

Phytoplankton Composition and trophic state of Guessabo lake (Upstream in Buyo lake, Ivory Coast)

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ABSTRACT

Objectives: The trophic quality of Guessabo Lake (Ivory Coast) under the influence of fishing and agriculture was analysed starting according from the phytoplankton composition.

Methodology and results: Sampling spatio-temporal in pelagic zone were made from a plankton net and supplemented with an integrated sampler. A total of three hundred and forty six species were identified. The groups, by orders of prevalence are, Chlorophyta with 164 species (48 %), Euglenophyta with 64 species (19 %), Cyanobacteria represented by 53 species (16 %), Bacillariophyta with 50 species (15 %) and, the other groups (Pyrrophyta, Chrysophyta and Xanthophyta) represented by 15 species (4 %). On the level of sampling stations of Guessabo Lake, 239 species were identified in Guemon sector against 231 and 257 species respectively in bed of the lake and Haut Sassandra sector. Concerning Guemon sector, 181 species were recorded at station G1 and 192 at station G5. In bed of lake (Channel), 154 species were listed at station G2, whereas 157 and 168 species were observed respectively at stations G4 and G6. For Haut Sassandra, stations G3 and G7 respectively presents 208 and 211 species. During this study, 103 species constant were listed against 66 species additional and 170 species accidental. In the floristic list, 73 species are common to all sampling stations. Sorensen index showed a similarity between the sectors and sampling stations. Trophic index based on Phytoplankton composition indicated that lake is from mesotrophy to eutrophy state.

Key words: Phytoplankton, lake, mesotrophy, eutrophy; Ivory Coast.

RÉSUMÉ

Objectif de la recherche : La qualité trophique du lac Guessabo (Côte d'Ivoire) sous l'influence de la pêche et de l'agriculture a été analysée à partir de la composition du phytoplancton.

Méthodologie et résultats : Des prélèvements spatio-temporels dans la zone pélagique ont été réalisés à l'aide d'un filet à plancton et d'un préleveur intégré. Au total, 340 taxons phytoplanctoniques ont pu être identifiés au niveaux spécifiques et infraspécifiques, Les embranchements, par ordres de prédominance sont, les Chlorophyta avec 164 taxons soit 48%, les Euglenophyta avec 64 taxons soit 19 %, les Cyanobacteria représentés par 53 taxons soit 16 %, les Bacillariophyta avec 50 taxons soit 15 %, et les autres embranchements (Pyrrophyta, Chrysophyta et Xanthophyta) représentés par 15 taxons soit 4 %.

Au niveau des stations de prélèvement du lac Guéssabo, 239 taxa ont été identifiés dans la partie Guémon contre 231 et 257 taxons respectivement dans le lit et dans la partie Haut Sassandra du lac. Concernant le secteur Guémon, 181 taxons ont été enregistrés à la station G1 et 192 à la station G5. Dans le lit du lac (chenal), 154 taxons ont été recensés au point G2, alors que 157 et 168 taxons ont été répertoriés respectivement aux stations G4 et G6. Pour la partie Haut Sassandra, les stations G3 et G7 regorge respectivement 208 et 211 taxons. Au cours de cette étude, 103 taxons constants ont été recensés contre 66 taxons accessoires et 170 taxons accidentels. Dans la liste floristique, 73 taxons sont communs à toutes les stations d'échantillonnage. Les valeurs de l'indice de Sorensen ont montré une ressemblance entre les secteurs et entre stations de prélèvement. Les indices de trophie basés sur la composition du peuplement montrent que l'état du lac tend de mésotrophe à eutrophe.

Mots clés : Phytoplankton, lac, mésotrophe, eutrophe, Côte d'Ivoire.

INTRODUCTION

The population explosion and the extension of cities have led, for decades, considerable pressure on aquatic environments by different activities (fishing, agriculture, urbanization). Freshwater ecosystems, especially lakes, are a receptacle for organic and mineral substances from agricultural, domestic and urban wastewater discharges. Thus, these environments are increasingly threatened by eutrophication and / or pollution. According to SCOR WG, 2007 and Collins *et al.*, 2014, the phenomena are related to a nutrient enrichment of the environment whose elements most important are nitrogen and phosphorus. Sources of primary production, their presence and quantity in water generally contribute to the development of aquatic plants and in particular micro-algae. Phytoplankton communities are considered as true sentinels or reflections of environmental conditions (SCOR WG, 2007; MerMex Group *et al.*, 2011). These organisms have a turn-over of a few days allowing them to evolve rapidly when environmental conditions change. However, some authors (Smith, 2003; Chomerat, 2005; Carmichael *et al.*, 2010) affirm that, depending on the conditions of stability and enrichment of useful nutrients, the development of an algal population may be beneficial or detrimental to man. Indeed, some phytoplankton species can secrete toxins into the water and affect heterotrophic species with public health risks (Anderson *et al.*, 2002; Glibert *et al.*, 2005). Faced with such situation, the use of water resources is questioned. Hence the need to envisage a policy of protecting water resources in order to preserve the ecological balance of aquatic

ecosystems (Dziocket *et al.*, 2006; Cabecinha *et al.*, 2009). Therefore, knowledge of ecological status and monitoring of aquatic ecosystems is imperative. In this context, according to Ngansoumana, (2006), knowledge of the phytoplankton composition makes it possible to establish real tools for diagnosis and pollution assessment. In addition, indices of biotic integrities were developed from the composition of the phytoplankton community to assess the trophic status of lakes (Thunmark, 1945 and Nygaard, 1949). In Côte d'Ivoire, Guessabo Lake located upstream of the Buyo Dam and on the Sassandra River is under the influence of several human activities. Main fishing lake, fishing on this lake is becoming more intense and badly practiced. Conventional standards are no longer respected by the use of chemicals for fishing. In addition, the exploitation of the banks for agricultural purposes during periods of low water. According to Kra, (2016), during this period, dry areas are used by farmers for food crops (rice, corn, peanuts, potatoes, bananas). Cocoa and rubber plantations are also activities in the vicinity of the lake. However, the use of agro-pharmaceutical products (weed killer, insecticide, fertilizer ...) in the treatment of crops and soil fertilization is a problem. Nutrient inputs, in particular phosphorus and nitrogen in the lake, are the source of the invasion of aquatic plants (*Salomonina molesta*, *Eicchornia crassipes*,) on its surface. Biological data available on this hydrosystem are the results of the works of Mambo *et al.*, (2001) and Kra, (2016). These data are only on the zooplanktonic

diversity, chlorophyll a and on fish production in the lake. Based on this observation and the economic interest of this lake, the knowledge of its current state is important through its phytoplankton organisms because this biology compartment has

not been exploited and remains unknown. The objective of this study is to determine the phytoplankton composition of the lake to assess its trophic status.

MATERIALS AND METHODS

Study Area: located in western Ivory Coast, 3 kilometres from Guessabo and 40 kilometres from Duekoue. Guessabo lake is to upstream of Buyo dam on the Sassandra stream and constitutes the administrative boundary between the regions of Haut-Sassandra and Guemon. The hydrological regime of lake is influenced by the rains and the closure of Buyo dam. The annual rainfall varies between 1300 mm and 1800 mm. The level of low water to high water is governed by the contributions of Sassandra and its tributaries. Four hydrological seasons are defined (Mambo *et al.*, 2001). Sassandra water level drops from December to January corresponding to a period of dry and hot season. From February to May, a period of low water corresponding to the period of low water during a

small dry season. Wet season cumulating rainfall and Sassandra flood during August to mid-December completely fill the lake every year. The study is done on the portion of Guessabo lake between 6.72° and 6.775° North latitude and between 6.96° and 7.00° West longitude which seven sampling points were retained (Figure 1). These sampling stations are selected in the following way, G1-G5 in the Guemon part, G2-G4-G6 in the bed of the lake (Channel) and G3-G7 in Haut Sassandra so as to take into account the cumulated effect of the activities to which are subjected Guessabo lake. Geographical coordinates of sampling points and the various carried on activities in the vicinity are consigned in the table below (Table1).

