

Phytochemical composition and insecticidal effects of native plant extracts against the mango mealybug, *Rastrococcus invadens* Williams (Homoptera: Pseudococcidae)

§,1,3Elias Mano, 2Désiré Dramane Abdoul Khader Sanou, 3Alizèta Sawadogo, 1Rasmané Abdou Ouédraogo, 4Abdoulaye Sidibé, 3Ouattara Boubakar, 3Pierre Alexandre Eric Djifaby Sombié, 3Anogmain Rémy Dabiré, 2Fernand Sankara & 3Souleymane Nacro

¹Institute of Research in Applied Sciences and Technologies (IRSAT)/Laboratory of Natural Substances and Technologies of Natural Products and the Environment (LABTECH-PRONE)/Ouagadougou/Burkina Faso

²Nazi Boni University / Institute of Rural Development (IDR), Bobo Dioulasso / Burkina Faso.

³Institute of the Environment and Agricultural Research (INERA), Central Horticulture Laboratory of the Regional Center of Excellence in Fruits and Vegetables of Bobo-Dioulasso / Burkina Faso.

⁴National Agricultural Training School of Matourkou (ENAF), Bobo Dioulasso / Burkina Faso

Corresponding author: Elias Mano. Phone number/Email: +226 71 04 02 91 / manoe2005@gmail.com

Abstract

The mango mealybug *Rastrococcus invadens* Williams causes enormous damage to fruit trees in Burkina Faso, with losses of up to 90% for mango trees and 53% for citrus fruits. The objective of this study was to evaluate the insecticidal effects of native plant extracts against this insect pest. The insecticidal activity and biological effectiveness of the extracts were evaluated on Petri dishes and mango leaves previously contaminated by spraying them with the native plant extracts. The experimental was a completely randomized Fischer block, and compared six doses of the hydroalcoholic extracts of *A. conyzoides* and *H. suaveolens* with abamectin 18 EC and water (control). These treatments were repeated up to ten times. The determination of the efficiency coefficients was achieved using Abbott's formula (1925). Both extracts of *A. conyzoides* and *H. suaveolens* were effective, but the one based on *A. conyzoides* at 50 gl⁻¹ was the most effective with 100% mortality in 48 hours compared to 95.00% for *H. suaveolens*' extract in 72 h. Their respective LC50 were 12.50 gl⁻¹ and 17.49 gl⁻¹, showing that *A. conyzoides*' extract is more toxic to *R. invadens*. The high content in phenols, flavonoids, alkaloids, etc. of leafy stems of *A. conyzoides* and of *H. suaveolens* could help to sustainably control *R. invadens* in an integrated approach and effectively protect mango orchards.

Keywords: *Rastrococcus invadens*, *Ageratum conyzoides*, *Hyptis suaveolens*, bioinsecticide, biological effectiveness.

Received November 9, 2023; **Revised** December 10, 2023; **Accepted** December 8, 2023

<https://dx.doi.org/10.4314/br.v21i3.15> This is an Open Access article distributed under the terms of the Creative Commons License [CC BY-NC-ND 4.0] <http://creativecommons.org/licenses/by-nc-nd/4.0>.

Journal Homepage: <http://www.bioresearch.com.ng>.

Publisher: Faculty of Biological Sciences, University of Nigeria, Nsukka, Nigeria.

INTRODUCTION

The mealybug *Rastrococcus invadens* Williams (Homoptera: Pseudococcidae) is a major insect pest of fruit trees including mango and citrus. This mealybug causes enormous damage to the mango tree, up to 90% loss of mangoes and 53% loss of fruit yield in citrus fruits (FAO, 2022). Currently, this insect pest is distributed in over 27 countries in Africa, Asia, and French Guiana in Southern America (CABI, 2022). Nymphs and females feed on fruit and leaves leading to severe damage. These produce honeydew which causes sooty mold, leading to a reduction in production. *Rastrococcus invadens* is a polyphagous insect pest attacking more than forty-five plant species in twenty-two families, including economically important crops such as mango and citrus, *Citrus* spp. (Nébié *et al.*, 2018, Fall *et al.* 2019).

