



Comparative Analysis of Oil Obtained from Moringa and Neem Seeds

Garba, L.1*, Mahmoud, A.A.1, Boryo, D. E. A.1, Abubakar, A.1 and Abdullahi Z. A.2

¹Department of Chemistry, Abubakar Tafawa Balewa University Bauchi, Nigeria

²Department of Chemistry, Gombe State University Gombe, Nigeria

ABSTRACT

This study reports the extraction of oil from moringa and neem seeds using soxhlet extractor and the production of biodiesel from the two oils extracted and compared with fossil diesel in terms of physico-chemical properties and cost of production. The biodiesels were produced by transesterification reaction with alcoholic KOH at 90°C temperature. From the results obtained it was found that the yield of biodiesel from moringa seed oil (48%) is less when compared with that neem seed oil (52%). It was found that the chemical properties of moringa and neem oils were, such as saponification, (196, 182) iodine value, (62.51, 83.4) specific gravity (0.71, 0.94) peroxide value, (1.22, 1.07) acid value, (0.62, 0.47) density, (0.72, 0.81) viscosity, (2.3, 4.2) when compared with the ASTM D 445 standard limit. The physical properties of the raw extracted oil and the biodiesel produced for moringa, neem and fossil diesel for flash point (149°C, 111°C, and 97°C), pour point (8°C, 5°C and 7°C) and cloud point (16°C, 9°C and 7°C) comparatively with the standard fossil fuel. The FT-IR result revealed the presence of SP³ C-H (stretch) at 2923.81 cm⁻¹, C=O stretching vibration at 1746.67 cm⁻¹ indicating the presence of carbonyl group, stretch vibrations at 3007.32 cm⁻¹ showing the presence of aromatic SP² C-H. Also HC=O stretching at 2853.67 cm⁻¹ was observed indicating the presence of aldehyde, the presence of HC =CH bend at 667.24 cm^{-1} indicates the presence of alkenes. The peak observed at 3564.19 cm⁻¹ indicates the presence of O-H, the peak observed at 1195.50 cm⁻¹ was assigned to C-O bend indicating the presence of carboxylic acid/esters. The moringa biodiesel contain more of fatty acids and the neem biodiesel contain more of esters due to esterification of the neem oil as revealed by through GC-MS spectrum of the neem oil. The neem oil cannot contain fatty acid due to the absent of the O-H (stretch) band on its FT-IR spectrum. The relative quantification was obtained from the chromatogram of both oil in GC-MS spectrum. The result showed that neem oil could be uses as an alternative to fossil diesel if large scale of production is employ.

Keywords: FTIR, Biodiesel, Oil, Fossil

INTRODUCTION

In this age of climate change, a lot of attention has been paid to environmental degradation, global warming, and the ever-depleting fossil fuel resources. These have developed to become important global concerns, and Numerous techniques have been proposed to decrease the negative repercussions of emissions from fossil fuels (Moyo *et al.*, 2021; Abubakar *et al.*, 2022). The most effective fuel to replace diesel is thought to be biodiesel, which is clean and renewable (Latchubugata et al., 2018). Nowadays, biodiesel may be utilized as a sustainable option that considers both environmental and economic aspects. It is a less damaging, biodegradable fuel that comes from renewable sources (Yunus et al., 2013). This biofuel has several advantages over conventional diesel fuel, including a high cetane number, low smoke and particle carbon dioxide emissions, low and hydrocarbon emissions (Kumar and Kant, 2013). Carbon (IV) oxide, smoke, and unburned hydrocarbon emissions are all decreased. Vegetable oil's high density,

Bima Journal of Science and Technology, Vol. 8(1B) Apr, 2024 ISSN: 2536-6041



DOI: 10.56892/bima.v8i1.624

viscosity, low calorific value, and nonvolatility cause atomization issues, pumping challenges, and poor combustion inside the diesel engine's combustion chamber (Abubakar *et al.*, 2022).

By chemically reacting vegetable or animal fat with an alcohol like methanol, biodiesel is created as a substitute fuel for diesel engines (Abubakar *et al.*, 2020). Because it can be used in any compression ignition engine without modification, biodiesel has recently been recognized as the best alternative for replacing diesel fuel. Compared to diesel fuel, biodiesel has several advantages: it is liquid, portable, renewable, and contains less sulfur and aromatics. It is also more biodegradable (Demirbas, 2009; Abubakar *et al.*, 2022).

A tropical plant that thrives throughout the tropics, moringa is a member of the Moringaceae family. Out of the thirteen species in the genus Moringa, only one, Moringa oleifera, has received research and development attention. In antiquity, it held great worth. The oil from the seeds was extracted by the Romans, Greeks, and Egyptians, who used it for skin lotion and perfume. Plantations growing Moringa in West India exported oil to Europe in the 19th century for use in industrial lubricants and perfumes. Native to northern India's sub-Himalayan regions, M. oleifera is also known as the "drumstick tree" or "orseradish tree." Because of its many uses in agriculture, industry, and medicine, moringa is a versatile tree with enormous economic value (Yisehak et al., 2011).

Neem seed, derived from the neem tree (Azadirachta indica), is well-known for its versatile applications in a variety of industries, particularly because of its high oil content (Eid, & Elmarzugi, 2017). Neem oil, extracted from the seeds, contains a variety of biologically active compounds, including

azadirachtin, nimbin, and nimbidin, all of which have potent pesticidal, insecticidal, and medicinal properties (Kumar, et al., 2018). Neem oil has been shown in studies to have significant antimicrobial and antifungal properties, making it a valuable ingredient in agricultural practices and pharmaceutical formulations (Eid, & Elmarzugi, 2017). Furthermore, its high fatty acid content, which includes oleic acid, linoleic acid, and palmitic acid, makes it ideal for skincare and hair care products (Gupta, et al., 2019). The oil's diverse array of bioactive constituents highlights its significance in both traditional medicine and modern scientific research, positioning neem seed oil as a versatile resource with enormous potential across various domains (Gupta, et al., 2019). Neem trees can grow up to 800 meters above sea level and tolerate mean annual rainfall of 450-1200 mm along with typical temperatures of 25-35 °C. The objective of this study is to synthesize, evaluate, and contrast biodiesel derived from the oils of trees (Azadirachta indica) and neem drumsticks (Moringa oleifera).

