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# Phytoremediation of Heavy Metals Polluted Soil from the Etelebou Dumpsite in Yenagoa, Bayelsa State, Nigeria

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The uncontrolled man's action has resulted into ecological Armageddon, and natures Submission: 11/06/2024 requite in a myriad form; of such man's action is the contamination of the soil with heavy Accepted: 22/08/2024 metals. The understanding that man is the precipice of his actions, account phytoremediation a costless, and ecofriendly process for the restoration of the shrinking soils. The adoption of Banana, Guava and Pawpaw fruits for the decontamination of heavy metal polluted soil from the Etelebou dumpsite form the bases of this study. The study recorded the presence of Pb, Cd, Cr, Ni, Fe, cu and Zn in the soil and in the fruits respectively. The account of the element in the fruit is in a multifold of the desire level due to the contaminated soil. The plants poses the ability to bio accumulates, translocation of heavy metals in varied quantities in the plants aerial parts (fruits). The extractions and accumulation of the metals were higher in the fruit peels than the edible parts, serving a good hypercummulator to the metals. The study demonstrated hyperaccumulative ability of plant, whose fruits should not be considered for consumption for the purpose of these exercise. Theplants via its fruits, if successfully applied can served cost effective, ecofriendly and easy decontamination purposes for heavy metal polluted soils.

Keywords: Contaminated soil; Heavy Metals; Hyperaccumulator; Phytoremediation; Rhizosphere.

## Introduction

Research article

The 21<sup>st</sup> century is a generation enjoying unprecedented Technological, Scientific, financial and environmental resources, yet it is perhaps the first generation to take the world to the brink of an environmental system breakdown, best described as the ecological Armageddon; a breakdown underscored by massive extinctions, biodiversity loss and environmental degradations with pollution. It is so disappointed that not all the advantages of intellect and education, social and political class do any inoculate humanity against environmental degradations.

The contamination of the biosphere with heavy metals has been on the headline discussion among experts and professionals in the build environment, yet no measurable changes to the needs of humankind because of bad inclinations, quest for fast profit, greed, unsatisfactory material/financial advancement, policy poor implementations etc (Ebuete & Bariweni, 2019). The lithosphere, which is the soil, is man's survival livewire but has been infuriated by waste generation and disposals, the leading driving force of heavy metals contamination in the soil; resulting into agricultural crops losses and contamination, surface and underground water contamination with an adverse human health (Ekere, Ukoha & Ekere, 2017). Nyiramigisha, Komariah and Sajidan (2023) reported that landfills is the persecutor to soil contamination, but still, there is no promising way proposed for the waste disposal management in some developing countries while many with an elaborate management proposals lack implementations. Several cleanup approaches involving the use of

Several cleanup approaches involving the use of flora Technically known as Pytoremediation spanning through the edges of phytodegradation, phytostabilization, phytoaccumulation and phytovolatizations, each with huge advantages and challenges has been introduced at low cost, environmentally friendly and ease practices. Researchers like McGrath and Zhao (2003), Raccio and Navari-Izzo (2011), Kaushik (2011), Gunasundari Kumar and (2018),Sivaramakrishnan et.al. (2018), Devi and Kumar (2020), Jilani et.al. (2021), Ozvigit and Dogan (2021), has utilized conventional and engineered ephemeral, biannual, deciduous plants, crops and vegetables for the decontamination of heavy metal polluted soils with an appreciable results, however, this study advance towards the used of annual and permanent fruiting plants for the decontamination of heavy metal polluted soil from the Etelebou dumpsite via their fruits This was underscore by the fact that the indigenous plants are capable to inactivate metals in the rhizosphere and translocate them in the aerial parts (edible fruits and peels) (Wang et.al., 2018).

The Etelebou dumpsite among other unregulated dumpsites within Yenagoa are threat to the soil, crops, aquatic organisms, and the communities living around the dumpsites. The report of Simon and Ayotamuno (2022) clearly shows that leachate generated from the Etelebou dumpsite has an index value of 17.004 as against standards (Indian standards for leachate disposal on Inland Waters) of 7.378; hence it is contaminated and therefore requires immediate remediation measures. Based on this calling, this paper is made to suggest the use of indigenous fruiting plants (phytoaccumulation) in the remediation of heavy metals polluted soils from the Etelebou Dumpsite. The process involving the use of indigenous guava (Psidium guajava), Banana (Musa acuminata) and Pawpaw (Carica Papaya) edible part and peels for the decontamination of heavy metals polluted waste soil. These plants have the capacity to inactivate metals in the rhizosphere and translocate them in the aerial parts (fruits peels); a measure that could encourage implement the year 2024 Global World Environment Day theme (accelerating land restoration, drought resilience and desertification) (Ajayi, 2021).