Table 1: Geographic coordinates and description of the 7 selected sampling stations

Sampling points	Longitude(X) dd	Latitude(Y) dd	Description of sampling stations
G1	-6,997	6,752	Located in the Guemon area near Dibobli village, presence of aquatic plants in these sampling points and: cocoa plantation, hevea plantation, artisanal fishing, corn culture, rice growing, banana plantation are activities practiced in the vicinity of these.
G5	-6,999	6,740	
G2	-6,991	6,768	Located in the bed of the lake (channel), the activities carried out on these points are artisanal fishing, navigation
G4	-6,985	6,752	
G6	-6,987	6,724	
G3	-6,971	6,769	Located in the Haut Sassandra area near Guessabo village, presence of aquatic plants in these sampling points and: cocoa plantation, hevea plantation, artisanal fishing, corn culture, rice growing, banana plantation are activities practiced in the vicinity of these.
G7	-6,972	6,738	

dd: decimal degree;

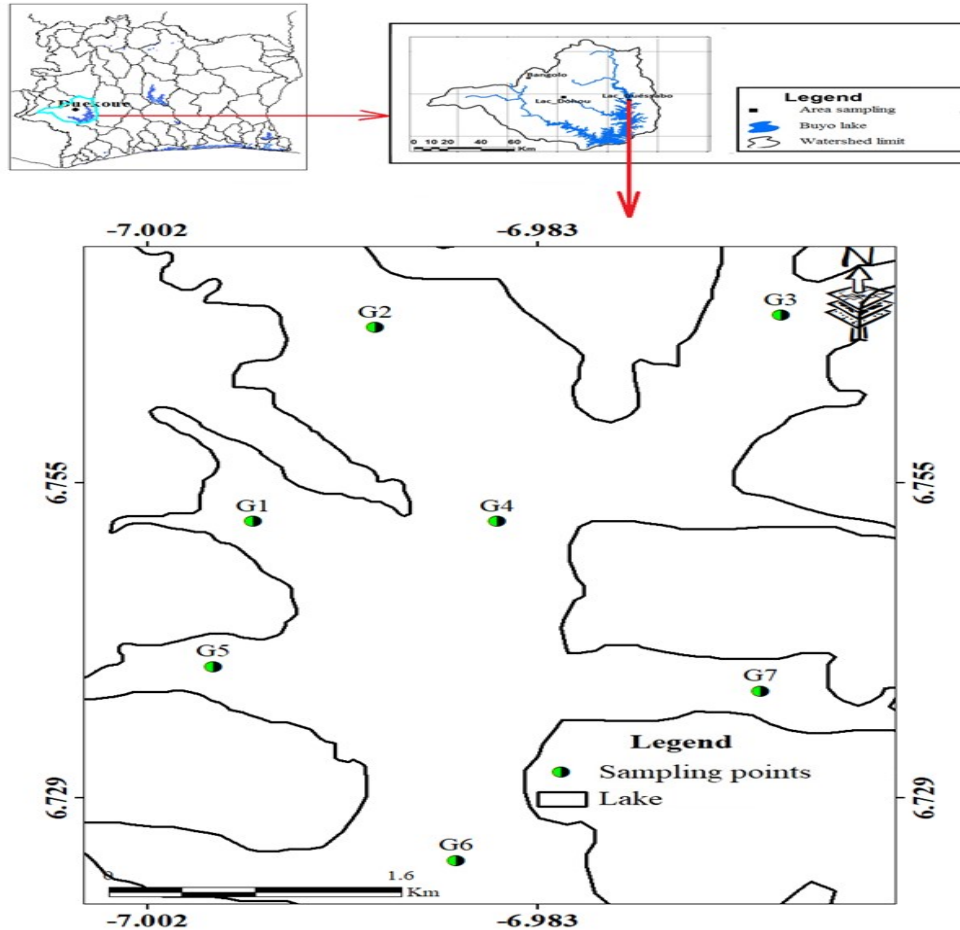


Figure 1: Map showing position of the sampling points on the Buyo Lake (Guessabo Area)

Sampling: Phytoplankton was sampled monthly at each of the seven points between October 2017 and September 2018. Phytoplankton sampling was performed using the recipient of 10 liter and plankton net (50 μm mesh size)(Ouattara, 2000). These harvests were supplemented by phytoplankton collection with sampler integrated in order to collect algae of very small sizes which pass through the mesh of the net to plankton. The samples were fixed with alcohol 90% and formalin 5% into collection bottles.

Observation and identification: In the laboratory, the samples were observed between slide and the cover glass under an optical microscope, LEICA with objective x 40. The various phytoplanktons were identified by consulting work of Förster1964; Philipose, 1967; Compère, 1977; Couté & Ittis, 1981; Komárek & Fott, 1983 and Uherkovich, 1995. The classification suggested in the key established by the editors of "Süßwasserflora" (Starmach, 1985) was used in this study.

Data Analysis: The ecological status of Guessabo Lake has been evaluated from the use of:

- Species richness, which is the number of taxa to which the taxa belong (Alliaume *et al.*, 1990)
- Occurrence frequency (F), which is the ratio (in percentage) between numbers of times that species appears in the samples (N) and total number of samples (Nt) (Dajoz, 2000). Depending on the Occurrence frequency, the groups of distinguished species are constant species ($F \geq 50\%$), accessory species ($25\% \leq F < 50\%$), accidental species ($F < 25\%$).
- Similarity index of Sorensen (Cs) which is the ratio of the number of species common to 2 stations (station a and station b) on number of species present in station a and in station b. Sorensen similarity index varies from 0 (no similarity) to 1 (identical station) (Throvald, 1949)
- Compound index (index A) is the report of sum of species number belonging to groups of Cyanobacteria, Chlorococcales, Euglenophyta and Centrale on number of species of Desmidiaceae (Nygaard, 1949). If this value < 1, water is oligotrophy, if

value is between 1 and 2.5 water is mesotrophy and if value > 2.5 water is eutrophy.

- Chlorophycean quotient (index B), which is the quotient of the number of Chlorococcales and Desmidiaceae species present in a water body (Thunmark, 1945). When the value of B < 1, the water is oligotrophy and when the value of B > 1, the water is eutrophy.

- Diatom index (index C) expresses the report of number of Centrales species on Pennales species (Nygaard, 1949) If the index C value is located between 0 and 2 water is oligotrophy and when the index C value is between 0, 2 and 3 water is eutrophy.

Phytoplankton composition: A total of 340 species divided into 115 genus, 53 families, 25 orders, 11 classes and 7 branches were identified. The branches, in orders of predominance are Chlorophyta with 164 species (48 %), Euglenophyta with 64 species (19 %), Cyanobacteria represented by 50 species (15 %), Bacillariophyta with 47 species (14 %), the other branches (Pyrrophyta, Chrysophyta, Rhodophyta and Xanthophyta) represented by 15 species (4 %) (Figure 2). The Branch of Chlorophyta is constituted of three classes, Chlorophyceae, Zygnematophyceae and Oedogoniophyceae. Class of Zygnematophyceae is richest and comprises three orders, Zygnematales, Volvocales and Chaetophorales. Zygnematales are diversified with mainly the family of Desmidiaceae. In this family, the genera most represented are *Staurastrum* (20 species), *Cosmarium* (17 species), *Scenedesmus* (12 species), *Euastrum* (11 species) and *Pediastrum* (9 species). Concerning Euglenophyta, this group is only constituted by the class of Euglenophyceae. This class is also composed one order, that of Euglenales with only one family

(Euglenaceae). The principal genera are *Trachelomonas* (25 species), *Phacus* (12 species), *Euglena* (10 species) and *Strombomonas* (10 species). As for Cyanobacteria, the group consists of only one class that of Cyanophyceae. This class gathers five orders which are Synechococcales, Chroococcales, Spirulinales, Nostocales and Oscillatoriales. The first two orders are richest. Synechococcales are composed of more families. These families are Merismopediaceae, Coelosphaeriaceae, Synechococcaceae and Pseudanabaenaceae. Among those, Merismopediaceae are represented by *Merismopedia* (4 species) and *Aphanocapsa* (4 species). For Chroococcales, the listed families are Microcystaceae, Chroococcaceae and Gomposphaerioidaceae. In these families, Microcystaceae are more represented with *Microcystis* genus (8 species). On the level of Bacillariophyta, two classes are observed, Coscinodiscophyceae and Fragilariophyceae. The most diversified class is that of Fragilariophyceae. This class is primarily constituted of the following orders, Naviculales, Eunotiales, Fragilariales and Bacillariales. Among these orders, Naviculales are richest. In this order two families are represented, Pinnulariaceae and Eunotiaceae with respectively the genera *Pinnularia* (9 species) and *Eunotia* (6 species). Other groups are the less diversified with 8 species for Xanthophyta, 6 species for Pyrrophyta and 1 specie for Chrysophyta. In Guessabo Lake, 73 species were common for all sampling stations. Concerning the lake Guessabo, the list of the phytoplankton composition regroups 103 species constant against 66 species additional and 170 species accidental.