MATERIALS AND METHODS

Chemicals and Reagents

All of the chemicals and reagents utilized in this study were of analytical grade. N-hexane, Acetic acid, Chloroform, hydroxide, Potassium iodide, Sodium thiosulphate, Iodine and Phenolphthalein

Sample Collection and Preparation

Fresh seeds of Neem and Moringa were collected from Arawa Gombe in Gombe State and were identified at the Department of Botany by a botanist in Gombe State University, Gombe. After three rounds of distilled water washing, the samples were dried for three weeks in the shade and then stored for later use.





Soxhlet Extraction

With minor modifications, the procedure outlined by Petal et al. (2011) and Akbar et al. (2009) was employed. Weighed and put into a Soxhlet extractor was 500.00 g of the ground seed. With n-hexane serving as the solvent,

Production of Biodiesel Fuel

One gram of potassium hydroxide was weighed and placed in a 250cm³ conical flask. A 120 cm³ of methanol was added to the KOH pellets and then stirred until all the solid catalysts were dissolved using a magnetic stirrer while heating at about 60 °C on a hot plate. About 20.00 cm³ of the oil was measured using a measuring cylinder and heated to 60 °C for about 10 minutes. Then the 10 cm^3 potassium methoxide was quantitatively added into the heated oil. The mixture was agitated vigorously for about one minute in conical flask. After agitation, the mixture was continuously and gently stirred

Properties of Free Fatty Acids Methyl Esters (FAMEs)

Jauro, A. and Haruna, A.M. (2011) and AOAC (2023) methods were adopted with slight modification for the analysis of FAMEs unless otherwise stated.

Determination of free fatty acid (FFA)

A 1.00 g of Moringa and Neemseed oils were measured separately and placed into a 250 $FFA = \frac{Titre value x M x 28.2}{Titre value x M x 28.2}$

Weight of sample

Where: $M = \text{concentration of the base in mol/dm}^3$ of the base, 28.2 is a constant

Determination of the peroxide value of the oil

One (1.00 g) of Moringa and Neemseed oil were weighed separately into a 250 cm³ conical flask and 30.00 cm³ of glacial acetic

acid/chloroform (3:2 V/V) was added. The mixtures were shaken until they dissolved. Saturated potassium iodide (1 cm³) was added then followed by the addition of 0.50 cm³ starch indicator.

the oil extraction process for eight hours. To eliminate any potential residual solvent in the oil extracted, the extracts were heated to 70 °C in an oven and allowed to air dry. The oil was weighed, and using a dry matter method, the percentage of oil yield was determined as follows:

Percentage of oil yield $=\frac{1}{\text{weight of sample on dry matter basis}}$ weight of oil - ×100 -1

with a magnetic stirrer for 2 hours. The mixture was then transferred into a separating funnel that was clamped on a retort stand for 24 hours. After separation the lower layer was decanted off leaving the top biodiesel layer. The next step was washing of the biodiesel. This was carried out to remove excess methanol, KOH, soap and escaped glycerol. Distill water was progressively swirled into the crude product. This was allowed to stand for two hours. The lower water layer was run off, resulting in a clear biodiesel product. The resulting biodiesel was golden yellow in colour. The biodiesel yield was calculated using:

Oil recovery = $\frac{\text{Volume of biodiesel collected} \times 100}{\text{Volume of biodiesel collected} \times 100}$ 2 Volume of sample oil

cm³ conical flask and warmed. Methanol (10.00 cm³) was added thorough stirring followed by 2 drops of phenolphthalein indicator. The content was titrated with 0.20 mol/dm³NaOH until a light pink colour which persisted for 1 minute was seen.

The end point was recorded and used in calculating the FFA as follows:

-1



This was titrated with 0.20 mol/dm³ Na₂S₂O₃ until the dark blue colour just disappeared. Blank determination was also carried out and the peroxide value was calculated as follows:

 $P.V = \frac{(S-B)x 1000 x C}{1000 x C}$

_ -2

Where S= titre volume of sample, B= titre volume of blank, C= concentration in mol/dm³ of sodium thiosulphate solution, W= weight of oil sample, 1000 is a constant.

Determination of refractive index

The refractive index of the oils were determined using refractor meter by exposing the oil to the meter.

Determination of specific gravity of biodiesel

A makeshift specific gravity bottles were cleaned, soaked in acetone, and dried in an oven. The bottles were placed in a desiccator

Specific grav

Where; W_1 = bottle weight + oil, W_2 = empty bottle weight, W_3 = equal volume of water + bottle weight.

Determination of density of the biodiesel

A makeshift bottle for specific gravity was cleaned and rinsed with acetone before being oven dried. In a desiccator, the bottles were allowed to cool to room temperature, and the

> (weight of bottle+oil)-weight of empty bottle Density =

Volume ofoil Determination of kinematic viscosity of the biodiesel

An innovative approach was utilized to determine this parameter as reported by

Kinematic viscosity =

Determination of pH of the biodiesel

The biodiesel (10.00 cm³) was measured into a 50 cm^3 beaker, the electrode of the meter was cleaned and dried, it was then placed into the sample and the measurement was recorded. (Schinas et al., 2008).

Determination of flash point

This determination was made using an improvised method. A 50 cm³ beaker was filled to a certain level (10.00 cm³) with and let to cool to room temperature. After, the weight of the washed bottles were observed. The weights of the water-filled bottles were calculated. After that, the water was transferred into a beaker, acetone was then used to cleaned the bottle, and it was dried. The specific gravity of the biodiesel sample was measured by repeating the same procedure

$$vity = \frac{W1 - W2}{W3 - W2} - -3$$

empty bottle's weight was calculated using a weighing scale. The weight of the bottle containing biodiesel was recorded. The volume of the oil in the bottle was also measured, and the density was calculated as follows: (Ugah et al., 2007).