The means of control undertaken throughout Africa, such as the destruction of the attacked organs or insecticide treatments, have gradually proven to be ineffective with toxicological risks and more growing invasion due to the great dispersion of host plants and to climate change (Germain *et al.*, 2015, Azrag *et al.*, 2023). Treatment with synthetic insecticides is not always effective and safe and leads the mango orchards owners to alternative practices, mainly the usage of botanicals.

Synthetic insecticides, such as Thiametoxam (neonicotinoid) and Chlorpyrifos-ethyl (organophosphorus) evaluated in real environments, have proven effective against the insect pest, but remain incompatible with the action of parasitoids associated with *R. invadens* (Nébié, 2011). Furthermore, the resistance to insecticides has been demonstrated for many years by several authors and the use of these pesticides is proving to be dangerous for ecosystems and human health (Ahmad *et al.*, 2010; Abd El-Rahman *et al.*, 2021). These pesticides even annihilate the action of parasitoids used in biological control programs against this insect pest in West Africa (Mano *et al.*, 2022, Mohamed *et al.*, 2022).

However, organic insecticides based on extracts and essential oils from indigenous plants could constitute a sustainable alternative. Numerous studies have previously revealed the insecticidal potential of plant extracts which, according to Mano and Nana (2022) also preserve natural enemies. For example, neem and Mahogoni extracts were reported to exhibit good control of the mango leafhopper *Idioscopus clypealis*, (Shawan *et al.*, 2018). The same was shown with neem oil

against the same insect pest on the mango tree (Ferdous and Jahan, 2020). The use of plant extracts in plant protection is non-polluting and less toxic compared to synthetic insecticides. This justifies the importance of the present study, which aimed at evaluating the insecticidal effects of native plant extracts against the mango mealybug, *R. invadens*.

MATERIALS AND METHODS

Insects and insecticides

The mealybugs were collected from untreated mango tree plants using bags in the city of Bobo Dioulasso in April 2023. They were bred in the laboratory (T° : 38 ± 2 °C; RH: 70%; PP : 12/12h) until the adults of the F1 generation were obtained. Identification was made by possible using the Williams (1989) and Williams and Granara De Willink (1992) identification keys. The native insecticide used were abamectin 18 EC and hydroalcoholic extracts of leafy stems of *Ageratum conyzoides* and *Hyptis suaveolens*.

Preparation of extracts

Powders from the leafy stems of *A. conyzoides* and *H. suaveolens* were used as solid-liquid extraction substrate. For this purpose, 50 g of each leafy stem powder were extracted by maceration with 0.5 L of 80% ethanol in 1 L Erlenmeyer flasks under constant mechanical stirring using a laboratory shaker for 24 h. An additional volume of 125 ml of the extracting solvent was added under a percolator and filtered through cotton. The total extracts obtained were dried in an oven (40 ± 1 °C) to 100 g l^{-1} and used as insecticide products.

Evaluation of insecticidal activity

The host mango tree leaves were used as a trophic support during the various tests in the laboratory from April to June 2023. The insecticidal activity of the extracts was evaluated under laboratory conditions in a Fischer block with four treatments [Abamectin (manufactured by the company SAVANA based in Burkina Faso), water control, extracts from *A. conyzoides* and *H. suaveolens*] repeated ten times. Mango leaves were previously cut into leaf discs and placed in Petri dishes. Each box received 10 adult mealybugs. and foliar contamination of 0.6 ml of extract. The number of dead or weakened individuals was counted under a binocular microscope after 24; 48 and 72 hours of exposure. As for dose effects,

concentrations of 12.5; 25 and 50 gL⁻¹ of the most active extracts were tested on Petri dishes as previously.

Evaluation of the biological effectiveness of insecticides

Four treatments including the extracts of *A. conyzoides* 50 SL and *H. suaveolens* 50 SL were compared with abamectin 18 EC and water as a control in the same device above. The infestation was recorded from the average number of *R. invadens* per mango leaf. The mortalities' means of the mealybug after 72 hours of treatment were calculated and submitted to the Abbott formula (1925) for the determination of the efficiency coefficient.

Phytochemical screening

The chemical screening of the extracts was carried out by the qualitative method described by Ciulei (1982). These are general and/or specific chemical reaction tests for coloring or complexation in a liquid medium. The aim is to highlight the bioactive phytochemical groups involved in the insecticidal activities observed. Quantitative analysis was performed following the standard procedure for phenols (Ainsworth and Gillespie, 2007), flavonoids (Chang *et al.*, 2002) and alkaloids (Puro *et al.*, 2018).

