



Studies on the efficacy of some preservatives used in packaged orange drinks

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ABSTRACT

The antibacterial activity of benzoic acid and sodium benzoate was investigated against bacterial isolates from packaged orange drinks using agar well diffusion and broth dilution methods. The antibacterial activity of the test agents against the standard NCTC bacteria species was also tested. The bacterial count from the packaged orange drinks ranged from 3.0×10^5 cfu/ml and 1.43×10^6 cfu/ml. The bacteria species detected consisted of *Bacillus subtilis*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Micrococcus sp.* The zones of inhibition of benzoic acid ranged from 19.0 mm – 31.5 mm while that of sodium benzoate ranged from 13.5 mm – 36.5 mm. The Minimum Inhibitory Concentration (MIC) of the preservatives against the test bacteria ranged between 0.156 μ g/ml and 0.625 μ g/ml while the Minimum Bactericidal Concentration (MBC) ranged between 0.313 μ g/ml and 500 μ g/ml. The preservatives were more effective against the Gram positive bacteria than the Gram negative bacteria. The preservatives at the concentration used in the examined drinks are inadequate to keep off indicator organisms and to ensure their safe consumption.

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Keywords: Benzoic acid, sodium benzoate, *Bacillus subtilis*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Micrococcus sp.*

INTRODUCTION

Antimicrobial preservatives are included in biological preparations to kill or inhibit the growth of microorganisms inadvertently introduced during manufacture or use which may cause microbial deterioration. The use of preservatives in products is to reduce the likelihood of microbial growth in aqueous products and also to minimize the chances of microbial survival in anhydrous products that may be contaminated (Baird, 2004). They are also added to industrial products which may, by their nature, support the growth of bacteria and moulds causing spoilage of the product and possibly infection of the user. Chemical

preservatives are frequently used in processed foods to prevent growth of bacteria, yeast or other microorganisms (Jegtvig, 2011). An ideal or satisfactory food preservative remains effective in a product until the product is consumed. Various steps are involved in the processing of orange drink; therefore there is need for preservation.

The commonly used preservatives in packaged orange drinks are benzoic acid and sodium benzoate. Sodium benzoate is a type of preservative commonly used in the fruit pies, jams, beverages, salads, relishes and sauerkraut, typically foods that have an acidic

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pH. Sodium benzoate and water produce benzoic acid, which is the active form of the preservative (Jegtvit, 2011).

Sodium benzoate is utilized in pharmaceutical industry as a diagnostic reagent for liver functions. Sodium benzoate and Benzoic acid are employed in a wide range of preservative applications because of its combination of bactericidal and bacteriostatic action with non-toxicity and tastelessness (Shree additives, 2010).

Benzoic acid would uncouple substrate transport and oxidative phosphorylation from the electron transport system by making the cytoplasmic membrane freely permeable to proton (Lou et al., 2007). Benzoates interfere with the utilization of acetate required for the function of energy rich compounds which results in blockage of cell metabolism (Olutimayin et al., 2001). The United States Food and Drug Administration has studied sodium benzoate extensively and found that it is safe when consumed in amounts found in normal diets (Jegtvig, 2011). The pH value of the products often affects the preservative action of sodium benzoate, the preservative action being much greater at low pH than at higher pH values. This study intends to ascertain whether the recommended concentration of the preservatives is able to meet the objectives of its inclusion in the packaged orange drinks.

MATERIALS AND METHODS

Sample collection

Samples of the packaged orange drinks were purchased from various outlets in Kaduna, Nigeria. The samples were worked on immediately or kept in the refrigerator at 4 °C.

Isolation and identification of bacteria

Each sample of the packaged orange drinks was diluted between 1:10 to 1:10⁵ in sterile normal saline. One milliliter (1.0 ml) of each dilution was added to each McCartney bottle containing the sterile nutrient agar and 3% Tween 80 and then mixed thoroughly. The

content of the bottle was poured into sterile petri dishes and allowed to set. The plates were incubated at 37 °C for 24 – 36 hours. After which the colonies which developed were counted using colony counter. The bacteria isolates were identified using their colonial morphology, cellular morphology and appropriate biochemical tests.