_4

(Abubakar et al., 2020). The biodiesel was poured into the clamped burette and allowed to drain freely into a beaker. The time of flow was observed, and the kinematic viscosity was determined as follows: volume of the burette

Time biodiesel, which was then heated on the hot plate at a consistent rate. The flash point was defined as the lowest temperature at which the test flame ignited the vapour above the sample. It was repeated three times, and the average was determined (Abubakar et al., 2020).

-5

Determination of cloud point

This parameter was estimated using an improvised method. A cylindrical test tube was filled with biodiesel to a certain level



(10.00 cm³) and clamped with a clamp containing a thermometer. The test tube was placed in the ice bath, and the setup was checked periodically for cloud formation. The cloud point was determined by the temperature at which a definite cloud developed at the bottom of the test tube (Abdulkareem *et al.*, 2012).

Determination of pour point

This parameter was estimated using an improvised method. A cylindrical test tube was filled with biodiesel to a certain level

Determination of cetane number (ASTM D 613)

 (10.00 cm^3) and secured with a clamp that

held the thermometer. In the ice bath, the

sample was allowed to drop to temperatures lower than 0 °C. At this point, it was removed

and tilted on the clamp while the setup was

observed periodically. The pour point was

observed by the lowest temperature at which

the biodiesel was flow (Abubakar et al., 2020).

Cetane Number: The cetane number was calculated as

$$CN = \frac{46.3 + 5458 \, \text{X I.V}}{\text{S.V} - 0.225} - -6$$

Where: CN = Cetane Number, SV = Saponification value, IV = Iodine value (Dagde, 2019).

FT-IR

SHIMADZU FTIR-8400S FTIR machine was used for the Infrared analysis of the fossil fuel and the biodiesel obtained.

Parameters	ysicochemical prope Neem Oil	Moringa Oil	Standard Values	Limit
Moisture content (%)	2.3 ± 0.33^{a}	2.6 ± 0.32^{a}	Standard values	Linit
Colour	Golden-yellow	Golden-black		
Oil Yield (%)	52 ± 0.2^{a}	48 ± 0.01^{b}		
Saponification value (mgKOH/g)	162 ± 0.3 ^a	$196\pm0.1~^{\rm b}$		189-198
Iodine value $(gI_2/100 g)$	$104\pm0.02~^{a}$	$98\pm0.04~^{\rm b}$	ASTM D4607max	≥130
FFA (mgKOH/g)	0.51 ± 0.09 $^{\rm a}$	1.09 ± 0.043 $^{\rm b}$		25
Refractive index (35 °C)	1.73 ± 0.01 a	1.28 ± 0.01 a		
Specific gravity	0.94 ± 0.01 $^{\rm a}$	0.71 ± 0.01 a	ASTM D6751	0.92
Peroxide value (meq/Kg)	1.07 ± 0.02 a	1.22 ± 0.02 a		2-10
PH value	6.79 ± 0.3 a	6.41 ± 0.2 a		
Acid value (mgKOH/g)	0.47 ± 0.2 a	0.62 ± 0.01 ^b	ASTM D6751	0-0.8
Density (g/cm ³)	0.80 ± 0.01 a	0.86 ± 0.01 $^{\rm a}$	ASTM D1298	0.86-0.90
Viscosity 35 °C (cm ³ /sec)	2.3 ± 0.1 a	4.9 ± 0.3 ^b	ASTM D445	1.9 - 6.0

RESULTS AND DISCUSSION

Key: a = significant difference, b = no significant difference.







Figure 1: Pictures of the obtained oil from Moringa seed



Figure 2: Pictures of the obtained oil from neem seed

Results of the physicochemical characteristics of Neem and Moringa seed oils were presented in Table 1. The colour of the neem seed oil is golden-yellow while Moringa seed oil is golden-black in colour as shown from the figure 1 and 2 above and had the viscosity of (2.3 mm²/s and 4.2 mm²/s) for neem and moringa oil. The percentage yield of the oils where 52% and 48% for Neem and Moringa seeds oil respectively, this is larger than the percentage yields of Z-spinachisti (Nabag) seed crude oil. (28.96) as reported by Saeed and Abbas (2016), J. gossvypiifolia and Jatropha curcas L. (23.04 and 31.00) respectively as reported by Jefferson et al. (2009), and mustard seed oil (44.67), canola seed oil (43.87) corn seed oil (4.23) cotton seed oil (17.83) as reported by Arif et al. (2012) and it is within the A.O.A.C 1990. This high percentage oil yield indicates its potential use in detergent/ soap making industries and edible purposes (Abubakar et al., 2020).

The saponification value of the Neem seed oil is 162.00 mgKOH/g, which is lower than that of Moringa oils 196 mgKOH/g. This is lower than Jatropha curcas seed oil (212.67 mgKOH/g) as reported by Abubakar et al. (2022). The Neem oil has showed a lower saponification value than Bebra seed oil (184.95 mgKOH/g) and Terminalia mantaly seed oil (196 mgKOH/g) as reported by Abubakar et al. (2020), and Z-spinachisti (Nabag) seed crude oil (183.39 mgKOH/g) as reported by Saeed and Abbas (2016). Moringa has a greater saponification value than sesame seed (192.70 mgKOH/g) and watermelon seed (192.09 mgKOH/g) as reported by Saeed and Shola (2015). Moreover, lower than bitter kola seed oil (229.45 mgKOH/g) and melon seed oil (247.0 mgKOH/g) as reported by Saeed and Shola (2015). This shows that there is a significant difference between mean values of Moringa and Neem oils. Hence, the lower the saponification, the higher the propensity of the biodiesel; conversely, the higher the saponification value, the lower the



CONTRACTOR OF THE OWNER

DOI: 10.56892/bima.v8i1.624

biodiesel; this suggests that moringa is preferable to neem in the creation of biodiesels.

