Pre-and post-vaccine measles antibody status in infants using serum and oral-fluid testing: an evaluation of routine immunization in Addis Ababa, Ethiopia

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Abstract

Background: Despite the use of measles vaccine, measles incidence in Ethiopia remains a serious public health concern. Progress towards the control of measles requires a national capacity to measure programme effectiveness. This includes evaluation of vaccine effectiveness in infants attending the routine immunization.

Objective: To evaluate the effectiveness of the measles routine immunization activities in Addis Ababa.

Methods: This study evaluated pre- and post-vaccine antibodies in children attending for routine measles immunization in Addis Ababa. Infants who presented to 3 health centres between September-November, 1998 for routine measles vaccination were enrolled in the study. In total 296 infants (median age 9 months) provided blood and oral-fluid samples, of which 230 (77%) returned to provide post vaccine samples (median interval of 15 days). Screening of sera was undertaken using commercial indirect ELISA kits, and of oral fluids using an in-house IgM-capture ELISA.

Results: Pre-vaccination serology showed 1.4% IgM positive, 2.0% IgG positive, and 97.0% seronegative. Post-vaccination seroprevalence of IgM and IgG was 91.3% and 85.0%, respectively, and 92.9% overall. The seroconversion rate was 92.6% (95%CI 88.2-95.7). Based on oral fluid results, 87.3% (95% CI 82.0-91.4) of children showed specific IgM antibody conversion.

Conclusion: These results are in support of the recommended age for measles vaccination in Addis Ababa, and show the merit of oral-fluid IgM screening as a non-invasive alternative to blood for assessing vaccine effectiveness. [Ethiop.J.Health Dev. 2003;17(3):149-155]

Introduction

Worldwide, it is estimated that measles kills some 880,000 children annually, a toll more than any other vaccine-preventable disease. The global plan, established by the World Health Organization (WHO) and United Nations

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Children's Fund (UNICEF), is to cut this burden by two-thirds between 2000 and 2005, and thereafter to prevent 600,000 measles fatalities annually (1). Half of the total deaths are concentrated in three African countries (Congo, Ethiopia and Nigeria) and one Asian country (India). Progress towards the control of measles requires that countries develop national capacity to measure programme effectiveness by which to assess and refine immunization policy. This includes evaluation of vaccine effectiveness in infants attending routine immunization clinics.

The Expanded Programme on Immunization (EPI) in Ethiopia was launched in 1980 (2,3), and offers a single dose of measles vaccine at 9

months of age. In 1999 the national measles coverage was 53% with uptake ranging from 7-88% (the highest 88% recorded for Tigray region the lowest 7% recorded for Somali region) in the different administrative regions (Source: Department of Family Health, MOH, 2000).

Research has demonstrated oral fluid to yield detectable levels of immunoglobulins (IgG and IgM antibodies) against a wide variety of infections (4-14) using sensitive and specific antibody capture assays. The detection of measles specific IgM antibodies in oral fluid by antibody capture ELISA (MACELISA) has previously been reported in a study of children who received measles vaccine (14). The position of non-invasive antibody testing for measles grows stronger as emphasis on vaccine programme surveillance increases. accompanied by technical developments in oral-fluid assays (13, 14) and favourable evaluations under a variety of settings (14-17).

The aim of this study is to evaluate pre- and post-vaccine antibody in children attending for routine measles immunization in Addis Ababa. Furthermore, we aim to evaluate the use of oral-fluid testing for measles specific IgM as an alternative to serum assays in estimating vaccine response and effectiveness. For this an MACELISA enhanced measles that incorporates amplification an stage is developed.

Methods

Study population: Addis Ababa, the capital of Ethiopia, a population of 2,570,004 (density of 4.847.8/km²) (18) settled at 2000-2800m altitude above sea level. Administratively there are 6 'Zones' (each divided 4-7 Weredas), and 28 'Weredas' (=district) each with a population of 45,277-153,688. In 1998/99 measles vaccination was given in 7 hospitals, 17 health centres, 9 health clinics, 20 health posts, 104 out reach sites under the Region 14 Health Bureau. The estimated measles vaccination coverage in 1998/99 (1991 E.C.-Ethiopian Julian Calendar) was 86% with variation in the

different 'Zones' and' 'Weredas' (63-100%). Measles vaccine consumption for the year was 10,250 vials given to 32,548 children aged under one year (Source: Addis Ababa City Government Health Bureau).

