

Etiologies of bloodstream infection and antimicrobial resistance: A cross sectional study among patients in a tertiary hospital, Northern Tanzania

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ABSTRACT

Background: Bloodstream infections are important causes of morbidity and mortality in people of all age groups, especially in sub-Saharan Africa. In Tanzania, a recent report indicates that case fatality rate of 37% is attributed to bloodstream infections. The aim of this study was to determine the prevalence and factors associated with bloodstream infections as well as to determine resistance pattern of bacterial isolates among patients visiting Kilimanjaro Christian Medical Centre (KCMC).

Methods: A cross-sectional study was conducted from April to June 2019 at KCMC. A total of 200 patients were included in the study. Blood samples were collected for culture, malaria rapid test, typhoid and brucella tests. Clinical features, co-morbid conditions and patients' hospitalization data were recorded in the questionnaire. Logistic regression was used to examine the factors associated with bloodstream infections. Predictors of the outcome were considered significant at $p < 0.05$.

Results: The prevalence of bloodstream infections was 52(26%). Participants with stomachache had less odds of having bloodstream infections as compared to other patients with symptoms (AOR=0.22, 5.33, 95%CI=0.05-0.97; $p=0.04$). Of the 41 identified isolates, *Staphylococcus aureus* showed the highest rates of resistance for Meropenem 8 (88.8%), Cefotaxime 6 (66.6%), Amikacin 6 (66.6%), Gentamicin 6 (66.6%) (and Imipenem 6 (66.6%). The lowest level of resistance was observed in Ceftriaxone 1(11.1%).

Conclusion: Bloodstream infections were highly prevalent in this sample (26%). *Staphylococcus spp* was the most commonly isolated organism and exhibited a high resistance rate to most antibiotics. This calls for increased and coordinated efforts to improve the identification, treatment and management of bloodstream infections and antimicrobial resistance, thereby improving clinical practice.

INTRODUCTION

Bloodstream infections (BSIs) are important causes of morbidity and mortality among children and adults in most African Countries¹⁻³. In a meta-analysis on community-acquired BSI in Africa, the mean mortality rate was reported to be 18.1%². In 2007, a study done at Muhimbili National Hospital, Dar es Salaam among a pediatric population, reported mortality rates of 20.2%, 43.5% and 16.7% for malaria, gram-negative BSIs and gram-positive BSIs respectively⁴. More recently findings from the same hospital indicated a case mortality rate of 37% among patients of all age groups⁵. The prevalence of BSIs

ranges from 13% to 14.5% in different parts of the Tanzania^{4,6,7}.

There are increased reports of antimicrobial resistance (AMR) among patients with BSI⁷⁻¹⁰. More importantly, rates of BSI are increasing owing to multidrug-resistant extended-spectrum beta-lactamase (ESBL)^{8,11,12}, methicillin-resistant *Staphylococcus aureus* (MRSA)¹³, and Vancomycin resistance¹⁴.

In Tanzanian, most of BSIs are misdiagnosed as malaria infections¹. This is due to the lack of capacity to properly identify the cause of febrile illness¹⁵. Consequently, Clinicians often rely on clinical features

to guide the treatment of patient who present with febrile illness¹⁶ which is less accurate in identifying BSIs, hence risking poor clinical outcomes and promotion of AMR¹⁷.

Various factors have been found to be associated with BSI, including; socio-demographics, comorbid conditions, prior hospitalization and recent exposures⁷. Comorbid conditions found to be associated with BSI include: acute and chronic renal failure, hepatic disease, diabetes, hypertension, congestive heart failure, and intravenous drug abuse¹⁸.

However, there is limited information about BSIs in Northern Tanzania. Specifically, the prevalence, level of resistance and factors associated have not been documented. Notably, the extent of AMR in most BSIs has not been studied. Understanding BSI at KCMC, resistance patterns, and the factors associated with BSIs will guide clinical management and appropriate antibiotic use. Therefore, the present study was designed to document prevalence, factors associated with BSI and resistance patterns of bacteria isolated among patients with BSI.

METHODS

Study design and area

This was a hospital-based cross-sectional study. The study was conducted at KCMC referral hospital, located in the foothills of the snowcapped Mount Kilimanjaro in Tanzania (<http://www.kcmc.ac.tz/>). KCMC has a 650-bed capacity and the second largest consultant referral university teaching hospital in the country serving over 15 million people from northern and central regions of Tanzania, attending more than 800 – 1000 outpatients daily.