## Materials and methods

## Description of the Study Location

The Etelebou Central waste dumpsite is the study focus. The Etelebou dumpsite lies within latitude  $4^{\circ}59^{I}28.320^{II}N - 5^{\circ}00^{I}6.342^{II}N$  and longitude  $6^{\circ}19^{I}38.346^{II}E - 6^{\circ}20^{I}18.942^{II}E$  covering an area of about 1.5km2 within a low swampy land (5meters below sea level) (Ebuete, Wodu & Ebuete, 2022). It lies along the Yenagoa-Amassoma road in Yenagoa Local Government Area of Bayelsa State.

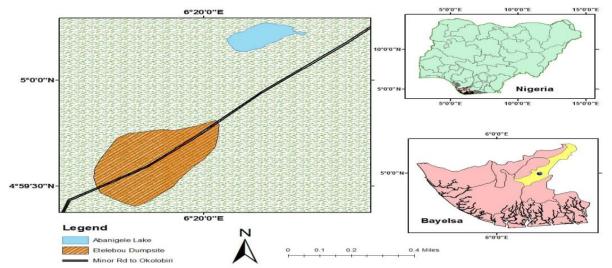


Figure. 1: Map Showing the Etelebou Central Waste Dumpsite, Yenagoa. Sources: Ebuete et. al. (2022).

# Sample Collection and Treatments

#### **Collection and Treatment of Fruits Samples**

A total of nine (9) samples of fruits (semi rip) were carefully picked with a sterilized fickle knife

(not allow hitting ground) from Banana plants (*Musa Acuminata*), Guava tree (*Psidium Guajava*) and Pawpaw tree (*Carica Papaya*), packed into clean polythene bags. The semi ripe fruits were

washed under running tap and later rinsed in distilled water, peel out their hulls from the edibles, each (edible and peels) air-dried at ambient temperature and pulverized into fine powder (composite) (Yaqub, *et.al.*, 2021).

# **Collection and Treatment of Soil Samples**

The soil samples were collected using Soil Push Probe at the base of each plant within the dumpsite. 16 soil samples were collected during the wet season at a depth of 30cm; visible remains of debris where removed. 30g of the Soil sample was oven dried to a constant weight of 23.5g at  $105^{\circ}$ C for six hours, grinded and mechanically passed through a 2 mm sieve and stored at  $4^{\circ}$ C for further analysis (Yagub *et al.*, 2021).

All reagents and chemicals were of analytical grade. Glass and plastic items (Merck, Germany) were carefully clean with 10% HNO<sub>3</sub> before been washed with distilled water (Markmanuel, Godwin & Ebuete, 2023).

## Sample Digestion and FAAS Analysis

All digestions followed the expression of Markmanuel, Godwin and Ebuete (2023), where one gram (1g) of each sample (fruit and soil) was digested with 20 mL solution of concentrated  $HNO_3/H_2SO_4$  containing 3:1; 5 mL of HCLO<sub>4</sub> was also added to each content. After digestion, a clear solution was obtained. The digest were cooled and diluted with 20 mL distilled water, filtered into 100 mL flask and distilled was added to make up the mark.

In triplicates, heavy metals concentrations (mg/kg) were analyzed using FAAS (Flame Atomic Absorption Spectrophotometer (S4-71096 model, Air-Acetylene Flame) (*Ibid*, 2023).

# Expression of Metals Translocations in the Fruit Plants

The inactivating of metals and translocations through plants rhizophere is expressed using the following equations:

a. Metal Transfer Factors (T.T.) = MCF / MCS......(Equ 1) Aralu and Okoye (2020).

Where: MCF is Metal Concentration in fruits samples (the sum concentration in fruits and peels) in mg/kg

MCS is Metal Concentration in Soil per mg/kg.

- b. Metals Translocation Factor (M.T) = MCef x MCfp / MCs .....(Equ 2) Primandani, Sajidan & Masykuri (2020)
- Where: MCef is Metals concentrations in mg/kg per the dry weight of edible fruits samples in mg; MCfp is Metal concentrations in mg/kg per the dry weight of Peels.

MCs is metals concentrations in Soil per mg/kg.