Susceptibility testing

Exactly 19.0 ml of sterile nutrient agar was inoculated with 0.1 ml of 24-hour broth culture which has been diluted to 0.5 McFarland, which is about 10⁶ cfu/ml. The mixture was properly but gently shaken, poured into sterile petri dishes and allowed to set. A sterile cork borer (No. 4) was used to bore about five equidistant cups into the agar plate. One drop of the molten agar was used to seal the bottom of the bored hole, so that the preservative will not sip beneath the agar. Different concentrations of the preservatives were added to fill the bored holes. One hour pre-diffusion time was allowed, after which the plates were incubated at 37°C for 24 hours. The zones of inhibition were then measured in millimeter. Control plates were prepared and incubated appropriately.

Determination of the Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC)

Ten test tubes of 2.5 ml nutrient broth were set in rows. The first tube contained double strength broth. To the first was added 2.5 ml of the preservative and thoroughly but gently mixed, 2.5 ml of the mixture was withdrawn and to the second tube and mixed properly, this dilution continued serially to the last tube, after mixing, 2.5 ml was withdrawn from the last tube and discarded. Each of the tubes was inoculated with 0.1 ml of the standardized inoculum. Three controls were set up to show the sterility of the media, the preservatives and to ascertain the growth promoting property of the media. The tubes were incubated at 37 °C for 18 hours. The lowest concentration of the preservative in the test tubes that showed no growth was

considered as the M. I. C. of the preservative against the organisms.

After incubation, a loopful from the tubes containing the least concentration of the preservative which prevent growth was streaked on sterile nutrient agar plates containing inactivating agents 3% $\frac{v}{v}$ Tween 80 and incubated at 37 °C for 24 hours. The least concentration of the preservative in the test agar plates that showed no growth was considered as the M. B. C. of the preservative against the test organism (Onaolapo et al., 1993).

RESULTS

The total count of the bacteria from the various orange drinks range from 3.0×10^5 cfu/ml – 1.43×10^6 cfu/ml (Table 1). The bacterial species isolated from the drinks included pathogens that are officially not permitted in drinks such as *Escherichia coli* and *Pseudomonas aeruginosa*. The bacterial load of *Micrococcus* sp and spore forming *Bacillus subtilis* isolated from the drinks was also higher than the officially permitted bacterial load for such organisms (Table 1).

From the result of the susceptibility testing, the preservatives showed more

antibacterial activity against the Gram positive bacteria than the Gram negative bacteria (Tables 2 and 3). The test bacteria species were more susceptible to the test preservatives than the standard bacteria species except *E. coli* (Tables 2 and 3).

The MIC values of the preservatives were higher against the Gram negative bacteria isolates; *E. coli* and *Ps. aeruginosa*, than the Gram positive bacteria isolates; *Micrococcus* sp. and *B. subtilis*, which means the preservatives showed more bacteristatic activity against the Gram positive bacteria than the Gram negative bacteria (Figure 1). The MBC values of the benzoic acid against the type bacteria species were relatively higher than those against the bacteria isolates from the drinks, which means that the benzoic acid was more active against the bacteria isolates from the drinks than the type bacteria. However, the MBC values of sodium benzoate against the bacteria isolates from the packaged orange drinks were relatively higher than those against the type NCTC bacteria, that is, sodium benzoate showed more bactericidal activity against type bacteria than bacteria isolates from the drinks (Figure 2).