Study population and inclusion criteria

The study included all out- and inpatients suspected with BSI. The study included patients of all age suspected of having bacterial or malarial infections. All patients who were critical ill, mentally unfit, and not able to communicate were excluded from the study.

Sample Size Estimation

Samples was calculated based on the following formula:

$$N = \frac{Z_p^2 (1-P)}{\epsilon^2}$$

Where: N = Sample size, Z = Level of confidence (1.96), P =proportion previous prevalence (13.4%) 6 and ϵ = Margin error (5%), plus 10% non-response. Sample size = $1.96^2 * 0.134 * (1-0.134) / (0.05)^2$. The minimum sample size was $178 + 10\% = 196$, we rounded up to give a sample size approximation of 200 patients.

Data collection

Interview

Data was collected by three research assistants who were experienced in data collection. All information was collected during face-to-face interviews using questionnaire in Swahili language. The interview lasted between 30 and 45 minutes. The interviews for Out-patients were conducted in a private room where no one other than the research assistant and the patients and/or guardian were allowed to be present. For inpatient, bedside interviews were conducted. The children's information's were given by their parents/guardians.

The questionnaire had three sections. Section 1: recorded patient's socio-demographic characteristics such as age, sex, occupation, level of education, income per month and residence. Section 2: assessed patient's clinical features and conditions. Other patients' information such as clinical outcomes were retrieved from the patient's medical records. Section 3: collected details on antibiotic usage, and lastly section 4 collated the laboratory investigations

Pre-testing of the questionnaire

To maximize its validity, the questionnaire was pretested on appropriate respondents before distribution. Interviews were conducted in five (5) patients to examine how patients understood and responded to the questions. In addition to the pilot, two experts in the field of survey design approved the quality of the questionnaire. After the pretest, adjustments in phrasings were made as necessary so that the questionnaire was simple and easily understood.

Blood sample collection and Blood culture

Venous blood was drawn aseptically from each patient. A total of 2-5ml of blood was collected in BD BACTEC bottles from pediatrics patients (BD BACTEC Peds PlusTM/F Culture Vials, Becton Dickinson and Company) and 8-10ml from adults (BD BACTEC Plus Aerobic/F Culture Vials, Becton Dickinson and Company). Blood samples were immediately transported at room temperature to the KCMC clinical laboratory and incubated in the BACTEC machine for further investigation. Blood samples were incubated in BD BACTEC machine for a maximum of 5 days. Positive blood cultures were inoculated on Blood agar, Chocolate agar (both from HI Media Laboratories, Mumbai, India), and MacConkey agar (Becton Dickinson and Company, Cockeysville, MD, USA) and incubated for 18–24 h at 37°C. Standard microbiological technique were conducted to identify bacteria including colony morphology, Gram stain, and biochemical tests (Oxoid). Gram-positive cocci were identified based on their gram reaction, catalase and coagulase test results. Gram-negative rods were identified by performing a series of biochemical tests such as Kligler Iron Agar (KIA), Simon's citrate agar, Indole, urea, and motility. Blood collected for malaria and serology were analyzed within 30 minutes in their

respective sections after collection. Samples were collected and processed by qualified laboratory technologist.

Susceptibility Test

Antimicrobial susceptibility testing was performed using disc diffusion on Müller–Hinton Agar according to Clinical Laboratory Standards Institute guidelines¹⁹. Bacterial isolates were tested against amoxicillin-clavulanic acid (30 µg), Amikacin (30 µg), Ceftriaxone (30 µg), Gentamicin (10 µg), Imipenem (30 µg), Trimethoprim-sulfamethoxazole (23.75 µg/1.25 µg), Ciprofloxacin (5 µg), Clindamycin (2 µg), Erythromycin (15 µg), Meropenem (10 µg), Vancomycin (30 µg), Tetracycline (30 µg), Cefotaxime (30 µg), Penicillin (10 µg) and Chloramphenicol (30 µg). All antibiotic discs were from Oxoid, ThermoFisher, and Scientific, USA. Antimicrobial sensitivity was reported as resistant, intermediate, and sensitive according to the Clinical Laboratory Standard Institute¹⁹. The choice of antibiotic agents varied depending on the range of antibiotics available to the laboratory.

Case definition

BSI was defined as having either positive blood culture, malaria parasites, a positive serological result or any co-infections.