The ratio in metal concentration in plant soil to fruit edible and fruit peels represent the translocation factor while the ratio in metal concentration in the soil to the fruits represents the bioaccumulation of the plants. Any plants having bioaccumulation or translocation factor of one (1) or more (>1) is termed as hyperaccumulator plant (Sharma, Tripathi & Chandra, 2020).

# **Statistical Analysis:**

All data were reported as means for triplicate samples to confirm the data variability and stability using Student's t-test (P < 0.05). The mean concentration of heavy metals in the soil, fruits, edibles and peels were subjected to further analyses using the analysis of variance (ANACOVA).

# **Results and Discussion**

Plants are inimitable creatures which encompass exceptional metabolic as well as immersion abilities and conveyance systems that can absorb nutrients plus pollutants specifically from the resources surrounding (Verma *et.al.*, 2022). The laboratory analysis of the soil, edible parts and peels were presented in Table 1 and Figure 1 and 2 which revealed an appreciable level of concentration (mg/kg) of heavy metals (Lead, Cadmium, Chromium, Nickel, Iron, Copper and Zinc) in the edible parts and peels from the cultivated fruiting plants.

Wodu and Ebuete and (2024)

Metals				Media (Samples)									
	Banana Edible	Banana Peels	M.T.	T.T.	Paw paw Edible	Paw paw Peels	M.T.	T.T.	Guava Edible	Guav a Peels	M.T.	T.T.	limits
	Mean	Mean			Mean	Mean			Mean	Mean			WHO mg/kg
Pb Soil	0.337	1.783 4.794	1.430	0.337	0.589	1.579 8.665	0.250	0.107	0.127	0.155 2.044	0.138	0.010	0.43
Cd Soil	0.016	0.019 0.993	0.035	0.0003	0.015	0.029 0.300	0.147	0.005	0.011	0.023 0.316	0.108	0.001	0.02
Cr Soil	0.186	0.794 9.493	0.103	0.016	0.836	1.019 13.042	0.142	0.065	0.157	0.314 9.810	0.049	0.005	1.3
Ni Soil	0.218	0.119 0.615	0.548	0.042	0.018	0.029 0.997	0.047	0.001	0.111	0.322 1.470	0.295	0.024	1.63
As Soil	0.019	0.036 0.616	0.089	0.001	0.115	0.415 1.348	0.393	0.035	0.058	0.209 0.467	0.572	0.026	0.1
Fe Soil	2.885	3.143 5.390	1.118	1.682	3.524	1.999 6.29	0.878	1.120	1.536	3.068 7.564	0.609	0.623	20
Cu Soil	0.296	1.498 3.780	0.475	0.117	0.315	1.518 6.091	0.301	0.079	0.615	1.22 3.018	0.608	0.249	10
Zn Soil	1.436	1.677 3.314	0.939	0.727	1.858	1.519 4.451	0.759	0.634	0.915	1.358 2.873	0.791	0.433	50

Table 1. Heavy metals concentrations in soils and Plants Rhizosphere (Edible fruits and Fruits peels)

Source: Researcher, 2024. Where, WHO variables were adopted from Bibi. gt. al. (2023).All samples in mg/kg.

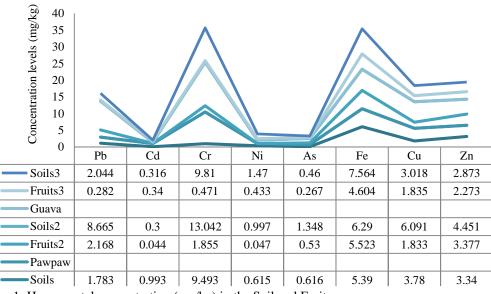
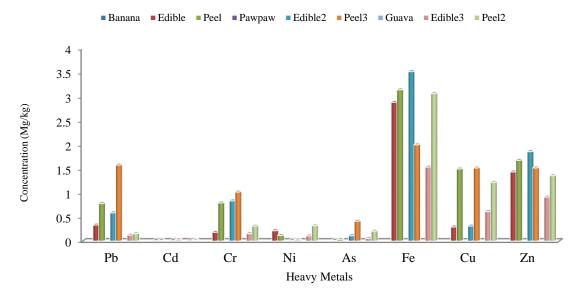