Statistical analysis of data

R software version 4.0.3 was used for data analysis. The Shapiro test verifying the

normality of data and that of Fligner-Killeen attesting the homogeneity of variances allowed the analysis of variance of mortalities. The means were compared using the Tukey test at 5% threshold. Lethal concentrations LC50 and LC90 were determined using logistic regression by probit analysis.

RESULTS

Extraction yield

The yields of hydroalcoholic extracts were higher in *A. conyzoides*. A mass of extract of 16.50 g or 8.25% was obtained with *A. conyzoides* compared to 12.86 g or 6.43% for *H. suaveolens*.

Insecticidal activity of the extracts on *R. invadens*

As demonstrated by the analysis of variance (Table I), the organic extracts studied induced mortality on adults of *R. invadens* with very highly significant differences ($P < .001$; $F \geq 244.10$). The extracts increased mortality proportionally to their doses. The mortalities induced by the extract of *A. conyzoides* varied from 50.00% with 12.5 g/L to 92.50% with 50 g/L in 24 hours and 50.11% with 12.5 g/L to 100% with 50 g/L in 72 hours. As for those of *H. suaveolens* extracts, the transition from 12.5 g/L to 50 g/L resulted in mortality rates varying from 37.50 to 90.00% in 24 hours, from 47.50% to 92.50% in 48 hours and 50.00% to 95.00% in 72 hours.

Table I: Analysis of mortality of *R. invadens* caused by treatments

Treatments	Concentration (g/L)	Means \pm SE of mortality (%)		
		24h	48 hours	72 hours
Control (water)	-	0.00 \pm 00 f	0.00 \pm 00d	4.75 \pm 0.63 f
Abamectin 18 EC	-	62.50 \pm 2.03 bc	75.00 \pm 2.17 b	80.00 \pm 2.09c
<i>H. suaveolens</i>	12.5	37.50 \pm 1.90 e	47.50 \pm 1.98c	50.00 \pm 2.04 e
	25	70.00 \pm 2.61 b	85.00 \pm 2.03 a	87.50 \pm 2.23 b.c.
<i>A. conyzoides</i>	50	90.00 \pm 2.04 a	92.50 \pm 2.05 a	95.00 \pm 2.11 ab
	12.5	50.00 \pm 2.00 d	50.02 \pm 2.04 c	50.11 \pm 2.33 e
	25	55.00 \pm 2.08 cds	55.00 \pm 1.89c	65.00 \pm 2.21 d
	50	92.50 \pm 2.05 a	92.50 \pm 2.06 a	100.00 \pm 0.01 a
F. value (24)		244.10	266.70	267.40
Pr		2.2e-16	2.2e-16	2.2e-16
Meaning		***	***	***

NB: Means in the same column, assigned with the same letter, are not significantly different at the 5% threshold (Tukey test); ***: Highly significant

Table II: Analysis of extracts toxicity on *R. invadens*

Extract	Concentration (gl ⁻¹)		CL90 ± SE	95% CI	R ²
	LC50 ± SE	95% CI			
<i>A. conyzoides</i>	12.50 ± 1.71	11.35 - 15.00	47.67 ± 1.01	43.12 – 50.56	0.863
<i>H. suaveolens</i>	17.49 ± 2.01	14.83 – 19.69	50.00 ± 0.14	47.78 – 58.21	0.998

