

## EFFECT OF PUBESCENCE ON THE OVIPOSITION AND FEEDING BEHAVIOUR OF *M. VITRATA* (FAB.) ON *VIGNA* SPECIES

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### Abstract

The ovipositional and feeding behaviour of *Maruca vitrata* (Fab.) were studied in the screenhouse and field on two wild *Vigna* species, namely *V. vexillata* (Rich.) (accession TVnu 72) and *V. oblongifolia* (Rich.) (accession TVnu 42), one wild cowpea, *V. unguiculata* ssp. *dekindtiana* (Harms) Verdcourt (accession TVnu 863), one cowpea, *V. unguiculata* ssp. *unguiculata* (L. Walp) landrace (TVu 13731) and two cowpea breeding lines (IT84S-2246 and IT91K-180) at the International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria. The density and length of the trichomes on the stem, leaves and pods of the *Vigna* species were studied using scanning electron microscopy. The effect of the trichomes on oviposition and feeding by *M. vitrata* was studied in the screen-house and field, respectively. Significant ( $P < 0.05$ ) differences were observed on the density of glandular and non-glandular trichomes as well as length of the non-glandular trichomes, on the stems, leaves and pods of the *Vigna* species. Gravid *M. vitrata* females laid significantly ( $P < 0.05$ ) fewer eggs on TVnu 42 plants under free choice and TVu 13731 under no choice condition compared with other treatments. The density of glandular trichomes on leaves reduced oviposition and accounted for 70.89 per cent of viable eggs laid under free choice. Flower damage was highest on IT84S-2246, IT91K-180 and TVnu 863 while pod load was significantly ( $P < 0.007$ ) higher on TVnu 72 compared with IT84-2246 and TVnu 863. Similarly, TVnu 863, IT84S-2246 and IT91K-180 had significantly ( $P < 0.0001$ ) higher pod damage compared with TVnu 72 and TVu 13731. Pod load and pod damage were dependent on flower damage, accounting for almost 100 per cent of viable eggs but pod damage was also influenced by the density of glandular trichomes on the pods. The pubescence of *Vigna* species did not affect damage to the flower by *M. vitrata* larvae. The implication of these findings in the development of resistant cowpea varieties against *M. vitrata* damage is discussed.

### Résumé

OIGIANGBE O. N., JACKAI L. E. N., EWETE F. K., LAJIDE L. & HUGHES J. D'A.: Effets de la pubescence sur la ponte et la manière de s'alimenter de *M. vitrata* Fabricius sur l'espèce *Vigna* Les manières de ponte et de s'alimenter de *Maruca vitrata* Fabricius étaient étudiées en cage de Faraday et sur le terrain sur deux espèces de *Vigna* sauvages à savoir *V. vexillata* (A. Richard) (accession TVnu 72) et *V. oblongifolia* (A. Richard) (accession TVnu 42), un dolique sauvage *V. unguiculata* ssp. *dekindtiana* [Harms Verdcourt] (accession TVnu 863), un dolique [*V. unguiculata* ssp. *Unguiculata* (L. Walp)] Variété de pays (TVu 13731) et deux lignées de variété de dolique (IT84S-2246 et IT91K-180) à l'Institut International d'Agriculture Tropicale (IITA), Ibadan, Nigéria. La densité et la longueur de trichomes sur la tige, les feuilles et les cosses des espèces de *Vigna* étaient étudiées en employant la microscopie électronique à balayage. L'effet des trichomes sur la ponte et l'alimentation par *M. vitrata* était étudié respectivement en cage de Faraday et sur le terrain. Des différences considérables ( $P < 0.05$ ) étaient observées de la densité des trichomes glandulaires et non-glandulaires ainsi que de la longueur de trichomes non-glandulaires sur les tiges, les feuilles et les cosses de l'espèce *Vigna*. Les femelles gravides de *M. vitrata* pondaient considérablement ( $P < 0.05$ ) moins d'œufs suivant le régime de plante TVnu 42 étant libre de choisir et de TVu 13731 sous la condition de choix nul comparé aux autres traitements. La densité de trichomes glandulaires sur les feuilles réduisait la ponte et représentait 70.89 pour cent de la variabilité en nombre d'œufs pondus étant libre de choisir. Les dégâts causés aux fleurs étaient plus élevés sous IT84S-2246, IT91K-180 et TVnu 863 alors que la charge de cosse était considérablement ( $P < 0.007$ ) plus élevée sous TVnu 72 comparée à IT84-2246 et TVnu 863. De la même façon, TVnu 863, IT84S-2246 et IT91K-180 avaient des dégâts causés aux cosses considérablement ( $P < 0.0001$ ) plus élevés comparés avec TVnu 72 et TVu 13731. La charge de cosse et les dégâts causés aux cosses dépendent en grandes parties de dégâts causés à la fleur représentant.

