

## Phytochemical Screening and Fourier Transform Infrared Spectroscopy (FT-IR) Analysis of *Vernonia amygdalina* Del. (Bitter leaf) Methanol Leaf Extract

\*<sup>a</sup>Bashir, R. A., <sup>a</sup>Mukhtar, Y., <sup>b</sup>Chimbekujwo, I. B., <sup>c</sup>Aisha, D. M., <sup>c</sup>Fatima, S. U. & <sup>c</sup>Salamatu, S. U.

<sup>a</sup>Department of Plant Biology, Faculty of Life Sciences, Bayero University, Kano, Nigeria.

<sup>b</sup>Department of Plant Biology, Faculty of Life Science, Modibbo Adama University of Technology, Yola, Nigeria.

<sup>c</sup>Chiroma Ahmad Academy, No. 39 Lamido Abba Road, Yola Town, Adamawa State.

\*Correspondence email: [rbashir.bashir@gmail.com](mailto:rbashir.bashir@gmail.com)

### Abstract

*Vernonia amygdalina* is a tropical African plant of the *Asteraceae* family and is occasionally cultivated for its medicinal uses, which include treatment of diarrhea, skin wounds, fever mastitis and worms infection. The aim of this study is to test for the phytochemical constituents and FT-IR analysis of the leaf extract of *V. amygdalina*. The result of qualitative phytochemical analysis revealed the presence of important metabolites such as saponin, alkaloid, tannin, phenol, flavonoid, glycoside and terpenoids. The present FT-IR results confirmed the presence of alkanes, alkenes, amines, carboxylic acids and alcohols on the basis of identification of functional groups of OH, C-H and C=O which signifies the presence of these compounds in the methanol extracts of *V. amygdalina*.

**Keywords:** *Vernonia amygdalina*, Phytochemicals, FT-IR Analysis, Methanol Leaf Extract

### INTRODUCTION

The search for natural products to cure diseases has received considerable attentions in which medicinal plants have been the most important source (Okwu, 2001). These plants are believed to be an essential source of new chemical substances with potential therapeutic effects (Kuhn and Winston, 2000) and due to the crucial role that plant-derived compounds have played in drug discovery and treatment of several diseases, the isolation of new bioactive compounds from medicinal plants based on ethnomedicinal data appears to be a very promising approach (Newman, 2008). Traditional medicine products have played and are still playing greater roles in the lives of the people across the world in the face of the global upsurge of drug resistance, toxicity, adverse effects and increasing costs of synthetic products (Mbi and Bilikha, 1998). Poly-herbal therapies have the synergistic and antagonistic pharmacological agents within themselves that work together in a dynamic way to produce therapeutic efficacy with minimum side effects (Tiwari and Rao, 2002; Alexandros, 2007; Ebong, 2008).

The *V. amygdalina* Del, commonly known as bitter leaf is a shrub of 2-5 m tall with petiolate green leaves of about 6mm diameter. It belongs to the family *Asteraceae* and it is the most widely cultivated species of the genus *Vernonia* which has about 1,000 species of shrubs (Muanya, 2013). It is vegetatively cultivated by stem cutting at an angle of 450° and popular in most of West Africa countries including Nigeria, Cameroon, Gabon and Congo Democratic Republic. The plant is frequently found in gardens (Schiffers, 2000).

True to its name, bitter leaf is bitter to taste but surprisingly delicious in meals (Abosi and Raseroka, 2003). Bitter leaf is called Omjunso in East Africa especially Tanzania, Onugbo in Igbo-Eastern Nigeria, Ewuro (Yoruba), and Shuwaka (Hausa). The branches are brittle and break off easily. It has grey or brown coloured bark; the bark has a rough texture and is flaked. The leaves are elliptical and up to 20cm long. The leaves are oblongate, lance-like in shape, veined are bear, soft hairs on the underside. The bark is green or brownish dark with rough surface. The leaves are green with characteristic odour and bitter taste. The flowers are white, small, tiny and clustered. The small fruits have slightly hairy small nuts inside. It can reach 23ft in height when fully grown (Mbinglo, 1998; Remison, 2005; Muanya, 2013). It is a multipurpose plant that has a lot of potential valuable uses yet to be harnessed by the rural population of Nigeria. The leaves are used for popular bitter leaf soup and all parts of the plant are pharmacologically useful (Ojiako and Nwanjo 2006). The roots and the leaves are used in ethnomedicine to treat fever, hiccups, kidney problems and stomach discomfort (Igwe, 2015).