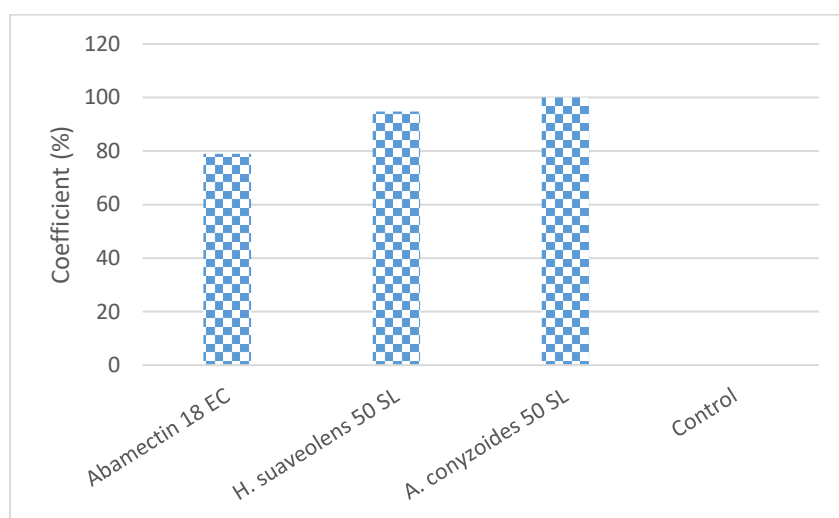


Figure 1: Biological effectiveness of treatments using the Abbott formula

Extracts toxicity on *R. invadens*

The toxicity analysis (Table II) shows that the extract of *A. conyzoides* is more toxic on *R. invadens* with LC 50 of 12.50 ± 1.71 gl⁻¹ and CL 90 of 47.67 ± 1.01 gl⁻¹ (R²= 0.998) against LC 50 of 17.49 ± 2.01 gl⁻¹ and an LC 90 of 50.00 ± 0.14 gl⁻¹ (R²= 0.863) for *H. suaveolens*.

Biological effectiveness of plant extracts on *R. invadens*

The data presented in Figure 1 showed variable biological effectiveness coefficients following the different treatments. The highest biological effectiveness (100.00%) was recorded with the extract of *A. conyzoides* 50 SL, followed by that of *H. suaveolens* 50 SL (94.75%) which is in close proximity to Abamectin 18 EC (79.00%).

Phytochemical composition of extracts

The chemical characterization test of the two active extracts showed the presence of polyphenols, flavonoids, leucoanthocyanins, tannins, alkaloids, saponins, quinones and anthraquinones and essential oils (Table III). Leucoanthocyanins, quinones and anthraquinones were absent in *A. conyzoides* while sterols and terpenes were not detected in *H. suaveolens*. Quantitative analysis indicated phenolic, flavonoid and alkaloid contents respectively of 0.50; 143.29; 125.48 mg/g of dry matter of *H. suaveolens* versus 7.54; 29.25 and 2.33 mg/g of dry matter of *A. conyzoides*.

Table II: Chemical composition of extracts

	Chemical groups	Hydro alcoholic extracts	
		<i>A. conyzoides</i>	<i>H. suaveolens</i>
Qualitative analysis	Sterols And terpenes	+	(-)
	Polyphenols	+	+
	Flavonoids	+	+
	Leucoanthocyanins	(-)	+
	Catechical tannins	+	+
	Tannins gallic	+	+
	Alkaloids	+	+
	Saponins	+	+
	Quinones and anthraquinones	(-)	+
	Essential oil	+	+
Quantitative analysis (mg/g)	Phenols	7.5 4± 0.36	0.50 ± 0.06
	Flavonoids	29.25 ± 0.87	125.48 ± 3.66
	Alkaloids	2.33 ± 0.90	143.29 ± 2.80

Note: += present; (-) = absent

DISCUSSION

An insecticidal activity causing mortalities of 50.00% to 95.00% of *R. invadens* in 72 hours was obtained with the hydroalcoholic extracts of *H. suaveolens*. Toxicological studies also reported an LC 50 of 17.49 gl^{-1} . These results confirmed those of several authors. According to Thiam *et al.*, (2017) the aqueous extract of *H. suaveolens* at 187.5 gl^{-1} is effective against *H. armigera*. Extracts from this plant help to reduce losses (Johnson *et al.*, 2018). Some authors exhibited larvicidal activity against the larvae of the yellow fever mosquito *Aedes aegypti* (L) and *Aedes albopictus* (Conti *et al.*, 2012 and Tennyson *et al.*, 2018). But what explains these biological activities detected in *H. suaveolens*? In this study, a wide range of chemical compounds were isolated from this plant. These were polyphenols, flavonoids, tannins, alkaloids, saponins, leucoanthocyanins, quinones and anthraquinones and essential oil. Previous studies indicated that the biological activities are due to compounds such as alpha-pinene, beta-pinene, sabinene, terpinolene, betacaryophyllene, and 4-terpineol (Conti *et al.*, 2012). Phenolic compounds for example, which are the most abundant secondary metabolites synthesized by plants subjected to biotic and abiotic stresses to adapt and survive, are potentially toxic for many pests (Dai and Mumper, 2010). Furthermore, this study reports that extracts of *A. conyzoides* caused mortalities of 50.11% and 100% in 72 hours at 50 gl^{-1} with a CL 50 of 17.49 gl^{-1} . These data confirmed the results of several studies which have shown that the variety of *Ageratum* species has excellent biological properties. Arya *et al.* (2011) and Vasantharani *et al.* (2022) reported various biological properties, such as antifungal, cytotoxic, nematicidal,