The iodine value of Neem and Moringa were 104 $gI_2/100g$ and 98 $gI_2/100g$ respectively, which is higher than bitter kola 53.99 gI₂/100g, Terminalia Catappa (54.567 gI₂) /100g) and 44.4 gI₂/100g cashew nut oil as reported by Aremu et al. (2006), and J. curcas L. (40.91 gI₂/100g) as reported by Abubakar et al. (2020). On the other hand, the Iodine value of this study is lower than the range of soyabean (128-143 $gI_2/100g$) as reported by Karmakar et al. (2017). The iodine value could be used to assess the quantity of double bond present in the oil, indicating the oil's vulnerability to oxidation. Oils with iodine value less than 100 $gI_2/100g$ of oil are nondrying oils, correspondingly, Aremu et al. (2006a), A good drying oil should have an iodine value of 130 or higher (Ebenezer, 2015). Biodiesel made from the same oil should have identical iodine values (Encinar 2010). It is related to the chemical composition of the fuel. A higher iodine value suggests more unsaturated fats and oils. The standard iodine value for biodiesel is 120, according to European EN 14214 regulations. This requirement is limited by the standard limits of linolenic acid methyl ester composition for biodiesel. The limiting of unsaturated fatty acids is important because heating more unsaturated fatty acids causes polymerization of glycerides. This can result in deposits or impairment of the lubricating property. Fuels with this property are also likely to form thick sludges in the sump of the engine, as fuel leaks down the sidewalls of the cylinder into the crank (Refaat 2010). As a result, there is a significant discrepancy in the average values of Neem and Moringa oils.

Moringa oil had a higher viscosity $(2.3 \text{ cm}^3/\text{sec})$ than neem oil $(4.9 \text{ cm}^3/\text{sec})$. Viscosity describes a fluid's internal resistance to flow and can be interpreted as a measure of fluid friction in optics to determine the rheological properties of these oils. When the temperature rises, the viscosity drops exponentially. Viscosity increased with molecular weight, but declined with rising unsaturation level and temperature (Akbar *et al.*, 2009).

In this study, the acid value of Neem oil was 0.47 mgKOH/g, which is greater than 0.62 mgKOH/g for Moringa and lower than 13.76 mgKOH/g for J. curcas as reported by Abubakar et al (2022). This is higher than T. mantaly seed oil (0.56 mgKOH/g) and Bebra seed oil (0.052 mgKOH/g), as reported by Abubakar et al (2020). The acid value of Neem seed obtained from this study is higher than that of Terminalia belerica seed oil (3.69 mgKOH/g) and Sunflower oil (0.5-5.0 mgKOH/g), but similar to Lin seed oil (6.0 mgKOH/g) by Hossain et al (2007). The acid value of Neem surpasses the limit of A.O.A.C. Standard of <4.0, but Moringa is within the range. According to Aremu et al. (2015), high acid value in oil indicates that the oil may not be suitable for use in cooking (edibility), but could be beneficial for the creation of paints, biodiesel, liquid soap, and shampoo (Aremu et al., 2006a).

The peroxide value of Neem oil was found to be 1.07 mEq/Kg in this study, which is lower than 1.22 mEq/Kg for Moringa seed oil and lower than that of T. mantaly (4 mEq/Kg) and Terminalia catappa (2.60 mEq/Kg), as reported by Abubakar et al (2020). And Malaysian and Indian Jatropha (1.93 mEq/Kg) and (3.70 mEq/Kg), and similar to (1.08 for Thailanian mEq/Kg) Jatropha, respectively reported by Emil et al. (2010), and also lower than that of Moringa seed (5.82 mEq/Kg), sesame seed (8.33 mEq/Kg). Bitter kola (10.22 mEq/Kg), Melon seed (7.19 mEq/Kg), and water melon seed (13.41



mEq/Kg). The value meets the AOAC 1990 standard (Saeed and Shola, 2015).

The peroxide value is the most prevalent biomarker of lipid oxidation. Unrefined vegetable oils have higher PV than refined oils. High PVs indicate high levels of oxidative rancidity in the oils, as well as the lack or low quantities of antioxidants (Aremu et al., 2015).

In this study, the density of Neem seed oil is 0.81 g/cm3, which is greater than that of Moringa oils (0.72 g/cm3), but lower than the density of Jatropha podagrica seed oil (0.90 g/cm3) reported by Premjet et al. (2021), Tropical almond seed oil (0.90 g/cm3) reported Orhevba et al. (2016) Terminalia mantaly seed oil (0.92 g/m3) reported by Abubakar et al. (2020); Jatropha curcas seed oil (0.90 g/cm3) reported by Emil *et al.* (2010)

soybean and sunflower (0.91 g/m3 and 0.92 g/m3, respectively), as well as the Bebra seed oil density of 0.94 g/m3 reported by Akpan et al. (2007). Vegetable oil has a lower density than water, and the variances amongst vegetable oils are minimal, especially among the most common vegetable oils. Generally, the density of oil decreases with molecular weight while increasing with unsaturation level (Gunstone, 2004; Ebenezer, 2015).

Neem oil has a specific gravity of 0.94, which is greater than Moringa seed oil's 0.71, but lower than that of sorrel and okra seed oils (1.00 and 0.90, respectively) as reported by Umar *et al.*, (2020), and similar to Bebra seed oil (0.93) and cashew nut (0.96) reported by Aremu *et al.*, (2006) and castor seed oil (0.95) reported by Andualem and Gassesse (2014) but similar to that of Tropical almond oil (0.89) reported by Orhevba *et al.* (2016).

Table 2: Physico-chemical	l properties of moringa	biodiesel. Neem biodi	esel and fossil diesel.

Properties	Moringa biodiesel	Neem biodiesel	Fossil diesel	Standard Values	Limit
Flash point (°C)	149 ± 7.0 ^a	111 ± 4.0 ^b	97 ± 4.0 ^b	ASTMD 93	100 - 170
Pour point (°C)	8 ± 0.5 a	5 ± 0.3 b	3 ± 1.1 b	ASTMD 97	15 - 16
Cloud point (°C)	11 ± 0.1 a	9 ± 0.2 b	7 ± 1.21 ^b	ASTMD 2500	3 - 12
Calorific value	39.7 ± 3.5 $^{\mathrm{a}}$	25.1 ± 2.7 ^b	$46.4\pm2.9~^{\rm a}$	-	-
(mg/kg)					
Viscosity at 35°C (cm ³ /sec)	$4.8\pm0.3~a$	$4.1\pm0.1\ b$	3.0 ± 1.0 c	ASTMD 445	1.9 - 6.0
Density (g/cm ³)	0.87 ± 0.6 ^a	$0.86{\pm}~0.8$ a	0.89 ± 0.7 a	ASTMD 1298	0.88-0.90
Carbon residue	7.1 ± 0.3 $^{\rm a}$	5.0 ± 0.2 $^{\rm a}$	10.0 ± 4.2 b	ASTMD 524	15 - 10
(wt%)					
Cetane number	39 ± 0.5 a	43 ± 0.4 a	30 ± 0.1 b	ASTMD 613	47min

a = significant difference, b = no significant difference.