Presque 100 pour cent de la variabilité, mais les dégâts causés aux cosses étaient aussi influencés par la densité de trichomes glandulaires sur les cosses. La pubescence des espèces de *Vigna* n'a pas eu un effet sur les dégâts causés à la fleur par les larves de *M. vitrata*. L'implication de ces résultats pour le développement de variétés de dolique résistantes aux dégâts de *M. vitrata* est discutée.

### Introduction

The behaviour of phytophagous insects is dependent on several plant characteristics, including chemical, colour, shape and pubescence. Those characteristics which elicit or inhibit oviposition play important roles in the survival of Lepidopterans, because the neonate larvae are immobile and rely on the right choice of the food plant by the adult female (Renwick & Chew, 1994). Two stage are, therefore, critical in the successful exploitation of food resource by this group of insects. The first is the ability of the female to lay eggs where the larvae can easily find food and the second is the ability of the larvae to feed on the available food. Ramaswamy (1988) and Gall (1990) reported that majority of moth species prefer hairy or rough surfaces as oviposition sites, but Navasero & Ramaswamy (1991) suggested that trichome type (i.e. glandular or non-glandular) may not influence oviposition. The resistance of a maize cultivar to *Chilo partellus*, however, was reported to be due to the high density of trichomes on the leaf surface (Kumar, 1992).

*Maruca vitrata* Fab. (Lepidoptera: Pyralidae) is a major pest of cowpea in sub-Saharan Africa (Taylor, 1978; Jackai *et al.*, 1985; Singh *et al.*, 1990). The larvae damage young stems, flower buds, flowers and pods (Taylor, 1967; Jackai, 1982) causing yield losses of 20-80 per cent (Singh & Jackai, 1985; Singh *et al.*, 1990) of all the cultivated cowpea varieties. High levels of resistance have been found in some wild *Vigna* species, notably *V. vexillata* (A. Richard) and *V. oblongifolia* (A. Richard) (Singh *et al.*, 1990; Jackai, Padulosi & Ng, 1996). There is need to develop resistant cowpea varieties with the possible use of genes

from wild *Vigna* species with the aid of new biotechnological techniques (Fatokun, Perriho & Ng, 1997).

The importance of pubescence on the stems, leaves and pods of *Vigna* species in conferring resistance against *M. vitrata* attack had been highlighted by Jackai & Oghiakhe (1989), Oghiakhe *et al.* (1992b) and Oghiakhe (1995, 1996), but little is known of the mechanism(s) by which trichomes reduce oviposition and larval damage to flowers and pods. Experiments were conducted in the screen house and field to test the hypothesis that trichome types and position affect the ovipositional behaviour of *M. vitrata* and the consequent flower or pod damage.

### Experimental

#### Source of seeds

Two wild *Vigna* species, namely *V. vexillata* (A. Rich.) (accession TVnu 72) and *V. oblongifolia* (A. Rich.) (accession TVnu 42), one wild cowpea, *V. unguiculata* ssp. *dekindtiana* [(Harms) Verdcourt] (accession TVnu 863), one cowpea, *V. unguiculata* ssp. *unguiculata* (L. Walp) landrace (TVu 13731) and two cowpea breeding lines (IT84S-2246 and IT91K-180) were used in the experiments. The wild *Vigna* and cowpea seeds were obtained from the International Institute of Tropical Agriculture (IITA) Germplasm Bank while seeds of the cowpea landrace and breeding lines were obtained from the IITA Breeding Programme.

#### Trichome study

Stem, leaf and pod samples were collected from plants, grown in pots filled with top-soil (collected from the IITA Farm Yard) at the podding stage and

fixed immediately in formo-acetic acid (FAA). Dehydration, critical point drying and coating of the samples were done using the methods of Oigiangbe *et al.* (2002). The samples were dehydrated briefly through a series of FAA, ethanol and amyl-acetate mixtures and dried in a critical point dryer CPD 030 (Bal-Tec) for 12 h.. Araldite<sup>®</sup> adhesive was used to mount each sample on stubs. The samples, adhesive and stubs were coated with a thin layer of gold-palladium alloy for 5 min to obtain a thickness of 380 nm in a Polaron SC 7610 Sputer Coater (Fisons Instruments). Each sample was scanned with a Philips 20XL scanning electron microscope at 10 kilovolts accelerating voltage. The density of glandular and non-glandular trichomes and length of the non-glandular trichome were taken from five different areas of each sample.

