphate. *J. Fish Biol.* 55: 301-315.
- Delistraty, D; Stone, A (2007). Dioxins, metals, and fish toxicity in ash residue from space heaters burning used motor oil. *Chemosphere* 68(5): 907-914.
- Demirak, A; Yilmaz, F; Levent-Tuna, A; Ozdemir, N (2006). Heavy metals in water, sediment and tissues of *Leuciscus cephalus* from a stream in southwestern Turkey. *Chemosphere* 63: 1451-1458.
- Dhanakumar, S; Solaraj, G; Mohanraj; R (2015). Heavy metal partitioning in sediments and bioaccumulation in commercial fish species of three major reservoirs of river Cauvery delta region, India. *Ecotoxicol. Environ. Saf.* 113: 145-151.
- Drevnick, PE; Sandheinrich, MB; Oris, JT (2006). Increased ovarian follicular apoptosis in fathead minnows (*Pimephales promelas*) exposed to dietary methylmercury. *Aquat Toxicol* 79: 49-54.
- Dural, DA; Bakare, OK; Ayodele, IA (2007). Trade in the wild mammalian Species for Traditional medicine in Ogun State, Nig. *Global J. Med. Res.* 12 (3): 6-21.
- Dural, M; Goksu, MZI; Ozak, AA; Derici, B (2006) Bioaccumulation of some heavy metals in different tissues of *Cicentrarchus labrax* L., 1758, *Sparus aurata* L., 1758 and *Mugil cephalus* L., 1758 from the Camlik Lagoon of the Eastern Coast of Mediterranean (Turkey). *Environ. Monit. Assess.* 118: 66-74.
- El-Badawi, AA (2005). Effect of lead toxicity on some physiological aspects of Nile tilapia fish, *Oreochromis niloticus*. In: *Inter. Conf. Vet. Res. Div.*, 2005, NRC, Cairo, Egypt.
- Erdogru, O; Erbilir, F (2007). Heavy metals and trace elements in various fish samples from Sir Dam Lake, Kahramanmaras, Turkey. *Environ. Monit. Assess.* 130 373-379.
- Erhabor, CU; Obasohan, EE; Eguavoen, OI (2010). Ecotoxicological studies: Heavy metals contaminating water and fishes from Ibiekuma Stream, Ekpoma, Edo State, Nigeria. *Nig. J. Fisheries.* 7(1&2): 81-90.
- Farag, AM; May, T; Marty, GD; Easton, M; Harper, DD; et al. (2006). The effect of chronic chromium exposure on the health of Chinook salmon (*Oncorhynchus tshawytscha*). *Aquat. Toxicol.* 76: 246-257.
- Farombi, EO; Adelowo, OA; Ajimoko, YR (2007). Biomarkers of oxidative stress and heavy metals levels as indicators of environmental pollution in African catfish (*Clarias gariepinus*) from Ogun River. *Int. J. Environ. Res. Public Health*, 4(2): 158-165.
- Filazi, A; Baskaya, R; Kum, C; Hismiogullar, SE (2003) Metal concentration in tissues of the Black



- sea fish *Mugil auratus* from Sinop –Icliman, Turkey. *Human Exp. Toxicol.* 22, 85-87.
- Garcia, JC; Martinez, DST; Alves, OL; Leonardo, AFG; Barbieri, E (2015). Ecotoxicological effects of carbofuran and oxidized multiwalled carbon nanotubes on the freshwater fish Nile tilapia: Nanotubes enhance pesticide ecotoxicity. *Ecotoxicol. Environ. Saf.*, 111: 131-137.
- Giardina A; Larson BE; Wisner B; Wheeler J; Chao M (2008) Long-term and acute effects of Zinc contamination of a stream on fish mortality and physiology. *Environ. Toxic. Chem.* 28(2): 287-295.
