# Molecular Identification of *Onchocerca* Species among Residents of Benue and Cross River States, Nigeria, Using Known Microsatellites and Mitochondrial DNA

#### Odama Richard Ikani, Agada Ali Samuel<sup>1</sup>, Amechi Nmesomachi Joy<sup>2</sup>, Kenechukwu Chibuike Onyekwelu<sup>2</sup>, Akighir John<sup>3</sup>, Ezeh Charles Ogbonna<sup>2</sup>

Department of Biochemistry, Faculty of Basic Medical Sciences, College of Medicine, University of Calabar, <sup>1</sup>Department of Medical Biochemistry, College of Health Sciences, Benue State University, Makurdi, <sup>2</sup>Department of Medical Biochemistry, College of Medicine, University of Nigeria, Enugu Campus, Enugu, <sup>3</sup>Department of Biochemistry, Pharmacological Biochemistry/Toxicological Unit, Federal University Wukari, Traba State, Nigeria

## Abstract

Background: Human onchocerciasis is a neglected tropical disease caused by Onchocerca volvulus and is transmitted by the species Simulium damnosum. Currently, onchocerciasis is estimated to have infected over 37 million people in tropical Africa, Yemen and Latin America, resulting in a debilitating eye and skin disease in more than 5million people with over 1.2 million cases of visual impairment or blindness. Also, an estimate of about 120 million people is at risk of contracting the disease due to the breeding habit of the vector. Community-directed ivermeetin administration has greatly reduced the infection burden in different parts of the world but they are persistent cases of onchocerciasis infection in Benue and Cross River States, Nigeria due to the terrain, relapse/poor ivermectin coverage, and COVID-19 outbreak. Aims and Objectives: To investigate the prevalence of onchocerciasis in these localities, this study reported the use of mitochondrial DNA and microsatellite markers for molecular identification studies of Onchocerca species amongst residents of Benue and Cross River State, Nigeria. Materials and Method: Three hundred (300) patients from Benue and Cross River State, Nigeria, 150 patients each from both states were screened using SD Bioline onchocerciasis test strip from South Korea with batch no: 61ADE002B for the detection of IgG4 antibodies against Ov16 in onchocerciasis. 25 were positive from Cross River State and 20 from Benue state. Six (6) from each state were sent for DNA extraction and PCR amplification (L1-6=Benue State, L6-12= Cross River) using polymerase chain reaction (PCR) amplification of mitochondrial cytochrome C oxidase subunit I (COXI) genes using HC02198: 5'-TAAACTTCAGGGTGACCAAAAAATCA-3' primers and the primers of the respective microsatellites; (GT)AT(GT)AT(GT)10, (GT)GC(GT)10, (CAG)2(CAA)10(CAG), and (GT)11TT(GT). Results: The results showed amplification of COX 1 subunit of the mtDNA of onchocerca volvulus at 344bp DNA sequence and amplification of (GT)AT(GT)AT(GT)10 microsatellites at 193bp, (GT)GC(GT)10 at 180bp, (CAG)<sub>2</sub>(CAA)<sub>10</sub>(CAG) at 209bp, and (GT)11TT(GT) 195bp respectively. Conclusion: This study clearly showed that active transmission of human onchocerciasis infection is still ongoing in Wanikade and Igede communities of Benue and Cross River State as evident by the skin lesions and depigmentation presented by a 55 years old woman and the molecular parasitological evidence of the incidence Onchocerca volvulus using the parasites' genetic materials. Therefore, we recommend intensifying community-directed ivermeetin intervention in these states.