antifeedant and larvicidal activities. These occurring activities could be related to the presence of various phytochemicals, such as bromophenols, polysaccharides, tannins, flavonoids, phenols, phenolic acids and carotenoids (Chandraker *et al.*, 2019; Puro *et al.*, 2018).

Phytochemical screening of leafy stem powder of *A. conyzoides* indicated the presence of secondary metabolites such as sterols and terpenes, polyphenols, flavonoids, catechic and gallic tannins, alkaloids and saponins. These results are not very similar to those recently reported by Chabi-Sika *et al.* (2023) but very similar to those of Okereke *et al.* (2017) and Ndacnou *et al.* (2020). These authors demonstrated in *A. conyzoides* the presence of various phytochemical compounds such as terpenoids, alkaloids, flavonoids, chromenes, coumarin, saponins, tannins, glycosides, phenols and resins. Variations in phytochemical profile would be due to the screening methodology, the nature, polarity and concentration of solvents, the collection area, the nature of the soil and the stage and organs of the plant used (Nawaz *et al.*, 2020, Lefebvre *et al.*, 2021). Phenolic compounds produced by plants contribute to their antioxidant capacity compared to other natural resources. Polyphenolic compounds such as flavonoids highlighted in this study, possess various biological activities including the formation of complexes with the extracellular wall (Ichino *et al.*, 2014).

CONCLUSION

This study aimed at evaluating the insecticidal effects of native plant extracts against the mango mealybug, *R. invadens*. Hydroalcoholic

extracts based on leafy stems of *A. conyzoides* and *H. suaveolens* at 50 gl⁻¹ have effective control against *R. invadens*. The one from *A. conyzoides* is more toxic to the mealybug. These plant extracts are rich in phytochemical compounds such as phenols, flavonoids, alkaloids, etc. which are natural resources with insecticidal properties. These active ingredients could contribute in effectively controlling mango mealybug in an integrated approach. Their application in mango orchards could increase fruit yields while preserving the ecosystems and environment of tropical areas. Not having been able to handle the young stages of the mealybug and deepen the chemical analysis, further studies should focus on the mechanism of action of each extract. Knowledge of the mechanism of action will help in the development of complexes of irresistible insecticide principles against *R. invadens* and other related homopteran insect pests.

Author contribution

Conceived, designed, and critically revised the study (M.E., S.F., S.A., D.A.R., N.S.); Collection and culture maintenance (S.A.K.D.D., S.A.); data analysis, interpretation and manuscript drafting (O.R.A., S.P.A.E.D.).

Conflict of interest

The authors declare that they have no conflict of Interests.

