



**PREVALENCE AND ANTIBIOTIC RESISTANCE PROFILE OF *Staphylococcus aureus* AND *Escherichia coli* AMONG PATIENTS ATTENDING UROLOGY CLINIC OF DALHATU ARAF SPECIALIST HOSPITAL (DASH) LAFIA, NASARAWA STATE, NIGERIA**

<sup>1</sup>Ramalan, A. S., <sup>1</sup>Joseph, F. N., <sup>1</sup>Owoseni, M. C., <sup>1,2\*</sup>Ya'aba Y. and <sup>1</sup>Chuku A.

<sup>1</sup>Department of Microbiology, Federal University of Lafia, P.M.B 146 Lafia, Nasarawa State, Nigeria.

<sup>2</sup>Department of Microbiology and biotechnology, National Institute for Pharmaceutical Research and Development (NIPRD) Abuja, Nigeria

\*Corresponding Author E-mail: [yakyabnig@yahoo.com](mailto:yakyabnig@yahoo.com), Phone: +2348033326544.

**ABSTRACT**

**An infection of any kind attacking any part of the urinary system is termed Urinary Tract Infection (UTI) which has continued to be a serious healthcare challenge globally. This research work therefore, was aimed at determining the prevalence and antibiotic resistance profile of *Staphylococcus aureus* and *Escherichia coli* among patients attending Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria. These two microorganisms of interest were isolated and identified from midstream urine samples of 202 patients suspected of having UTIs through the use of microbiological techniques (culture, microscopy, and biochemical tests). Antimicrobial Susceptibility Testing of the isolates was carried out in accordance with the Clinical and Laboratory Standards Institute (CLSI). The prevalence of UTI cases were found to be 51.49% of the total samples analyzed. The *Staphylococcus aureus* had high prevalence of 59.62% compared to *E. coli*, which had a prevalence of 40.38%. The findings revealed that UTIs are more prevalent among males than in females. The highest prevalence of UTIs was found within the age group of 30-39 years (59.62%) followed by patients within the age group of 70 years and above with prevalence rate of 26.92%. *E. coli* had the lowest mean average (78.35%) of resistance compared to *Staphylococcus aureus* which had the highest (80.77%). Resistance to antibiotics ranges from 64.29% (Levofloxacin) to 89% (Nalidixic acid) for *E. coli* and 74.19% (Levofloxacin) to 93.55% (Rifampicin) for *S. aureus*. This high rate of resistance could be associated with self-prescription, incomplete dosage and indiscriminate use of antibiotics. Bacteria strains resistance to multiple antibiotics exists among patients suffering from UTIs.**

**Keywords: *Staphylococcus aureus*, *Escherichia coli*, Urine samples, Antibiotic resistance, UTI.**

**INTRODUCTION**

An infection of any kind attacking any part of the urinary system is termed Urinary Tract Infections (UTIs) (Anejo-Okopi *et al.*, 2015; Derbie *et al.*, 2017; Sule *et al.*, 2018). UTIs have continued to be a serious healthcare challenge, affecting over 250 million individuals globally each year (Derbie *et al.*, 2017; Sule *et al.*, 2018). This is attributed to the ability of the pathogenic microorganism to successfully colonize the urinary tract and cause a disease condition (Otajevwo, 2013). The microorganisms often associated with UTIs include; bacteria, fungi, yeast or protozoa, virus or even parasites (Ajao and Yakubu, 2015). However, bacteria have been reported as the major uropathogen affecting the urinary tract (Khoshbakht *et al.*, 2013). These infections are also reported to be the cause of most hospital visits and the most associated nosocomial

infection of the humans (Timothy *et al.*, 2014; Sule *et al.*, 2018).

Epidemiologically, uropathogenic infections are in the increase with associated cause of most morbidity and mortality worldwide (Timothy *et al.*, 2014; Tahira *et al.*, 2020). It is estimated that over 250 million people worldwide are infected with UTI yearly; which are mostly caused during hospital visits (Sule *et al.*, 2018; Tahira *et al.*, 2020), which is seen to be a burden to the nations and individuals affected (Akingbade *et al.*, 2014). The prevalence of UTI in Nigeria varies across geo-political zones. Among the semi-urban settlers in South-West Nigeria, the prevalence rate was reported to be 11.96%, 2.7% prevalence in South-East urban settlers, 13.7% prevalence in North-East (Akinsete *et al.*, 2018), 35.5% North-Central Nigeria (Muhammed, 2015).

Pathogenic UTIs cases is drastically changing with several etiologic agents of UTIs been reported (Alo *et al.*, 2015). Gram-positive organisms like *Staphylococcus aureus* and *Streptococcus pyogenes* and other Gram-negative species such as *Escherichia coli* are the predominantly reported organisms (Sule *et al.*, 2018), others include; *Klebsiella pneumonia* and *Proteus mirabilis* (Reis *et al.*, 2016). *Staphylococcus aureus* and *Escherichia coli* are documented to be the leading cause of human and animal infections, causing variety of infections; from the squamous and tissue infections, soft tissue infections, surgical site infection, bone infections, joint infections, and urinary tract infections to enteric and systemic infections. *Staphylococcus aureus* is known to be the commonest cause of hospital-acquired infection and bacteremia. It has also been established to be the cause of most hospital acquired respiratory infections (Onanuga and Awhowho, 2012). Other pathogens such as *Klebsiella* species, *Enterobacter* species, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Citrobacter sp.*, *Staphylococcus sp.*, *Neisseria gonorrhoea* and *Candida sp.*, are reported to be the cause of most UTIs (Timothy *et al.*, 2014; Ekwealor *et al.*, 2016; Derby *et al.*, 2017; Tahira *et al.*, 2020). However, some previous studies have reported the increasing prevalence of *S. aureus* in UTIs (Onanuga and Awhowho, 2012).