Moringa biodiesel has a flash point of 149 °C, which is greater than that of neem biodiesel and fossil diesels, which are 111 °C and 97 °C, respectively, which is lower than the 152 °C reported for J. curcas biodiesel by Abubakar et al (2020). Moringa has a greater flash point than Lageneria sicerria seed oil, according to Mahmoud et al. (2020), high-speed diesel (60-80 °C). Non-edible based seed oils have a greater flash point than fossil diesel (Kannahi and Arulmozhi, 2013). This demonstrates that there is a significant difference in the mean values of Neem and Moringa biodiesels, but no significant difference between Neem and fossil diesel at the 95% confidence level.

In this study, the pour point of Neem biodiesel was 8°C, higher than Moringa and Fossil biodiesels, which were 5°C and 3°C, respectively. This is greater than 0°C for J. curcas biodiesel, as reported by Lagenaria



siceria biodiesel, 0°C, and -6°C and -9°C for castor oil biodiesel as reported by Mahmoud et al. (2020). The pour points of all biodiesels were almost within the desired range, although the conventional ranged from (Abubakar et al., 2020). This demonstrates that there is a significant difference in the mean values of Neem and Moringa biodiesels, but no difference between Neem and fossil biodiesels at the 95% confidence level.

Neem biodiesel, Moringa biodiesel, and Fossil biodiesel had cloud points of 11°C, 9°C, and 7°C, respectively, Abubakar et al. (2020) recorded a temperature of 4.56°C for J. curcas biodiesel, while Mahmoud et al. (2020) reported a temperature of 4°C for L. siceria biodiesel. All of the cloud points were within the standard, while the usual runs from -3 to -12, (Abubakar et al., 2020). This indicates that there is a significant difference between the mean values of Neem and Moringa biodiesels, but no difference between Neem and fossil biodiesels at the 95% confidence level.

The viscosity of Neem in this study is 2.1 cm³/sec, which is lower when compared to that of Moringa and fossil diesel 4.8 cm³/sec and 3.0 cm³/sec respectively, and these are lower than 6.50 cm^3/sec as reported for L. siceria biodiesel as reported by Mahmoud et al. (2020) and higher than the viscosity of conventional D2 (at 40 °C) is between 1.9 and 6.0 cm³/sec (ASTM, 2012). This demonstrates that there is a significant difference in mean values between Neem, Moringa, and fossil biodiesels at the 95% confidence level.

The density of Neem was determined to be 0.86 g/cm3, which is lower than that of Moringa and fossil diesels (0.87 g/cm3 and 0.89 g/cm3, respectively). This is lower than 0.96 g/cm³ for T. catappa biodiesel reported by Orhevba et al., (2016), but close to 0.89 g/cm³ reported by Jauro and Haruna (2011). It has a density similar to diesel (0.845 g/cm3) but is less dense than water. It can be used as an alternative fuel because it meets the ASTM criteria for biodiesel (0.86 - 0.92) g/cm³. This indicate that there is no significant difference in mean values between Neem, Moringa, and fossil biodiesels at the 95 percent confidence level.

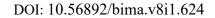
The cetane number (CN) determined in this study was 63 for Neem biodiesel, which is higher than 59 and 41 for Moringa and Fossil biodiesels, respectively. Moringa values are greater than those of avocado biodiesel 52.20 published by Dagde (2019), while Neem and Moringa values are higher than 51.70 reported for T. catappa biodiesel by Orhevba et al (2016). This demonstrates that there is no significant difference between the mean values of Neem and Moringa, but there is a significant difference between Neem and fossil biodiesels, as well as Moringa and fossil diesel, at the 95% confidence level. The cetane number is a measure of the fuel's ignition delay. Higher cetane levels suggest shorter periods between fuel injection and ignition.

FT-IR ANALYSIS

Neem and moringa biodiesel samples were analysed using FT-IR. The following functional categories were discovered in neem and moringa.



Table 3: Functional groups present in neem biodiesel			
Wave number (cm ⁻¹)	Types of vibration	Nature of functional Group	
3500.04	O-H (Bonded)	Carboxylic/alcohol	
1746.67	C=O (stretch)	Aldehyde/ketone /ester	
2853.67	H-C=O (stretch)	Aldehyde	
2923.81	C-H (Stretch)	Alkanes	
3007.32	C-H (stretch)	Aromatic	
1463.79	C-H (bend)	Alkanes	
1163.48	C-O (stretch)	Esters / alcohol / ethers	
667.24	C-H (bend)	Alkynes	