REFERENCES

- Abbott, S.W. (1925) A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology*, **18**, 265–267.
- Abd El-Rahman A. A., Aml, A. E., Rehab, E. M. E. S., Salah, M. E., & Helmy, E. H. (2021). Resistance of white mango scale by pesticide spraying using a mist blower and the effect of spraying process on fruits quality. *International Agricultural Engineering Journal*, **30**(2), 19-30.
- Ahmad, S. F., Ahmed, S., Khan, R. R., & Nadeem, M. K. (2010). Evaluation of insecticide resistance in two strains of fruit fly, *Bactrocera zonata* (Saunders) (Tephritidae: Diptera), with fruit dip method. *Pakistan Entomologist*, **32** (2), 163-167.
- Ainsworth, E. A., & Gillespie, K. M. (2007). Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin–Ciocalteu reagent. *Nature protocols*, **2**(4), 875-877.
- Akinyemi, K. O., Oladapo, O., Okwara, C. E., Ibe, C. C., & Fasura, K. A. (2005). Screening of crude extracts of six medicinal plants used in South-West Nigerian unorthodox medicine for anti-methicillin resistant *Staphylococcus aureus* activity. *BMC complementary and alternative medicine*, **5**, 1-7.
- Arya, N., Chaurasia, S., Shakya, A., Bharti, M., & Sahai, N. (2011). Efficacy of *Ageratum conyzoides* against the control of mosquitoes. *International Journal of Pharmaceutical Sciences and Research*, **2** (12), 3235.
- Azrag AGA, Mohamed SA, Ndlela S and Ekesi S (2023) Invasion risk by fruit trees mealybug *Rastrococcus invadens* (Williams) (Homoptera: Pseudococcidae) under climate warming. *Frontiers in Ecology and Evolution*. **11**:1-14.
- CABI (2022). "Rastrococcus invadens," in Invasive species compendium (Wallingford, UK: International). Available at: <https://www.cabi.org/iscCAB>.
- Chabi-Sika, K., Sina, H., Boya, B., Salami, H. A., A Dossou, G., Mama-Sirou, I., ... & Baba-Moussa, L. (2023). Ethnobotanical Survey and Some Biological Activities of *Ageratum conyzoides* Collected in Southern-Benin. *International Journal of Biochemistry Research & Review*, **32**(1), 9-25.
- Chandraker, S.K.; Lal, M.; Shukla, R. (2019). DNA-binding, antioxidant, H₂O₂ sensing and photocatalytic properties of biogenic silver nanoparticles using *Ageratum conyzoides* L. leaf extract. *Royal Society of Chemistry Advances*, **9**, 23408–23417.
- Chang, C. C., Yang, M. H., Wen, H. M., & Chern, J. C. (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *Journal of Food and Drug Analysis*, **10**(3).
- Ciulei I. (1982). Methodology for analysis of vegetable drug. Practical manual on industrial utilization of medicinal and aromatic plants, Bucharest, Ministry of Chemical industry, 67 p.
- Conti, B., Benelli, G., Flamini, G., Cioni, P. L., Profeti, R., Ceccarini, L., Macchia, M., & Canale, A. (2012). Larvicidal and repellent activity of *Hyptis suaveolens* (Lamiaceae) essential oil against the mosquito *Aedes albopictus* Skuse (Diptera: Culicidae). *Parasitology Research*, **110**, 2013-2021.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, **15**(10), 7313-7352
- Fall, A., Toure, M., Seye, F., Ndione, Rd, Seydou, T., Badiane, Bw, & Ndiaye, M. (2019). Evaluation of losses caused by *Rastrococcus invadens* (Williams, 1986) (Homoptera, Pseudococcidae) on mango in Senegal