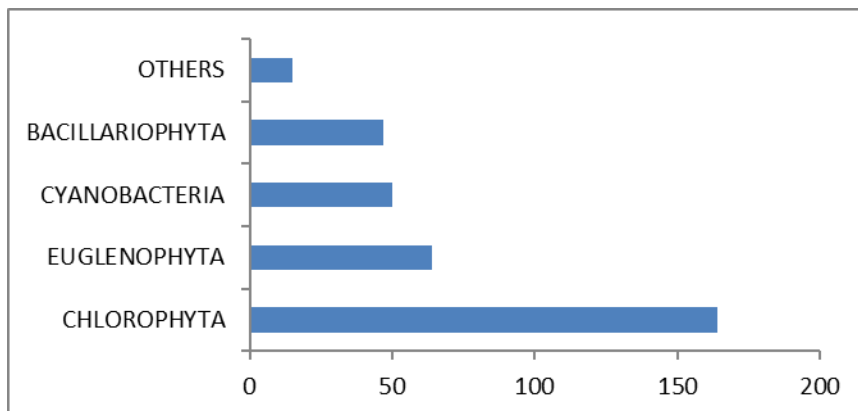


Figure 2: Proportion of phytoplankton groups in Guessabo Lake

Space variation of the composition: On the level of the sampling stations of the lake Guessabo (Figure 3), 234 species were identified in Guemon sector against 232 and 255 species respectively in bed and Haut Sassandra sector of the lake. Concerning Guemon sector, 179 species were recorded at G1 station and 192 at G5 station. In bed of the lake (channel), 154 species were listed at the G2 point, whereas 157 and 168 species were identified respectively at the G4 station and the G6 station. For sector Haut Sassandra,

G3 stations and G7 present respectively 208 and 211 species. The algae composition is similar for all stations except G3 and G7. Thus Chlorophyta varies from 37 % (G2) to 50 % (G3) and Euglenophyta from 20.83 % (G5) to 24.86 % (G1). As for Cyanobacteria, the proportions are between 14.36 % (G3) and 19.74 % (G4). For Bacillariophyta, the relative richness oscillates between 12.5 % (G4) and 16.88 % (G2), the other groups represent that only proportions going from 4.16 % (G6) to 5.84 % (G2)(Figure 4).

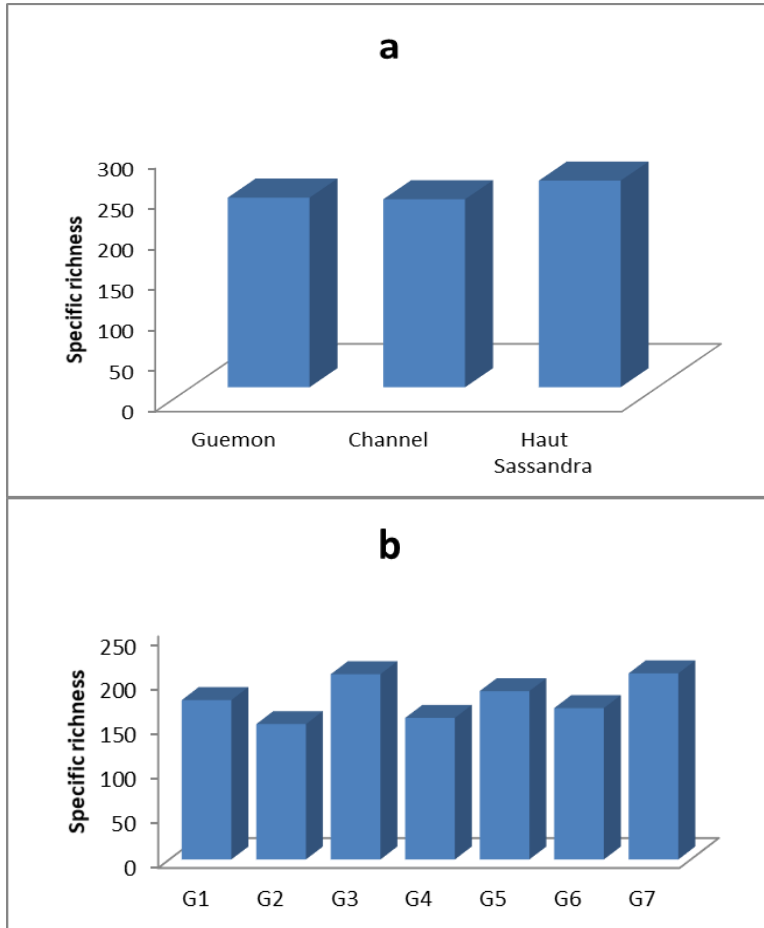


Figure 3: Space variation of specific richness in Guessabo Lake a: richness of the various sectors; b: richness of the various sampling stations; G: Guessabo; 1 to 7: number of sampling point

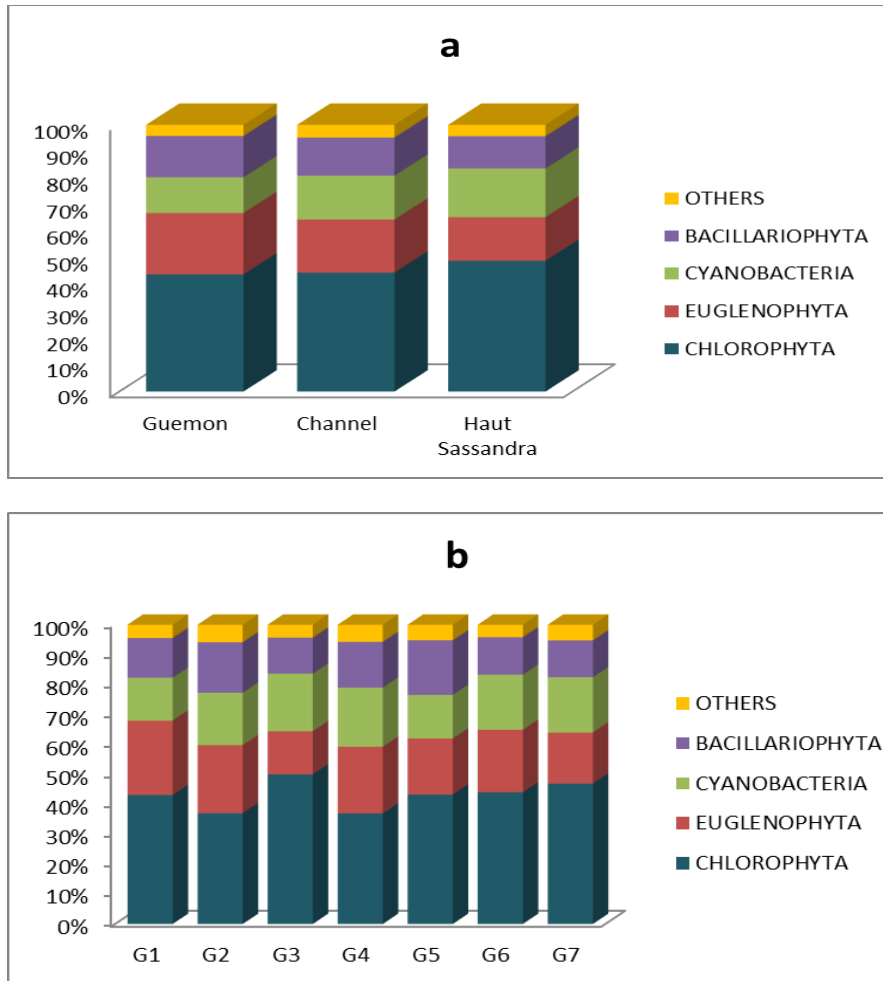


Figure 4: Space variation of the relative richness in Guessabo Lake. a: richness of the various sectors; b: richness of the various sampling stations; G: Guessabo; 1 to 7: number of sampling point.