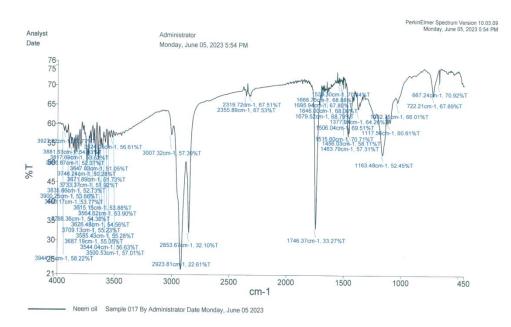


Figure 3: FTIR spectrum of Neem biodiesel

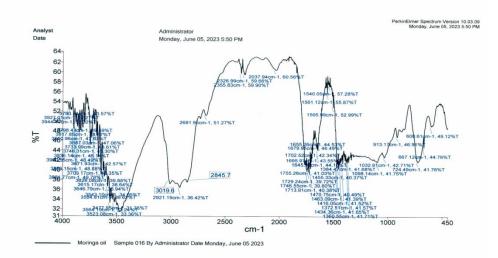
The FT-IR results were given in table 3 and the FTIR spectrum on figure 3. The FT-IR measurements show the presence of C-H (stretch) at 2923.81 cm⁻¹ for alkanes. The C=O stretching vibration at 1746.67 cm⁻¹ indicates the existence of esters/ketones. C-H stretch vibrations at 3007.32 cm⁻¹ indicate the existence of aromatics. C-H bending at 1463.79 cm⁻¹ suggests the existence of alkanes. Aldehyde is indicated by HC=O stretching at 2853.67 cm⁻¹, while alkenes are indicated by HC=CH bend at 667.2 cm⁻¹. This result was consistent with the literature results C-H stretch (~2900 cm-1), C=O stretch (~1700 cm-1), broad O-H stretch (~3400 cm-1) and C-O stretch (~1100cm-1) 886 cm⁻¹, 1436 cm⁻¹ and 1644 cm⁻¹ as reported by Snezana *et al.*, (2020). Also absorption were observed at 2923.81 cm⁻¹ indicating alkyls (CH₃, CH₂, CH), 1722.68 cm⁻¹ indicating ketone, 1598.10 cm⁻¹ indicating alkenes and 1374.83 cm⁻¹Methyl CH₃ (Ogwuche and Edema, 2020). And also similar to what has been reported by Elkady *et al.*, (2015) 721 cm⁻¹ -CH₂ rocking, 1373.7 cm⁻¹ Bending vibrations of CH₂ groups, 1745 cm⁻¹ C=O ester stretch, 1163 cm⁻¹ C-O stretching, 1456 cm⁻¹ Bending vibrations of the CH₂, 2925 -CH₂ stretching.

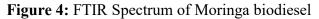




Table 4:	Functional	groups	present	in <i>moringa</i>	<i>v</i> biodiesel

Wave number (cm ⁻¹)	Types of vibration	Nature of functional group
3564.19	O-H (stretching)	Phenol/ alcohol
2681.95	HC=O (stretch)	Alkanoic acid/ aldehyde
3019.6	C-H (stretch)	Alkanes
1755.26	C=O (stretch)	Esters
1463.09	C-H (bend)	Alkane
1372.51	C-H (rock)	Alkanes
1169.52	C=C (stretching)	Alkenes
2845.7	C-H (stretch)	Alkanes



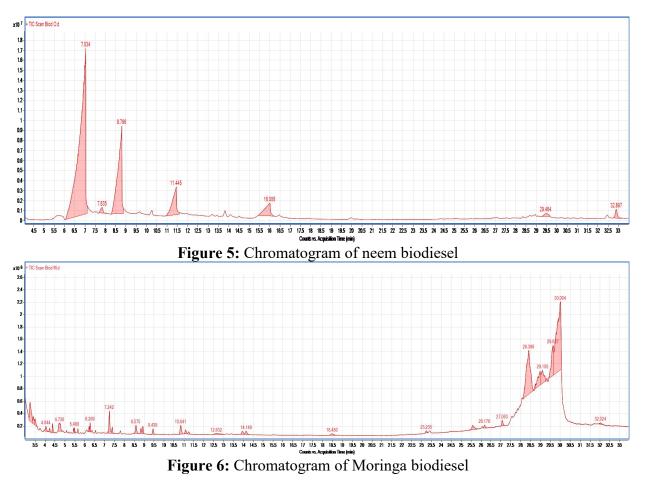


The FT-IR data were presented in Table 4 and the FTIR spectrum on figure 4. The signal at 3564.19 cm⁻¹ indicates the existence of Stretching phenol/alcohol O-H. The HC=O (stretching) at 2681.95 cm⁻¹ indicates the existence of aldehyde. The C-H stretching vibration at 3019.6 cm⁻¹ suggests the existence of alkanes. C-H bending vibrations at 1463.09 cm⁻¹ indicate the existence of alkanes, whereas C-O bend at 1195.50 cm⁻¹ indicates the presence of carboxylic acid, esters/alcohol. C=C stretch at 1169.52 cm⁻¹ indicates the presence of alkenes; C-H (rock) stretch at 1372.52 cm⁻¹ indicates the existence of alkanes; and C=O stretch at 1755.26 cm⁻¹ indicates the presence of ester. This result is consistent with previous results for C-H stretch (~2900 cm⁻¹), C=O stretch (~1700 cm⁻

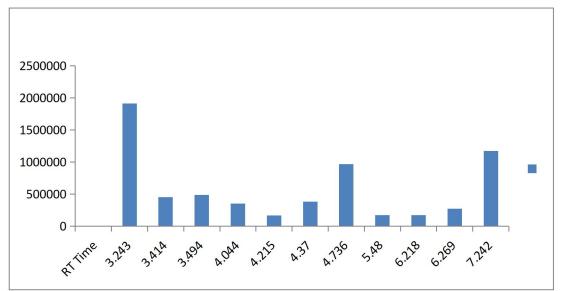
¹), broad O-H stretch (~3400 cm⁻¹), and C-O stretch (~1100 cm⁻¹) as reported Snezana *et al.*, (2020).

The FT-IR results show that moringa biodiesel contains more fatty acids than neem biodiesel, which contains more esters due to esterification of the fatty acid in neem oil. Neem oil cannot contain fatty acids because it lacks the O-H (stretch) band on its FT-IR spectrum. So, the GC-MS results of both the neem and moringa biodiesel indicated the presence of undecanoic acid in moringa oil but not in neem oils.

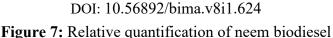


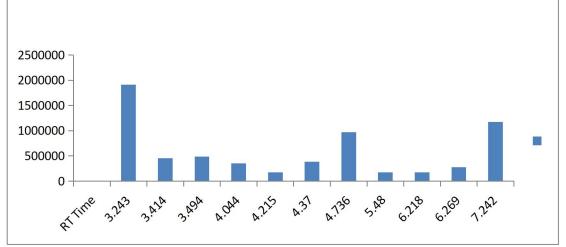


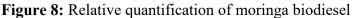
The chromatogram for both neem and moringa biodiesel were presented on figure 5 and 6 respectively they all have many peaks but there relative quantification were shown below.