#### *Oviposition study*

Oviposition by *M. vitrata* was assessed in the screenhouse using potted plants under free choice and no-choice conditions. Under the free choice condition, six pots containing one or two plants at the flowering stage of each of the six *Vigna* species were randomly arranged in a nylon-mesh cage measuring  $77.61 \times 77.61 \times 77.61$  cm<sup>3</sup> and replicated three times. Thirty *M. vitrata* females, reared on artificial diet (Jackai & Raulston, 1988) were released at the centre of the cage in the evening. Under the no choice condition, one pot containing one or two plants of a particular *Vigna* species was put in a nylon-mesh cage measuring  $56.42 \times 65.42 \times 61.00$  cm<sup>3</sup> and replicated six times. Ten gravid *M. vitrata* females were released in each cage in the evening. In both choice situations, female were allowed to lay eggs on the plants for two nights. The number of eggs laid on the vegetative and floral parts of the plants was recorded. Data were log transformed and analysed using the analysis of variance (ANOVA) and means were separated with Student-Newman-Keuls (SNK) test, where the F-value was significant. The relationships between the

variables were assessed using correlation and regression analysis.

#### *Field study*

The field experiment was conducted between April and July 1997 at IITA, Ibadan. The *Vigna* species were each planted in plots arranged in a randomized complete block design. Each plot had five rows of 5 m long with a spacing of 1.50 m between plots, 0.75 m between rows, and 0.20 m within rows. Thinning to one seedling per stand was done 2 weeks after planting (WAP). Two weeks prior to planting the test materials, a highly susceptible cultivar (IT86D-889) was planted in a perpendicular manner to every range of rows within the plot to serve as multiplication foci (spreader row) for the insect. Monocrotophos (Nuvacron<sup>®</sup> 40EC) was sprayed at the onset of flower bud initiation at a rate of 200 g a.i./ha to control leaf feeding beetles, aphids, thrips and pod sucking bugs later during the podding. This was to ensure that these pests did not mask the effect of *M. vitrata* on the crop. Monocrotophos had been reported to be non-toxic to *M. vitrata* (Jackai, 1983).

Flower damage was evaluated twice weekly for 3 weeks from one week after flower bud initiation, using the rapid visual evaluation (RVE) method of Oghiakhe, Jackai & Makanjuola (1992a). This method involves a random selection of 20-50 flowers per plot, opening them immediately and recording the number of damaged flowers (with *M. vitrata* larvae or frass). The average flower damage for each week was calculated from the two records. Scoring the treatments at pod filling stage on pod load and pod damage (Jackai & Singh, 1988; Jackai, 1995) was used to assess pod production and damage. Both scores were based on a scale of 1-9, one the reverse of the other (Jackai, 1995). The data were log (for pod load) or square root (for pod damage) transformed and analyzed using the analysis of variance (SAS System for Windows<sup>®</sup>). Means were separated by SNK test where the F-value was significant. Correlation, as

TABLE 1

Mean density ( $\text{mm}^2$ ) of glandular and non-glandular trichomes and length ( $\mu\text{m}$ ) of non-glandular trichomes of stems of wild and cultivated *Vigna* species

| Vigna species                                 | Accession No. | Density             |                         | Length of non-glandular trichomes |
|---|---------------|---------------------|-------------------------|-----------------------------------|
|   |               | Glandular trichomes | Non-glandular trichomes |                                   |
| <i>V. vexillata</i>                           | TVnu 72       | 1.00 $\pm$ 0.24b    | 2.60 $\pm$ 0.11c        | 2064.00 $\pm$ 63.33a              |
| <i>V. oblongifolia</i>                        | TVnu 42       | 5.40 $\pm$ 0.30b    | 7.80 $\pm$ 0.36a        | 455.40 $\pm$ 25.30b               |
| <i>V. unguiculata</i> ssp. <i>dekindtiana</i> | TVnu 863      | 7.80 $\pm$ 0.38b    | 5.20 $\pm$ 0.30b        | 119.90 $\pm$ 4.64c                |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | IT84S-2246    | 1.80 $\pm$ 0.17b    | 0.00d                   | 0.00c                             |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | IT91K-180     | 21.60 $\pm$ 1.80a   | 0.00d                   | 0.00c                             |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | TVu 13731     | 7.60 $\pm$ 0.30b    | 1.80 $\pm$ 0.17c        | 80.20 $\pm$ 2.33c                 |

Means followed by the same letter(s) in columns are not significantly different ( $P < 0.05$ ; SNK).