The predisposing factors to UTIs includes: sexual intercourse with infected person (Tahira *et al.*, 2020). In relation to gender, females are most predisposed (Tahira *et al.*, 2020). Other predisposing factors include inheritance, disease condition such as diabetes, stoppage of menses, poor nutrition, urination after sex, kidney stones, old age, utilization of urinary catheter, pregnancy, prostrate syndrome among males, diaphragm use, abnormalities in the urinary tract as well as poor hygiene practices (Dash *et al.*, 2013; Anejo-Okopi *et al.*, 2015; Ekwealor *et al.*, 2016; Tahira *et al.*, 2020). Also, previous exposure to UTIs is known to be a predisposing factor (Moges *et al.*, 2003).

Antibiotic resistance (AR) by microorganisms has become a major threat to public health with more microorganisms becoming resistant multiple antibiotics (Ochada *et al.*, 2015). This resistance could be natural, acquired or clinical and can be defined as the ability of microorganisms such as pathogenic bacteria to resist the effect of antibiotic or antimicrobial agents (Nolte, 2014). The natural or intrinsic resistance is an inborn caution for bacteria to resist antibiotics without previously exposed to antibiotic or without horizontal gene transfer while the acquired resistance is cause by

intrinsic gene mutation before exposure to some mutagens, antibiotic or through horizontal genetic transfer (Hollenbeck and Rice, 2012).

Diagnosis of UTIs by doctors and physicians in the out-patient department is principally through the presentation of signs and symptoms without laboratory test result obtained; because an immediate treatment is needed to be administered. Laboratory diagnosis on the other hand is based on urinalysis, dipstick leukocyte esterase test for detecting blood in urine (pyuria) with the ability to detect 10 white blood cells in millimeter cube of urine sample (10 WBC/mm<sup>3</sup>) (Sobel, 2014). The presence of Nitrite in a urine sample is an indication of UTIs (Helen and Jennifer, 2018). However, urine microscopy, culture and sensitivity (M/C/S) are the widely used and most preferred methods. Treatment of UTIs is mainly by the administration of antibiotics through various means; cutaneous, venous or muscularly.

In Nigeria, most UTIs cases in symptomatic patients usually indulged in indiscriminate use of antibiotics before consulting the physicians or clinicians, when they could no longer treat or control the symptomatic conditions. On the other hand, the physicians usually prescribe and treat the patients with wide broad-spectrum antibiotics without any microbiological findings (Aboderin *et al.*, 2009; Oli *et al.*, 2017). These deep indiscriminate use and inappropriate prescription of antibiotics in the treatment of UTIs are vital contributing factors to the emergence and widespread of bacterial resistance to the most commonly used antimicrobial agents (Sule *et al.*, 2018). The situation is deteriorating with the increase prevalence of substandard and fake antimicrobial agents marketed in the country (Nigeria) (Oli *et al.*, 2017).

Antibiotic resistant pathogens have degraded the choices of administering an effective antibiotic treatment (Cassir *et al.*, 2014; Moroh *et al.*, 2014). Furthermore, as the number of resistant strains increase in clinical environment, broad spectrum antibiotics becomes the paramount choice, but the incorporation of resistance to broad spectrum antibiotics in multidrug resistant strains degrades the chances of selecting an effective empirical treatment (Hirsch and Tam, 2010). While the prevailing crisis of AR has been reported elsewhere, the resistance status towards commonly used antibiotics at Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria is not yet known. There is a limited report of AR in Lafia, Nasarawa State, Nigeria on commonly used antibiotics in the hospitals. The changing spectrum of microorganisms involved in UTIs necessitates

the need for continuous and regular antibiotic resistance (AR) surveillance of these organisms in order to guide empirical treatment in UTIs.

This cross-sectional designed research work was aimed at determining the prevalence and AR profiles of *S. aureus* and *E. coli* strains from patients with UTIs attending Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria.

## **MATERIALS AND METHODS**

### **Study Location**

The study was conducted among patients attending Urology Clinic in Dalhatu Araf Specialist Hospital (DASH) Lafia, Nasarawa State, Nigeria. Lafia is the state capital of Nasarawa state situated in the North central part of Nigeria lying at latitude 8°29'30" North of the equator and longitude 8°31'0" East of Greenwich Meridian (Akwa *et al.*, 2007). Lafia has a total inhabitant of 330, 712 (Census, 2006; Akwa *et al.*, 2007). It is currently reported that Lafia has a population of 127, 236 (WPR, 2019).

### **Ethical Approval**

Ethical approval for the study was secured from Dalhatu Araf Specialist Hospital ethical committee, in accordance with the code of ethics for biomedical research involving human subjects. The subjects were recruited after they were given health talk on the aim and importance of the study.

### **Consent**

The participants were consented and a form was given to them to fill and sign in other for them to participate fully in this study. A questionnaire was used to obtain demographic data of participants such as age, sex, use of catheter as well as risk factors. The information obtained from the participants was assured to be confidential and kept out of third-party disposal.

A total of approximately 194 samples were calculated based on prevalence rate of 14.8%. However, 202 patients were selected for the study to allow for non-response.