From the above data obtained in the GC-MS analysis as from in figure 5, 6, 7 and 8. The compound with the highest concentration is the oleic acid for the undecanoic acid. While the compound with the highest concentration is the methyl-12-methyltetradecanoate ester. The relative quantification is obtained from the chromatogram of both biodiesel in the GC-MS spectrum.

CONCLUSION

This study describes the synthesis, characterisation, and comparison of biodiesels made from Neem and Moringa seed oils, with the goal of obtaining the highest output possible. Moringa and Neem biodiesel yielded 69.80% and 63.40%, respectively. The study found that neem seed oil might be used as a substitute for fossil oil. Producers must carefully analyze and alter their operations to preserve biodiesel quality and efficiency in the face of feedstock changes.

REFERENCES

Abdulkareem, A. S., Jimoh, A., Afolabi, A.S., Odigure, J.O. and Patience, D. (2012). Production and Characterization of Biofuel from Non-edible Oils: An Alternative Energy Source to Petrol Diesel. *Energy Conversion*, 171-196.

- Abubakar, A., Mahmoud, A.A., D.A. Ajiya., Hassan, U.F., Muhammad, K.Y., Sallau, S. et al. (2022). Optimization Process of Parameters for the Transesterification of Jatropha curcas Seed Oil using Response Surface Methodology (R.S.M.). *International Journal of Research and Innovation in Applied Science*, 7, 14-22 (2022).
- Abubakar, A., Mahmoud, A.A., Yakubu, H., Yushau, S. (2020). Extraction and Physicochemical Characterization of Seed Oil from Terminalia mantaly Seed. *International Journal of Research and Innovation in Applied Science*, 5(1), 60-64.
- Abubakar, H., Jauro, A., Abubakar, I.M., Abubakar, A., Yushau, S. and Sallau, S. (2020). Production and Biodegradability of Biodiesel from Lagenaria siceraria Seed Oil. International Journal of Research and Innovation in Applied Science, 5(3), 130-134.
- Akbar, E., Yaakob, Z., Kamarudin, S. and Ismail, M. (2009). Characteristics and Composition of Jatropha curcasOil Seed from Malaysia and its Potential as



Biodiesel Feedstock. *European Journal* of Scientific Research, 29, 396-403.

- Akpan, U.G. and Mohammed, J.A. (2007) Extraction, Characterization and Modification of Castor Seed Oil. *Leonardo Journal of Sciences*, 8, 43–52.
- Aremu, M.O., Ibrahim, H. and Bamidele, T.O. (2015). Physicochemical Characteristics of the Oils Extracted from some Nigerian Plants Foods. (A Review). *Chemical and Engineering Research*, 32, 36 52.
- Aremu, M.O., lonisakin, O, Bako, D.A. and Madu, P.C. (2006). Compositional Studies and Physicochemical Characteristics of Cashew nut (Anarcadium *occidentale*) Flour. Pakistan Journal of Nutrition, 5(4), 328-333.
- Aremu, M.O., Olaofe, O. and Akintayo, E.T. (2006a). Chemical Composition and Physicochemical Characteristics of Two Varieties of Bambara Groundnut (*Vigna subterrenea*) Flours. Journal of Applied Sciences, 6(9), 1900 - 1903.
- Arif, N.M., Masood, T. and Syed, S. S. (2012). Evaluation of Oil Seeds for their Potential Nutrients. *ARPN Journal of Agricultural and Biological Science*, 7(9), 730-734.
- Andualem, B. and Gassesse, A. (2014). Proximate Composition Mineral Content and Antinutritional Factors of Bebra (*Milletiaferruginea*) Seed Flour as well as Physicochemical Characterization of its Seed Oil. A springer Open Journal, 3, (298), 1-10.
- AOAC (2023). Official Method of Analysis. 22th edition, Association of Oficial Analytical Chemists, Washinton DC. https://www.aoac.org
- Dagde, K. K. (2019). Extraction of Vegetable Oil from Avocado Seeds for Production of Biodiesel. *Journal of Applied Sciences Environmental Management*, 23(2), 215– 221.

- Demirbas, A. (2009). Biodiesel Fuels from Vegetable Oils Via Catalytic and Noncatalytic Supercritical Alcohol Transesterification and other Methods: A Survey. *Energy Conversion and Management*, 44, 2093-2109.
- Ebenezer, K. S. (2015). Extraction and Physicochemical Characterization of Oil Extract from the Seed of Umbrella Tree (*Terminalia mantalis*). International Journal of Scientific and Engineering Research, 6(10), 144-147.
- Elkady, M.F., Ahmed, Z. and Ola, B. (2015). Production of Biodiesel from Waste Vegetable Oil via KM Micromixer. *Journal of Chemistry*, 9, 1-9.
- Emil, A., Zahira Y., Kumar, M. N. S., Jahim, J. M., Salimon, J. (2010). Comparative Evaluation of Physicochemical Properties of Jatropha Seed Oil from Malaysia, Indonesia and Thailand. *Journal of American oil Chemists' Society*, 87, 689–695.
- Encinar, J. M., Gonzalez, J. F. and Rodriguez,
 R. A. (2007). Ethanolysis of Used Frying
 Oil. Biodiesel Preparation and
 Characterization. *Fuel Processing Technology*, 88, 513-522.
- Gunstone, F.D. (2004). Rapeseed and Canola Oil: Production, Processing, Properties and Uses. London: Blackwell Publishing Ltd. Pp.217-287.
- Hossain, M. M.D.T., Alam, M.T. and Islam, M.T.U. (2007). Physico-Chemical and Nutritional Studies of *Terminalia belericaRoxb*. Seed Oil and Seed Kernel.*Journal of Bio-Sciences*, 15, 117-126.
- Jauro, A. and Haruna, A.M. (2011). Production and Biodegradability of Biodiesel from Balanites aegyptiacaSeed Oil. *Journal of the Korean Chemical Society*, 55, (4), 680-684.
- Jefferson, S. De, O., Polyanna, M. L., Lincoln, B., De S., Vinicius, M. et al. (2009).