Keywords: Benue and Cross River states, microsatellites, mitochondrial DNA, Nigeria, Onchocerca species

Received on: 24-08-21 Review completed on: 08-12-21 Accepted on: 25-12-21 Published on: \*\*\*

## INTRODUCTION

Onchocerciasis, as a serious neglected tropical disease (NTD), is caused by the filarial nematode *Onchocerca volvulus*, which can lead to blindness and chronic disability.<sup>[1,2]</sup> The filariae are a group of tissue-dwelling nematodes of vertebrates that are spread by blood-sucking arthropods, Onchocerca

Access this article online		
Quick Response Code:	Website: www.atpjournal.org	
	<b>DOI:</b> 10.4103/atp.atp_15_21	

Address for correspondence: Dr. Odama Richard Ikani, Department of Biochemistry, Faculty of Basic Medical Sciences, College of Medicine, University of Calabar, Calabar, Nigeria. E-mail: ikanirichard@yahoo.com

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow\_reprints@wolterskluwer.com

How to cite this article: Ikani OR, Samuel AA, Joy AN, Onyekwelu KC, John A, Ogbonna EC. Molecular identification of *Onchocerca* species among residents of Benue and Cross River states, Nigeria, using known microsatellites and mitochondrial DNA. Ann Trop Pathol 2022;XX:XX-XX.

volvulus.<sup>[2,3]</sup> Currently, onchocerciasis is estimated to have infected about 37million people in tropical Africa, Yemen, and Latin America.<sup>[3,4]</sup> resulting in a debilitating eye and skin disease in more than 5 million people,<sup>[3,5]</sup> with over 1.2 million cases of visual impairment or blindness.<sup>[2]</sup> Also, an estimate of about 120 million people is at risk of contracting the disease due to the breeding habit of the vector. The World Health Organization (WHO) recommends annual or biannual mass administration of ivermectin<sup>[2,6]</sup> as an eradication approach alongside vector control,<sup>[7,8]</sup> due to its significant human health and socioeconomic burden since the 1970s.[9-11] Clinical symptoms of onchocerciasis reflect the developmental stage of the parasites, and the immune response by the host and are highly variable. The clinical symptoms of onchocerciasis are often present after the L3 stage of the parasite matures into an adult worm between 9 months and 2 years and these symptoms include the formation of subcutaneous nodules (onchocercomata), generalized pruritus in early infection,<sup>[11]</sup> and skin rashes, extreme itching, eye cataract, itching of the eyes, light sensitivity, loss of skin elasticity, loss of vision [Figure 1]<sup>[12,13]</sup> in severe cases and change of the skin pigmentation [Figure 2]. These occur as a result of the inflammatory response to dying microfilaria and Wolbachia antigen (Punctuate keratitis/Snowflake opacities),<sup>[12,13]</sup> corneal fibrosis or opacification, and other ocular manifestation such as iridocyclitis, glaucoma, choroiditis and optic atrophy.[11,14]

Over the years, the WHO recommends ivermectin or moxidectin as the drugs of choice for treating onchocerciasis due to its micro-filaricidal properties,  $[^{7,11,13,15]}$  and it is administered at least once yearly for dosing interval of 3–12 months for a period of 10–12 years  $[^{7,16]}$  followed by vector control as elimination strategies.

However, there is a limitation to the use of Ivermectin in areas where Onchocerca volvulus co-exists with loa loa as it can result in severe adverse neurological events<sup>[7,11,16,17]</sup> and the sub-optimal (or atypical) responses to Ivermectin,<sup>[18]</sup> as well as the effect of COVID-19 pandemic, has resulted in the resurgence of onchocerciasis in the rural part of Benue and Cross River State, Nigeria. Also, in these tropical African regions, this illness is diagnosed through clinical manifestation and microscopic examination of skin samples for the presence of microfilaria. The limitation with this technique is that there is a high incidence of false-negative as in most cases microfilaria are not present in the skin samples of infected patients under examination and this has impaired researchers from avidly quantifying the prevalence of this illness in several regions.

Therefore, this study exploited the advances in molecular diagnosis, making use of genetic materials (mitochondrial deoxyribonucleic acid [Mt DNA] and microsatellites) peculiar to the concerned organism to cut down the cases of false negatives, and also scaled up quantifying the prevalence of this illness in these regions. The molecular identification of this parasite in Benue and Cross River State of Nigeria using known microsatellites and mitochondrial deoxyribonucleic acid (DNA) is essential<sup>[3]</sup> as this will support future basic and translational onchocerciasis research with particular relevance to the ongoing onchocerciasis elimination program and boost efforts to intensify community-based ivermectin intervention in these localities.