Similarities between sampling points: Similarity between the sampling stations according to the inventoried species was presented in table 2. The values of the Sorensen index calculated between the sampling points reveal values greater than 0.5. The specific richness of phytoplankton is substantially the

same in all points. However, this similarity is stronger on the one hand between G3 and G7 points (Cs: 0.78) of Haut Sassandra sector and on the other hand between G1 and G5 points (Cs: 0.73) of Guemon sector. Also a strong similarity is observed between sectors.

Table 2: Values of Sorensen similarity index between sampling points (a) and between sectors (b).

a)	G1	G2	G3	G4	G5	G6
G2	0.66					
G3	0.70	0.62				
G4	0.67	0.72	0.64			
G5	0.73	0.63	0.72	0.62		
G6	0.72	0.71	0.66	0.70	0.64	
G7	0.72	0.65	0.78	0.65	0.66	0.64

b)	Guemon	Channel
Channel	0.73	
Haut Sassandra	0.74	0.73

The trophy status of Guessabo Lake: Table 3 presents the trophic index starting from the phytoplankton quotients at the sampling points. In the whole of the stations of the Guessabo lake, the results of the compound index A and the index B (Chlorococcales/ Desmidiaceae) vary respectively from 3 to 4.21 and 1.21 to 1.68. These two indexes

would suggest a eutrophic state of the whole lake stations. As for the index C representing the diatom quotient, it oscillates from 0.18 to 0.33 and thus provides two quite distinct types (oligotrophic and eutrophic) an intermediate type (mesotrophic). The trophic state of the whole of the stations of the lake evolves of a mesotrophic state to a eutrophic state

Table 3: Values of trophic index in sampling stations.

Index	G1	St	G2	St	G3	St	G4	St	G5	St	G6	St	G7	St
A	3,77	E	3,96	E	3	E	4,21	E	3,56	E	3,47	E	4	E
B	1,29	E	1,21	E	1,28	E	1,29	E	1,38	E	1,25	E	1,68	E
C	0,33	E	0,18	O	0,25	E	0,2	O	0,21	E	0,31	E	0,18	O
		E		M		E		M		E		E		M

G: Guessabo; 1 to 7: number of sampling point; St: trophic state; E: eutrophic; M: mesotrophic; O: oligotrophic A: compound index; B: Chlorophycean index; C: diatom quotient

DISCUSSION

In the present study, 340 species were observed in the whole sampling stations of Guessabo Lake. This high taxonomic richness would be due to the combination of two complementary methods. The algae were initially collected by filtration with the plankton net. These collections were supplemented by taking away with the sampler integrated in order to collect the algae of very small sizes which pass through the meshes of plankton net. In comparison with work of Ouattara, 2000 on Ayamé Lake and Grogga, 2010 on Kossou Lake in Ivory Coast, Guessabo Lake is most diversified. That could be explained by the high number of samples resulting of sampling stations. Chlorophyta with 164 species either 48 % and Euglenophyta with 64 species or 19 %, represent the most diversified groups. This prevalence of Chlorophyta and Euglenophyta would be characteristic of an environment rich in putrescible matter organic. *Staurastrum* (20 species), *Cosmarium* (17 species) and *Trachelomonas* (25 species) genera are most represented of these groups. Adon, (2013) and Kouassi-Blé, (2013) observed this same taxonomic diversity on Azopé reservoir in Ivory Coast. Moreover Ouattara, (2000) and Njine *et al.*, (2004) had observed respectively a greater diversity of Chlorophyta on Ayamé lake in Ivory Coast and a composition mainly constituted of Euglenophyta on Yaoundé municipal lake in Cameroun. The small proportions of the other groups (Chrysophyta, Xanthophyta, Pyrrophyta) justify eutrophic state of environments subjected to this study. According to Nicholls, (1995), these groups of phytoplankton develop better in oligotrophic aquatic ecosystems. On level of sampling stations of Guessabo lake, stations of Haut Sassandra sectors and Guemon

represents more species that in the bed of lake. This richness would be related to their lentic character which could thus support the complete cycle of reproduction and development of the algae. Indeed, phytoplankton development depends closely on stability in ecosystem fonctionment faced with environmental disturbances (Gonzales and Descam, 2004). Similarity coefficients between the various sampling stations shows a resemblance on algae flora inventoried in seven sampling stations. This resemblance could be explained by the great species number which various stations share commonly and shows that stations have the same environmental characteristics. Ecological conditions of aquatic environment can be evaluated from knowledge of relative species number representing various groups taxonomic in a phytoplankton sample (Ngansoumana, 2006). Thus, the compound index A most largely used because of its appreciations range of all trophy degrees in lakes, indicate eutrophic state of Guessabo lake (A varies from 3 to 4.21). Also, the quotient provides by the two groups taxonomic of Chlorophyceae (Chlorococcales/ Desmidiaceae) in lake stations is higher than 1, translating aneutrophic conditions. That is justified by the fact that in eutrophic lakes the number of species of Chlorococcales would be more significant than the number of Desmidiaceae, and the conditions opposite in oligotrophic lakes (Rawson, 1956). As for the index C based on the quotient of the diatoms (Centrales/Pennales), it oscillates between 0.18 to 0.33 and provides a mesotrophic state. All these quotients highlight only qualitative aspect of algae community. Indeed, by considering biological approach, concept of

specific richness with biotic index used, also testifies an

eutrophication process of Guessabo Lake.

CONCLUSION

The results obtained starting from the specific composition of the phytoplankton in Guessabo Lake indicates a total of 346 species divided in seven groups. These groups represented by orders of prevalence are, Chlorophyta (48 %), Euglenophyta (19 %), Cyanobacteria (16 %), Bacillariophyta or diatoms (14 %) and the other groups (Pyrrophyta, Chrysophyta, and Xanthophyta) with 4 %. The trophic state of the whole

of the stations of lake evolves from mesotrophy state to eutrophy state This state deserves giving the lake a detailed attention by sensitizing the users and the actors of various activities in the vicinity which to sanitary risks they are exposed to. Quantitative studies of the community of phytoplankton and physicochemical parameters could be undertaken to confirm or change this state of lake.

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Annexe: phytoplanktonic composition of Guessabo Lake. G1 to G7: sampling stations

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
CYANOBACTERIA								
CYANOPHYCEAE								
SYNECHOCOCCALES								
	Synechococcaceae							
<i>Synechococcus aqualitis</i> Sauvageau		x	x	x	x	x	x	64,29
<i>synechococcus</i> sp				x		x		17,85
	Merismopediaceae							
<i>Aphanocapsa pulchra</i> (Kützing) Rabenhorst		x	x	x	x	x	x	85,71
<i>Aphanocapsa thermalis</i> Brügger		x	x	x	x	x	x	100
<i>Aphanocapsa conferta</i> (W.West. & G.W.West) Komárkova-Legnerova		x	x	x	x	x	x	100
<i>Aphanocapsa incerta</i> (Lemmermann) Cronberg & Komárek		x	x	x	x	x	x	85,71
<i>Merismopedia punctata</i> Meyen		x	x	x	x	x	x	85,71
<i>Merismopedia tenuissima</i> Lemmermann		x	x	x	x	x	x	100
<i>Merismopedia glauca</i> (Ehrenberg.) Kützing			x	x		x	x	85,71
<i>Merismopedia elegans</i> Braun			x	x	x		x	35,71
	Coelosphaeriaceae							
<i>Coelosphaerium micoporum</i> Nägeli		x		x		x		85,71
<i>Coelosphaerium</i> sp.1		x	x	x	x	x	x	85,71
<i>Coelosphaerium</i> sp.2				x	x		x	14,29
<i>Snowella atomus</i> J. Komárek & Hindák		x	x	x	x	x	x	100
	Pseudanabaenaceae							
<i>Pseudanabaena</i> cf. <i>limnetica</i> (Lemmermann) Komárek		x	x	x	x	x	x	64,29
<i>Pseudanabaena catenata</i> Lauterborn				x		x		11,90
CHROOCOCCALES								
	Chroococcaceae							
<i>Chroococcus minimus</i> (Keissler)		x	x	x	x	x	x	100
<i>Chroococcus minutus</i> (Kützing) Nägeli		x	x	x	x	x	x	100
<i>Chroococcus limneticus</i> Lemmermann		x	x	x	x	x	x	64,29
<i>Chroococcus turgidus</i> (Kützing) Nägeli			x	x		x		64,29
<i>Chroococcus dispersus</i> (Keissler) Lemmermann		x	x	x		x	x	64,29
<i>Chroococcus aphanocapsoides</i> Skuja						x		4,76
	Microcystaceae							
<i>Microcystis flos-aquae</i> (Wittrock) Kirchner				x		x	x	21,24
<i>Microcystis aeruginosa</i> (Kützing) Kützing		x	x	x	x	x	x	85,71
<i>Microcystis wesenbergii</i> (Komárek) Komárek		x	x	x	x	x	x	85,71
<i>Microcystis incerta</i> (Lemmermann) Lemmermann		x		x		x	x	47,61
<i>Microcystis robusta</i> (Clark) Nygaard							x	2,38
<i>Microcystis densa</i> W.West. & G. W.West.				x			x	4,76
<i>Microcystis</i> sp.1							x	2,38