### **Sample Collection**

A total of 202 urine samples were collected from patients attending Urology Clinic of the hospital between July 2019 and September 2019. The recruitment of the study patients was randomized, as only patients attending the clinic were used irrespective of their complaints or whether they have symptoms associated with UTIs. The patients were told how to collect urine sample. Mid-stream urine specimens were collected from each of the patient between 6a.m. and 9a.m. into sterile bottles. Their caps were tightly closed then stored in iced packed

containers for onward transport to the laboratory. To avoid cross-contamination during sampling, sterile gloves and forceps were used. The samples were immediately transported to the Microbiology laboratory of Federal of University, Lafia for culture and other microbiological analysis.

### **Isolation and Identification of Test Organisms**

Isolation of *S. aureus* and *E. coli* were carried out with the aid of a standard wire loop (0.001 ml), where the urine from the sample container were aseptically streaked onto Mannitol Salt Agar (MSA) and Eosin Methylene Blue Agar (EMBA) media plates and incubated at 37°C for 24hrs as described by Cheesbrough, (2006). Greenish metallic sheen colonies that grew on EMBA were selected as presumptive *E. coli* while those appearing yellow with color change on MSA plates were selected as presumptive for *S. aureus*. The presumptive isolates were identified through microscopy (Gram stain) and minimum selective biochemical tests notably catalase, coagulase, indole and citrate tests respectively. The isolates were then stored in a nutrient agar slant for subsequent analysis.

### **Antibiotic Profile:**

The antibiotic resistance profile test for *S. aureus* and *E. coli* were carried out as prescribed by Sule *et al.* (2018) following standard protocols laid by Clinical and Laboratory Standard Institute (CLSI), (2014). The pure isolates were standardized to turbidity of 0.5 McFarland standards before surface inoculation on Muller Hinton agar (MHA) using a sterile swab stick. The inoculated plates were allowed for 15 minutes to ensure the surface of the culture plate is dried and ready for antimicrobial testing. Antimicrobial discs of known concentration were placed on the surface of the inoculated MHA respectively (i.e. Gram-negative *E. coli* and Gram-positive *S. aureus*) using sterile forceps. The plates were then incubated in the incubator for 37°C for 18hrs, after which the zone of inhibition was measured using a metre rule. The antibiotics discs used include; Ofloxacin (10µg), Levofloxacin (10µg), Nalidixic acid (30µg), Ciprofloxacin (10µg), Augmentin (30µg), Gentamycin (10µg), Cotrimoxazole (30µg), Streptomycin (30µg), Cefalexin (10µg), and Ampicillin (30µg) while Gram positive disk contained Ciprofloxacin (10µg), Norfloxacin (10µg), Gentamycin (10µg), Streptomycin (30µg), Amoxicillin (20µg), Erythromycin (30µg), Rifampicin (20µg), Chloramphenicol (30µg), Ampicillin (20µg), and Levofloxacin (20µg) (Godwin, *et al.*, 2018).

**Statistical Analysis**

The data obtained in this research work were subjected to simple statistical analysis and analysis of variance (ANOVA). A p-value of less than or equal to 0.05 is considered to be statistically significant ( $P \leq 0.05$ ).

**RESULTS**

Of all of the 202 patients recruited for this study completed their questionnaire correctly giving a response rate of 100%, this was achieved through the entry point used in administering the questionnaires. There were 153 (75.74%)

males and 49 (24.26%) females. Out of which 88 (84.62%) and 16 (15.38%) male and female have significant bacteriuria respectively (Table 1). The use of catheter was observed in 65.35% participants with 67.31% having significant bacteriuria. The antimicrobial profile of *S. aureus* and *E. coli* from the result of this research work (Table 2), showed that Levofloxacin had the highest rate of sensitivity against both *S. aureus* (74.19%) and *E. coli* (64.29%). However, *S. aureus* was highly resistant (93.55%) to Rifampicin whereas *E. coli* was most resistant (89.00%) to Nalidixic acid.

**Table 1:** Socio-demographic characteristics and Anthropogenic factors associated with Urinary Tract Infections.

<b>Risk Factors</b>	<b>No. of Samples Collected (n =202)</b>	<b>Samples with Sig. Bacteriuria</b>	<b>% of Sig. Bacteriuria</b>
<b>Age group (yrs)</b>			
1 – 29	5	2	1.92
30 – 39	35	60	57.69
40 – 49	10	6	5.77
50 – 59	21	5	4.81
60 – 69	55	3	2.88
≥70	76	28	26.92
<b>Gender</b>			
Male	153	88	84.62
Female	49	16	15.38
<b>Use of catheter</b>			
Yes	132	70	67.31
No	70	34	32.69
<b>Risk factors</b>			
Antibiotic admin.			
Present	11	8	7.69
1 week before	21	16	15.38
2 weeks before	32	20	19.23
1 month before	50	40	38.47
Not on drugs	88	20	19.23
Total	202	104	100

N= total number of samples, No.= number, yrs= years, admin= administration, Sig.= Significant.