Many authors describe the medicinal and biological activities of *V. amygdalina* (Ijeh and Ejike, 2011; Atangwho, 2010). The leaves of *V. amygdalina* are crushed to make a paste to be rubbed on the body of livestock for destroying ectoparasites (Asawalam, 2008; Alawa, 2006). It is one of the traditionally used antifertility plants in Ethiopia. Preliminary study also confirmed that the plant has antifertility effect (Asawalam, 2008). The plant is well known for its antidiabetic and antihypertensive properties. The common and documented medicinal uses include the treatment of schistosomiasis, amoebic dysentery, and gastrointestinal problems. It is also used in the treatment of headache, fever, malaria, venereal diseases, wounds, hepatitis, and diabetes (Oyugi, 2011; Ayoola, 2008). Plants are composed of secondary metabolites which are believed to be responsible for their various therapeutics and toxicological properties and can be detected through phytochemical screening. These active constituents are organic compounds which are not involved in the process of growth, development, reproduction of an organism, or other primary functions. Therefore in the absence of these compounds, a plant will not die immediately, but will result in long term impairment in other functions, such as defensive mechanisms or aesthetic appearance, or in some cases of impact. Where a primary metabolites are necessary for functioning of the plant. This study is aimed at screening the phytochemical constituents of this plant material and conduct FT-IR analysis in order to have better understanding of its chemical profile which is very crucial in drug development.

## MATERIALS AND METHODS

### *Collection and Identification of Plant Material*

Fresh sample of the leaves of *V. amygdalina* were collected from the field in July, 2019 from Adamawa state, Nigeria. The identity of the plant was confirmed by a botanist at the plant science Department of Modibbo Adama University of Technology, Yola.

### *Preparation of Plant Material*

The samples of the plant parts were thoroughly washed, chopped into pieces and air dried at room temperature of about 33°C for a period of two weeks before powdering. The dried leaves were coarsely pounded using pestle and mortar and stored in a tightly covered glass jars for further studies.

***Extraction of Plant Material***

Exactly 50g of the powdered material of the leaves was soaked in 500ml of methanol for 72 hours at room temperature. Extracts were first filtered through Whatman No. 1 filter paper. After filtration, the extract was concentrated to dryness and kept in a refrigerator.

***Phytochemical Screening***

Qualitative preliminary phytochemical analysis of all the solvent extracts was conducted in accordance with the standard procedure (Harborne, 1992) to detect the nature of phytoconstituents and their presence in plant parts. The tests for saponins, tannins, flavanoids, glycoside, alkaloids and phenols were carried out by standard qualitative methods (Trease, 2002; Sood and Kaur, 2012).

***Fourier Transform Infrared Spectrophotometer (FT-IR)***

Dried powder of the leaves extract of *V. amygdalina* was used for FT-IR analysis. 10 mg of the powder was encapsulated in 100 mg of KBr pellet, in order to prepare translucent sample discs. The powdered sample of the extracts was loaded in FT-IR spectroscope, with a Scan range from 400 to 4000  $\text{cm}^{-1}$  with a resolution of 4  $\text{cm}^{-1}$  (Kalsi, 2007).

**RESULTS AND DISCUSSION**

Result of phytochemical screening of methanol leaf extract of *V. amygdalina* shows the presence of the following constituents: Saponin, Alkaloid, Tannin, Phenol, Flavonoid, Glycoside and Terpenoid. The FT-IR analysis data interpretation are shown in table 1.

Table 1: FT-IR analysis data interpretation of the methanolic leaf extract of *V. amygdalina*.