Figure 1: Heavy metal concentration (mg/kg) in the Soil and Fruits



#### Wodu and Ebuete and (2024)

Figure 2: Heavy Metals Concentration (mg/kg) in fruit Edible and Peels

The mean Lead (Pb) concentration in the soil were 1.783, 8.665 and 2.044 mg/kg respectively around Banana, Pawpaw and guava plants as against 1.121 (0.337 in edible and 0.783 peels), 2.168 (0.589 in fruits and 1.579 in peels) and 0.282 mg/kg (0.127 in edible parts and 0.155 in peels) fruits (table 1). The differences accounted for about 18, 80 and 76% respectively, representing a conventional way for fruiting plants to absorbed lead from the waste leachate through the soil and transfer to the edible aerial parts. The peels accumulate more lead than the edible part, which accounted for 40, 45 and 10% in Banana, Pawpaw and Guava fruits respectively that is in support with the report of Ashraf et.al. (2011). from the foregoing, Banana and Pawpaw were recognized as hyperaccumulator of lead from the soil; bioaccumulation more than 1 (<1) (Table 1). Sharma, Tripathi and Chandra (2020) made similar report.

The mean concentrations of Cadmium in the edible fruits were 0.016, 0.15 and 0.011 mg/kg for Banana, Pawpaw and guava fruits respectively, while in the peels were 0.035, 0.044 and 0.034 mg/kg for Banana, pawpaw and guava plants (table 1). The percentage concentration difference between the peels (0.019, 0.029 and 0.023 mg/kg)and edible parts (0.016, 0.015 and 0.011 mg/kg) accounted for 8, 32 and 36% respectively representing significant accumulations of cadmium in the plants rhizosphere. The

background concentration difference of cadmium between the soils and plants represents the translocation mechanism of the plants, which accounted for 94, 74 and 80% for Banana, Pawpaw and Guava respectively (Table 1 and Figure 2). The transfer and translocation of cadmium in the plants were slower in all the fruits because of external factors such as species and soil pH. Amusan, Ige and Olawale (2005) observed similar outcome in their study; Kpee and Edori, (2017) reported Nd-0.010 mg/kg, Anyanwu, *et.al.* (2022) 0.431-0.815 mg/kg.

The mean Chromium concentrations in the soil were 9.493, 13.042 and 9.810 mg/kg while in the fruits were 0.98, 1.855 and 0.471 mg/kg in Banana, pawpaw and Guava respectively (table 1). This accounted for 82, 76 and 90% difference between soil and fruits. On the other hand, the bioaccumulation of Chromium among fruit parts were Banana (0.186 in edible fruit and 0.794 mg/kg in peel), pawpaw (0.836 in edible fruit and 1.019 mg/kg in peels) and guava (0.157 in edible and 0.314 mg/kg in peels) which accounted for 62, 10 and 34% differences respectively. Chromium is not a nutritional compositions among the studied fruits but it presents is a function of heavy metal contamination in the soils; the peels store more of such metals in the fruit peels. However, its insignificant amount is function of а evapotranspiration that supported the view of Kaushik (2011); Manzoor, Sharma and Wani (2018) that evapotranspiration promote the transfer of contaminants into the plant shoots and fruits.

Nickel (Ni) is a micro nutrient element required by plants for their normal metabolism at 1.63mg/Kg (Bibi et al., 2023). The mean concentrations of Ni in the soil were 0.615, 0.997 and 1.470 mg/kg while in the fruits were 0.337 mg/kg (0.218 edible and 0.119 in peels), 0.047 mg/kg (0.018 fruit and 0.026 in peels) and 0.433 mg/kg (0.111 edible and 0.322 mg/kg in peels) in Banana, Pawpaw and Guava respectively (Table 1). The concentration difference of Ni in the soil and fruits accounted for 28, 92 and 54% respectively while among fruit parts were 30, 24 and 52% respectively. The trace element mobility and transfer of Ni in plants aerial parts was higher in Banana with an established description as hyperaccumulator (>1) but lower in pawpaw and guava (Table 1). This report was similar with the study of Kassaye, et.al.(2012), Anyanwu, et.al. (2022), Wani et.al. (2023).

The mean concentration of As in the soil were 0.616, 1.348 and 0.467 mg/kg while in the fruits were 0.055, 5.523 and 0.267 mg/kg which accounted for 84, 44 and 28% difference in Banana, Pawpaw and guava respectively (table1). The translocation and bioaccumulation of arsenic (As) in the fruit parts were 0.019 in edible and 0.036 mg/kg in peel; 0.115 in edible and 0.415mg/kg in peel, and 0.058 in edible and 0.209 mg/kg in peel; which accounted for 32, 96 and 56% respectively (table 1). Sedrah and Alshamary (2023) demonstrated similar positive result on the effectiveness of banana peels to Arsenic removal. fruit demonstrated The guava to be hyperaccumulator of Arsenic for its soil transfer (bioaccumulation) was equal to 1 (table 1). Maduforo et.al. (2020) reported similarly on the Red Dacca banana variety (0.0030 mg/kg); Patel et. al., (2023) recorded 0.003-1.3mg/kg for banana.