TABLE 2

Mean density ( $\text{mm}^2$ ) of glandular and non-glandular trichomes and length ( $\mu\text{m}$ ) of non-glandular trichomes of leaves of wild and cultivated *Vigna* species ( $\pm$  SE)

| Vigna species                                 | Accession No. | Density             |                         | Length of non-glandular trichomes |
|---|---------------|---------------------|-------------------------|-----------------------------------|
|   |               | Glandular trichomes | Non-glandular trichomes |                                   |
| <i>V. vexillata</i>                           | TVnu 72       | 6.60 $\pm$ 0.36d    | 2.40 $\pm$ 0.23bc       | 447.40 $\pm$ 37.56a               |
| <i>V. oblongifolia</i>                        | TVnu 42       | 21.00 $\pm$ 0.40a   | 5.60 $\pm$ 0.36a        | 398.80 $\pm$ 26.11ab              |
| <i>V. unguiculata</i> ssp. <i>dekindtiana</i> | TVnu 863      | 7.40 $\pm$ 0.23cd   | 3.80 $\pm$ 0.18b        | 295.50 $\pm$ 28.88bc              |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | IT84S-2246    | 9.40 $\pm$ 0.11c    | 1.40 $\pm$ 0.27c        | 89.30 $\pm$ 2.46d                 |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | IT91K-180     | 13.40 $\pm$ 0.54b   | 1.20 $\pm$ 0.17c        | 99.52 $\pm$ 13.01cd               |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | TVu 13731     | 15.00 $\pm$ 0.14b   | 5.20 $\pm$ 0.33a        | 77.80 $\pm$ 5.39d                 |

Means followed by the same letter(s) in columns are not significantly different ( $P < 0.05$ ; SNK).

well as simple and stepwise regressions, was used to investigate the relationship between the variables.

### Results and discussion

Significant variations ( $P < 0.05$ ) were found in the density of glandular and non-glandular trichomes, as well as the length of the non-glandular trichomes of stems, leaves, and pods of the *Vigna* species (Tables. 1, 2 and 3). Significantly ( $P < 0.05$ ) fewer eggs were laid on TVnu 42 and TVu 13731 under free choice and TVu 13731 under no choice

compared with other treatments (Table 4). The density of glandular and non-glandular trichomes on leaves had significant negative correlations with the number of eggs laid under free choice (Table 5) with glandular trichomes accounting for 73.78 per cent of viable eggs (Fig. 1).

Flower damage was highest on IT84S-2246, IT91K-180 and TVnu 863 (Fig. 2) while pod load was significantly ( $P < 0.007$ ) higher on TVnu 72 compared with IT84S-2246 and TVnu 863 (Fig. 3). Pod damage was significantly ( $P < 0.0001$ ) higher on TVnu 863, IT84S-2246 and IT91K-180 (which

TABLE 3

Mean density ( $\text{mm}^2$ ) of glandular and non-glandular trichomes and length ( $\mu\text{m}$ ) of non-glandular trichomes of pods of wild and cultivated *Vigna* species

| <i>Vigna</i> species                          | Accession No. | Density             |                         | Length of non-glandular trichomes |
|---|---------------|---------------------|-------------------------|-----------------------------------|
|   |               | Glandular trichomes | Non-glandular trichomes |                                   |
| <i>V. vexillata</i>                           | TVnu 72       | 8.60 $\pm$ 0.36a    | 5.00 $\pm$ 0.19a        | 1082.60 $\pm$ 29.98a              |
| <i>V. oblongifolia</i>                        | TVnu 42       | 7.00 $\pm$ 0.24ab   | 4.40 $\pm$ 0.18a        | 683.40 $\pm$ 31.37b               |
| <i>V. unguiculata</i> ssp. <i>dekindtiana</i> | TVnu 863      | 2.60 $\pm$ 0.11c    | 10.80 $\pm$ 0.11a       | 68.14 $\pm$ 0.98c                 |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | IT84S-2246    | 5.00 $\pm$ 0.31bc   | 9.00 $\pm$ 0.20a        | 80.78 $\pm$ 4.68c                 |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | IT91K-180     | 7.40 $\pm$ 0.18ab   | 9.00 $\pm$ 0.28a        | 71.38 $\pm$ 2.60c                 |
| <i>V. unguiculata</i> ssp. <i>unguiculata</i> | TVu 13731     | 7.60 $\pm$ 0.41ab   | 9.80 $\pm$ 0.36a        | 58.52 $\pm$ 1.77c                 |

Means followed by the same letter(s) in columns are not significantly different ( $P < 0.05$ ; SNK).