- (Casamance). *Journal of Applied Biosciences*, **136**, 13854-13860.
- FAO. (2022). Start of the biological control program against the mango mealybug. <https://www.fao.org/burundi/actualites/detail-events/en/c/1492468/>
- Ferdous, G., & Jahan, M. (2020). Efficacy of insecticides and botanicals against the mango hopper *Idioscopus clypealis*. *Indian Journal of Entomology*, **82**(1), 13-15.
- Germain, J.-F., Laplace, D., Devarieux, A., Boavida, C., Gachet, E., & Goergen, G. (2015). *Rastrococcus invadens* Williams discovers America. *Phytoma* N° 686 août-septembre 2015, 4p
- Ichino, T., Fuji, K., Ueda, H., Takahashi, H., Koumoto, Y., Takagi, J., ... & Hara-Nishimura, I. (2014). GFS 9/TT 9 contributes to intracellular membrane trafficking and flavonoid accumulation in *A. rabidopsis thaliana*. *The Plant Journal*, **80**(3), 410-423.
- Johnson, F., Oussou, RK, Kanko, C., Tonzibo, FZ, Foua-Bi, K., & Tano, Y. (2018). Bioeffectiveness of Essential Oils of Three Plant Species (*Ocimum gratissimum*, *Ocimum canum* and *Hyptis suaveolens*), of the Labiaceae Family in the fight against *Sitophilus zeamais*. *European Journal of Scientific Research*, **150** (3), 273-284.
- Lefebvre, T., Destandau, E., & Lesellier, E. (2021). Selective extraction of bioactive compounds from plants using recent extraction techniques: A review. *Journal of Chromatography A*, 1635, 461770.
- Mano, E., & Nana, J. (2022). Bio-effectiveness and safety of bioinsecticides based on powders of *Balanites aegyptiaca* L. Drel. and *Moringa oleifera* Lam. on *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae) and its natural enemies. *Journal of Applied Biosciences*, **175** (1), 18171-18181.
- Mano, E., Simde, R., Kambou, G., Ouedraogo, B., & Somda, I. (2022). Potential for natural regulation of the moth *Helicoverpa armigera* (Hübner 1808) (Lepidoptera: Noctuidae) by its procession of parasitoids under organic tomato cultivation conditions. *Journal of Applied Biosciences*, **173** (1), 17953-17962.
- Mohamed, S. A., Dubois, T., Azrag, A. G., Ndlela, S., & Neuenschwander, P. (2022). Classical biological of key horticultural pests in Africa: successes, challenges, and opportunities. *Current Opinion in Insect Science*, **53**, 100945.
- Nawaz, H., Shad, M. A., Rehman, N., Andaleeb, H., & Ullah, N. (2020). Effect of solvent polarity on extraction yield and antioxidant properties of phytochemicals from bean (*Phaseolus vulgaris*) seeds. *Brazilian Journal of Pharmaceutical Sciences*, **56**, e17129.
- Ndacnou MK, Pantaleon A, Tchinda JS, Mangapche ELN, Keumedjio F, Boyoguemo DB. (2020). Phytochemical study and anti-oomycete activity of *Ageratum conyzoides* Linnaeus. *Industrial Crop Products*, **153**:112589.
- Nébié, K., Nacro, S., Otoïdobiga, L. C., & Somda, I. (2018). Host plants of the mango mealybug *Rastrococcus invadens* Williams (Homoptera: Pseudococcidae) in Western Burkina Faso. *International Journal of Agriculture and Environmental Research*, **4**(4), 891-901.
- Okereke, S. C., Chukwudoruo, C. S., & Nwaokezie, C. O. (2017). Phytochemical screening using GC-FID and sub-chronic assessment of Hydroethanolic leaf extract of *Ageratum conyzoides* Linn. on albino rats. *Journal of Medicinal Plants Studies*, **5**, 282-287.
- Puro, K.N.; Mazumder, M.U.; Khazeo, P.; Jyrwa, R.; Jamir, N.; Sailo, L. (2018). Qualitative and quantitative analysis of phytochemicals of crude extracts of *Ageratum conyzoides* L. leaves. In Proceedings of the Mizoram Science Congress (MSC 2018), Aizawl, India, 4–5 October 2018; pp. 164–168.
- Shawan, S. I., Rashed, R. U., Mitu, A. S., & Jahan, M. (2018). Efficacy of different chemical and botanical insecticides in controlling mango hopper (*Amritodusatkinsoni* L.). *Advances in Plants & Agriculture Research*, **8**(2), 127-131.
- Tennyson, S., Arivoli, S., Raveen, R., Selvakumar, S., Jayakumar, M., & Kumar, L. (2018). Bioefficacy of *Catharanthus roseus* (L.) G. Don (Apocyanaceae) and *Hyptis suaveolens* (L.) Poit (Lamiaceae) ethanolic aerial extracts on the larval instars of the dengue and chikungunya vector *Aedes aegypti* Linnaeus 1762 (Diptera: Culicidae). *International Journal of Mosquito Research*, **5**, 7-18.
- Thiam, A., Mbodji, M., Samba, S. A. N., Bopp, S., & Diatta, M. (2017). Effect of the aqueous extract of *Hyptis suaveolens* on *Lycopersicon esculentum* M. for the control of *Helicoverpa armigera* in the hot off-season in Senegal. *Africa Science*, **13** (6), 76-86.
- Vasantharani, S., Thirugnanasampandan, R., & Bhuvaneshwari, G. (2022). Qualitative and quantitative analysis of Precocene II, estimation of enzymatic, nonenzymatic antioxidant, and cytotoxic potentials of methyl jasmonate-elicited shoot culture of *Ageratum conyzoides* Linn. *Journal of Applied Biology and Biotechnology*, **11**(1), 93-99.
- Williams D.J. et Granara DE Willink M.C., (1992). Mealybugs of Central and South America. CAB International, London, England. 635 pp.
- Williams D.J., (1989). The mealybug genus *Rastrococcus* Ferris (Hemiptera: Pseudococcidae). *Systematic Entomology*, **14**: 433-486.