The mean concentrations of iron (Fe) were 6.028 mg/kg (2.885 in edible fruit; 3.143 mg/kg in peel), 5.523 mg/kg (3.524 in edible fruit and 1.999 mg/kg in peel) and 4.604 mg/kg (1.536 for edible and 3.068 mg/kg) in Banana, Pawpaw and guava fruiting plant respectively. This was above the recommended nutrient composition of iron (0.10–1.3 mg/100g) in Pawpaw by Jones and Layne (2022). The peels of the banana and pawpaw fruits stored more of the said metal than the edible fruits

(4 and 28%); however this is contrary to guava fruits were the edible fruits stored more of the iron than the peel (34%) (Table 1). The soils around the plants contain 5.390, 6.290 and 7.564 mg/kg of iron for Banana, Pawpaw and guava respectively, such that the bioaccumulation and translocation of the metals from soil to fruits parts account for 36, 8 and 24% in Banana, pawpaw and Guava fruits, indicating а higher metal transfer. The translocation factor of the fruiting plants indicated hyperaccumulator for iron (>1) and recommended for the cleanup of iron contaminated soils. The result was in contrast with the view of Anyanwu, et.al. (2022) that iron (Fe) is not easily mobilized in Pawpaw plant, however, it was similar with the report of Nwofia, Ojimelukwe and Eji (2012) 2.31 - 2.67 mg/kg and Kurade et. al. (2021).

The mean concentration of Cu were 1.794 mg/kg (0.296 in edible fruit and 1.498 mg/kg in peel), 1.833mg/kg (0.315 in edible fruit and 1.518 mg/kg in peel) and 1.835 mg/kg (0.615 in edible and 1.22 mg/kg in peels). This was in contrast with the concentration in the soil 3.780 mg/kg (68%), 4.451 mg/kg (66%) and 2.873 mg/kg (32%) in respective plants. The record shows that Cu is easily mobilized in the fruiting plants; thus, pawpaw and guava plants are hyperaccumulator of copper since the tested translocation and bioaccumulation values were equal to 1 and above (<1) respectively (table 1). Similarly, Kpee and Edori, (2017), Kafle. et.al. (2022), Sedrah and AIshamary (2023) has demonstrated the used of banana peels for the management of copper polluted soil.

The mean concentration of Zn in the soil was 3.314, 4.451 and 2.873 mg/kg while in the fruits were 3.113 mg/kg (1.436 in edible part and 1.677 mg/kg in peel), 3.377 mg/kg(1.858 in edible part and 1.519 mg/kg in peel) and 2.273 mg/kg (0.915 in edible part and 1.358 mg/kg in peel) for Banana, Pawpaw and Guava fruits respectively. The records were above the 0.01-0.2 mg/100g recommended for zinc composition in Banana groaned in a free and uncontaminated soil by Dotto, Matennu and Ndakidemi (2019) Vaganan, Ravi and Mustafa (2022). The results shows that fruit plants has high bioaccumulation capacity to translocate Zinc between peels and edible part in respective account 0.727 (8%), 0.634 (10%) and 0.433 mg/kg (20%). The concentration difference between soil and fruits were just 4%, 14% and 12% respectively; indicating a smooth and faster

transfer mechanism of the said metals. Wang *et.al.* (2019) affirm that increased Zn phytoremediation could be achieve by over expressing SaNramp1, SaIRT1 of Pseudomonas fluorescens under metal stress in Sedum alfredii. This report is similar with the report of Kpee and Edori, (2017) Zn 3.84-6.32 for pawpaw.

Statistically, at the 0.05% degree of freedom, the difference between heavy metal concentration in the approximate soil and fruits was insignificant, so also between the fruit edible part and fruit peels.

#### Conclusion

Man has destroyed the pristine ecosystem with its activities, such that he is direly looking for a cost effective, eco-friendly and feasible approach for solutions. One of such approach is through the introduction of plant biomass into the environment and allowing the plants to assimilate/degrade the unwanted pollutants in their aerial and subsurface portions known as phytoremediation. The nutritional values of fruit are largely dependent on its adjoining composition such that the present of unwanted composite and high concentration of these composite signify danger. In this study, we adopted the use of ephemeral and permanent indigenous fruiting plants with the understanding that these plants over their fruiting time will absorb, bioaccumulate, translocate and store hazardous metals into their fruits.