Type of Chemical bond	Absorption Frequency ( $\text{cm}^{-1}$ )	Intensity	Remark an assignment
OH group (alcohol)	3436.69	S, B	OH stretching, H-bonded
CH Alkanes	2928.20	M	C-H stretching alkanes
CH Alkanes	2861.80	M	C-H stretching alkanes
Ester group	1743.75	S	C=O ester stretching
Aromatic C=C group	1644.48	S	C=C stretching
Methylene group	1462.26	M	C-H bending
OH group (alcohol)	1376.92	M	OH stretching
C-O Carboxylic Acid	1212.97	S	C-O ester stretching

Key: S=Strong, B=Broad, M=Medium.

***FT-IR Spectral Analysis***

The bitter leaf spectra show the identity of O-H stretching of intermolecular bonded alcohol at 3436  $\text{cm}^{-1}$ . The double peak at 2928  $\text{cm}^{-1}$  and 2861  $\text{cm}^{-1}$  indicate the presence of C-H stretching of ester takes place at 1743  $\text{cm}^{-1}$ . The peak at 1644  $\text{cm}^{-1}$  elucidate the occurrence of C=C stretching. The fingerprint region of the spectra accounts for the presence of methylene C-H bending, O-H bending of alcohol, and C-O stretching at 1462  $\text{cm}^{-1}$ , 1376  $\text{cm}^{-1}$  and 1212  $\text{cm}^{-1}$  respectively.

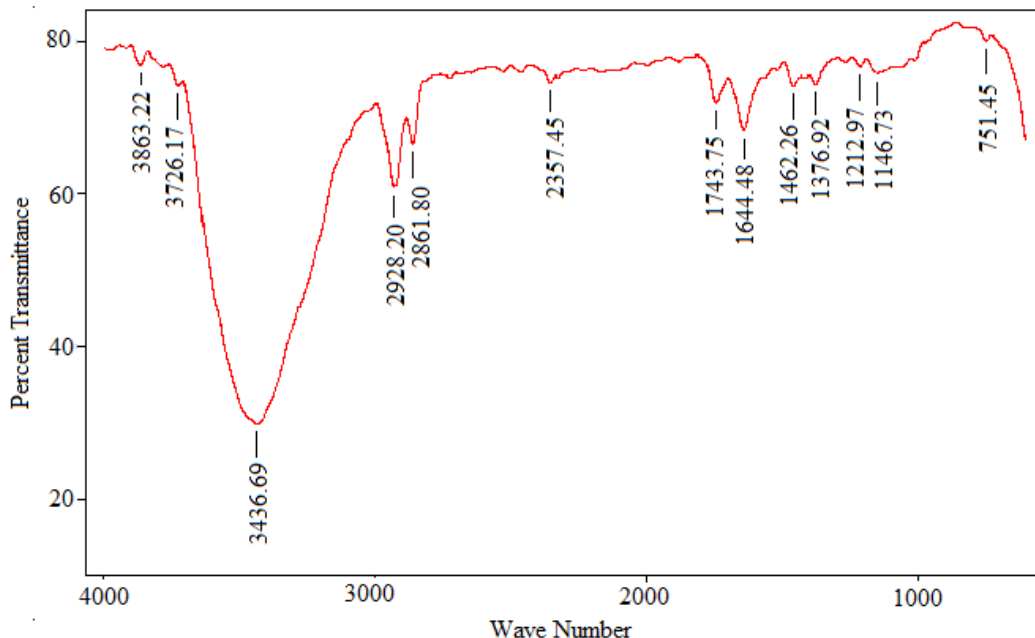


Figure 1: FT-IR spectrum of methanolic leaf extract.

The therapeutic effect of medicinal plants is attributed to the possession of important chemical constituents known as phytochemical (Nyamai and Arika, 2016). The outcome of this study revealed the presence of saponin, alkaloid, tannin, phenol, flavonoid, glycoside and terpenoids which is in agreements with the work of Aguru (2017), Maku (2007) and Wakirwa (2013). These observations therefore support the use of *V. amygdalina* in herbal care remedies. A wide array of phytochemical, including anti-nutritional factors, has been shown to be present in *V. amygdalina* (Awe, 1999; Oboh, 2009; Yeap, 2010).