**Table 2:** Antibiotic resistance profile of *Staphylococcus aureus* and *Escherichia coli* (N = 62; n = 42)

Antibiotics	Disk content ( $\mu\text{g}$ )	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>
CPR	10	13(79.03)	13(69.04)
LEV	20	5(74.19)	15(64.29)
RD	20	4(93.55)	
S	30	10(83.87)	6(85.71)
NB	10	14(77.42)	
ERY	30	8(87.10)	
CN	10	7(88.29)	6(85.71)
CH	30	13(79.03)	
APX	20	18(70.97)	
AMX	30	16(74.20)	
OFL	10		12(71.43)
NA	30		11(89.00)
AUG	30		6(85.71)
SXT	30		10(76.19)
CEP	10		11(89)
PN	30		14(66.67)

**Key:** N = number of positive *S. aureus*, n = number of positive *E. coli*, CPR = ciprofloxacin, LEV = levofloxacin, RD = rifampicin, S = streptomycin, NB = norfloxacin, ERY = erythromycin, CN = gentamycin, CH = chloramphenicol, APX = ampicillin, AMX = amoxicillin, SXT = cotrimoxazol, PN = ampicillin, OFL = ofloxacin, CEP = cefalexin, NA = nalidixic acid, AUG = augmentin.

## DISCUSSION

Urinary tract infections (UTIs) have been documented to be caused mostly by Gram-negative enterobacteria with *E. coli* being the most prevalent (Onanuga and Awhowho, 2012; Moroh *et al.*, 2014; Tahira *et al.*, 2020). However, there is an increasing prevalence of *S. aureus* as a UTIs etiologic agent with a fearful rate of developing antimicrobial resistance (Onanuga and Awhowho, 2012). Resistance empowers bacteria to escape the attack by antibiotics and in turn slows the ability to treat infections (Spellberg *et al.*, 2008). Thus, antibiotics resistance has been considered one of the vital threats to medicine (Walker *et al.*, 2009).

Urinary Tract Infection is a major cause of morbidity and mortality. Among a total number of 202 patients analyzed in this study, the prevalence of 51.49% positive isolates was recorded. This is closely related to a previous study carried out by Otajevwo and Amedu, (2015) which they recorded a prevalence rate of 59.2% in Evbuobanosa, Edo State. However, this prevalence was more than those reported by Iregbu and Nwajobi, (2013) and Aiyegoro *et al.* (2007) in Abuja and Ile-Ife both in Nigeria which they recorded a prevalence of 13% and 11.9% respectively. The variations established in the studies could be attributed to variations in study populations and/or in the criteria employed by various centers in selecting urine samples for culture.

The prevalence of *S. aureus* in this study was 62 (30.69%) which is not far from that reported by Martin *et al.* (2019) while that of *E. coli* recorded was 42 (20.79) which shows a great variation with the work conducted by Martin *et al.* (2019) who reported a prevalence rate of 36/86 (41.9%) and also the work reported by Mayanga *et al.* (2005) who reported an estimated prevalence range between 40 to 46% of *E. coli*. The infection with *S. aureus* indicates contamination during instrumentation and also possibly due to contamination with the normal flora of the hands (Iregbu and Nwajobi, 2013). The findings in this research revealed that UTIs are more prevalent among males compared to their female counterparts. This higher prevalence in males may be attributed to the fact that more males attend the clinic than females as well as inadequate awareness of UTIs with emphasis on the danger it carries to them and the general public. This is in contrast with reports of Derby *et al.* (2017) in which they started the nature of female anatomy being a major factor contributing to the high prevalence in females than their male counterparts. However, the finding in this research was in accordance with that reported by Otajevwo and Amedu, (2015). In their study, they reported a prevalence rate of 57.1% (compared to 51.49% obtained in this research work) in males without stating clearly the reason for that. However, this may be attributed to lack of circumcision, receptive anal intercourse and HIV infection among the study participants.

This research work revealed that Gram positive bacteria isolates were more prevalent (61.54%) than Gram negative bacteria (40.38%). This is however, contrary to the study carried out by Tosin *et al.* (2018) in a tertiary hospital in South West, Nigeria, who reported high prevalence rate of 96.2% among Gram negative bacteria than in Gram positives bacteria of 3.8%.

From this research work, it was also revealed that Levofloxacin had the highest inhibition against the bacterial isolates which is in agreement with the work carried out by (Chibuike *et al.*, 2014; Agbagwa and Jirigwa, 2015; Ajao and Yakubu, 2015).

Considering age groups in this study, UTIs occurred highest in the age group of 30-39 years having a prevalence rate of 60 (57.69%). This could be attributed to the fact that, at this age, people are sexually active. This prevalence rate was followed by people in the age brackets of 70 years and above with prevalence rate of 28 (26.92%). This could be as a result of weakened immune system, less sexual activity and weakened urogenital system (possibly due to prostate). This finding is similar to the study done by Tosin *et al.* (2018) who reported higher incidence rate in the age range between 30 – to 49 years with prevalence rates of 18 (40%). Urethral catheterization has been established to be a great risk factor for UTIs and was reported to have a prevalence rate of 70 (67.31%) in this research work. This is however far from the value obtained by Tosin *et al.* (2018), who reported a prevalence rate of 32.5% among catheterized patients.

The finding from this work revealed multiple drug resistance in the two organisms (*S. aureus* and *E. coli*). This may possibly indicate that some of the resistance were as a result of antibiotic misuse or abuse or possibly that the isolates were carrying plasmids for drug resistance. This is similar to the finding of Eugene *et al.* (2013) who reported that resistance shown to antibiotics by the bacteria might be due to various reasons such as gross misuse of these drugs, over production of target sites, inactivation of the antibiotic, alteration of membrane permeability and spontaneous mutation through DNA transfer amongst others.