Three of 19 Government health centres were selected for the study: one from the centre of the city (Arada), one in the outer city (Akaki), and the third from an area between the centre and outer city (Wereda 25). All infants who presented to these health centres between September-November, 1998 for routine measles vaccination were eligible for enrolment in the the children study. All had received vaccinations of BCG and OPV at birth, and 3 doses of DPT and OPV according to the recommended schedule of the EPI Global . Advisory Group (6, 10 and 14 weeks)(19). Informed consent was obtained from guardians by explaining title of the research project, basic information about the disease and objective of the study, information about coordinators of the project and information about how the work was carried out. Permission for this work was obtained from Addis Ababa City Government Health Bureau (No.1370/172) in 29th August 1998.

Sample collection: Blood samples were collected by finger prick using Safety Flow Lancet into Microtainers (Becton Dickinson. Oxford, England). Oral-fluid specimens were collected and processed as described previously (11) by sponge swab collection devices (Oracol: Malvern Medical Developments, Worcester, England) from infants before they received live attenuated measles vaccine strain. SmithKline (Schwartz Beecham Biologicals, Rixensart, Belgium). Mothers were requested to return with their infants two weeks after vaccination for a second oral-fluid and blood sample as recent study demonstrates that the presence of considerable amounts of haemagglutinin-specific antibodies in the serum within the early days (5-10 days) after the onset of rash, which are readily detectable with an ELISA (20). Samples collected were processed at the virology laboratory of the Ethiopian Health and Nutrition Research Institute.

(EHNRI). Specimens were transported on dry ice to the UK for laboratory analysis.

measles IgM/IgG determination: Serum Serum samples were screened for measles virus (MV) specific IgM and IgG by using a commercial ELISA kit (Enzygnost for IgM, and IgG; Behring Diagnostic, Marburg, Germany). In the IgG ELISA optical density (OD) readings of <0.100, >0.200 and between 0.100-0.200 obtained from 1:231 IgG serum dilution considered as negative, positive and equivocal respectively. The limit of the IgG detection of the test is 150 mIU/ml, equivalent to an OD of 0.100. Similarly, in the IgM ELISA OD readings of <0.100, >0.200 and between 0.100-0.200 obtained from 1: 42 IgM dilution were considered as negative, positive and equivocal respectively. Serum screening was undertaken at EHNRI.

Oral-fluid MV IgM detection: MV specific IgM in oral fluid was determined by MACELISA, which was a modification of the .FITC/anti-FITC IgG ' capture ELISA (GACELISA) described previously (13). A description of these modifications follows. Wells of microtitre plates (Maxisorb "U" wells, Life Technologies, Paisley, UK) were coated with 100µl of a 11.2 g/L solution of rabbit antihuman IgM serum (Dako, Ely, UK) diluted 1:3000 (as this dilution factor was found optimal during developmental (optimisation) stage of the assay) in 0.05Mcarbonate/bicarbonate buffer, pH 9.6. Plates were then incubated with 100 ul undiluted oral fluid samples after blocking with 200 µl of 5% Sol-u-pro (Dynagel Inc., Calumet City, IL). After addition of measles antigen, anti-measles haemagglutinin monoclonal antibody-FITC conjugate (Chemicon Inc., Temicula, CA., USA) diluted 1:3000 (instead of 1:4000 for GACELISA). Anti-FITC horseradish peroxidase conjugate and TMB were then added and the reaction stopped by adding 100 ul of 0.5M HCl. Oral-fluid MV IgM detection was carried out at Central Public Health Laboratory (CPHL), UK.

OD results from the MACELISA were expressed as T/N ratios (test sample OD /negative control OD) in order to have results fall, from negative to strong positive, represents the "dynamic range" of an assay. The receiver operating characteristics (ROC) curve was generated (21) to determine the appropriate cutoff T/N value (set to the maximum sensitivity and specificity relative to serum results). For assays designed for determining immunity in the population, a more appropriate and accurate cut-off can be set using a statistical techniques such as ROC. Relative to serum IgM results (excluding equivocal results), the MACELISA was 93.6% (190/203) sensitive and 93.4% (284/304) specific, using a T/N ratio of 1.14. T/N ratios less than the cutoff value (1.14) were considered negative in the analyses.