The fruiting plants have demonstrated heavy metals consumption and absorbtion via the cortical tissues present in roots because of their correspondence to some necessary micronutrients (such as zinc) as well as implemented to reach the xylem transport system for symplastic and/or apoplectic pathway into their fruits (edible and peels). The study understand the ability of these plants vacuole to immobilized metalloid and metal ions intracellular through the vascular tissue of the plant roots revealed that the concerned heavy metals are presents in varied concentration levels

#### References

Ajayi, O. A. (2021): Comparative assessment of heavy metals uptake of vegetables cultivated on a dumpsite at Abraham Adesanya Polytechnic Ijebu-Igbo, Ogun State, Nigeria. *Journal of Chemical Society of Nigeria*, **46** (4):0651 – 0655 in the soil and fruits of Banana, Pawpaw and Guava. It also demonstrated that plants vascular tissue received metaloid from the soil. bioaccumulate and transfer metals into the plants rhizosphere. The fruit peels store more of these metals than the edible part especially of banana because of it hyper accumulative ability. The plants are noninvasive and eco-friendly to demonstrate hyperaccumulator with the following elements (Fe, Zn, Ni, As and Pb). The plants possess the ability to assimilate, transfer into the fruits parts and can provide a safe, easy, ecofriendly and cheap method of soil treatment, provided the fruits remained inconsumable but incinerated for energy recovery against potent future health challenges. The plants also aid reduces soil erosion and metal leaching by stabilizing heavy metals against further media contamination.

#### Recommendations

The cultivation of the sampled plants should be encouraged in heavy metal polluted soil due to its hyperaccumulation potential, ease and ecofriendly. Due to the hyperaccumulator and translocation property of the plants with heavy metals for the purpose of this study, the fruits should not be considered for direct consumption. There should be a proper management of the plants fruits since, the fruits serves as hyperaccumulator to these metals.

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#### **Conflicts of Interest**

The authors declare no conflict of interest.

- Amusan, A.A., D.V. Ige and Olawale, R. (2005), Characteristics of soils and crops uptake of metals in municipal waste dumpsites in Nigeria. *Journal of Human Ecology.*, **17**(3):167-171.
- Anyanwu J.C., Rufus L.N., Duluora, J.O., Egbuawa O.I., Nwobu E.A. and

Onwuagba C.G. (2020): Comparative analysis of heavy metals in Carica Papaya (pawpaw) fruit and soil in selected urban, rural and forestland in owerri, imo state, *International Journal in Advance Resources*, **8**(03):10-20; http://dx.doi.org/10.21474/IJAR01/10592.

- Aralu, C.C. and Okoye, P.A.C. (2020): Assessment of heavy metals levels in soil and vegetables in the vicinity of unlined waste dumpsite in Nnewi, Anambra State Nigeria Journal of Chemical Society of Nigeria 45(4):687-696.
- Ashraf, A.M.; A. Wajid; K. Mahmood; J.M. Maah and Yosoff, I. (2011): Low cost biosorbent banana peel (musa sapientum) for the removal of heavy metals, *Scientific Research and Essays*, **6**(19):4055-4054. https://doi.org/10.5897/SRE11.303.
- Bibi, M., F. Samiullah, F. Behlil, S. Afzal and Sahifa, G. (2023): Essential and nonessential heavy metals sources and impacts on human health and plants; *Pure* and Applied Biology, **12**(2):835-847, http://dx.doi.org/10.19045/bspab.2023.120 083.
- Devi, P. and Kumar, R. (2020): Concept and Application of Phytoremediation in the fight of heavy metal toxicity, *Journal of Pharmaceutical Sciences and Research*, 12: 795- 805.
- Dotto, J., A.O. Matennu and Ndakidemi, P.A. (2019): Nutrient Composition and Selected physicochemical properties of fifteen Mchan Cooking Banana: A study conducted in Northern Tanzania, *Scientific Africa*, **6**, Nov. 2019, e.00150.
- Ebuete, A.W. and Bariweni, P.A. (2019).Water Quality Index of Kolo Creek, Bayelsa State, Nigeria, *Journal of Applied Science and Environmental Management*, **23**(11):1923-1927, D O I :https://dx.doi.org/10.4314/jasem.v23i11. 3.
- Ebuete, A.W., D. P. Wodu and Ebuete, E. (2022): Dumping on Waters: The Lacunae in Waste Management in the Niger Delta, Nigeria; *American Journal of Environment and Climate* 1(2):100-109. DOI: https://doi.org/10.54536/ajec.v1i2.602