- Golovanova, IL (2008). Effects of heavy metals on the physiological and biochemical status of fishes and aquatic invertebrates. *Inland Wat. Bio.* 1(1):93-101
- Javed, M; Usmani, N; Ahmad, I; Ahmad, M (2015). Studies on the oxidative stress and gill histopathology in *Channa punctatus* of the canal receiving heavy metalloaded effluent of Kasimpur Thermal Power Plant. *Environ. Monit. Assess.* 187: 4179.
- Jeziarska, B; Witeska, M (2006). The metal uptake and accumulation in fish living in polluted waters. *NATO Science Series*, Netherlands: Springer.
- Kehinde, AS; Adalakun, KM (2019). Occurrence of heavy metals in selected fish species of River Oli, Kainji Lake National Park, Nigeria. *SINET: Ethiop. J. Sci.*, 42(1):18-24.
- Kennedy, CJ; (2011). The toxicology of metals in fishes. Academic Press, San Diego, California, USA.
- Kumolu-Johnson, CA; Ndimele, PE; Akintola, SL; Jibuike, CC (2010). Copper, zinc and iron concentrations in water, sediment and *Cynothrissa mento* (Regan, 1917) from Ologe Lagoon, Nigeria: a preliminary survey. *Afr. J. Aquat. Sci.*, 35(1): 87-94.
- Kusemiju, V; Fadiya, AA; Aderinola, OJ; Akintola, SL (2001). Comparative analysis of heavy metals in water, sediments and tissues of *Lumbricus violaceus* from Agbara and Iba streams. *Nig. J. Res. Rev. Sci.*, 2: 114-117.
- Mahboob, S; Al-Ghanim, KA; Al- Balawi, HFA; Al-Misned, F; Ahmed, Z (2016). Heavy metals in *Clarias gariepinus* (Burchell, 1822) from Wadi-Hanefah, Saudi Arabia. *Pak. J. Zoo.* 48 (3): 875-880.
- Monteiro, SM; Mancera, JM; Fontainhas-Fernandes, A; Sousa, M (2005). Copper induced alterations of biochemical parameter in the gill and plasma of *Oreochromis niloticus*. *Comp. Biochem. Physiol. C.* 141: 375- 383.
- Murtala, BA; Abdul, WA; Oyebanjo, W; Akinyemi, AA (2012). Bioaccumulation of heavy metals in fish (*Hydrocynus forskahlii*, *Hyperopisus bebe occidentalis* and *Clarias gariepinus*) organs in downstream Ogun coastal water, Nigeria. *Trans. J. Sci. Tech.* 2 (5): 119-133.
- Nhiwatiwa, T; Barson, M; Harrison, AP; Utete, B; and Cooper, RG (2011). Metal concentrations in water, sediment and sharptooth catfish *Clarias gariepinus* from three peri-urban rivers in the upper Manyame catchment, Zimbabwe. *Afr. J. Aquat. Sci.* 36: 243–252.
- Novozamsky, I; Houba, VJG; van Eck, R; van Vark, W (1983). A novel digestion technique for multi-element plant analysis. *Com. Soil Sci. Plant Anal.* 14: 239-248
- Obianime, AW; Odili, O; Olorunfemi, OJ; Wokoma, TO; Chuemere, AN (2017). Air and soil carcinogenic risk evaluation and impact on the health status of automechanics: The Nigeria Environment. *Inter. J. Med. Res. Applica.* 1 (1): 26-34.
- Obodo, GA (2002). The bioaccumulation of heavy metals in fish from the lower reaches of River Niger. *J. Chem. Soc. Nig.* 27(2):173–176.
- Okafo, CN; Yakubu, SE; Ameh, JB; Okuofu, CA (2018). Heavy metal levels in some fish species harvested from River Kaduna, Nigeria. *Afr. J. Nat. Sci.* 21: 9-16.
- Oliveira-Ribeiro, CA; Vollaie, Y; Sanchez-Chardi, A; Roche, H (2005). Bioaccumulation and the effects of organochlorine pesticides, PAH and heavymetals in the eel (*Anguilla anguilla*) at the Camargue Nature Reserve, France. *Aquat. Toxicol.* 74: 53–69.