## MATERIALS AND METHODS

## **Study area**

Benue State; one of the North Central states in Nigeria (latitude 4°N and 14°N and longitude 2° and 15°E)<sup>[19]</sup> has a population of about 4,253,641 in the 2006 census, located at coordinate 7°20'N 8°45'E. It is inhabited predominantly by the Tiv, Idoma, Igede, and Etulo natives respectively with its capital in Makurdi. The territory was initially known as Munshi Province until 1918 when the name of its dominant geographical feature, the 'Benue River' was adopted. Cross River on the other hand is a state in South-South Nigeria that is located at coordinate 5°45'N 8°30'E,[20] bordering Cameroon to the east. Its capital is Calabar and its name is derived from the Cross River (Oyono), which passes through the state. English and French are the major foreign languages of the state, while Bekwarra, Bette people, Ejagham, and Efik<sup>[21]</sup> are major indigenous languages of this state. These states are made up of several streams which empty into these major rivers, making them a perfect breeding ground for black flies. After a careful literature review, Benue State and the Cross River State of Nigeria was chosen because of the high incidence of onchocerciasis in these states and these states are bounded by River Benue and Cross River State River Basin respectively and its tributaries which serve as a reservoir for the breeding of black flies which are the sole host of Onchocerca volvulus. According to Onekutu, et al., (2018)<sup>[22]</sup> in the community of Kuhe and Gube of Benue state of Nigeria, research on 546 persons showed a high prevalence of onchocerciasis at 61% and 71% respectively. Also, Opera, et al., (2006)<sup>[23]</sup> show that certain patients were blind due to onchocerciasis and were positive to onchocerciasis test in the lower Cross River basin.

### Screening test and sample collection

Three hundred patients with varying skin lesions from Benue and Cross River States, Nigeria, were recruited for this study. 150 each from both states and were screened using SD Bioline onchocerciasis test strip [Figure 3]<sup>[24]</sup> from South Korea with batch no: 61ADE002B for the detection of IgG4 antibodies against Ov16 in onchocerciasis. 25 were positive from Cross River state and 20 from Benue state. Six (6) from each state were sent for DNA extraction and PCR amplification (L1-6=Benue State, L6-12= Cross River).<sup>[24]</sup> The SD BIOLINE contains a membrane strip, which is coated with recombinant Ov16 capture antigen on the test line region. The anti-human IgG4-gold colloid conjugate and the specimen move along the membrane chromatographically to the test region (T) and form a visible line as the antigen-antibody gold particle complex is formed with a high degree of sensitivity and specificity.<sup>[24,25]</sup> Both test and control windows are clearly labelled "T" and "C" respectively.

#### Mitochondrial deoxyribonucleic acid extraction

Whole-cell DNA was extracted from fresh blood samples frozen at -20°C after collection of the specimen using QIAGEN DNeasy Blood & Tissue kit from Southern Cross Biotechnology Ltd, South Africa.<sup>[25]</sup> Samples were first lysed using proteinase K with buffering conditions adjusted to provide optimal DNA binding, and the lysate is loaded onto the DNeasy Mini spin column and centrifuged. During centrifugation, DNA is selectively bound to the DNeasy membrane as contaminants pass through. The remaining contaminants and enzyme inhibitors are removed in two efficient wash steps and DNA is then eluted in the buffer ready for use.

#### Polymerase chain reaction amplification

Onchocerciasis infection identification was done using PCR amplification of mitochondrial cytochrome C oxidase subunit I (COXI) genes using HC02198: 5'-TAAACTTCAGGGTGACCAAAAAATCA-3' primers. 25  $\mu$ l reaction mixtures contained 2.5  $\mu$ l of Super Taq 10x buffer, 0.5  $\mu$ l of HC02198 forward primer, 0.5  $\mu$ l of HC02198 reverse primer, 0.2  $\mu$ l of 100 mM dNTP, 1  $\mu$ l of DNA polymerase, 14.55  $\mu$ l of distilled water, 0.75  $\mu$ l of MgCl<sub>2</sub>, and 5  $\mu$ l of sample DNA. The PCR condition involved an initial denaturation step at 95°C for 2 min, followed by 37 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s with a final extension step at 72°C for 5 min.<sup>[25]</sup> The expected band size is 344 bp.