Statistical analysis: Analysis was conducted using STATA V7.0 (Stata Corp, College Station, Texas, USA). Comparison between median ages uses the non-parametric Kruskall-Wallace test (with ties). Estimates of the proportions serpositive for either IgM or IgG excludes samples with equivocal results. Overall proportions based on serum specific IgM or IgG results exclude only samples equivocal by both tests. Estimates of seroconversion are derived from the proportion of individuals specific-antibody negative prevaccine whose status changes to specificantibody positive post-vaccination. Proportions seropositive are compared using Fisher's Exact test. Exact binomial confidence intervals are calculated for prevalence estimates.

Results

A total of 296 infants were recruited with median age 9 months (m, range 5-40 m) all providing a blood and an oral-fluid sample. The number and median age of individuals attending Akaki, Arada and Wareda 25 clinics were, 86, 9m, 41, 9m and 169, 10m, respectively (median ages did not differ between clinics: $\chi^2_{(2)}$ 5:235, P=0.073). Of the 296 recruits, 230 (77%) returned to provide post-vaccine samples (median interval of 15 days, range 14-46 days) oral-fluid and blood.

Results of the measles-specific IgG and IgM tests on serum samples from pre- and post-vaccinated children are shown in Table 1. Seroprevalence estimates were made excluding

equivocal results. Pre-vaccination, 1.4% of children were IgM seropositive and 2.0% IgG positive. Post-vaccination seroprevalence of IgM and IgG was 91.3% and 85.0%.

Table 1: Seroprevalence of measles antibody status (IgM and IgG) in the pre- and post-vaccinated children in Addis Ababa, Ethiopia 1998.

| / | IgM | | | lgG |
|-------------------------|-----------|-------------|-----------|------------|
| Results | Pre-vacc. | ·Post-vacc. | Pre-vacc. | Post-vacc. |
| Positive | 4 | 199 | 6 | 170 |
| Negative | 285 | 19 | 290 | 30 |
| Equivocal | 6 | 9 | 0 | 28 |
| NTa | 1 | 3 | 0 | 2 |
| Total | 296 | 230 | 296 | 230 |
| Proportion ^b | 1.38 % | 91.3 % | 2.03 % | 85.0% |

a NT = Sample collected but not tested because of insufficient serum

Of the 4 pre-vaccination IgM positive samples, 3 were IgG negative (2 aged 9m and 1 age 10m) and one high titre IgG positive (age 9m). All the 4 samples were repeated by Behring serum IgM ELISA and found to remain positive. Of 6 pre-vaccination IgM equivocal results all were IgG negative. Of 6 pre-vaccine sera that were measles-specific IgG positive, 5 had high titres (>900mIU/ml), three of which were aged 9m, one 10m and one 11m. One of the 5 was IgM positive, and one (age 10m) had low level antibody (296mIU/ml) for which the paired post-vaccine sample showed a greater than fourfold rise in titre (ie suggesting this individual had residual maternal antibody).

Of 19 children specific-IgM negative post-vaccination, in pre-vaccination samples 5 were IgG positive and one other was IgM positive. Of 9 post-vaccination IgM equivocal samples, all were IgM and IgG negative pre-vaccine and, post-vaccine, 2 were IgG equivocal and 2 were IgG positive. The time between the pre- and post-sample collection for the 2 IgG positive specimens was 15 and 9 days. Post-vaccine,

there were 30 samples negative and 28 equivocal for measles specific-IgG; results which should be interpreted in relation to the short interval (median of 15 days) between vaccination and second sample collection. Of the 30 samples IgG negative post-vaccination, pre-vaccination, none were IgG positive, 1 was IgM positive and 2 had equivocal IgM status.

Overall prevalence and antibody conversion rate for the sera (based on IgG and IgM results) and oral fluid are presented in Table 2. Exact binomial 95% confidence intervals (CI) are shown. There was no significant difference between the three clinics in seroprevalence prevaccination (Fisher's Exact P=0.066) or postvaccination (P=0.752). Compared with serum estimates, oral-fluid prevalence was nearly 4% higher pre-vaccine, and 6% lower post-vaccine, although these differences were not significant (95% confidence intervals on prevalence estimates overlap). The overall antibody conversion rate for serum was 92.6% (200/216),which was higher (but not significantly higher) than for oral-fluid, 87.3% (185/212).