TAXONS	Sampling stations							
	G1	G2	G3	G4	G5	G6	G7	FO
<i>Microcystis</i> sp.2			x					2,38
Gomphosphaerioidaceae								
<i>Gomphosphaeria aponina</i> Kützing	x		x	x			x	23,8
<i>Gomphosphaeria</i> sp								
SPIRULINALES								
Spirulinaceae								
<i>Spirulina</i> sp.1		x					x	4,76
<i>Spirula</i> sp.2				x				3,57
NOSTOCALES								
Nostocaceae								
<i>Anabaena affinis</i> Lemmermann	x	x	x	x	x	x	x	58,33
<i>Anabaena variabilis</i> (Kützing) ex Bornet & Flahault					x			3,57
<i>Anabaena circinalis</i> (Rabenhorst) ex Bornet & Flahault						x		3,57
<i>Anabaena spiroide</i> Klebahn	x	x	x	x	x	x	x	85,71
<i>Nostoc piscinale</i> (Kützing) ex Bornet & Flahault			x	x		x		9,52
Scytonemataceae								
<i>Scytonema</i> sp				x				3,57
OSCILLATORIALES								
Oscillatoriaceae								
<i>Planktolyngbya contorta</i> (Lemmermann) Anagnostidis & Komárek	x	x	x	x	x	x	x	100
<i>Planktolyngbya lemnetica</i> (Lemmermann) Komárkova-Legnerova	x	x	x	x	x	x	x	100
<i>Lyngbya major</i> (Vaucher) Hansgirg							x	3,57
<i>Oscillatoria lutea</i> Agardh ex Gomont				x	x		x	21,43
<i>Oscillatoria sancta</i> Kützing							x	5,95
<i>Oscillatoria limosa</i> (Dillwyn) Agardh	x		x		x	x		28,57
<i>Phormidium articulatum</i> (Gardner) Anagnostidis & Komárek		x	x	x	x	x	x	42,86
<i>Phormidium granulatum</i> (Gardner) Anagnostidis			x			x		14,28
<i>Plectonema puteale</i> (Kirchner) Hansgirg	x	x	x	x	x	x	x	85,71
Microcoleaceae								
<i>Planktothrix compressa</i> (Utermöhl) Anagnostidis & Komárek			x	x				9,52
Gomontiellaceae								
<i>Komvophoron constrictum</i> (Szafer) Anagnostidis & Komárek		x	x			x	x	23,81
EUGLENOPHYTA								
EUGLENOPHYCEAE								
EUGLENALES								
Euglenaceae								
<i>Euglena acus</i> var. <i>acus</i> Ehrenberg	x	x	x	x	x	x	x	58,33
<i>Euglena excavata</i> Schiller		x						3,57
<i>Euglena charkowiensis</i> Svirenko	x							9,52
<i>Euglena gracilis</i> Klebs	x	x	x	x	x	x	x	95,52
<i>Euglena caudata</i> Hübner	x	x		x		x		28,57
<i>Euglena polymorpha</i> Dangeard		x	x			x	x	28,57

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
<i>Euglena sanguinea</i> Ehrenberg	x	x		x	x	x		42,85
<i>Euglena oxyrus</i> Schmarda	x				x		x	17,85
<i>Euglena</i> sp.1					x			3,57
<i>Euglena</i> sp.2	x	x				x		17,85
<i>Trachelomona rugulosa</i> Stein			x		x	x	x	28,57
<i>Trachelomonas varians</i> Skvortzov	x	x	x	x	x			35,71
<i>Trachelomonas abrupta</i> Svirenko				x	x			12,29
<i>Trachelomonas amata</i> var. <i>amata</i> E. Stein					x			3,57
<i>Trachelomonas amata</i> (Ehrenberg) Stein var. <i>gordeievi</i> Skvortzov	x							3,57
<i>Trachelomonas amata</i> var. <i>heterospina</i> Svirenko	x							3,57
<i>Trachelomonas bernardinensis</i> var. <i>africana</i> Vischer	x		x				x	28,57
<i>Trachelomonas conica</i> fo. <i>Punctata</i> Deflandre	x	x	x	x	x	x	x	33,33
<i>Trachelomonas cylindrica</i> Ehrenberg	x	x	x	x	x	x	x	64,28
<i>Trachelomonas dastuguei</i> Balech var. <i>africana</i> Couté & Iltis				x		x		9,52
<i>Trachelomonas dubia</i> Svirenko			x					3,57
<i>Trachelomonas dubia</i> var. <i>lata forma</i> Deflandre		x			x	x		14,29
<i>Trachelomonas lacustris</i> var. <i>ovalis</i> Drezepolski	x	x	x	x	x	x	x	64,28
<i>Trachelomonas oblonga</i> Lemmermann	x	x	x	x	x	x	x	64,29
<i>Trachelomonas klebsii</i> Deflandre.	x		x	x	x	x	x	53,57
<i>Trachelomonas oblonga</i> var. <i>attenuata</i> Playfair					x			3,57
<i>Trachelomonas robusta</i> Svirenko	x	x	x	x	x	x	x	58,33
<i>Trachelomonas similis</i> Stockes	x	x	x	x	x		x	42,59
<i>Trachelomonas similis</i> var. <i>hyalina</i> Skvortzov	x		x		x	x	x	28,59
<i>Trachelomonas volvocina</i> Ehrenberg	x	x	x	x	x	x	x	100
<i>Trachelomonas volvocinospis</i> Svirenko	x	x	x	x	x	x	x	64,29
<i>Trachelomonas sydneyensis</i> Playflair			x				x	7,14
<i>Trachelomonas hispida</i> var. <i>coronata</i> Lemmermann	x	x	x	x	x	x	x	83,33
<i>Trachelomonas</i> sp.1		x				x		7,14
<i>Trachelomonas planctonica</i> Svirenko			x		x		x	28,57
<i>Strombomonas acuminata</i> var. <i>deflandreana</i> W. Conrad	x						x	14,29
<i>Strombomonas deflandrei</i> (Roll) Deflandre	x	x		x	x	x	x	42,86
<i>Strombomonas girardina</i> (Playfair) Deflandre	x	x				x	x	28,58
<i>Strombomonas verrucosa</i> var. <i>verrucosa</i> (Daday) Deflandre	x		x	x	x	x		41,67
<i>Strombomonas verrucosa</i> var. <i>elongata</i> (Skvortzov) Deflandre	x	x		x		x	x	41,67
<i>Strombomonas fluviatilis</i> var. <i>levis</i> (Lemmermann) Deflandre	x	x		x			x	28,58
<i>Strombomonas</i> cf. <i>triquetra</i> (Playflair) Deflandre	x	x		x		x	x	35,71
<i>Strombomonas gibberosa</i> Playfair	x			x		x		21,43
<i>Strombomonas verrucosa</i> (Daday) Deflandre var. <i>zmiewika</i> (Svirenko) Deflandre	x	x		x	x			23,80
<i>Strombomonas</i> sp.1	x							3,57
Phacaceae								
<i>Lepocinclis marssonii</i> Lemmermann.	x		x	x			x	47,62
<i>Lepocinclis oxyuris</i> fo <i>charkowiensis</i> Hüber-Pestalozzi	x		x		x		x	47,62