Amplification of *Onchocerca* microsatellites [Table 1] was done using PCR-based microsatellites' respective primers. All amplifications were performed using 25  $\mu$ l reaction mixtures containing 2.5  $\mu$ l of Super Taq 10x buffer, 0.5  $\mu$ l of the forward primer, 0.5  $\mu$ l of the reverse primer, respectively, 0.2  $\mu$ l of 100 mM dNTP, 1  $\mu$ l of DNA polymerase, 14.55  $\mu$ l of distilled water, 0.75  $\mu$ l of MgCl<sub>2</sub>, and 5  $\mu$ l of sample DNA. The PCR condition involved an initial denaturation step at 95°C for 2 min, followed by 37 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s with a final extension step at 72°C for 5 min. Then, 5  $\mu$ l of PCR product each was mixed with standard loading dye and electrophoresed alongside 50 bp DNA

Table 1: Onchocerca microsa	tellites, primer sequence,
and amplified band size	

Microsatellites	Primer sequence	Band sizes (bp)
(GT)AT(GT)	CCCATTTGCCAGTTGAGGTGA-F	193
AT(GT) <sub>10</sub>	CCCGTCAACATTGTGGCTACG-R	
(GT)GC(GT) <sub>10</sub>	GACGCATACCGAGTCCTTGT-F	180
	TACGCACACATTTTTTCTATTTC-R	
(GT) <sub>11</sub> TT(GT)	CGCTAACGCTGTGCAATATTG-F	195
	TGACGAACTTTGGGACGACA-R	
(CAG) <sub>2</sub> (CAA) <sub>10</sub>	CGACAACGTGTCTCGACAAA-F	209
(CAG)	CGAAAACAACATACGAAGGG-R	

ladder in 1.5% agarose stained with ethidium bromide (0.5  $\mu g/$  ml). The products were visualized and photographed under ultraviolet illumination.

## RESULTS

Twenty-five people were positive from Cross River state and 20 from Benue state using the onchocerciasis SD BIOLINE test strip. Six samples from each state were subjected to PCR amplification (L1–6 = Benue state and L 6–12 = Cross River).

On visualization, the results show amplification of COX 1 subunit of the mtDNA of onchocerca volvulus across L1-12 using PCR *HC02198: 5'-TAAACTTCAGGGTGACCAAAAAATCA-3* 9 [Figure 4].

Marked amplification of (GT)AT(GT)AT(GT)10 microsatellite across sample L1, L2, L3, L5, and L6 from Benue and sample L7, L8 and L11 from Cross River State using CCCATTTGCCAGTTGAGGTGA-F, CCCGTCAACATTGTGGCTACG-R microsatellites primer sequence were observed.

Furthermore, there was an amplification of (GT)  $GC(GT)_{10}$  microsatellite in the sample L3 and L5 from Benue State and sample L12 from Cross River State using GACGCATACCGAGTCCTTGT-F, TACGCACACATTTTTCTATTTC-R microsatellite sequence [Figure 5], as well as amplification of (CAG) <sub>2</sub>(CAA) <sub>10</sub>(CAG) O. volvulus microsatellites in the sample L1-L6 from Benue and L7, L9, L10, and L12 from Cross River state using CGACAACGTGTCTCGACAAA-F; CGAAAACAACATACGAAGGG-R microsatellite sequence [Figure 6] followed by amplification of (GT) <sub>11</sub>TT (GT) O. volvulus microsatellites in sample L3, L5, and L6 from Benue and L7-L12 from Cross River states using CGCTAACGCTGTGCAATATTG-F, TGACGAACTTTGGGACGACA-R microsatellite sequence, respectively [Figure 7].