TABLE 4

Mean number of eggs laid by *M. vitrata* females on wild and cultivated *Vigna* species

| <i>Vigna</i> species | Free choice          | No choice            |
|----------------------|----------------------|----------------------|
| TVnu 72              | 434.00 $\pm$ 20.83 a | 420.00 $\pm$ 22.53a  |
| TVnu 42              | 35.75 $\pm$ 4.790b   | 283.00 $\pm$ 49.98ab |
| TVnu 863             | 492.70 $\pm$ 77.31a  | 838.00 $\pm$ 115.97a |
| TVu 13731            | 91.30 $\pm$ 29.73b   | 50.30 $\pm$ 5.97bc   |
| IT91K-180            | 374.30 $\pm$ 56.02a  | 366.70 $\pm$ 43.54a  |
| IT84S-2246           | 608.70 $\pm$ 75.10a  | 793.30 $\pm$ 21.32a  |

Means followed by the same letter(s) in rows are not significantly different ( $P < 0.05$ ; SNK).

TABLE 5

Correlation coefficients of the relationship between oviposition of *M. vitrata* and pubescence of wild and cultivated *Vigna* species

| Variable | Free choice | No choice |
|----------|-------------|-----------|
| DGTP     | -0.4787     | 0.1543    |
| DNGTP    | 0.3633      | -0.4921   |
| LNGTP    | -0.1700     | 0.5529    |
| DGTL     | -0.8590*    | -0.4589   |
| DNGTL    | -0.8157*    | -0.6018   |
| LNGTL    | -0.1586     | 0.3841    |
| DGTS     | -0.1175     | -0.3385   |
| DNGTS    | -0.5050     | 0.2585    |
| LNGTS    | -0.0601     | 0.0321    |

\* = Significant at  $P = 0.05$ .

DGTP = density of glandular trichomes on pods; DNGTP = density of non-glandular trichomes on pods; LNGTP = length of non-glandular trichomes on pods; DGTL = density of glandular trichomes on leaves; DNGTL = density of non-glandular trichomes on leaves; LNGTL = length of non-glandular trichomes on leaves; DGTS = density of glandular trichomes on stem; DNGTS = density of non-glandular trichomes on stem and LNGTS = length of non-glandular trichomes on stem.

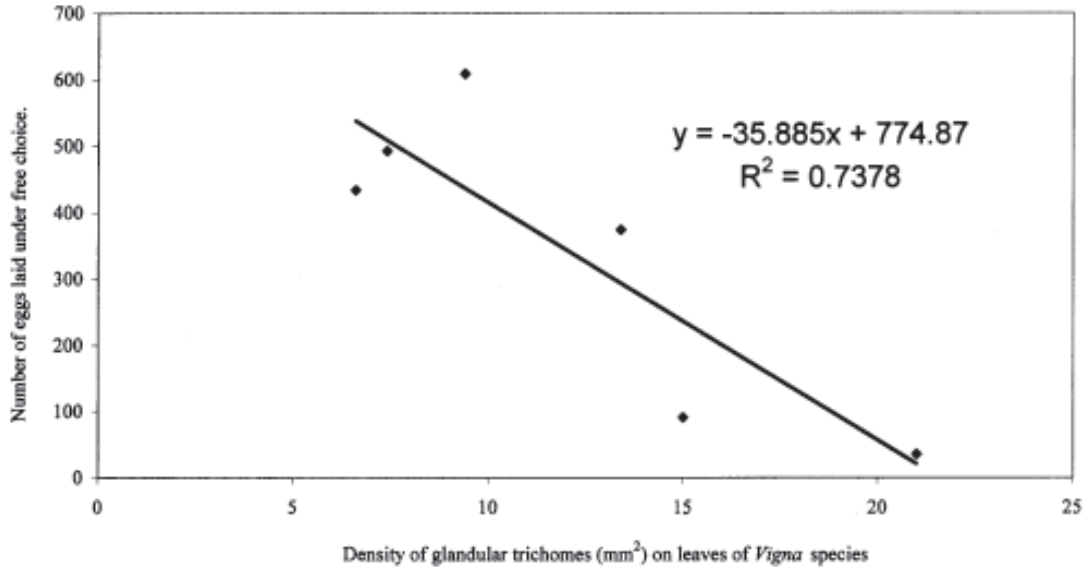


Fig. 1. Regression of number of eggs laid by *M. vitrata* females against density of glandular trichomes on leaves of wild and cultivated *Vigna* species