## REFERENCES

- Aboderin, O. A., Abdu, A., Odetoyin, B. W. and Lamikanra, A. (2009). Antimicrobial resistance in Escherichia coli strains from urinary tract infections. *J Natl Med Assoc*, **101**:1268-73.
- Abulreesh, H. H., and Organji S. R. (2011). The prevalence of multi-drug resistant staphylococci in food and the

## CONCLUSION

There is emergence of multiple drug resistance *S. aureus* and *E. coli* causing UTIs among patients visiting DASH Lafia. Their prevalence was high among male patients compared to their female counterpart and the occurrence was more among patients within the age group of 30-39 years. This high rate of resistance could be associated with self-prescription, incomplete dosage and indiscriminate use of antibiotics. Bacteria strains resistance to multiple antibiotics exists among patients suffering from UTIs.

## Limitation and Recommendation

The findings in this study are only limited to two hundred and two (202) samples, a national antibiotic resistance surveillance of these isolates is recommended for further study. Public awareness should be increased on the impacts caused by indiscriminate use and self-prescription of antibiotics. Development and strict adherence to antibiotic policy should be implemented to guide the sale, prescription and use of antibiotics by the masses and health care workers.

## Contributions of Authors

This work was carried out in collaboration between all authors. Authors ASR and FNJ designed the study, help in sample collection, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Authors MCO and YY managed the analyses of the study. Authors ASR and YY managed the literature searches and wrote the final manuscript. Author CA proofread and edited the final manuscript. All authors read and approved the final manuscript.

## Competing Interests

Authors have declared no competing interests.

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environment of Makkah, Saudi Arabia. *Research Journal of Microbiology*, **6** (6):510-523.

- Agbagwa, O. E and Jirigwa, C. E. (2015). Antibiotics resistance and plasmid profile of Staphylococcus aureus from wound swabs in Port Harcourt Nigeria. *Current Research in Bacteriology*, **8**:70-76.

- Aiyegoro, O. A., Igbinsola, O. O., Ogunmwonyi, I. N., Ojadjaro, E., Igbinsola, O. E., and Okoh, A. I. (2007). Incidence of urinary tract infections (UTI) among children and adolescents in Ile-Ife, Nigeria. *African Journal of Microbiology Research*, **1**:13-19.
- Ajao, T. and Yakubu, S. E. (2015). Identification, characterization and plasmid profiling of multi drug resistant nocomial pathogens isolated from selected hospitals in Ilorin metropolis. *British Microbiology Research Journal*, **5**(1):33-43.
- Akingbade, O. Balagun, S., Ojo D., Akinduti, P., Okerent Ugba P. O., Nwanze, J. C. and Okonko, O. I. (2014). Resistant plasmid profile analysis of multi-drug resistant *Escherichia coli* isolated from urinary tract infections in Abeokuta, Nigeria. *African Health Sciences*, **14**(4):821- 828.
- Akinsete, A. M., and Ezeaka, C. (2018). Prevalence and risk factors of asymptomatic bacteriuria among children living with HIV in Lagos, Nigeria. *Pan African Medical Journal*, **31**:181.
- Akortha, E. E. and Ibadin, O. K. (2008). Incidence and antibiotic susceptibility pattern of *Staphylococcus aureus* amongst patients with urinary tract infection (UTIS) in UBTH Benin City, Nigeria. *Afr J Biotechnol*, **7**:1637-40.
- Akwa, V. L., Bimbol, N. L., Samaila, K. L. and Macus, N. D. (2007). *Geography Perspective of Nassarawa State*. Onaivi Printing and Publishing Company, Keffi, Nigeria. 3-5.
- Alo, M. N., Saidu, A. Y., Ugah, U. I., and Alhassan, M. (2015). Prevalence and Antbiogram of Bacterial Isolates Causing Urinary Tract Infections at Federal Teaching Hospital Abakaliki I (FETHA I). *British Microbiology Research Journal*, **8**(2): 403-417.
- Anejo-Okopi, A. J., Okwori A. E. J., Eze, M. I., Onaji, A. I., Ali. M., Adekwu, A., Ejiji, I. S. (2015). Prevalence and Antibiotic Resistance Pattern of Urinary Tract Bacterial Infections among Symptomatic Patients Attending University of Maiduguri Teaching Hospital, North East Nigeria. *European Journal of Advanced Research in Biological and Life Sciences*, **3**(3): 31-41.
- Anejo-Okopi, J. A., Okojokwu, O. J., Ramyil, S. M.-C., Bakwet, P. B., Okechalu, J., Agada, G., Bassi, P. A., and Adeniyi, S. D. (2017). Bacterial and antibiotic susceptibility pattern of urinary tract infection isolated from asymptomatic and symptomatic diabetic patients attending tertiary hospital in Jos, Nigeria. *Trends in Medicine*, **17**: 1-5.
- Bashir, M. F., Qazi, J. I., Ahmad, N. and Riaz, S. (2008). Diversity of urinary tract pathogens and drug resistant isolates of *Escherichia coli* in different age and gender groups of Pakistanis. *Trop J Pharm Res*, **7**:1025-31.
- Blair, K. A. (2007). Evidence based Urinary Tract Infection across the life span: Current Updates. *Journal of Nursing Practice*, **3**:629-632.
- Cassir, N., Rolain, J. M. and Brouqui, P. (2014). A new strategy to fight antimicrobial resistance: the revival of old antibiotics. *Frontiers in microbiology*. **5**:551. <https://doi.org/10.3389/fmicb.2014.00551> PMID: 25368610
- Census, (2006). Retrieved from: "<https://en.wikipedia.org/w/index.php?title=Lafia&oldid=917958901>"
- Centre for Disease Control and Prevention (CDC) (2015). Urinary tract infection. Archived from the original on February 2016. 10-15.
- Chamoun, K., Farah, M., Araj, G., Daoud, Z., Moghnieh, R. and Salameh, P. (2016). Surveillance of antimicrobial resistance in Lebanese hospitals: retrospective nationwide compiled data. *International journal of infectious diseases*. **46**:64-70. <https://doi.org/10.1016/j.ijid.2016.03.010> PMID: 26996458
- Cheesbrough, M. (2006). Antibiotics. *District laboratory practice in tropical countries, Cambridge University United Kingdom, Part 2*. 63-70.
- Chibuikwe, I., Reginald, A. O. and Solomon, C. U. (2014). Antibiotic resistance patterns and plasmid profiles of methicillin resistant *Staphylococcus aureus* isolates from human samples. *British Microbiology Research Journal*, **4**(2):185-194.
- Clinical Laboratory Standards Institute (CLSI), (2016). Performance Standards for antimicrobial susceptibility testing. 26 ed. Wayne PA: Clinical and laboratory standards institute; 2016. 252.
- Dash, M., Padhi, S., Mohanty, I., Panda, P., Parida, B. (2013). Antimicrobial resistance in pathogens causing urinary tract infections in a rural community of Odisha, India. *Journal of Family and Community Medicine*, **20**(1): 20-26.