- Omwenga, JM (2003). Heavy metals and nutrient retention in the Kerenga Wetland, Kericho, Kenya. Unpublished M.Sc. Thesis.

- Pacheco, M; Santos, MA; Pereira, P; Martínez, JI; Alonso, PJ; Soares, MJ; Lopes, JC (2013). EPR detection of paramagnetic chromium in liver of fish (*Anguilla anguilla*) treated with dichromate (VI) and associated oxidative stress responses-Contribution to elucidation of toxicity mechanisms. *Comp. Biochem. Physiol. C*. 157: 132-140.
- Rajappa, B; Manjappa, S; Puttaiah, ET (2010). Monitoring of Heavy metal in groundwater of Hakinaka TaluK, India. *Contemp. Eng. Sci.* 3(4): 183 – 190.
- Rashed, MN; (2001). Monitoring of environmental heavy metals in fish from Nasser Lake. *Environ. Int.* 27: 27-33.
- Reid, SD (2011). Molybdenum and chromium. Academic Press, New York, USA.
- Sepe, A; Ciaralli, L; Ciprotti, M; Giordano, R; Fumari, E; Costantini, S; (2003). Determination of cadmium, chromium, lead and vanadium in six fish species from the Adriatic Sea. *Food Addit. Contam.* 20: 543-552.
- Sfakianakis, DG; Renieri, E; Kentouri, M; Tsatsakis, AM (2015). Effect of heavy metals on fish larvae deformities: A review. *Environ. Res.* 137: 246-255.
- Shukla, V; Dhankhar, M; Prakash, J; Sastry, KV (2007) Bioaccumulation of Zn, Cu, and Cd in *Channa punctatus*. *J. Environ. Biol.*, 28, 395-397.
- Sivaperumal, P; Sankar, TV; Viswanathan-Nair, PG (2007). Heavy metal concentrations in fish, shellfish and fish products from internal markets of India vis-a-vis international standards. *Food Chem.* 102: 612-620.
- Steel, RGD; Torrie, J.H (1980). *Principle and procedures of statistics*. McGraw-Hill New York, USA, pp 633.
- Storelli, MM; Barone, G; Storelli, A; Marcotrigiano, GO (2006). Trace metals in tissues of Mugilids (*Mugil auratus*, *Mugil capito* and *Mugil labrosus*) from the Mediterranean Sea. *Bull. Environ. Contam. Toxicol.* 77: 43-50.
- Tirkey, A; Shrivastava, P; Saxena, A (2012). Bioaccumulation of heavy metals in different components of two Lakes ecosystem. *Current World Environ.* 7(2), 293-297.
- Uysal, K; Kose, E; Bulbul, M; Donmez, M; Erdogan, Y; Koyun, M; Omeroglu, C; Ozmal, F (2009). The comparison of heavy metal accumulation ratios of some fish species in EnneDarne Lake (Kutahya, Turkey). *Environ. Monit. Assess.* 157: 355-362.
- Vera-Candioti, J; Soloneski, S; Larramendy, ML; (2011). Acute toxicity of chromium on *Cnesterodon decemmaculatus* (Pisces: Poeciliidae). *Theoria* 20: 81-88.
- Vinodhini, R; Narayanan, M (2008). Bioaccumulations of heavy metals in organs of fresh water fish *Cyprinus carpio* (Common carp). *Int J Environ Sci. Tech.* 5: 179-182.
- Wangboje, OM; Ekundayo, OT; Iwu, HU (2013). Heavy metal concentrations in two economically important fish species and water of Ovia River, Southern Nigeria. *Afr. J. Environ. Pollut. Health.* 10:26-32.
- World Health Organization, WHO (2008). Guidelines for Drinking Water Quality. 3rd Edn. Health Criteria and Supporting Information. WHO, Geneva, pp: 668. Retrieved from: [http://www.who.int/water\\_sanitation\\_health/dwg/fulltext.pdf](http://www.who.int/water_sanitation_health/dwg/fulltext.pdf).