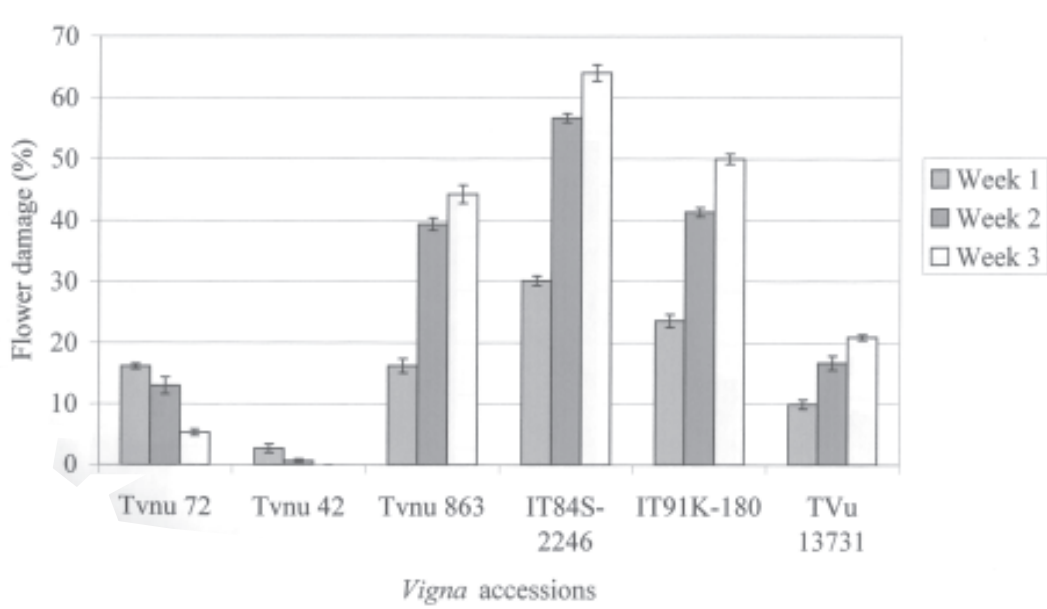


Fig. 2. Mean flower damage of wild and cultivated *Vigna* species by *M. vitrata*

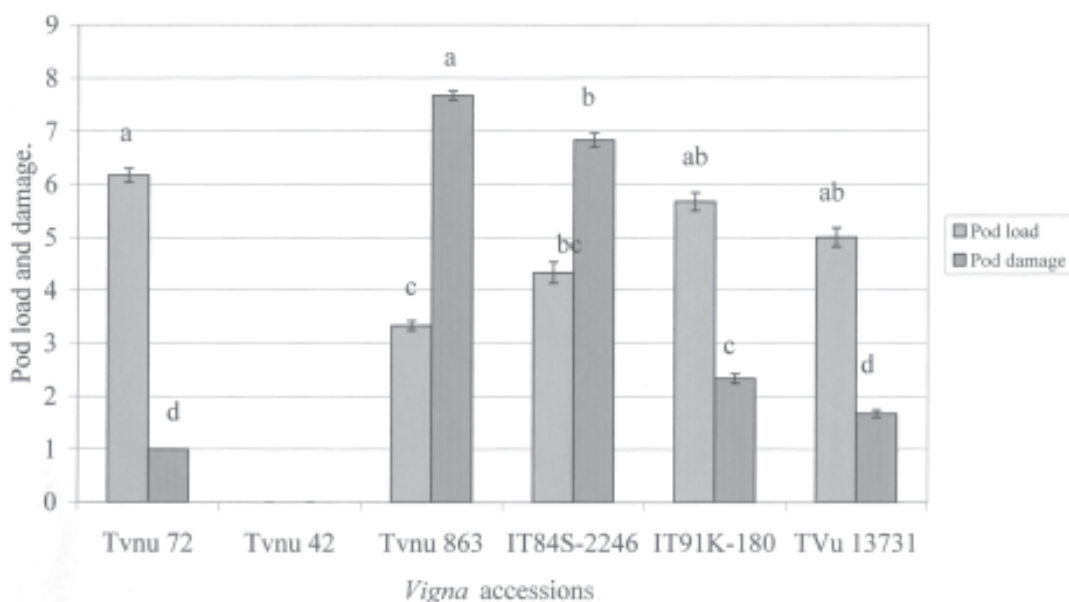


Fig. 3. Mean pod load and damage by *M. vitrata* of wild and cultivated *Vigna* species

TABLE 6

Stepwise regression procedures for the dependent variable mean pod load of wild and cultivated *Vigna* species under field infestation by *M. vitrata*

| Variable | Model R-squared | F-value | Probability of F |
|----------|-----------------|---------|------------------|
| FDWK1    | 0.0120          | 0.01    | 0.9302           |
| FDWK2    | 0.9168          | 11.01   | 0.1863           |
| FDWK3    | 0.9998          | 4580.12 | 0.0094**         |
| DGTS     | 0.6821          | 2.15    | 0.3814           |
| DNGTS    | 0.6345          | 1.74    | 0.4133           |
| LNGTS    | 0.6411          | 1.79    | 0.4090           |
| DGTL     | 0.0002          | 0.00    | 0.9905           |
| DNGTL    | 0.1680          | 0.20    | 0.7312           |
| LNGTL    | 0.0989          | 0.11    | 0.7964           |
| DGTP     | 0.9308          | 13.46   | 0.1694           |
| DNGTP    | 0.8006          | 4.01    | 0.2947           |
| LNGTP    | 0.6500          | 1.86    | 0.4030           |