Data analysis plan

Data were analyzed using IBM SPSS Statistics for Windows, Version 22.0 (IBM Corp, Armonk, NY, USA). Cross-tabulation of categorical variables was calculated using Chi-square test (χ^2) while Fisher's exact test was used in cases when expected counts were ≤ 5 . The association between categorical predictors and the BSI was presented as odds ratio (OR) with 95% confidence intervals (95% CI) using logistic regression. Predictors significantly associated with BSI in the bivariate analysis were selected for multivariable analysis in the final model. A significance level ≤ 0.05 was used throughout.

Ethical Consideration

Ethical approval to conduct this study was obtained from the Kilimanjaro Christian Medical University College Research and Ethics Review Committee (CRERC) with ethics certificate number 2472. Written consent was signed before filling the questionnaire. Permission to conduct the study was obtained from KCMC administration. Though all measures to protect the privacy and confidentiality was considered that neither name nor registration number was mentioned during data collection.

RESULTS

Demographic characteristics of the study population

A total of 200 patients were enrolled in the study, giving a response rate of 100%. The modal age group was 0-5 years with a frequency of 72 (36%). More

than half of the participants were female 110 (55%). The majority of the participants lived in urban 119 (59.5%). Most participants were self-employed 65 (32.5%). Lastly, 58 (29%) and 62 (31%) had primary and secondary education respectively (*Table 1*).

Prevalence and common etiologies of BSI infections

Overall, 52/200 (26%) of the participants were evident of having BSI infections. A total of 123 blood cultures were performed, 76/123 (61.8%) from children (0-17 years) and 47/123 (38.2%) from adults >17 years. Positive bacterial growth was observed in 41/123 (33.3%) isolates with 18/123 (14.6%) being significant for antimicrobial susceptibility testing. The number of pathogens recovered in the study period is presented in *Figure 1*. A total of 67 samples were subjected to serological tests. Out of these, 27(40.3%) were tested for Widal, 20(29.9%) tested for Brucella and the remaining were tested for syphilis. The decisions for specific tests on samples were based on the clinical symptoms presented by the patient and the tests ordered by the attending clinician. A total of 10(37.0%) were positive for Widal while 2(10.5%) were positive for Brucella. Two of the participants had co-infection (typhoid and Brucellosis), *Figure 2*. A total of 51 participants were tested for malaria, only 1(2.0%) was found positive.

Antimicrobial susceptibility patterns of bacterial isolates

Antibiotic-resistance patterns of bacteria against the antimicrobial agents is shown in *Table 2*. Coagulase Negative Staphylococcus (CoNS) showed highest resistance for Gentamicin 24 (96%), Trimethoprim/sulfamethoxazole 24 (96%), Imipenem 24 (96%), Chloramphenicol 24 (96%), Ciprofloxacin 23 (92%), Clindamycin 23 (92%) and Gentamycin 22 (88%). Staphylococcus aureus showed the highest resistance for Meropenem 8 (88.8%), Chloramphenicol 8 (88.8%), Cefotaxime 6 (66.6%), Tetracycline 6 (66.6%), Amikacin 6 (66.6%), Gentamicin 5 (66.6%) and Imipenem 6 (66.6%). The lowest level of resistance was observed in Ceftriaxone 1 (11.1%).