Alkaloids are synthesized form of amino acids. They have multiplicity of host-mediated biological activities, including antimicrobial, cytotoxic, analgesic and antipyretic activities (Tylor, 2011; Semwal *et al.*, 2011; Hanita, *et al.*, 2013). Alkaloids have a wide range of pharmacological activities including antimalarial (e.g., quinine), anticancer (e.g., homoharringtonine), antibacterial (e.g., chelerythrine) and antihyperglycemic activities (e.g., piperine) (Namadina, *et al.*, 2019). Flavonoids are phenolic compounds and possessed the properties of antibacterial, antioxidant, anticancer, anti-inflammatory, antipyretic and analgesic activities (Ali *et al.*, 2013; Magdalena *et al.*, 2013; Emad, 2013; Elzbieta *et al.*, 2013). The biological function of flavonoid includes protection against allergies, inflammation, free radicals, platelet aggregation, microbes, ulcers, hepatotoxins and tumors (Okwu, 2004). Saponins are glycosidic triterpenoids and they are widely found in plants. They have the pharmacological properties like anti-inflammatory and antipyretic activities (Emmanuel *et al.*, 2012; Adiukwu *et al.*, 2013; Atangwho, *et al.*, 2010). The tannin-containing plants extracts can be used as astringents, anti-diarrhea, diuretics, treatment of stomach and duodenal tumours and as anti-inflammatory, antiseptic, antioxidant and haemostatic pharmaceuticals (Mamta, 2013). Tannin is one of the major active ingredients found in plant based medicines (Haslam, 1996). They possess antiviral, antibacterial, and antitumor activity (Haslam, 1996; Khanbabaee and Van Ree, 2001). Tannin has been reported to selectively inhibit Human Immune Virus (HIV) replication (Kashiwada *et al.*, 1992). Phenols exhibit antimicrobial,

anthelmintic, antiapoptotic and antidiarrhoeal activities (Cowan, 1999). Terpenes have been reported to have antimicrobial, antioxidant, anticancer and Antiparasitic activities (Sankhadip *et al.*, 2010; Chonthicha *et al.*, 2013; Oboh and Enobhayisobo, 2009; Ifedaya *et al.*, 2013; Ayoola *et al.*, 2008).

Fourier Transform Infrared Spectrophotometer (FT-IR) is perhaps the most powerful tool for identifying the types of chemical bonds/functional groups present in the phytochemicals. The wavelength of light absorbed is salient feature of the chemical bond as can be seen in the annotated spectrum. By interpreting the infrared absorption spectrum, the chemical bonds in a compound can be determined (Geethu, *et al.*, 2014). The FT-IR spectrum showed in figure-1 of the methanolic leaf extract of *V. amygdalina* has bands and wave numbers of  $3436.69\text{ cm}^{-1}$  to  $2861.80\text{ cm}^{-1}$  as the prominent peaks. The peak at a frequency of  $3436.69\text{ cm}^{-1}$  to  $1146.73\text{ cm}^{-1}$  were strong, broad and medium. The present FT-IR results confirmed the presence of alkanes, alkenes, amines, carboxylic acids and alcohols in the methanol extracts of *V. amygdalina* and is also in agreement with previous studies conducted on other plants (Muruganantham, 2009; Meenambal, 2012; Vijayalakshmi, 2012; Janakiraman, 2011). Many works conducted revealed that the FT-IR spectrum is an effective tool for differentiating, classifying and discriminating closely related plants and other organisms.

## CONCLUSION

The leaves of *Vernonia amygdalina* is well known for its ethnomedicinal properties. The common and documented medicinal uses include the treatment of schistosomiasis, amoebic dysentery, and gastrointestinal problems. From the present study, the leaf extracts showed an abundant production of phytochemicals as secondary metabolite. Through FT-IR analysis, it was found out that the methanol extracts of the plant material contained 8 peaks based on the differences of polarity. The identification of functional groups of OH, C-H and C=O confirmed the presence of alcohol, alkanes and ester compounds in this plant specimen. Further extensive study of chemical composition of this herbal species may pave way for identification and isolation of significant bioactive compounds with pronounced antioxidant and other therapeutic activities for drug development.