\*\* = Significant at  $P = 0.01$

FDWK1 – 3 = Flower damage at weeks 1 – 3 after the initiation of flowering; DGTS = Density of glandular trichomes on stem; DNGTS = Density of non-glandular trichomes on stem; LNGTS = Length of non-glandular trichomes on stem; DGTL = Density of glandular trichomes on leaves; DNGTL = Density of non-glandular trichomes on leaves; LNGTL = Length of non-glandular trichomes on leaves; DGTP = Density of glandular trichomes on pods; DNGTP = Density of non-glandular trichomes on pods; LNGTP = Length of non-glandular trichomes on pods.

TABLE 7

Stepwise regression procedures for the dependent variable mean pod damage of wild and cultivated *Vigna* species under field infestation by *M. vitrata*

| Variable | Model R-squared | F-value | Probability of F |
|----------|-----------------|---------|------------------|
| FDWK1    | 0.1818          | 0.22    | 0.7195           |
| FDWK2    | 0.9985          | 681.55  | 0.0244*          |
| FDWK3    | 0.9033          | 9.34    | 0.2013           |
| DGTS     | 0.3574          | 0.56    | 0.5921           |
| DNGTS    | 0.9021          | 9.21    | 0.2026           |
| LNGTS    | 0.3164          | 0.46    | 0.6197           |
| DGTL     | 0.1149          | 0.13    | 0.7798           |
| DNGTL    | 0.0083          | 0.01    | 0.9419           |
| LNGTL    | 0.0001          | 0.00    | 0.9929           |
| DGTP     | 0.9958          | 237.19  | 0.0413*-         |
| DNGTP    | 0.4915          | 0.97    | 0.5054           |
| LNGTP    | 0.3251          | 0.48    | 0.6137           |

\* = Significant at  $P = 0.05$

FDWK1 – 3 = Flower damage at weeks 1 – 3 after the initiation of flowering; DGTS = Density of glandular trichomes on stem; DNGTS = Density of non-glandular trichomes on stem; LNGTS = Length of non-glandular trichomes on stem; DGTL = Density of glandular trichomes on leaves; DNGTL = Density of non-glandular trichomes on leaves; LNGTL = Length of non-glandular trichomes on leaves; DGTP = Density of glandular trichomes on pods; DNGTP = Density of non-glandular trichomes on pods; LNGTP = Length of non-glandular trichomes on pods.

TABLE 8

Stepwise regression procedures for the dependent variable mean flower damage at week 3 of wild and cultivated *Vigna* species under field infestation by *M. vitrata*

| Variable | Model R-squared | F-value | Probability of F |
|----------|-----------------|---------|------------------|
| DGTS     | 0.4425          | 1.59    | 0.3348           |
| DNGTS    | 0.0237          | 0.05    | 0.8461           |
| LNGTS    | 0.2870          | 0.80    | 0.4643           |
| DGTL     | 0.2553          | 0.69    | 0.4947           |
| DNGTL    | 0.0132          | 0.03    | 0.8850           |
| LNGTL    | 0.2021          | 0.51    | 0.5504           |
| DGTP     | 0.7314          | 5.45    | 0.1448           |
| DNGTP    | 0.8626          | 12.56   | 0.0712           |
| LNGTP    | 0.6128          | 3.16    | 0.2172           |

DGTS = Density of glandular trichomes on stem; DNGTS = Density of non-glandular trichomes on stem; LNGTS = Length of non-glandular trichomes on stem; DGTL = Density of glandular trichomes on leaves; DNGTL = Density of non-glandular trichomes on leaves; LNGTL = Length of non-glandular trichomes on leaves; DGTP = Density of glandular trichomes on pods; DNGTP = Density of non-glandular trichomes on pods; LNGTP = Length of non-glandular trichomes on pods.

were also significantly different from each other) compared with TVnu 72 and TVu 13731 (Fig. 3). There was no data for TVnu 42 because the plants remained vegetative throughout the period,

perhaps, due to day-length sensitivity or photo-periodism. The pod load depended significantly ( $P < 0.05$ ) on the flower damage by *M. vitrata* during the third week of flowering accounting for