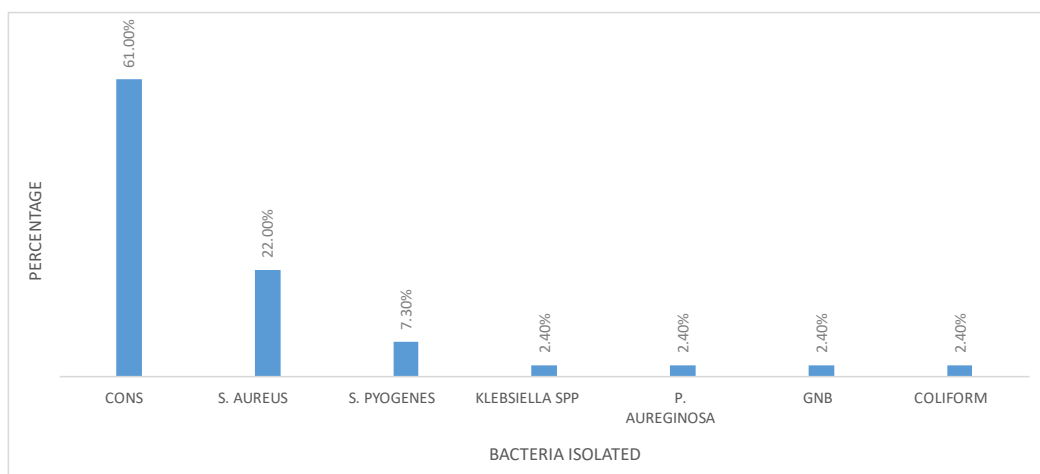
Risk factors associated with BSI

The results of the bi- and multivariate analyses of the association between selected predictors and BSI are shown in *Table 3*. In the bivariate analysis: age of the participants, education level, infection risk, admission, length of admission and hospitalization showed an association with the outcome and were then selected to be included in the multivariate model. Only two factors remained independently associated with the occurrence of BSI after adjusting for confounding. Participants who reported having a diploma as their highest level of education had a higher odds of having BSIs compared to those with a degree (AOR= 5.33, 95%CI=1.39-20.38; P=0.01). Participants with a stomachache had lower odds of having BSI compared to other patients with symptoms (AOR=0.22, 5.33, 95%CI=0.05-0.97; P=0.04).

TABLE 1: Socio-demographic characteristics of the respondents (N =200)

Variable	n (%)	BSI infection	
		Positive n (%)	Negative n (%)
Age group (years)			
0-5	72 (36.0)	26 (49.1)	46 (31.3)
6-17	7 (3.5)	1 (1.9)	6 (4.1)
18-45	69 (34.5)	17 (32.1)	52 (35.4)
More than 45	52 (26.0)	9 (17.0)	43 (29.3)
Locality			
Urban	119 (59.5)	31 (58.5)	88 (59.9)
Rural	81 (40.5)	22 (41.5)	59 (40.1)
Sex			
Male	90 (45.0)	26 (49.1)	64 (43.5)
Female	110 (55.0)	27 (50.9)	83 (56.5)
Education level			
Illiterate	4 (2.0)	0 (0.0)	4 (2.7)
Primary level	58 (29.0)	19 (35.8)	39 (29.5)
Secondary level	62 (31.0)	17 (32.1)	45 (30.6)
Diploma level	32 (16.0)	2 (3.8)	30 (20.4)
Degree level	32 (16.0)	11 (20.8)	21 (14.3)
Others	12 (6.0)	4 (7.5)	8 (5.4)
Occupation status			
Employed	61 (30.5)	13 (24.5)	48 (32.7)
Unemployed	43 (21.5)	12 (22.6)	31 (21.1)
Student	31 (15.5)	7 (13.2)	24 (16.3)
Self-employed	65 (32.5)	21 (39.6)	44 (29.9)
Income			
Less than 300,000/=	62 (31.0)	18 (34.0)	44 (29.9)
300,000/= to 1,000,000/=	57 (28.5)	14 (26.4)	43 (29.3)
More than 1,000,000/=	4 (2.0)	1 (1.9)	3 (2.0)
No income	77 (38.5)	20 (37.7)	57 (38.8)