^b Proportion = Number Positive / Number (Positive + Negative)

Table 2: Overall antibody prevalence and conversion rate in vaccine recipients in Addis Ababa. Ethiopia 1998.

| Sample type | Pre-vaccine % (positive/total) 95% CI | Post-vaccine % (positive/total) 95% CI | Antibody conversion % (converters/total) 95% CI |
|-------------|---|---|---|
| Serum | 3.04% (9/296) | 92.9% (208/224) | 92.6% (200/216) |
| | 1.40%-5.69% | 88.7%-95.9% | 88.2%-95.7% |

86.9% (199/229)

81.8%-91.0%

Discussion

Oral fluid

Prestatus and post-vaccination childrenBI: Assessment of vaccine recipient children (median age 9 months) at three representative vaccine clinics in the city of Addis Ababa, showed an absence of measlesspecific IgM and IgG antibody in 99% and 98% of serum samples, respectively. Considering either antibody class, the data suggest that 97% (95% CI 94-98%) of the children attending for routine (9 months of age) measles vaccination would respond to vaccine. Following measles vaccination the seroconversion rate (based on either antibody class) was 93% (95% CI 88-96). Children attending for routine measles vaccination had a median age of 9m; only 5% were less than 9m and 10% were 1 year or above. Together these results indicate a highly successful routine immunization programme. The present seroconversion rate study confirms vaccinating children at age 9 months in the Ethiopia setting is in accordance to the WHO recommendation (22)previous and mathematical model studies made for developing countries (23-25).

6.76% (20/296)

4.18%-10.2%

A full assessment of whether or not the present age at immunization is optimal should consider the prevalence of residual maternal antibodies and proportion with evidence of recent infection in samples from infants attending for In this study, of 15 routine vaccination. samples that were positive or equivocal for serum measles-specific antibody prior to vaccination, only 1 appeared to indicate the presence of residual maternal antibody (low specific-IgG titre with subsequent four fold rise post-vaccination). In contrast there were 8 prevaccine serum samples specific-IgM positive specific-IgG titre positive, and/or high

indicative of recent infection (6 aged 9m, and 1 each aged 10 and 11m). These results suggest that at the routine target vaccination age a negligible proportion of infants have residual maternal antibody, but a small but significant proportion (8/296=2.7%) have been exposed to measles virus. This is, at least, an indication of the need to discourage any delay in bringing children for measles vaccination beyond the appointed age.

87.3% (185/212)

82.0%-91.4%

The present study was primarily designed to investigate the development of measles virus specific IgM antibodies with an average interval of 15 days between receiving vaccination and the collection of second samples. Consequently the results yield a high proportion of serologically negative and eguivocal IgG results post-vaccination, compared to IgM data, as IgM antibodies are produced initially followed by IgG. Taking IgM serological data alone yields a serconversion estimate of 90%, only marginally lower than that based on IgG and IgM results.

Serum/oral fluid comparison: The detection of oral fluid specific IgM for measles virus by antibody captures ELISA in pre- and postvaccinated infants has been previously described (14). The present work a new MACELISA test is described that employs a FITC/anti-FITC amplification system (26) to enhance the sensitivity of the oral fluid assay to a level comparable to that of the corresponding 'gold standard' serum testing using commercial kit. Using this amplification system, adequately sensitive and specific measles and rubella virus specific IgG oral fluid assay methods have been developed (12, 13). The IgM assay developed in this study

demonstrated 93.6% sensitivity and 93.4% specificity, which is comparable to the results of 91% sensitivity and 95% specificity found in a previous oral-fluid testing study (14). Based on oral-fluid samples screened using the MACELISA, an estimate of specific antibody conversion of 87% (82-91) was obtained. Although lower than the overall serconversion rate obtained by Behring ELISA, the result is not significantly lower. With such performance the oral-fluid testing method (MACELISA) would be of use for measuring measles antibody prevalence in pre- and post-vaccine infants.

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References

- 1. Brown P. A plan to reduce measles deaths. Nature Medicine 2000;6(12):1305.
- Ministry of Health (MOH). Guidelines for EPI in Ethiopia, Addis Ababa. 1981.
- 3. Ministry of Health (MOH). Annual Report of EPI in Ethiopia, Addis Ababa. 1981.
- Parry JV, Perry KR, Panday S, Mortimer PP. Diagnosis of hepatitis A and B by testing saliva. J Med Virol 1989;28:255-260.
- Parry JV. Simple and reliable salivary tests for HIV and hepatitis A and B virus diagnosis and surveillance. Ann NY Acad Sci 1993; 694: 216-233.