- Derbie, A., Hailu, D., Mekonnen, D., Abera, B., Yitayew, G. (2017). Antibiogram profile of uropathogens isolated at Bahir Dar Regional Health Research Laboratory Centre, Northwest Ethiopia. *Pan African Medical Journal*, **26**, 134.
- Dielubanza, E. J., and Scheaffer, A. J. (2011). Urinary tract infections in women. *The Medical clinic of North America*, **95**(1):27-41.
- Dugdale, C. D. (2011). Female urinary tract. Retrieved from: *MedlinePlus Medical encyclopedia*. <https://www.medlineplus.org/female-urinary-tract/24467>.
- Ehinmidu, J. O. (2003). Antibiotic susceptibility pattern of urine bacterial isolates in Zaria, Nigeria. *Tropical Journal of Pharmaceutical Research*, **2**(2):223-228.
- Ekwealor, P. A., Ugwu, M. C., Ezeobi, I., Amalukwe, G., Ugwu, B. C., Okezie, U., Stanley, C., and Esimone, C. (2016). Antimicrobial Evaluation of Bacterial Isolates from Urine Specimen of Patients with Complaints of Urinary Tract Infections in Awka, Nigeria. *International Journal of Microbiology*, Article ID 9740273. Available from: <http://dx.doi.org/10.1155/2016/9740273> (Accessed 2nd Sept. 2020).
- Eugene, O. T., Ekpo, M. A., Akinjogunla, O. J. and Idiong, D. F. (2013). Microorganisms associated with acute otitis media diagnosed in Uyo City, *Journal of Nigeria Science Researcher*, **4**(6):560-564.
- Eves, F. J., and Rivera, N. (2010). Prevention of urinary tract infections in persons with spinal cord injury in home health care. *Journal of Home Health Care Nurse*, **28**(4):230-241.
- Foxman, B. (2010). The epidemiology of Urinary tract infection. *Journal of natural and revised Urology*. **7**:653-660.
- Foxmann, B., Barlow, R., d-Arcy, H. (2000). Urinary tract infection: estimated incidence and associated costs. *Journal of Epidemiology*, **10**:509-15.
- Godwin, A. O., Ebele, U. U., Emmanuel, T. A., Aleruchi, C. and Peter, A. (2018). Incidence and antibiotic susceptibility pattern of *E. coli* and *Staph. aureus* isolated from meat pie sold in a Nigerian North Central town. *Journal of Medical Sciences*. **6**(1):21-28.
- Gould, C. V., Umscheid, C. A., Agarwal, R. K., Kuntz, G., and Pegues, D. A. (2010). Guideline for prevention of catheter associated urinary tract infections 2009. *Infections Control and Hospital Epidemiology*, **31**(4):319-26.
- Helen, S. L., and Jenifer, L. (2018). Urinary tract infections. *Journal of Infectious Disease*, **7**:28.
- Hirsch, E.B. and Tam, V. H. (2010). Impact of multidrug-resistant *Pseudomonas aeruginosa* infection on patient outcomes. *Expert review of Pharmacoeconomics & outcomes research*, **10**(4):441-51.
- Hollenbeck, B. L. and Rice, L. B. (2012). Intrinsic and acquired resistance mechanisms in enterococcus. *Virulence*. **3**(5):421-569. <https://doi.org/10.4161/viru.21282> PMID: 23076243
- Iregbu, K., and Nwajiobi, P. (2013). Urinary tract infection in a tertiary Hospital in Abuja, Nigeria. *African Journal of Clinical and Experimental Microbiology*, **14**(3):169-173.
- Kaper, J. B., Nataro, J. P., and Mobley, H. L. (2004). Pathogenic *Escherichia coli*. *National Revised Microbiology*, **2**(2):123-140.
- Khoshbakht, R., Salimi, A., Aski, H. S., Keshavarzi, H. (2013). Antibiotic Susceptibility of Bacterial Strains Isolated from Urinary Tract Infections in Karaj, Iran. *Jundishapur Journal of Microbiology*, **6**(1): 86-90.
- Kolawole, A. S., Kolawole, O. M., Kendaki Olukemi, T. T., Babatunde, S. K., Durowode, K. A., and Kolawole, C. F. (2009). Prevalence of Urinary Tract Infections (UTI) among patients attending Dalhatu Araf Specialist Hospital, Lafia, Nasarawa State, Nigeria. *International Journal of Medical Sciences*, **1**:163-167.
- Kose, Y., Abasiyanik, M. F. and Salih, B. A. (2007). Antibiotic resistance of *Escherichia coli* urinary tract isolates in Riza province, Turkey. *J Infect Dev Ctries*, **1**:147-50.
- Lane, D. R., and Takhar, S. S. (2011). Diagnosis and management of urinary tract infection and pyelonephritis. *Emergency Medicine Clinics of North America*, **29** (3):539-552.
- Mayanga, R., Kiggundu, C., and Kaddu-Mullindwa, D. (2005). The prevalence of asymptomatic bacteriuria and associated factors among women attending antenatal clinics in lower Mulago Hospital. *M.Ed. dissertation of Mayanga, Makerere University, Kampala, Uganda*, 1-200.