FIGURE 1: Isolated Bacteria



CoNS- Coagulase negative Staphylococcus
GNB- Gram negative bacilli

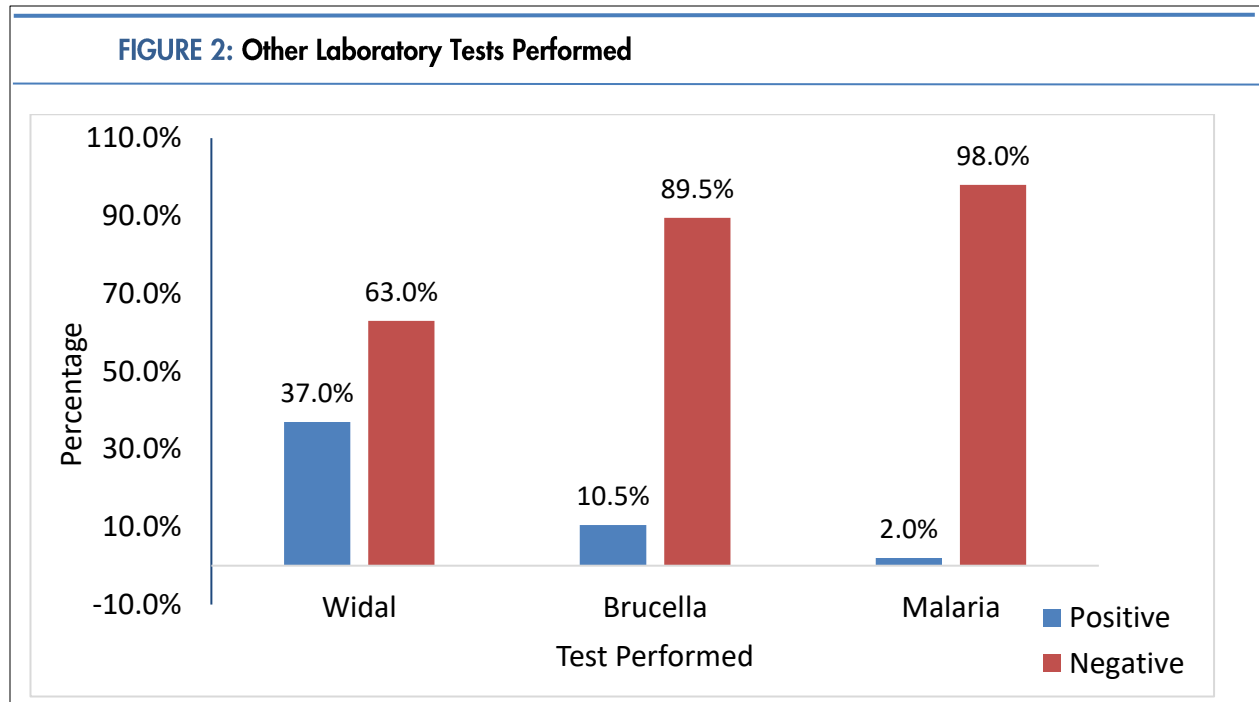


TABLE 3: Factors associated with BSI

Variable	COR	P-value	AOR	P-value
Age group (years)				
0-5	0.37 (0.15-0.87)	0.02	1.29 (0.17-9.41)	0.7
6-17	1.25 (0.13-11.74)	0.8	3.77 (0.25-55.05)	0.3
18-45	0.69 (0.27-1.72)	0.4	0.61 (0.20-1.89)	0.3
More than 45	Reference		Reference	
Locality				
Urban	1.10 (0.58-2.10)	0.7		
Rural	Reference			
Sex				
Male	0.84 (0.44-1.59)	0.6		
Female	Reference			
Education level				
Primary level	1.12 (0.45-2.73)	0.8	2.37 (0.69-8.07)	0.1
Secondary level	1.47 (0.60-3.61)	0.3	2.75 (0.89-8.53)	0.07
Diploma level	5.04 (1.44-17.58)	0.01	5.33 (1.39-20.38)	0.01
Degree level and above	Reference		Reference	
Occupation				
Employed	1.93 (0.86-4.32)	0.1	1.69 (0.59-4.79)	0.3
Unemployed	1.65 (0.78-3.47)	0.1	1.44 (0.59-3.49)	0.4
Self-employed	Reference			
Income				
Less than 300,000/=	0.91 (0.09-9.11)	0.9		
300,000/= to 1,000,000/=	1.02 (0.09-10.65)	0.9		
More than 1,000,000/=	Reference			
Symptoms				

	Fever	0.78 (0.35-1.75)	0.5	0.67 (0.27-1.69)	0.4
	Headache	1.56 (0.44-5.52)	0.4	1.14 (0.27-4.84)	0.8
	Stomachache	0.44 (0.13-1.53)	0.1	0.22 (0.05-0.97)	0.04
	Diarrhea	0.44 (0.06-3.01)	0.4	0.22 (0.02-2.13)	0.1
	Others*	Reference			
Co-morbidities					
	Diabetes	1.78 (0.20-16.67)	0.6		
	Hypertension	0.89 (0.26-2.98)	0.8		
	Cancer	0.35 (0.49-2.60)	0.3		
	Others	1.06 (0.27-4.13)	0.9		
	None	Reference			
Infection risks (N=87)					
	Urinary catheter	0.59 (0.14-2.45)	0.4	0.80 (0.15-4.13)	0.7
	Intravascular catheter	0.43 (0.22-0.84)	0.01	0.74 (0.29-1.85)	0.5
	None	Reference		Reference	
Admission					
	Yes	0.32 (0.16-0.65)	0.02	0.12 (0.005-3.10)	0.2
	No	Reference		Reference	
Duration of hospitalization					
	Less than 7 days	0.41 (0.19-0.87)	0.02	1.35 (0.02-68.09)	0.8
	7 to 14 days	0.29 (0.11-0.73)	0.009	0.71 (0.01-39.24)	0.8
	More than 14 days	0.18 (0.04-0.82)	0.02	0.46 (0.006-34.71)	0.7
	Not admitted	Reference		Reference	
Ward					
	Medical	0.75 (0.26-2.15)	0.5	4.70 (0.35-62.17)	0.2
	Surgical	0.34 (0.07-1.57)	0.1	3.22 (0.17-59.01)	0.4
	Pediatric	0.32 (0.15-0.67)	0.02	1.31 (0.07-22.70)	0.8
	Others	Reference		Reference	