## References

- Aguoru, C. U., Bashayi, C. G and Ogbonna, I. O. (2017). Phytochemical profile of stem bark extracts of *K. senegalensis* by Gas Chromatography-Mass Spectrometry (GC-MS) analysis. *Journal of Pharmacognosy and Phytotherapy*, 9(3): 35-43.
- Alawa, C. B. I. *et al.*, (2006). Nutritive value and haemolytic properties (in vitro) of the leaves of *Vernonia amygdalina* on human erythrocyte. *Journal of Nutrition and health*, 18(2): 151-160.
- Asawalam, E. F., Emosairue, S. O., and Hassanali, A. (2008). Contribution of different constituents to the toxicity of the essential oil constituents of *Vernonia amygdalina* (Compositae) and *Xylopii aetiopica* (Annonaceae) on maize weevil *Sitophilus Zea mays* Motschulsky (Coleoptera: Curculionidae). *African Journal of Biotechnology*, 7(16).
- Atangwho, I. J., Ebong, P. E., Eyong, E. U., and Eteng, M. U. (2010). Combined administration of extracts of *Vernonia amygdalina* (Del) and *Azadirachta indica* (A. Juss) mimic insulin in time-course body weight and glucose regulation in diabetic and non-diabetic rats. *Nigerian Journal of Biochemistry and Molecular Biology*, 25(1): 44-49.
- Awe, S. O., Makinde, J. M., and Olajide, O. A. (1999). Cathartic effect of the leaf extract of

- Vernonia amygdalina*. *Fitoterapia*, 70(2): 161-165.
- Ayoola, G. A. *et al.*, (2008). Phytochemical screening and antioxidant activities of some selected medicinal plants used for malaria therapy in Southwestern Nigeria. *Tropical Journal of Pharmaceutical Research*, 7(3): 1019-1024.
- Cowan, M. M. (1999). Plant products as antimicrobial agents. *Journal of Clinical microbiology reviews*, 12(4): 564-582.
- Geethu, M. G., Suchithra, P. S., Kavitha, C. H., Aswathy, J. M., Dinesh, B., and Murugan, K. (2014). Fourier-transform infrared spectroscopy analysis of Different solvent extracts of water hyacinth (*Eichhornia crassipes* mart solms.) An allelopathic Approach. *World Journal of pharmacy and pharmaceutical sciences*. Volume 3, issue 6, 1256-1266. Research article ISSN 2278 – 4357. SJIF impact factor 2.786.
- Harborne, J. B. (1992). Phytochemical methods. Chapman and Hall publications, London. pp.7-8.
- Haslam, E. (1996). Natural polyphenols (vegetable tannins) as drugs: possible modes of action. *Journal of Natural Products*, 59(2): 205–215.
- Igwe, K. K., Okafor, P. N., Ijeh, I. I., and Anika, S. M. (2015). Effect of *Vernonia amygdalina* Del ethanolic extract on serum prolactin in lactating and non-lactating female albino wistar rats. *Research Journal of Chemical Sciences*, Vol. 5(11): 40-45.
- Ijeh, I. I., and Ejike, C. E. (2011). Current perspectives on the medicinal potentials of *Vernonia amygdalina* Del. *Journal of Medicinal Plants Research*, 5(7): 1051-1061.
- Janakiraman, N., Satish, S. S., Johnson, M. (2011). UV-VIS and FT-IR spectroscopic studies on *Peristrophe bicalyculata* (Retz.) Nees. *Asian Journal of Pharmaceutical and Clinical Research*, 4(4): 125-129.
- Kalsi P. S. (2007). Spectroscopy of organic compounds. Edn 6, New Age International Publishers, New Delhi.
- Kashiwada, Y., Huang, L., Kilkuskie, R. E., Bodner, A. J and Lee, K. H. (1992). New Hexahydroxydiphenyl Derivatives as Potent Inhibitors of HIV replication in H9 lymphocytes. *Bioorganic and Medicinal Chemistry Letters*, 2(3): 235–238.
- Khanbabae, K. and Van Ree, T. (2001). Tannins Classification and Definition. *Natural Product Reports*, 18(6): 641–649.
- Lakshmi, S. P., and Bindu, R. N. (2014). Functional group analysis of *Cleome viscosa* L. and *C. burmanni* W. & A. (Cleomaceae) extracts by FT-IR. *Journal of Pharmacognosy and Phytochemistry*, 2(6): 120-124.
- Makut, M. D., Gyar S. D., Pennap, G. R. I. and Anthony, P. (2007). Phytochemical screening and antimicrobial activity of the ethanolic and methanolic extracts of the leaf and bark of *Khaya senegalensis*. *African Journal of Biotechnology*, 7(9): 1216-1219.
- Mamta, S., Jyoti, S., Rajeev N., Dharmendra, S., and Abhishek, G. (2013). Phytochemistry of Medicinal Plants. *Journal of Pharmacognosy and Phytochemistry*, 1(6): 168-182.
- Meenambal, M., Phugalendy, K., Vasantharaja, C., Prapakaran, S., Vijayan, P. (2012). Phytochemical investigation from FTIR and GCMS studies of methanol extract of *Delonix elata* leaves. *International Journal of Chemical and Analytical Science*, 3(6): 1446-1448.
- Muruganantham, S., Anbalagan, G., Ramamurthy, N. (2009). FTIR and SEM-EDS comparative analysis of medicinal plants, *Eclipta alba* Hassk and *Eclipta prostrata* Linn. *Romanian Journal of Bio physiology*, 19(4): 285-294.
- Namadina, M. M., Nuhu, A., Aliyu, B. S., and Sani, A. (2019). Physicochemical screening and acute Toxicity Study of *Anogeissus leiocarpa* Stem Bark. *Dutse Journal of Pure and Applied Sciences* (DUJOPAS), Vol. 5 No. 1b.