- Manikandan, S., Ganesapandian, S., Singh, M. and Kumaraguru, A. K. (2001). Antimicrobial susceptibility pattern of urinary tract infection causing human pathogenic bacteria. *Asian J Med Sci*, **3**:56-60.
- Martin, O., Adamu, A. A., Julius, T., Josephat, N. M., Eddie, W., Charles, D. K., Ezera, A., and Joel, B. (2019). Prevalence of bacterial urinary tract infection and associated factors among patients attending Hospitals in Bushenyi district, Uganda. *Hindawi International Journal of Microbiology*, **20**(19):1-8.
- Martinez, J. L. (2012). Natural antibiotic resistance and contamination by antibiotic resistance determinants: the two ages in the evolution of resistance to antimicrobials. *Frontiers in microbiology*, **3**:1-8. <https://doi.org/10.3389/fmicb.2012.00001> PMID: 22275914
- McLaughlin, S. P., and Carson, C. C. (2004). Urinary tract infections in women, *Medical Clinics of North America*. **88**(2):417-429.
- Mincey, B. A. and Parkulo, M. A. (2001). Antibiotic prescribing practices in a teaching clinic: Comparison of resident and staff physicians. *South Med J*, **94**:365-9.
- Moges, F., Genetu, A., and Mengstu, G. (2003). Antibiotic sensitivity of common bacterial pathogens in urinary tract infection at Gondat Hospital, Ethiopia. *East Africa Medical Journal*, **79**:140-142.
- Morphy, S. C. (2006). Malaria and Global infectious diseases: why should we care? *Journal of Virtual Mentor*, **8**(4):245-50.
- Moroh, J.-L. A., Fleury, Y., Tia, H., Bahi, C., Lietard, C., Coroller, L., Edoh, V., Coulibaly, A., Labia, R., and Leguerinel, I. (2014). Diversity and antibiotic resistance of uropathogenic bacteria from Abidjan. *African Journal of Urology*, **20**, 18-24.
- Morris, B. J., and Wiswell, T. E. (2013). Circumcision and lifetime risk of urinary tract infection: a systematic review and meta-analysis. *The Journal of Urology*, **189**(6):2118-2122.
- Muhammed, M. (2015). Urinary Tract Infections amongst pregnant women attending a medical Centre in Kaduna, Nigeria. *African Journal of Clinical and Experimental Microbiology*, **16**(10): 7-11.
- Nicolle, L. E. (2008). Uncomplicated Urinary tract infections in adults including uncomplicated pyelonephritis. *Urology Clinic of North American*, **35**(1):1-12.
- Nolte, O. (2014). Antimicrobial resistance in the 21st century: a multifaceted challenge. *Protein and peptide letters*, **21**(4):330-5. <https://doi.org/10.2174/09298665113206660106> PMID: 24164264
- Nwanze, P.I., Nwaru, L. M., Oranusi, S., Dimkpa, U., Okwu, M.U. and Babatunde, B.B. (2007). Urinary tract infection in Okada village: Prevalence and antimicrobial susceptibility pattern. *Sci Res Essay*, **2**:112-6.
- Ochada, N. S., Nasiru, I. A., Thairu, Y., Okanlowan, M. B., Abdulakeem, Y. O. (2015). Antimicrobial Susceptibility Pattern of Urinary Pathogens Isolated from two Tertiary Hospitals in Southwestern Nigeria. *African Journal of Clinical and Experimental Microbiology*, **16**(1): 12-22.
- Ojiegbe, G. C., and Nworie, W. C. (2000). Asymptomatic bacteriuria among school pupils in Enugu Urban Areas. *Journal of Medical Science*, **9**:42-46.
- Oli, A. N., Akabueze, V. B., Ezeudu, C. E., Eleje, G. U., Ejiofor, O. S., Ezebialu, I. U., Oguejiofor, C. B., Ekejindu, I. M., Emechebe, G. O., and Okeke, K. N. (2017). Bacteriology and Antibiogram of Urinary tract infection among female patients in a Tertiary Health Facility in South Eastern Nigeria. *The Open Microbiology Journal*, **11**: 292-300.
- Onanuga, A., and Awhowho, G. O. (2012). Antimicrobial resistance of *Staphylococcus aureus* strains from patients with urinary tract infections in Yenagoa, Nigeria. *Journal of Pharmacy and Bioallied Sciences*, **4**: 226-30.
- Otajewwo, F. D., and Amedu, S. S. (2015). Community acquired urinary tract infection prevalence in a tertiary institution based in Ewuobanosa, Edo state, Nigeria. *Global Journal of Medical Research*, **15**:52-63.
- Otajewwo, F. D. (2013). Urinary tract infection among symptomatic out-patients visiting a tertiary hospital based in Midwestern Nigeria. *Global Journal of Health Science*. **5**:17-99.
- Phipps, S., Lim, Y. N., McClinton, S., Barry, C., Rane, A., and N-Dow, J. (2006). Short term urinary catheter policy following urogenital surgery in adults. *Cochrane*