COR: Crude Odds Ratio, AOR: Adjusted Odds Ratio

* Other symptom such as Rashes and vomit/nausea

DISCUSSION

This study was conducted among patients who visited KCMC. The study aimed at determining the prevalence of BSIs, associated factors, and antibiotic susceptibility. The prevalence of positive blood culture in this study was found to be 14.6% which is comparable to other studies conducted in the same region (Zanzibar (14.0%)²⁰, Muhimbili-Tanzania (13.4%).⁶ A higher prevalence was observed in a study done in Malawi which reported a prevalence of 30%, also difference may be due to different study population whereby it was specific to adults only who presented with fever²². Coagulase Negative Staphylococcus (61.0%) was the most predominant bacteria isolated among participants with bacterial infections, followed by Staphylococcus aureus (22.0%). This pattern is similar to a study conducted in Dar Es Salaam Tanzania which also reported that Staphylococcus aureus as the most common cause of BSIs⁶. CoNS was also reported to be the most isolated bacteria in a study conducted in Ghana²³, while Staphylococcus aureus was the second most isolated bacteria among patients with BSI in Ghana²³ and Malawi²⁴. Salmonella typhi was the most common organism isolated among the positive participants. Different studies have also reported

Salmonella being a major cause of infection in Bangladesh (36.9%)²¹, Nepal (71%)²⁵ and Malawi²⁴.

Coagulase Negative Staphylococcus showed the highest rates of drug resistance to Trimethoprim/sulfamethoxazole, Imipenem, Chloramphenicol, Ciprofloxacin, Clindamycin and Gentamycin. Similarly, a study in India reported that Ciprofloxacin and Clindamycin was resistance to CoNS²⁶. Staphylococcus aureus showed the highest rates of resistance for gentamicin (66.6%). This finding is comparable to similar reports from Brazil²⁷, India²⁶ and Senegal²⁸. In contrary, a study conducted in Zanzibar reported high susceptibility of Staphylococcus aureus to Cefotaxime and Tetracycline²⁰. In general, our findings suggest that the that isolates found on the Tanzanian mainland are quite similar to those found elsewhere²⁶⁻²⁸. However, we noted small differences with a study conducted in Zanzibar²⁰. It is possible that the smaller population and more limited circulation of people reduces the transmission of some strains of the pathogen. Based on the observed results in this study, treatment should follow microbiology investigations and not be solely based on clinical symptoms. This will improve the accuracy of diagnosis and support the reduction in the occurrence of AMR. Further efforts

are needed to raise awareness amongst the community and healthcare workers as to the rapid increase in drug resistance and advocate on the antimicrobial stewardship program in these settings.

The education of individuals can help tackling the spread of disease, given that educated people may be more aware how diseases circulate in their community. The probability of having or dying from infections such as malaria is inversely related to income and education²⁹. However, this was not the case in this study, our results suggest that those with a higher level of education (diploma) were more likely to have a BSI. It is possible that this result is caused by a selection bias, the study was hospital based, so it is possible that those who are more educated may have better health seeking behavior and therefore more likely to attend hospital and then be included in this study. A study conducted in America showed that education level was a significant factor associated with BSIs for more than 30% of participants¹⁸. Though lacking statistical significance our findings suggest that participants with diabetes were 78% more likely to have BSIs compared to others. Our sample size was small, and this may have reduced our power to detect an association. Further studies should be conducted to investigate this association. Finally, our study suggests that increased efforts should be taken to reduce opportunities for infection.

The findings of this study should be considered with the following limitations in mind. Firstly, a single blood culture specimen was collected from each patient, therefore it was not possible to determine if the patients with CoNS isolation had true bacteremia or the finding was due to skin contamination. The data for this study were obtained for three months between April and June 2019, and therefore seasonal variations in the frequency of BSI and causative microorganisms could not be assessed.