- Ekere, J.N., I.P. Ukoha and Ekere, N.R. (2017): Ecological and human health risk assessment of heavy metal contamination in soil of a municipal solid waste dump in Uyo, Nigeria. *Journal of Environment and Geochemical Health*, **39**(3): 497–515. doi: 10.1007/s10653-016-9830-4.
- Jilani, G.; I.H. Shamsi, D. Zhang and Hina, K. (2021): Recent advances in heavy phytoremediation metalsof soils: review. contaminated Α In Bioremediation for Environmental Sustainability; Elsevier: Amsterdam, the Netherlands, 2021: 23–41.
- Jones, S.C. and Layne, D.R. (2022): Pawpaw Description and nutritional information, *Kentucky State University Cooperative Extension Program*. Access from https://www.kysu.edu/academics/collegeahnr/school-of-anr/pawpapw-/pawpawdescription-and-nutritionalinformation.php/#Table2.
- Kafle, A.; A. Timilsina; A. Gautam; K. Adhikari;
  A. Bhattarai and Aryal, N. (2022): Phytoremediation: Mechanisms, plant selection and enhancement by natural and synthetic agents, *Environmental Advancement.* 2022, 8:100203.
- Kassaye, Y.A., L. Skipperud, S. Meland, E. Dadebo, J. Einset and B. Salbu (2012): Trace element mobility and transfer to vegetation within the Ethiopian Rift Valley lake areas. *Journal of Environment and Evaluation Monitoring*, **14**(10):2698-2709.
- Kaushik, S. (2011): Phytoremediation—Use of green plants to remove pollutants. *Environmental Biotechnology, Biotech Articles,* 2011. Available online: <u>https://biotecharticles.com/Environmental-</u> <u>Biotechnology-Article/Phytoremediation-</u> <u>Use-of-green-plantsto-remove-pollutants-</u> <u>704.html</u>.
- Kpee, F. and Edori, O.S. (2017): Prevalence of Some Heavy in Mango and Pawpaw Found in Dumpsites of Obio/Akpo and Eleme Local Government Areas in River State, Nigeria. *Journal of Environment and Anal Chemistry* 4: 196. doi:10.41722380-2391.1000196

- Kumar, P.S. and Gunasundari, E. (2018): Bioremediation of heavy metals. In Bioremediation: Applications for Environmental Protection and Management; *Springer: Singapore*, 201Metals**8**: 165–195.
- Kurade, M.B.; Y.H. Ha; J.Q. Xiong, S.P. Govindwar,.; M. Jang, and Jeon, B.H. (2021): Phytoremediation as a green biotechnology tool for emerging environmental pollution: A step forward towards sustainable rehabilitation of the environment; Chemical Engineering Journal, 2021. 19. https://doi.org/10.1016/j.cej.2021.129040.
- Maduforo A.N., Okoro C.E., Ogbuabo D.A., Ezeh C.J., Bok I.S., Adesanmi A.R., Okorie J.N., Obiloma A.A., Aloysius-Maduforo M.C., and Okwara C.C. (2020): Heavy Metal in Banana (Musa Acuminata) Varieties Sold by Fruit Vendors in Enugu State, Nigeria; *Journal of Dietitians Association of Nigeria (JDAN)*; **11**(2):52-57. Access from www.jdan.org.ng
- Manzoor, J., M. Sharma and Wani, K. A. (2018): Heavy metals in vegetables and their impact on the nutrient quality of vegetables: A review, *Journal of Plant Nutrition*, 23(9):1-22; DOI: 10.1080/01904167.2018.1462382
- Markmanuel, D.P., Godwin J, and Ebuete, A.W. (2023): Potential cancer and non-cancer hazard of some heavy metals in children and adults via consumption of canned chicken luncheon and Turkey luncheon. *Discovery*, **59**: e56d1237
- McGrath, S.P. and Zhao, F.J. (2003): Phytoextraction of metals and metalloids from contaminated soils, *current Opinion in Biotechnology*, 14:277-282.
- Nwofia, G.E., P. Ojimelukwe and Eji, C. (2012): Chemical composition of leaves, fruit pulp and seeds in some Carica papaya (L) morphotypes, *International Journal in Medicine and Aromatic Plants*, **2**(1) :200-206.
- Nyiramigisha, P., C. Komariah and Sajidan, N. (2023): Harmful Impacts of Heavy Metal Contamination in the Soil and Crops Grown Around Dumpsites; *Reviews in*

*Agricultural Science*, **9**: 271–282, 2021 https://dx.doi.org/10.7831/ras.9.0\_271.