Characteristics and Composition of J. gossypiifoliaand J. curcas L. Oils and Application for Biodiesel Conversion. *Biomass and Bioenergy*, 33, 449-453.

- Karmakar, G., Ghosh, P. and Sharma, B.K. (2017). Chemically Modifying Vegetable Oils to Prepare Green Lubricants. Lubricants, 5, (4), 1-17.
- Kannahi, M. and Arulmozhi, R. (2013). Production of Biodiesel from Edible and Non-edible Oils using *Rhizopus oryzae* and *Aspergillus niger*. *Asian Journal of Plant Science and Research*, 3 (5), 60-64.
- Kumar, V. and Kant, P. (2013). Study of Physical and Chemical Properties of Biodiesel from Sorghum Oil. *Research Journal of Chemical Sciences*,3 (9), 64-68.
- Latchubugata, C. S., Kondapaneni, R. V., Patluri, K. K., Virendra, U. and Vedantam, S. (2018). Kinetics and Optimization Studies Using Response Surface Methodology in Biodiesel Production Using Heterogeneous Catalyst. Chemical Engineering Research and Design, 18, 1-30.
- Mahmoud, A.A., Kwada, S.I., Jauro, A. and Abubakar, A. (2020). Production and Biodegradability of Biodiesel from Lagenaria siceraria Seed Oil. *International Journal of Research and Innovation in Applied Science*, 5 (3), 94-98.
- Moyo, L.B., Iyuke, S.E., Muvhiiwa, R.F., Simate, G.S. and Hlabangana, N. (2021).
 Application of Response Surface Methodology for Optimization of Biodiesel Production Parameters from Waste Cooking Oil Using a Membrane Reactor. South African Journal of Chemical Engineering, 35, 1-7.
- Ogwuche, C. E. and Edema, M. O. (2020). GC-MS and FTIR Characterization of Essential Oil from the Fresh Leaves of Pandanus candalabrum Obtained from

Bayelsa State, Nigeria. Nigerian Journal of Chemical Research, 25 (1), 1-10.

- Orhevba, B.A., Adebayo, S. E. and Salihu, A.O. (2016). Synthesis of Biodiesel from Tropical Almond (Terminalia catappa) Seed Oil. Current Research in Agricultural Sciences, 3(4), 57-63.
- Petal, S., Petal, K., Nilkanth, F., Vasudev, T. and Shubramanian, R.B. L. (2011). Extraction and analysis of Jatropha curcasL. seed oil. BC`African Journal of Biotechnology, 10(79), 18210-18213.
- Premjet, D., Abraham K. O., Hah Y. Y., Seung W. K. and Premjet, S. (2021).
 Physicochemical Characterization of *Jatropha podagrica* Seed Oil for Potential Biodiesel Production and other Industrial Applications in Thailand. *Sains Malaysiana*, 50(1), 85-92.
- Refaat, A. A. (2010). Different Techniques for the Production of Biodiesel from Waste Vegetable Oil. *International Journal of environmental Science and Technology*, 7(1), 183-213.
- Saeed, M.D. and Shola, E.A. (2015). Extraction and Physico Chemical Properties of Some Edible Seed Oils Sampled in Kano Metropolis, Kano State. *Bayero Journal of Pure and Applied Sciences*, 8(2), 239 – 244.
- Saeed K. A. E. and Abbas S. S.M. (2016). Determination of the Physico-Chemical Properties of Z-spinachisti (Nabag) Seed Crude Oil. *Journal of Agricultural Research*, 2(4), 22-35.
- Schinas, P. G., Karavalakis, C., Davaris, G., Anastopolous, D., Karonis; F., Zanikos,
 S. et al.(2008). Pumpkin Seed Oil as an Alternative Feedstock for the Production of Biodiesel in Greece. National Technical University of Athens, 157-80 Athens, Greece.
- Snezana A.K., Petar, R., Vladimir, G., Tatiana M. L. and David W. M. (2020). Essential Oil Quality and Purity Evaluation via

Bima Journal of Science and Technology, Vol. 8(1B) Apr, 2024 ISSN: 2536-6041



DOI: 10.56892/bima.v8i1.624

FT-IR Spectroscopy and Pattern Recognition Techniques. *Applied sciences*, 10 (7294), 1-12.

- Ugah, I.A., Abayeh, O.J. and Onuoha, E.C.E. (2007). Transesterified the Vitianerifolia Seed Oil as a Biodiesel Fuel. *Global Journal of Environmental Research*, 1(3), 124-127.
- Umar, D. M. A., Jauro, A., Yushau, S., Abubakar, A., Abubakar, H., Abubakar, I.M., Sham'unu Aliyu, S. I. (2020). Characterization and Determination of Nutritional and Anti-Nutritional Values of the Seed Oils of Sorrel (*Hibbiscus* Sabdariffa) and Okro (*Abelmoschus* Esculentus). International Journal of Research and Innovation in Applied Science, 5(5), 1-7.
- Yisehak K, Solomon M. and Tadelle M. (2011). Contribution of Moringa (Moringa stenopetala, Bac0. a Highly Nutritious Vegetable Tree, for Food Security in South Ethiopia: A Review. *Asian Journal of Applied Science*, 4(5): 477-488.
- Yunus, M.M., Zuru, A.A., Faruq, U.Z. and Aliero, A.A. (2013). Assessment of Physicochemical Properties of Biodiesel from African Grapes (Lanneamicrocarpa). Nigerian Journal of Basic and Applied Science, 21(2), 127-130.
- Eid, A., Jaradat, N., & Elmarzugi, N. (2017). A Review of chemical constituents and traditional usage of Neem plant (Azadirachta Indica). *Palestinian Medical and Pharmaceutical Journal*, 2(2), 3.
- Kumar, R., Mehta, S., & Pathak, S. R. (2018).Bioactive constituents of neem.In Synthesis of medicinal agents from plants (pp. 75-103). Elsevier.
- Gupta, A., Ansari, S., Gupta, S., Narwani, M., Gupta, M., & Singh, M. (2019). Therapeutics role of neem and its

bioactive constituents in disease prevention and treatment. Journal of Pharmacognosy and Phytochemistry, 8(3), 680-691.