CONCLUSION

We identified a high prevalence of BSIs across all age groups and more commonly associated with education level. *Staphylococcus* spp was the most commonly isolated organism among patients with BSIs and exhibited high levels of AMR. We recommend that laboratory investigations and susceptibility tests of isolated bacteria should guide treatment protocols, and these should not be solely based on clinical symptoms.

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Conflict of interest

There was no conflict of interest

REFERENCES

- Crump A., Morrissey A., Nicholson W., et al. Etiology of severe non-malaria febrile illness in Northern Tanzania: a prospective cohort study. *PLoS Negl Trop Dis.* 2013;7(7):1-9.
- Reddy EA, Shaw A V, Crump JA. Community-acquired bloodstream infections in Africa: a systematic review and meta-analysis Elizabeth. *Lancet Infect Dis.* 2010;10(6):70072-70074. doi:10.1016/S1473-3099(10)70072-4.
- Nichols C, Cruz Espinoza LM, Von Kalckreuth V, et al. Bloodstream infections and frequency of pretreatment associated with age and hospitalization status in Sub-Saharan Africa. *Clin Infect Dis.* 2015;61(Suppl 4):S372-S379. doi:10.1093/cid/civ730
- Blomberg B, Manji KP, Urassa WK, et al. Antimicrobial resistance predicts death in Tanzanian children with bloodstream infections: A prospective cohort study. *BMC Infect Dis.* 2007;7:1-14. doi:10.1186/1471-2334-7-46
- Manyahi J, Kibwana U, Mgimba E, Id MM. Multi-drug resistant bacteria predict mortality in bloodstream infection in a tertiary setting in Tanzania. *PLoS One.* 2020;15(3):1-11. doi:10.1371/journal.pone.0220424
- Moyo S, Aboud S, Kasubi M, Maselle SY. Bacteria isolated from bloodstream infections at a tertiary hospital in Dar es Salaam, Tanzania - antimicrobial resistance of isolates. *South African Med J.* 2010;100(12):835-838. doi:10.7196/SAMJ.4186
- Seni J, Mwakyoma AA, Mashuda F, et al. Deciphering risk factors for blood stream infections , bacteria species and antimicrobial resistance profiles among children under five years of age in North- Western Tanzania : a multicentre study in a cascade of referral health care system. *BMC Pediatr.* 2019;19(32):1-11.
- Kajeguka DC, Nambunga PP, Kabissi F, et al. Antimicrobial resistance patterns of phenotype Extended Spectrum Beta-Lactamase producing bacterial isolates in a referral hospital in northern Tanzania. *Tanzan J Health Res.* 2015;17(3):1-8. doi:10.4314/thrb.v17i3.%c
- Moremi N, Claus H, Mshana SE. Antimicrobial resistance pattern: A report of microbiological cultures at a tertiary hospital in Tanzania. *BMC Infect Dis.* 2016;16(1). doi:10.1186/s12879-016-2082-1
- Kumburu HH, Sonda T, Mmbaga BT, et al. Patterns of infections, aetiological agents and antimicrobial resistance at a tertiary care hospital in northern Tanzania. *Trop Med Int Heal.* 2017;22(4):454-464. doi:10.1111/tmi.12836
- Seni J, Sweya E, Mabewa A, Mshana SE, Gilyoma JM. Comparison of antimicrobial resistance patterns of ESBL and non ESBL bacterial isolates among patients with secondary peritonitis at Bugando Medical Centre, Mwanza - Tanzania. *BMC Emerg Med.* 2016;16(1). doi:10.1186/s12873-016-0106-1
- Marando R, Seni J, Mirambo MM, et al. Predictors of the extended-spectrum-beta lactamases producing Enterobacteriaceae neonatal sepsis at a tertiary hospital, Tanzania. *Int J Med Microbiol.* 2018;308(7):803-811. doi:10.1016/j.ijmm.2018.06.012
- Kumburu HH, Sonda T, Leekitcharoenphon P, et al. Hospital Epidemiology of Methicillin-Resistant *Staphylococcus aureus* in a Tertiary Care Hospital in Moshi, Tanzania, as Determined by Whole Genome Sequencing. *Biomed Res Int.* 2018;2018:1-12. doi:10.1155/2018/2087693
- Blomberg B. Antimicrobial resistance in bacterial infections in urban and rural Tanzania. *BMC Public Health.* 2007. doi:10.1186/1471-2334-4-35
- Kajeguka DC, Kaaya RD, Mwakalinga S, et al. Prevalence of dengue and chikungunya virus infections in north-eastern Tanzania: a cross sectional study among participants presenting with malaria-like symptoms. *BMC Infect Dis.* 2016;16(183):1-9. doi:DOI 10.1186/s12879-016-1511-5
- Crump JA, Gove S, Parry CM. Management of adolescents and adults with febrile illness in resource limited areas. *BMJ.* 2011;343(d4847). doi:10.1136/bmj.d4847
- Kajeguka DC, Desrochers RE, Mwangi R, et al. Knowledge and practice regarding dengue and chikungunya: a cross-sectional study among Healthcare workers and community in Northern Tanzania. *Trop Med Int Heal.* 2017;22(5):583-593.