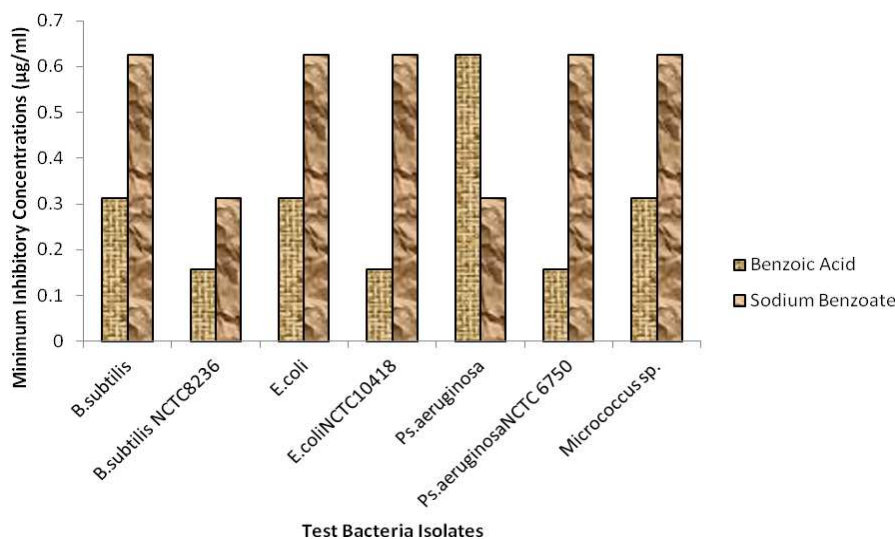


Figure 1: Minimum Inhibitory Concentrations of Benzoic Acid and Sodium Benzoate against the Bacteria Isolates from the Orange Drinks and Type NCTC Bacteria species.

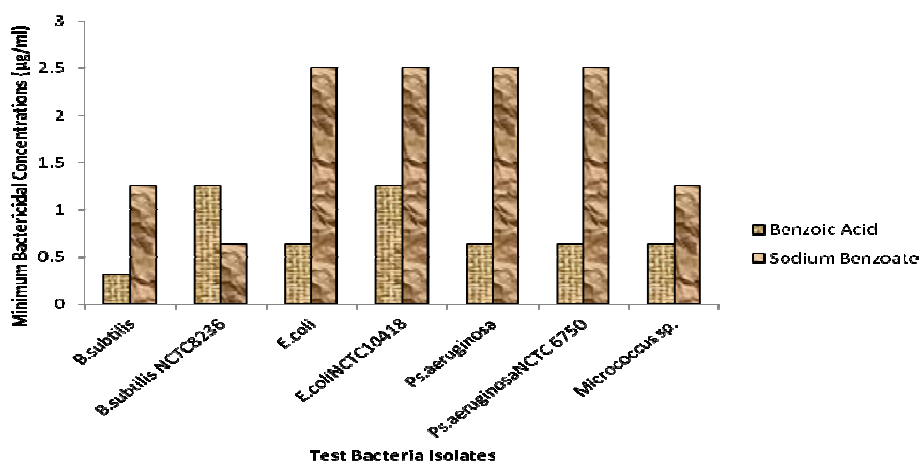


Figure 2: Minimum Bactericidal Concentrations of Benzoic Acid and Sodium Benzoate against the Bacteria Isolates from the Orange Drinks and Type NCTC Bacteria species.

Table 1: Bacterial viable count and distribution of bacteria isolated from the various packaged orange drinks.

Brand	Sample	Bacterial viable count (cfu/ml)	<i>B. subtilis</i>	<i>E. coli</i>	<i>Ps. aeruginosa</i>	<i>Micrococcus</i> sp.
A	10	1.43 x 10 ⁶	+	+	+	-
B	10	1.11x10 ⁶	+	+	+	+
C	10	1.01 x10 ⁶	+	+	-	-
D	10	9.3 x 10 ⁵	+	+	-	+
E	10	9.1 x 10 ⁵	+	-	-	+
F	10	8.1 x 10 ⁵	+	-	-	+
G	10	3.0 x 10 ⁵	+	+	+	-

+ = present, - = absent.

Table 2: Susceptibility of the bacteria isolates from the drinks and type bacteria species to benzoic acid.

Test Bacteria	Zones of Inhibition (mm)		
	0.4 µg/ml	0.2 µg/ml	0.1 µg/ml
<i>B. subtilis</i>	31.0	29.5	28.0
<i>B. subtilis</i> NCTC 8236	30.0	25.5	25.0
<i>E. coli</i>	27	26	22
<i>E. coli</i> NCTC 10418	31.5	31	24
<i>Ps. aeruginosa</i>	24.5	22	19
<i>Ps. aeruginosa</i> NCTC 6750	22	21	20.5
<i>Micrococcus</i> sp.	37	31	28

Table 3: Susceptibility of the bacteria isolates from the drinks and type bacteria species to sodium benzoate.