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
<i>Lepocinclis ovum</i> (Ehrenberg) Minkevich	x	x	x	x	x	x	x	100
<i>Lepocinclis ovum</i> cf. <i>texta</i> (Dujardin) Lemmermann	x	x	x	x	x	x	x	95,23
<i>Lepocynclis salina</i> Fritsch	x	x		x	x	x		47,62
<i>Lepocynclis</i> sp.1	x	x	x	x	x	x	x	59,52
<i>Lepocynclis</i> sp.2			x		x			11,90
<i>Phacus acuminatus</i> var. <i>acuticauda</i> Pochmann	x	x	x	x	x	x	x	64,29
<i>Phacus acuminatus</i> var. <i>discifer</i> (Pochmann) Hüber- Pestalozzi	x					x	x	41,67
<i>Phacus acuminatus</i> var. <i>granulatus</i> (Roll) Hüber-Pestalozzi				x				2,38
<i>Phacus angulatus</i> Pochmann		x		x				14,29
<i>Phacus glaber</i> (Deflandre) Pochmann	x		x					14,29
<i>Phacus longicauda</i> var. <i>insecta</i> Koczwara	x	x				x	x	23,81
<i>Phacus longicauda</i> var. <i>longicauda</i> (Ehrenberg) Dujardin	x	x		x		x	x	41,67
<i>Phacus platalea</i> Drezepolski	x			x		x		41,67
<i>Phacus sesquitortus</i> Pochmann		x		x			x	21,42
<i>Phacus suecicus</i> Lemmermann	x		x		x			10,71
<i>Phacus</i> sp.1					x			7,14
<i>Phacus</i> sp.2					x			7,15
CHLOROPHYTA								
CHLOROPHYCEAE								
CHLOROCOCCALES								
Golenkiniaceae								
<i>Golenkinia radiata</i> Chodat						x		10,71
<i>Golenkiniopsis chlorelloides</i> (Lund) Flott			x		x		x	21,42
Palmellaceae								
<i>Desmাত্রactum indutum</i> (Geitler) Pascher	x		x		x		x	64,29
<i>Treubaria quadrispina</i> (G.M.Smith) Flott & Kováčik		x						5,95
Hydrodictyceae								
<i>Pediastrum duplex</i> var. <i>duplex</i> Meyen	x	x	x	x	x	x	x	66,67
<i>Pediastrum duplex</i> var. <i>gracilimum</i> W.West & G.S.West	x		x		x		x	35,71
<i>Pediastrum tetras</i> var. <i>tetraedron</i> (Ehrenberg) Ralfs			x			x	x	7,14
<i>Pediastrum angulosum</i> var. <i>angulosum</i> (Ehrenberg) ex Meneghini					x			3,57
<i>Pediastrum boryanum</i> var. <i>longicorne</i> Reinsch	x	x	x		x	x	x	47,62
<i>Pediastrum biradiatum</i> var. <i>longicornitum</i> Gutwinski			x				x	5,95
<i>Pediastrum boryanum</i> var. <i>cornutum</i> (Raciborski) Sulek	x		x	x	x	x	x	57,14
<i>Pediastrum duplex</i> var. <i>rugulosum</i> Raciborski		x			x			16,57
<i>Pediastrum simplex</i> var. <i>simplex</i> Meyen					x			7,14
Micractiniaceae								
<i>Selenodictyum brasiliense</i> Uherk					x			5,96
<i>Dictyosphaerium pulchellum</i> Wood	x	x	x	x	x	x	x	57,14
<i>Dictyosphaerium tetrachotonum</i> Printz		x	x	x	x		x	64,49
<i>Dictyosphaerium elegans</i> Bachmann			x	x	x	x	x	64,49
<i>Botryococcus braunii</i> Kützing							x	5,96

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
<i>Micractinium pusillum</i> Fresenius.	x	x			x	x	x	23,81
<i>Micractinium valkanovii</i> Vodenicarov		x	x					14,29
<i>Micractinium bornhemiense</i> (Conrad) Korshikov							x	5,96
Chlorellaceae								
<i>Kirchneriella diana</i> var. <i>major</i> (Bohlin) Comas	x		x	x	x		x	64,49
<i>Kirchneriella obesa</i> (W.West & G.S. West) Schmidle							x	5,96
<i>Kirchneriella irregulari</i> (G.M.Smith) Korshikov s			x		x	x	x	28,57
<i>Ankistrodesmus gracilis</i> (Reinsch) Korshikov	x	x	x	x	x	x	x	100
<i>Ankistrodesmus fusiformis corda sensu</i> Korshikov	x		x		x	x		35,71
<i>Ankistrodesmus fusiformis</i> West & G.S.West	x	x	x	x	x	x	x	100
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs	x	x	x	x	x	x	x	100
<i>Ankistrodesmus bernardii</i> Komárek			x					7,11
<i>Ankistrodesmus spiralis</i> (Turner) Lemmermann			x		x			19,05
<i>Monoraphidium tortile</i> (West & G.S.West) Komárek-Legnerova	x	x	x	x	x	x	x	85,71
<i>Closterinospis acicularis</i> (Chodat) Belcher & Swale		x		x		x		21,42
<i>Quadrigula closterioides</i> (Bohl) Printz	x		x		x		x	21,42
<i>Chlorella vulgaris</i> Beijerinck	x	x	x	x	x	x	x	58,33
<i>Chlorococcum minutum</i> Starr		x	x	x		x	x	35,71
<i>Gleocystis vesiculosa</i> Nägeli							x	5,96
Oocystaceae								
<i>Oocystis lacustris</i> Chodat	x	x	x	x	x	x	x	100
<i>Oocystis borgei</i> Snow			x				x	10,71
<i>Oocystis solitaria</i> Wittrock	x	x	x	x	x	x	x	85,71
<i>Oocystis</i> sp			x				x	10,71
<i>Lagerheimia chodatii</i> Bernard	x		x	x		x	x	59,52
<i>Lagerheimia marssonii</i> Lemmermann	x		x				x	21,42
<i>Elakatothrix gelatinosa</i> Wille	x		x		x		x	10,05
Coelastraceae								
<i>Coelastrum microporum sphaericum</i> Nägeli	x	x	x	x	x		x	64,29
<i>Coelastrum pseudomicroporum</i> Korshikov	x		x	x	x		x	71,43
<i>Coelastrum reticulatum</i> (Dangeard) Senn	x	x	x	x			x	64,29
<i>Actinastrum hantzschii</i> Lagerheim		x	x	x			x	64,487
<i>Actinastrum gracillimum</i> Smith	x		x	x	x	x	x	85,71
Chlorococcaceae								
<i>Tetraedron arthrodesmiforme</i> f. <i>typicum</i> (West) Woloszyńska	x					x	x	9,52
<i>Tetraedron minimum</i> (Brébisson) Hansgirg			x	x	x	x	x	71,43
<i>Tetraedron minimum</i> fo. <i>apiculatum</i> (Reinsch) De Tony					x			3,57
<i>Tetraedron arthrodesmiforme</i> G.S. West			x					3,57
<i>Tetraedron regulare</i> Kützing							x	7,14
<i>Tetraedron trigonum</i> (Nägeli) Hansgirg	x		x		x	x	x	59,52
<i>Pseudostaurastrum hastatum</i> (Reinsch) Chodat			x		x	x	x	57,14
<i>Pseudostaurastrum lobulatum</i> (Pascher) Fott in Fott & Komárek	x		x	x		x		57,14