- doi:10.1111/tmi.12863
18. Rybak MJ, Zasowski EJ, Jorgensen SCJ. Risk Factors for Bloodstream Infections Among an Urban Population with Skin and Soft Tissue Infections: A Retrospective Unmatched Case-Control Study. *Infect Dis Ther*. 2018;1-11. doi:10.1007/s40121-018-0227-9
 19. CLSI. Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing: CLSI AST News Update. 2018;1-3.
 20. Onken A, Said AK, Melissa J, Jenum PA. Prevalence and Antimicrobial Resistance of Microbes Causing Bloodstream Infections in Unguja, Zanzibar. *PLoS One*. 2015;10(12):1-10. doi:10.1371/journal.pone.0145632
 21. Ahmed D, Nahid A, Sami AB, et al. Bacterial etiology of bloodstream infections and antimicrobial resistance in Dhaka. *Antimicrob Resist Infect Control*. 2017;6(2):1-11. doi:10.1186/s13756-016-0162-z
 22. Archibald LK, McDonald LC, Nwyanwu O, et al. A Hospital-Based Prevalence Survey of Bloodstream Infections in Febrile Patients in Malawi: Implications for Diagnosis and Therapy. *J Infect Dis*. 2000;181:1414-1420.
 23. Deku JG, Dakorah MP, Lokpo SY, et al. The Epidemiology of Bloodstream Infections and Antimicrobial Susceptibility Patterns: A Nine-Year Retrospective Study at St. Dominic Hospital, Akwatia, Ghana. *J Trop Med*. 2019;2019:1-10. doi:10.1155/2019/6750864
 24. Musicha P, Cornick JE, Bar-Zeev N, et al. Trends in antimicrobial resistance in bloodstream infection isolates at a large urban hospital in Malawi (1998–2016): a surveillance study. *Lancet Infect Dis*. 2017;17(10):1042-1052. doi:10.1016/S1473-3099(17)30394-8
 25. Bhandari P, Manandhar S, Shrestha B, Dulal N. Etiology of bloodstream infection and antibiotic. *Asian J Med Sci*. 2016;7(2):71-75. doi:10.3126/ajms.v7i2.13444
 26. Singh A, Venkatesh V, Singh R, Singh M. Bacterial and antimicrobial resistance profile of bloodstream infections: A hospital-based study. *CHRISMED J Heal Res*. 2014;1(3):140-144. doi:10.4103/2348-3334.138881
 27. Monteiro A de S, Pinto BLS, Monteiro J de M, et al. Phylogenetic and molecular profile of *Staphylococcus aureus* isolated from bloodstream infections in northeast Brazil. *Microorganisms*. 2019;7(210):1-14. doi:10.3390/microorganisms7070210
 28. Lakhe NA, Sylla K, Mbaye KD, et al. Bacteremia: Profile and Antibiotic Resistance at the Infectious and Tropical Diseases Clinic in Fann Hospital, Dakar, Senegal. *J Infect Dis Ther*. 2018;6(1):1-8. doi:10.4172/2332-0877.1000348
 29. Tusting LS, Willey B, Lucas H, et al. Socioeconomic development as an intervention against malaria: A systematic review and meta-analysis. *Lancet*. 2013;382:963-972. doi:10.1016/S0140-6736(13)60851-X
 30. Dinan TG, Cryan JF. Microbes Immunity and Behavior: Psychoneuroimmunology Meets the Microbiome. *Neuropsychopharmacology*. 2017;42(1):178-192. doi:10.1038/npp.2016.103

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