Test Bacteria	Zones of Inhibition (mm)				
	4.0 µg/ml	2.0 µg/ml	1.0 µg/ml	0.5 µg/ml	0.25 µg/ml
<i>B. subtilis</i>	36.5	34.5	31	20	16.5
<i>B. subtilis</i> NCTC 8236	25	23	17.5	NI	NI
<i>E. coli</i>	32	26	15.5	NI	NI
<i>E. coli</i> NCTC 10418	27.5	17	NI	NI	NI
<i>Ps. aeruginosa</i>	31.5	22.5	13.5	NI	NI
<i>Ps. aeruginosa</i> NCTC 6750	25	19	NI	NI	NI
<i>Micrococcus</i> sp.	33	24.5	19	NI	NI

NI – No Inhibition.

DISCUSSION

The high load of bacteria from the packaged orange drinks, despite the addition of preservatives, suggests low efficiency of the preservatives. The aim of preservation is to satisfactorily keep a product against microbial challenge while it is in trade channel and in analyzed samples. The official limit recommended for microbial contamination of drinking water, which also includes fruit drinks, requires the complete absence of some pathogens like *E. coli* and other coliforms, *Pseudomonas aeruginosa* and other organisms like *S. aureus*. Other permissible bacteria should not be more than 10^3 colony forming unit (cfu)/ml while mould/yeast should not exceed 10^2 spore forming unit (sfu)/ml (WHO, 2008). Benzoic acid and sodium benzoate had been found to permit the development of some bacteria (Gorman and Scott, 2004). Packaged orange drinks must have a preservative system that is capable of sterilizing the drinks if contamination should occur (Olutimayin et al., 2001).

The result of the test on the efficacy of the preservatives showed that they are more active against Gram positive bacteria, *B. subtilis* and *Micrococcus* sp than the Gram negative bacteria *E. coli* and *Ps. aeruginosa*. Benzoic acid and sodium benzoate have been reported to be less active against Gram negative bacteria (Olutimayin et al., 2001). Gram negative bacteria are known to be resistant to the action of most antimicrobial agents because of the presence of outer

phospholipids membrane with the structural lipopolysaccharide components, which make their cell wall impermeable to antimicrobial agents (Willey et al., 2008). *Pseudomonas aeruginosa* appeared to be less sensitive to the preservatives than all the other organisms. Strains of *Pseudomonas* sp. have been reported to be less sensitive to many antimicrobial agents (Wiley et al., 2008). Among other factors, the lack of sensitivity to antimicrobial agent by bacteria may be due to inability of the agent to diffuse into the cell and cellular impermeability which leads to a reduced concentration of the antimicrobial compound available at the target site so that the cell may escape injury (Denyer and Russell, 2004).

From the results of the MIC of the preservatives against the organisms, it can be suggested that the recommended concentrations of 0.05% - 0.1% is low and may not be active against the isolated bacteria species and this may account for the high bacteria load in the samples. The MIC values showed by sodium benzoate are higher than that of benzoic acid, signifying that benzoic acid is more active than sodium benzoate. Although, benzoic acid was more active against the bacteria isolates from the drinks than the type bacteria. However, the MBC values of sodium benzoate against the bacteria isolates from the packaged orange drinks were relatively higher than those against the type NCTC bacteria, that is, sodium benzoate showed more bactericidal activity against type bacteria than bacteria isolates from the drinks.

Benzoic acid has limited use in preservatives because it has pH 4.2 which is highly acidic for food products and development of resistance to it by some organisms involving in some cases of metabolism of the acid resulting in complete loss of activity (Gorman and Scott, 2004). This disadvantage is one of the reasons why the salt of benzoic acid, sodium benzoate, is preferred in the preservation of foods and drinks. Therefore, if sodium benzoate is preferred to benzoic acid in preservation, then it is evident from this result that the salt cannot curtail the proliferation of the contaminating bacteria. The result of the MBC of the preservatives against the bacteria sp. also showed that the in-use concentration of sodium benzoate is low if the orange drinks are to remain safe for consumption. When used as a preservative, sodium benzoate is typically added to foods in small amounts only. If too much is added, food may take on a very bitter taste (Ellis-Christensen, 2011).

Conclusion

Generally, the susceptibility of the bacteria isolates and standard NCTC bacteria to the tested preservatives was relatively good *in-vitro*; the used preservatives were not able to curtail the proliferation of bacteria in the tested packaged orange drinks *in-vivo*. It is therefore suggested that due to the fact that the preservatives at high concentration may have adverse effect on the drinks and on the consumers, the use of other preservatives of local sources can be considered for the preservation of the packaged drinks.

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