TABLE 9

Stepwise regression procedures for the dependent variable mean flower damage at week 2 of wild and cultivated *Vigna* species under field infestation by *M. vitrata*

| Variable | Model R-squared | F-value | Probability of F |
|----------|-----------------|---------|------------------|
| DGTS     | 0.2124          | 0.54    | 0.5392           |
| DNGTS    | 0.0501          | 0.11    | 0.7762           |
| LNGTS    | 0.0979          | 0.22    | 0.6871           |
| DGTL     | 0.4853          | 1.89    | 0.3034           |
| DNGTL    | 0.1257          | 0.29    | 0.6454           |
| LNGTL    | 0.0796          | 0.17    | 0.7179           |
| DGTP     | 0.6572          | 3.83    | 0.1893           |
| DNGTP    | 0.7134          | 4.98    | 0.1554           |
| LNGTP    | 0.3727          | 1.19    | 0.3895           |

DGTS = Density of glandular trichomes on stem; DNGTS = Density of non-glandular trichomes on stem; LNGTS = Length of non-glandular trichomes on stem; DGTL = Density of glandular trichomes on leaves; DNGTL = Density of non-glandular trichomes on leaves; LNGTL = Length of non-glandular trichomes on leaves; DGTP = Density of glandular trichomes on pods; DNGTP = Density of non-glandular trichomes on pods; LNGTP = Length of non-glandular trichomes on pods.

99.98 per cent of the variability (Table 6). Similarly, the pod damage was influenced by the flower damage during the second week of flowering and the density of the glandular trichomes on the pods accounting for almost 100 per cent of viable eggs (Table 7). Flower damage by *M. vitrata* larvae was not affected by the pubescence of the *Vigna* species (Tables 8 and 9).

The results of the oviposition experiment show clearly that pubescence can affect the ovipositional behaviour of gravid female *M. vitrata* on *Vigna* species. That density of the trichomes on leaves reduced oviposition disagrees with the report of Navasero & Ramaswamy (1991) on *Heliothis virescens* which indicated that trichome type did not influence oviposition, but agrees with that of Plourde, Goonewardene & Kwolek (1985) which suggested that leaf pubescence was an oviposition rather than a larval barrier. Similarly, the results agree with those of Malakar & Tingey (1991) which suggested that glandular trichomes were responsible for the reluctance of potato tuber moth, *Phthorimaea operculella* Zeller to oviposit on foliage of *Solanum berthaultii*. Haddad & Hicks (2000) showed that female *Papilio troilus* (L.) discriminated against pubescence *Sassafras*

*albidum* (Nuttall). It is possible that exudates from these glandular trichomes discouraged *M. vitrata* females from laying eggs on the leaves. Kellogg, Taylor & Krings (2002) noted that several authors have shown that secondary metabolites produced by glandular trichomes are effective in discouraging many insect pests due to their odour, taste, stickiness or toxicity. In most cases, such secretions inhibit feeding, but they can also prohibit oviposition or reduce larval development. Non-glandular trichomes can interfere with female insect movement and anchorage while laying eggs.

The field experiment shows that flower damage contributes significantly to pod damage and production. This is as a result of direct damage to flower buds and flowers as well as increased survival of the larvae, which migrate to the pods. The suitability of *Vigna* species for oviposition by female *M. vitrata* appears to be the major determinant of the flower damage as the latter was not affected by the pubescence of the plants. Barre *et al.* (2002) observed that although female insects discriminate between plant species, their choice was not linked with the development of their offspring. Thus, *M. vitrata* neonate larvae were able to move from the leaves or other oviposition

sites (after hatching) to the flowers. These findings are in agreement with previous reports by Rausher (1979) and Thompson (1988) which explained that the choice of an oviposition site was not related to the suitability of the host for larval development.

Although it was easy for the larvae to exploit the flowers as food resource, feeding became difficult with the appearance of pods 2 weeks into the flowering period on some of the treatments. The larvae, now at the second to fourth instar stages, were faced with some resistance from the glandular trichomes on the pods. Haddad & Hicks (2000) argued that pubescence appears to be effective in lowering larval survivorship in the first two instars since the increase in damage is exponential with larval development. *M. vitrata* larvae spend only little time on the surface of the pods as they feed on the developing seeds within the pod. Substances from the glandular trichomes may cause them to spend more time on the pod surface. The pods and flower buds of TVu 13731 were reported to show antibiosis to *M. vitrata* larvae by IITA (1995). It is possible to develop cowpea varieties with acceptable levels of resistance to *M. vitrata* if efforts are concentrated on the factors controlling the oviposition choice of the adult female and larval feeding.

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