TAXONS	Sampling stations							
	G1	G2	G3	G4	G5	G6	G7	FO
<i>Pseudostaurastrum gracile</i> (Reinch) Chodat ex. Bourrelly	x					x	x	21,43
Radiococccaceae								
<i>Coenocystis quadriguloides</i> Fott								
Scenedesmaceae								
<i>Crucigenia quadrata</i> Morren	x	x	x	x	x	x	x	71,43
<i>Crucigenia fenestrata</i> (Schmidle) Schmidle							x	10,71
<i>Crucigenia tetrapedia</i> (Kirchner) Kuntze	x	x		x	x	x		59,52
<i>Desmodesmus quadricaudata</i> (Turpin) Hegewald	x	x	x			x	x	64,29
<i>Desmodesmus regularis</i> (Swirenko) Hegewald & Vanormelinger			x					7,11
<i>Dimorphococcus lunatus</i> A. Braun						x		7,11
<i>Scenedesmus quadricauda</i> (Turpin) Brébisson	x	x	x	x	x	x	x	100
<i>Scenedesmus acuminatus</i> var. <i>javanensis</i> f. <i>globolus</i> Uherkovich					x			7,11
<i>Scenedesmus acuminatus</i> var. <i>acuminatus</i> (Lagerheim) Chodat	x	x	x			x	x	64,29
<i>Scenedesmus denticulatus</i> Lagerheim		x			x			23,80
<i>Scenedesmus acutiformis</i> Schröder	x		x	x			x	42,86
<i>Scenedesmus disciformis</i> (Chodat) Fott & Komárek	x	x	x	x	x	x	x	85,71
<i>Scenedesmus bicaudatus</i> (Hansgirg) Chodat	x			x	x	x	x	64,29
<i>Scenedesmus eornis</i> (Ehrenberg) Chodat			x					10,71
<i>Scenedesmus bernadii</i> Chodat			x				x	23,80
<i>Scenedesmus naegellii</i> Brébisson	x	x	x			x	x	64,29
<i>Scenedesmus magnus</i> Meyen	x	x				x	x	47,62
<i>Scenedesmus opoliensis</i> Richt	x		x	x	x	x	x	64,29
<i>Tetrastrum elegans</i> Playfair			x		x		x	21,42
<i>Tetrallentos lagerheimii</i> Teiling	x		x		x	x		42,86
<i>westella botryoides</i> (W.West) De Wild				x		x	x	21,42
ZYGNEMATOPHYCEAE								
ZYGNEMATALES								
Desmidiaceae								
<i>Cladophora holsatica</i> Kützing	x					x		7,14
<i>Hyalotheca dissiliense</i> Brébisson ex Ralfs				x				7,14
<i>Arthrodesmus convergens</i> (Ehrenberg) ex Ralfs	x		x	x	x	x	x	35,71
<i>Arthrodesmus curvatus</i> (Turner) Coesel & Van Geest	x							14,29
<i>Pleurotaenium ehrenbergii</i> var. <i>undulatum</i> (De Bary)	x							7,14
<i>Closterium ehrenbergii</i> Meneghini ex Ralfs							x	7,14
<i>Closterium lineatum</i> Ehrenberg						x		7,14
<i>Closterium dinae</i> var. <i>minus</i> Ehrenberg		x	x		x			32,14
<i>Closterium gracile</i> Brébisson ex Ralfs	x	x	x	x	x		x	85,71
<i>Closterium kuetzingii</i> Brébisson		x	x				x	42,86
<i>Closterium striolatum</i> Ehrenberg ex Ralfs				x				7,14
<i>Closterium subulatum</i> (Kützing) Brébisson				x			x	9,52
<i>Cosmarium vexatum</i> West	x			x				9,52
<i>Cosmarium depressum</i> var. <i>achondrum</i> (Boldt) West & West							x	7,14

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
<i>Cosmarium granatum</i> Brébisson ex Ralfs var. <i>granatum</i>	x		x		x			21,42
<i>Cosmarium granatum</i> var. <i>concovum</i> Lagerheim			x	x		x		28,57
<i>Cosmarium trilobulatum</i> Reinsch			x				x	14,29
<i>Cosmarium laeve</i> Rabenhorst						x		7,14
<i>Cosmarium depressum</i> (Nägeli) Lundell		x	x			x		21,14
<i>Cosmarium margaritatum</i> (Lundell) Roy et Bisset var. <i>margaritatum</i>			x					7,14
<i>Cosmarium binoculatum</i> (Brébisson) ex Ralfs				x	x	x		28,57
<i>Cosmarium obsoletum</i> (Hantzsch) Reinsch	x		x		x			9,52
<i>Cosmarium venustum</i> (Brébisson) Archer					x			7,14
<i>Cosmarium brébissonii</i> Ralfs		x						7,14
<i>Cosmarium contractum</i> Kirchner	x		x				x	8,33
<i>Cosmarium quadrum</i> Lundell							x	7,14
<i>Cosmarium moniliforme</i> var. <i>moniliforme</i> (Turpin) Ralfs	x	x	x		x		x	42,85
<i>Cosmarium succisum</i> West			x					5,85
<i>Cosmarium</i> sp			x	x		x		42,85
<i>Euastrum denticulatum</i> (Kirchner) Gay			x	x		x		42,85
<i>Euastrum denticulatum</i> var. <i>quadrifarium</i> Willi Krieger			x		x			28,57
<i>Euastrum evolutum</i> (Nordstedt) West. & G.S. West							x	7,14
<i>Euastrum sphyroides</i> (Nordstedt) var. <i>hieronymusii</i> (Schmidle) Krieger				x				7,14
<i>Euastrum elegans</i> (Turpin) Ralfs	x				x			7,14
<i>Euastrum dubium</i> Nægeli	x		x		x	x		42,85
<i>Euastrum germanicum</i> (Schmidle) Krieger	x		x		x		x	42,85
<i>Euastrum glaziovii</i> BØrges	x	x	x	x	x	x	x	42,85
<i>Euastrum pseudopectinatum</i> Schmidle	x	x		x		x		42,85
<i>Euastrum ivoirensis</i> Bourrelly	x		x			x		42,85
<i>Euastrum</i> sp	x		x					14,29
<i>Sphaeroszma granulatum</i> Roy & Bisset				x		x		14,29
<i>Sphaeroszma excavatum</i> Ralfs ex Ralf	x	x	x	x	x	x	x	100
<i>Sphaeroszma filiformis</i> Ehrenberg ex Ralfs			x			x		14,29
<i>Teilingia granulata</i> (Roy & Bisset) Bourrelly		x	x	x				7,14
<i>Spondylosium</i> sp			x			x		14,29
<i>Staurodesmus cuspidatus</i> (brébisson) Teiling	x	x	x		x	x	x	57,14
<i>Staurodesmus dejectus</i> (brébisson) Teiling	x	x	x		x	x		47,62
<i>Staurodesmus patens</i> (nordstedt) croasdale		x						7,14
<i>Staurodesmus triangularis</i> (brébisson) Teiling	x	x	x		x	x	x	57,14
<i>Staurodesmus mucronatus</i> var. <i>subtriangularis</i> West. & G.S. West					x			7,14
<i>Staurastrum caledonense</i> Hüber-Pestalozzi	x		x				x	28,57
<i>Staurastrum excavatum</i> West & G.S. West	x		x		x	x	x	41,67
<i>Staurastrum forficulatum</i> var. <i>minus</i> (Fritsch & Rich) Grönblad	x	x	x	x	x	x	x	100
<i>Staurastrum furcatum</i> (Ehrenberg) Brébisson		x	x		x		x	42,86
<i>Staurastrum gracile</i> Ralf ex Ralf	x	x	x	x	x	x	x	100
<i>Staurastrum gracile</i> var. <i>coronulatum</i> Boldt		x	x		x	x	x	41,67