- Database of Systematic Reviews*, **3**(2):43-74.
- Randrianirina, F., Soares, J. L., Carod, J. F., Ratsima, E., Thonnier, V. and Combe, P. (2007). Antimicrobial resistance among uropathogens that cause community-acquired urinary tract infections in Antananarivo, Madagascar. *J Antimicrob Chemother*, **59**:309-12.
- Raufu, A. (2002). Influx of fake drugs to Nigeria worries health experts. *BMJ*, 324:698.
- Reis, A. C. C., Santos, S. R. S., deSouza, S. C., Saldanha, M. G., Pitanga, T. N., and Oliveira, R. R. (2016). Ciprofloxacin resistance pattern among bacteria isolated from patients with community-acquired urinary tract infection. *Revisita do Instituto de Medicina Tropical de SaóPaulo*, 58.
- Siti, N. M. Z., Yus, A. Y., Norio, S., Koji, I., Masafumi, G., Motoo, U., Noraziz, O., Zuriati, Z. and Hirofumi, H. (2016). Phenotypic and genetic characterization of multidrug resistant *Staphylococcus aureus* in the Tropics of South-East Asia. *Journal of Microbiology*, **162**:2064-2074.
- Sobel, J. O. and Kaye, D. (2014). Urinary tract infections. In: Mandell, G. L., Bennett, J. E. eds. *Principles and practice of infectious diseases*, 8<sup>th</sup> ed. Philadelphia: Elsevier Saunders. 886-913.
- Spellberg, B., Gidos, R. and Gilbert, D. (2008). The epidemic of antibiotic-resistant infections: a call to action for the medical community from the Infectious Diseases Society of America. *Clin Infect Dis*, **46**:155-64.
- Stamm, W. E. and Norrby, S. R. (2001). Urinary tract infections: disease panorama and challenges. *Journal of Infectious Diseases*, **183**(1):1-4.
- Sule, H., Uba, A., and Kumurya, A. S. (2018). Antibiotic susceptibility pattern of uropathogens from some selected hospitals in Kano-Nigeria. *Journal of Microbiology and Experimentation*, **6**(2): 127-134.
- Tahira, F., Rafiq, S., Iqbal, A., and Husnain, S. (2020). Prevalence and Antibiogram of MDR *E. coli* Strains Isolated from UTI Patients-1-Year Retrospective Study at Nishtar Medical Hospital, Multan. *SN Comprehensive Clinical Medicine*, **2**: 423-431.
- Taylor, R. B., Shakoor, O., Behrens, R. H., Everard, M., Low, A. S. Wangboonskul, J. M. (2001). Pharmacopoeial quality of drugs supplied by Nigerian pharmacies. *Lancet*, **357**:1933-6.
- Timothy, O. O., Olusesan, F. J., Adesola, B. O., Temitayo, A. A., Fayomi Oluwaseun David, Oyekale Oluwatoyin Ige. (2014). Antibiotic resistance pattern of bacterial isolates from cases of urinary tract infections among hospitalized and out-patients at a tertiary health facility in South Western Nigeria. *Annals of Tropical Medicine and Public Health*, **7**(2): 130-135.
- Tosin, T. O., Akinwumi, A. A., Adeyemi, O., and Oluseyi, A. A. (2018). Urinary tract infections and antibiotic sensitivity pattern of uropathogens in a tertiary hospital in South West, Nigeria. *Sahel Medical Journal*, **21**(1):18-22.
- van Hoek, AHAM., Mevius, D., Guerra, B., Mullany, P., Roberts, A. P. and Aarts, HJM. (2011). Acquired antibiotic resistance genes: an overview. *Frontiers in microbiology*, **2**:203-207. <https://doi.org/10.3389/fmicb.2011.00203> PMID: 22046172
- Walker, B., Barrett, S., Polasky, S., Galaz, V., Folke, C., Engström, G., Ackerman, F., Arrow, K., Carpenter, S., Chopra, K., Daily, G., Ehrlich, P., Hughes, T., Kautsky, N., Levin, S., Mäler, K. G., Shogren, J., Vincent, J., Xepapadeas, T. and deZeeuw, A. (2009). Environment looming global-scale failures and missing institutions. *Science*, **325**(5946):1345-6.
- Warren, J. W., Abrutyn, E., Hebel, J. R., Johnson, J. R., Schaeffer, A. J. and Stamm, W. E. (1999). Guidelines for antimicrobial treatment of uncomplicated acute bacterial cystitis and acute pyelonephritis in women. *Clin Infect Dis*, **29**:745-58.
- World Population Review (WPR) (2019). *Population of Cities in Nigeria*. Retrieved from: <https://worldpopulationreview.com/countries/nigeriapopulation/cities/>. Tuesday, 15/10/2019:21;18.
- Wyngaarden, J. B., Smith, L. H., and Benneth, J. C. (2002). Hospital acquired infections. In: *Cecil Text-Book of Medicine*, 19<sup>th</sup> edition. By Saunders, W. B. 123-240.
- Yusuf, I., Arzai, A. H., Haruna, M., Sharif, A. A., and Getso, M. I. (2014). Detection of multidrug resistant bacteria in major hospitals in Kano, North-west, Nigeria. *Brazilian Journal of Microbiology*, **45**(3):791-798.