- Nyamai, D. W., Arika W., Ogola, P. E., Njagi, E. N. N, and Ngugi, M. P. (2016). Medicinally important phytochemicals: An updated research avenue. Research and Reviews: *Journal of Pharmacognosy*, 4(1): 35-49.
- Oboh, F. O. J., and Enobhayisobo, E. I. (2009). Effect of aqueous extract of *Vernonia amygdalina* leaves on plasma lipids of hyperlipidaemic adult male albino New Zealand rabbits, 10 (4).
- Ojiako, O. A. and Nwanjo, H. U. (2006). Is *Vernonia* hepatotoxic or hepatoprotective? Response from biochemical and toxicity studies in rat. *African Journal of Biotechnology*, 5(18): 1648-1651.
- Okwu, D. E. (2004). Phytochemicals and vitamins content of indigenous spices of South Eastern Nigeria. *Journal of Sustainable Agricultural Environment*, 6: 30-34.
- Oyugi, D. A. *et al.*, (2011). Biological activity and mass spectrometric analysis of *Vernonia amygdalina* fractions. *Journal of Biological science Technique*, 2(3): 287-304.
- Robards, K., Prenzler, P. D., Tucker, G., Swatsitang, P., Glover, W. (1990). Phenolic compounds and their role in oxidative processes in fruits. *Journal of Food Chemistry*, 66: 401-36.
- Vijayalakshmi, R., Ravindhran, R., (2012). Comparative fingerprint and extraction yield of *Diospyrus ferrea* (Wild.) Bakh. Root with phenol compounds (garlic acid), as determined by UV-Vis and FT-IR spectroscopy. *Asian Pacific Journal of Tropical Biomedicine*, S1367-S1371.
- Wakirwa, J. H., Idris S., Madu S. J., Dibal, M., and Malgwi, T. (2013). Assessment of the In-vitro antimicrobial potential of *Khaya senegalensis* ethanol leaf extract. *Journal of Chemical Pharmacognosy Resources*, 5(7): 182-186.
- Yeap, S. K. *et al.*, (2010). *Vernonia amygdalina*, an ethnoveterinary and ethno medical used green vegetable with multiple bioactivities. *Journal of Medicinal Plants Resources*, 4: 2787-2812.



© 2020 by the authors. License FUTY Journal of the Environment, Yola, Nigeria. This article is an open access distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).