- Ozyigit, I.I.; H. Can and Dogan, I. (2021): Phytoremediation using genetically engineered plants to remove metals: A review. *Environment and Chemical Letters.* **19**, 669–698.
- Patel, K.S.; K.P. Pandey, P. Martin-Ramos, W.T. Corns, S. Varol, P. Bhattacharya and Zhu, Y. (2023): A review on Arsenic in the Environment: Bioaccumulation, remediation and disporsal; *RSC Advance*, 13(22):14914-14929;
  doi:10.1020/d2ra02018c

doi:10.1039/d3ra02018e.

- Primandani, A., A. Sajidan and M. Masykuri (2020): Soil and banana crops (Musa paradisiaca L.) risk by chromium (Cr) accumulation through leachate and its health risk assessment, *Journal of Physics: Conference Series*, **1567** (2020):042058, IOP Publishing, doi:10.1088/1742-6596/1567/4/042058.
- Raccio, N. and Navari-Izzo, F. (2011): Heavy metal hyperaccumulating Plants: How and Why do they do it? And what makes them so interesting?, *Plant Science*, 180:169-181.
- Sedrah, Z.T. and Alshamary, E.I. (2023): Study of the use of Banana peels powder in removing some heavy metals, 2023 IOP Conf. Ser.: *Earth Environmental Science*. 1252 (2023) 012162. Doi:10.1088/1755-1315/1252/1/012162.
- Sharma, P., S. Tripathi and Chandra, R. (2020): Phytoremediation potential of heavy metal accumulator plants for waste management in the pulp and paper industry, *Heliyon* **6** (2020) e04559.
- Simon, C.E. and Ayotamuno, M.J. (2022): Leachate Pollution Index Determination at Yenagoa Central Waste Dump during the First Quarter of 2019, Bayelsa State, Nigeria; *Acta Scientific Environmental Science*, **1**(1): 23-28.
- Sivaramakrishnan, M.; N. Sivarajasekar; J. Vivek;
  P. Thanaraj; M.Naushad; J. Prakashmaran;
  V. Gayathri and Al-Duaij, O.K. (2018):
  Phytoremediation of heavy metals:
  Mechanism, methods and enhancements.
  Environmental Chemistry Letters, 16, 1339–1359.

- Vaganan, M.M., I. Ravi and Mustafa, M.M. (2022): Banana: A Nutritive and Nutraceuticals-Rich Fruit, Technical Folder No:4, *National Research Center for Banana*, India Council of Agricultural research.
- Verma, R.K., S.S. Mahipal, E. B. Ekta, K. Parihar, K. Kant and Awasthi, K.K. (2022): of Heavy Phytoremediation Metals Extracted from Soil and Aquatic Environments: Current Advances as well as Emerging Trends, **Biointerface** Research in Applied Chemistry, 12(4): 5486 5509. https://doi.org/10.33263/BRIAC 124.54865509 124.54865509.
- Yaqub, G., A. Khan, M. Z. Ahmad and Irshad, U. (2021): Determination of Concentration of Heavy Metals in Fruits, Vegetables, Groundwater, and Soil Samples of the Cement Industry and Nearby Communities and Assessment of Associated Health Risks. *Hindawi Journal of Food Quality*, 2021, Article ID 3354867:1-9,

https://doi.org/10.1155/2021/3354867

- Wang, C.; Na, G.; Bermejo, E.S.; Chen, Y.; Banks, J.A.; Salt, D.E. and Zhao, F.J. (2018): Dissecting the components controlling root-to-shoot arsenic translocation in Arabidopsis thaliana. *New Phytology*, **217**, 206–218.
- Wang, Q.; J. Ye; Y. Wu; S. Luo; B. Chen; L. Ma;
  F. Pan; Y. Feng and Yang, X. (2019): Promotion of the root development and Zn uptake of Sedum alfredii was achieved by an endophytic bacterium Sasm05. *Ecotoxicology and Environmental Safety*, 172, 97–104.
- Wani, Z.A.; Ahmad, Z.; Asgher, M.; Bhat, J.A.; Sharma, M.; Kumar, A.; Sharma, V.; Kumar, A.; Pant, S.; Lukatkin, A.S. *et al.* (2023): Phytoremediation of Potentially Toxic Elements: Role, Status and Concerns. *Plants*, 12:429. <u>https://doi.org/10.3390/plants12030429</u>.