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
<i>Staurastrum gracile</i> var. <i>elongatum</i> Scott & Prescott	x	x	x	x	x	x	x	100
<i>Staurastrum leptocladum</i> var. <i>cornutum</i> Wille	x	x	x	x	x	x	x	71,43
<i>Staurastrum pingue</i> Teiling		x	x	x	x	x	x	42,86
<i>Staurastrum subgracillimum</i> West. & G.S. West	x				x	x		35,71
<i>Staurastrum ginzbergeri</i> Grönblad	x					x		14,29
<i>Staurastrum setigerum</i> var. <i>occidentale</i> West & G.S. West					x	x		21,43
<i>Staurastrum setigerum</i> var. <i>setigerum</i> Cleve			x		x	x	x	28,57
<i>Staurastrum thomassonianum</i> Compère		x	x	x	x	x	x	47,62
<i>Staurastrum unicome</i> Turner					x			7,14
<i>Staurastrum subarcuatum</i> Wolle			x				x	23,80
<i>Staurastrum volans</i> West & G.S. West	x	x	x	x	x	x	x	100
<i>Staurastrum boreale</i> var. <i>quadriradiatum</i> Korshikov				x				11,90
<i>Staurastrum pentasterias</i> Grönblad				x				11,90
<i>Staurastrum circulus</i> Grönblad			x					7,14
<i>Desmidium Baileyi</i> (Ralfs) Nordstedt		x					x	10,71
<i>Desmidium aptogonum</i> Brébisson ex Kützing			x				x	11,90
<i>Ichthyodontum sachlanii</i> Scott & Prescott			x				x	10,71
<i>Mougeotia</i> sp							x	11,90
Zygnemataceae								
<i>Zygnema ramosum</i> Ahmad & Goldstein							x	3,57
<i>Spirogyra</i> sp	x				x			10,71
Mesotaeniaceae								
<i>Mesotaenium macrococcum</i> (Kützing ex Kützing) Roy & Bisset							x	3,57
<i>Netrium pseudactinotaenium</i> Coesel		x				x		10,71
VOLVOCALES								
Volvaceae								
<i>Eudorina elegans</i> Ehrenberg	x	x	x	x	x		x	85,71
<i>Eudorina uniccocca</i> Smith	x	x	x	x	x		x	85,71
<i>Pandoria morum</i> Bory	x	x	x	x	x	x	x	100
<i>Volvox aureus</i> Ehrenberg	x		x		x		x	14,29
<i>Volvox carteri</i> Stein	x		x		x		x	14,29
OEDOGONIOPHYCEAE								
OEDOGONIALE								
Oedogoniaceae								
<i>Oedogonium</i> sp	x				x			23,80
<i>bulbochaete</i> sp			x				x	02,38
PYRRPHYTA								
DINOPHYCEAE								
GYMNODINIALES								
Gymnodiniaceae								
<i>Gymnodinium rotundatum</i> klebs		x		x		x		28,57
<i>Karenia</i> sp	x		x		x		x	28,57

TAXONS	Sampling stations								FO
	G1	G2	G3	G4	G5	G6	G7		
<i>Alexandrium minutum</i> Halim	x	x	x	x	x	x	x	x	42,86
PERIDINIALES									
<i>Peridinium cinctum</i> (Müller) Ehrenberg	x	x	x	x	x	x	x	x	100
<i>Protoperidinium quinquecornu</i> (Abé) Balech	x	x	x	x	x	x	x	x	57,14
CRYPTOPHYCEAE									
CRYPTOMONADALES									
<i>cryptomonas</i> sp									02:38
BACILLARIOPHYTA									
COSCINODISCOPHYCEAE									
CHAETOCEROTALES									
<i>Acanthoceras zachariasii</i> (Brun) Simonsen									23,80
<i>Chaetoceros</i> sp	x						x		23,80
AULACOSEIRALES									
<i>Aulacoseira granulata</i> (Ehrenberg) Simonsen	x	x	x	x	x	x	x	x	100
<i>Aulacoseira granulata</i> var. <i>angustissima</i> (Müller) Simonsen	x	x	x	x	x	x	x	x	100
<i>Aulacoseira ambigua</i> (Grunow) Simonsen	x	x	x	x	x	x	x	x	100
<i>Aulacoseira ambigua</i> f. <i>japonica</i> Tuji & Williams	x	x	x	x	x	x	x	x	100
THALASSIOSIRALES									
<i>Cyclotella</i> sp	x					x			7,14
FRAGILARIOPHYCEAE									
FRAGILARIALES									
<i>Asterionella formosa</i> Hassall	x	x	x	x	x	x	x	x	85,71
<i>Synedra acus</i> Kützing	x	x	x	x	x	x	x	x	85,71
<i>Synedra ulna</i> (Nitzsch) Ehrenberg.	x	x	x	x	x	x	x	x	85,71
<i>Ulnaria ulna</i> (Nitzsch) Compère	x	x	x	x	x	x	x	x	42,86
<i>Fragilaria crotonensis</i> Kitton						x		x	14,28
EUNOTIALES									
<i>Eunotia bilunaris</i> (Ehrenberg) Mills						x			5,85
<i>Eunotia incisa</i> Smith & Gregory									14,28
<i>Eunotia mesiana</i> Cholnoky								x	5,85
<i>Eunotia monodon</i> var. <i>bidens</i> (Ehrenberg) Hustedt									5,85
<i>Eunotia pectinalis</i> (Kützing) Grunow								x	47,61
<i>Eunotia zasuminensis</i> (Cabejszekowna) Kömer						x			32,14
ACHNANTHALES									

TAXONS	Sampling stations							FO
	G1	G2	G3	G4	G5	G6	G7	
Achnanthes sp				x	x	x		32,14
CYMBELLALES								
Cymbellaceae								
<i>Encyonema sp</i>	x	x	x	x	x	x	x	85,71
Gomphonemataceae								
<i>Gomphonema affine</i> Kützing				x			x	14,28
<i>Gomphonema augur</i> Ehrenberg .var. <i>turris</i> (Ehrenberg) Lange-Bertalot							x	5,85
NAVICULALES								
Pleurosigmataceae								
<i>Gyrosigma spencerii</i> (Quekett) Griffith & Henfrey	x	x	x	x	x		x	85,71
Pinnulariaceae								
<i>Pinnularia acrosphaeria</i> Smith	x	x	x	x	x	x	x	85,71
<i>Pinnularia gibba</i> (Ehrenberg) Ehrenberg	x	x	x	x	x	x	x	85,72
<i>Pinnularia interrupta</i> Smith	x	x	x	x	x	x	x	85,71
<i>Pinnularia viridis</i> (Nitzsch) Ehrenberg			x		x			5,85
<i>Pinnularia micostauron</i> (Ehrenberg) Cleve					x			5,85
<i>Pinnularia neomajor</i> Krammer.			x	x				7,14
<i>Pinnularia divergence</i> Smith	x		x		x		x	06:57
<i>Pinnularia sp.1</i>	x	x						7,14
naviculaceae								
<i>Navicula cryptocephala</i> Kützing	x				x	x	x	38,09
<i>Craticula cuspidata</i> (Kützing) Mann					x			5,85
<i>Sellaphora pupula</i> (Kützing) Mereschkovsky	x		x	x	x	x	x	57,14
Neidiumaceae								
<i>Neidium ampliatum</i> (Ehrenberg) Krammer		x		x		x		17,85
<i>Neidium sp</i>	x				x			5,85
Caloneidaceae								
<i>Caloneis silicula</i> (Ehrenberg) Cleve					x			5,85
<i>Caloneis sp.1</i>					x			5,85
Stauroneidaceae								
<i>Stauroneis anceps</i> Ehrenberg	x	x		x	x	x		47,61
<i>Stauroneis phoenicenteron</i> (Nitzsch) Ehrenberg			x				x	7,14
BACILLARIALES								
Bacillariaceae								
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow		x				x		7,14
<i>Nitzschia palea</i> (Kützing) W.Smith	x	x	x		x		x	85,71
<i>Nitzschia cf. sigmaformis</i> Hustedt				x				2,38
<i>Nitzschia sp</i>	x	x	x		x	x	x	85,71
SURIRELLALES								
Surirellaceae								
<i>Surirella biseriata</i> De Brébisson		x		x			x	17,85

TAXONS	Sampling stations								FO
	G1	G2	G3	G4	G5	G6	G7		
<i>Surirella linearis</i> Smith		x				x		7,14	
<i>Surirella</i> sp	x	x	x	x	x		x	85,71	
CHRYSOPHYTA									
CHRYSOPHYCEAE									
CHROMULINALES									
		Dinobryaceae							
<i>Dinobryon sertularia</i> Ehrenberg.	x	x	x	x	x	x	x	85,71	
XANTHOPHYTA									
XANTHOPHYCEAE									
MISCHOCOCCALES									
		Centrtractaceae							
<i>Xanthonema</i> sp							x	2,38	
<i>Heterococcus fuornensis</i> Vicher	x	x	x	x	x	x	x	41,67	
<i>Centrtractus belonophorus</i> (Schmidle) Lemmermann	x	x	x	x	x	x	x	100	
		Ophiocytaceae							
<i>Ophiocytium capitatum</i> Wolle		x			x			14,28	
<i>Ophiocytium cochleare</i> (Eichwald) Braun				x			x	14,29	
		Pleurochloridaceae							
<i>Tetraplektron torsum</i> Turner	x		x	x	x		x	64,29	
<i>Tetraplektron</i> sp					x			5,85	
<i>Tetraedriella gigas</i> (Pascher) Smith			x					5,85	
<i>Isthmochloron</i> sp			x				x	2,38	
Total = 340	179	152	208	159	189	170	209		