- Hunt AJ, CJ Christofinis J, Parry JV, Weatherburn P, Hickson GCI, Coxon APM, Davies PM, MacManus TJ, Sutherland S. The testing of saliva samples for HIV-1 antibodies-reliablity in a nonclinical setting. Genitourin Med 1993;69: 29-30.
- Perry KR, Brown DWG, Parry JV, Panday S, Pipkin C, Richards A. Detection of measles, mumps and rubella antibodies in saliva using capture radioimmunoassay. J Med Virol 1993;40:235-240.
- Rice PS and Cohen BJ. A school outbreak of parvovirus B19 infection investigated using salivary antibodies assays. Epidemiol Infect 1996;116:331-338.
- George JR and Fitchen JH. Future applications of oral fluid specimen technology. Amer J Med 1997; 102:21-25.
- Nokes DJ, Nigatu W, Abebe A, Messele T, Dejene A, Enqueslassie F, Vyse A, Brown DWG, Cutts FT. A comparison of oral fluid and serum for the detection of rubella-specific antibodies in a community study in Addis Ababa, Ethiopia. Trop Med Int Health 1998a;3:258-267.
- 11. Nokes DJ, Enquselassie F, Vyse A, Nigatu W, Cutts FT, Brown DWG. An evaluation of oral fluid collection devices for the determination of rubella antibody status in a rural Ethiopian community. Trans Roy Soc Trop Med Hyg 1998b;9:679-684.
- Vyse AJ, Brown DWG, Cohen BJ, Samuel R, Nokes DJ. Detection of rubella virusspecific immunoglobulin G in saliva by an amplification-based enzyme-linked immunossorbent assay using monoclonal antibody to fluorescein isothiocynate. J Clin Microbiol 1999;37:391-395.
- Nigatu W, Nokes DJ, Enquselassie F, Brown DWG, Cohen BJ, Vyse AV, Cutts FT. Detection of measles specific IgG in oral fluid using an FITC/anti-FITC IgG capture enzyme-linked immonosorbet assay (GACELISA). J Virol Methods 1999;83:135-144.
- Helfand RF, Kebede S, Alxander JP, Alemu W, Heath JL, Gary HE, Anderson LJ, Beyene H, Bellini WJ. Comparative

- detection of measles-specific IgM in oral fluid and serum from children by an antibody-capture IgM EIA. J Infect Dis 1996;173:1470-4.
- 15. Nokes DJ, Enquselassie F, Nigatu W, Vyse A, Cohen BJ, Brown DWG, Cutts FT. Has oral fluid the potential to replace serum for the evaluation of population immunity levels? A study of measles, rubella and hepatitis B in rural Ethiopia. Bull Wld Hlth Org 2001;79(7):588-95.
- Vyse AJ, Cohen BJ, Ramsay ME. A comparison of oral fluid collection devices for use in the surveillance of virus diseases in children. Public Health 2001;115:201-207.
- 17. Ramsay ME, Li J, White J, Litton P, Cohen B, Brown D. The elimination of indigenous measles transmission in England and Wales. Journal of Infectious Diseases 2003;187(suppl):5198-5207.
- Central Statistical Authority (CSA).
 Federal Democratic Republic of Ethiopia Statistical Abstract 2000.
- WHO. Expanded Programme on Immunization, Global Advisory Group. Weekly Epidemiological Record 1985; 60:13-16.
- Bouche FB, Brons NH, Houard S, Schneider F, Muller CP. Evaluation of hemagglutinin protein-specific immunoglobulin M for diagnosis of measles by an

- enzyme-linked immunosorbent assay based on recombinant protein produced in highefficiency mammalian expression system. J Clin Micrbiol 1998;36:3509-3513.
- 21. Choi B. Slopes of a receiver operating characteristics curve and likelihood ratios for diagnostic test. Amer J Epidemiol 1998;148:1127-32.
- WHO: Expanded programme on immunization. Measles immunization. Wkly Epidemiol Rec 1979; 54:337-9.
- 23. McLean AR, Anderson RM. Measles in developing countries. Part II. The predicted impact of mass vaccination. Epidemiol Inf 1988;100:419-42.
- Nokes DJ, McLean AR, Anderson RM, Grabowsky M. Measles immunization strategies for countries with high transmission rates: interim guidelines predicting using a mathematical model. Inf J Eipdemiol 1990:19:703-10.
- 25. Williams BG, Cutts FT, Dye C. Measles vaccination policy. Epidemiology and Infection 1995; 115:603-621.
- Samuel D, Patt R J, Abuknesha R A. A sensitive method of detecting proteins on dot blot and western blots using a monoclonal antibody to FITC. J Immunol Methods 1998;107:217-224.