Figure 1: Loss of vision<sup>[12,13]</sup>

## DISCUSSION

Over the years, microsatellites and Mt DNA are regarded as molecular tools<sup>[26]</sup> for species identification and characterization, diagnostic target for infectious diseases, and estimation of divergence on evolutionary timescales and have become essential in many areas of evolutionary biology, including system biology, biochemistry, and molecular biology.<sup>[26]</sup> Therefore, in this study, we employed microsatellites and Mt DNA to carry out molecular identification of *Onchocerca* species among patients from Benue state and Cross River state of Nigeria to extrapolate the prevalence of onchocerciasis infection on the molecular level.

The PCR primers HC02198 (5' TAAACTTCAGGGT GACCAAAAAATCA 3') [Figure 4], and microsatellite primers (5'-CCCATTTGCCAGTTGAGGTGA-3'-F, 5'-CCCGTCAACATTGTGGCTACG-3'-R; 5'-GACGCA TACCGAGTCCTTGT-3'-F, 5'-TACGCACACATTT TTCTATTTC-3'-R; 5'-CGCTAACGCTGTGCA ATATTG-3'-F, 5'-TGACGAACTTTGGGACGACA-3'-R; 5'-CGACAACGTGTCTCGACAAA-3'-F; 5'-CGAAAACAACATACGAAGGG-3'-R) were designed to amplify the COX 1 portion of O. volvulus Mt DNA (GenBank accession number AF015193)<sup>[27]</sup> and (GT) AT (GT) AT (GT)  $_{10}$  [Figure 8], (GT) GC (GT)  $_{10}$  [Figure 6], (CAG)  $_{2}$ (CAA) <sup>10</sup>(CAG) [Figure 6], (GT) <sub>11</sub>TT (GT) [Figure 7] microsatellite portions, respectively, of O. volvulus (GenBank accession number AF015193)<sup>[27]</sup> for identification of O. volvulus among onchocerciasis-infected patients. Primer size for both forward and reverse primers were 50bp each with an annealing temperature of 55°C. Visualisation of the gel image performed on the obtained sequences showed amplification of 344bp DNA sequence (EMBL GenBank accession: (O. volvulus AF228575, AF228574, AF228565))<sup>[27,28]</sup> which codes for O. volvulus COX subunit 1 gene (GenBank accession number accession HG738213.2)<sup>[26]</sup> and 193bp, 180bp, 209bp and 195bp microsatellites DNA sequence respectively<sup>[29]</sup> (EMBL GenBank accession: O. volvulus AF228565)<sup>[26,27]</sup> coding O. volvulus microsatellite portions. (GenBank accession number accession HG738213.2)[30]

Three hundred patients examined presented with varying features of onchocerciasis as seen in the case of a 55-year-old woman from Barracks-Wanikade, North Ukelle [Figure 2], with skin lesion and depigmentation, which shows evidence of ongoing incidence of onchocerciasis in these localities. The result of this study showed 30% prevalence rate in Benue and 37% Cross River prevalence when expressed against the 150 patients from each state. Cases of infection tend to be higher among people of this community because these are riverine areas and favor the breeding of black flies.<sup>[30]</sup> This agrees with the studies reported in different parts of Nigeria by Oyibo & Fagbenro, (2003)<sup>[31,32]</sup> (40%) on the Effect of repeated community-based Ivermectin treatment on the intensity of onchocerciasis in Nigeria; Adesina, *et al.*,(2017)<sup>[33]</sup> on the Incidence of onchocerciasis and vector knowledge among



Figure 2: A 55-year-old woman from Wanikade, North Ukelle, of Cross River state of Nigeria with skin lesion and depigmentation



Figure 3: SD BIOLINE onchocerciasis IgG4 (61FK10) rapid test strip<sup>[24]</sup>



**Figure 4:** A representative agarose gel picture showing the resolution of the amplified products of COX1 using HC02198: 5'-TAAACTTCAGGGTGACCAAAAAATCA-3'

residents of some part of Ondo State, Nigeria (75%); Onekutu, *et al.*,(2018)<sup>[22]</sup> on the prevalence and distribution of human onchocerciasis and dermatological features in Kwande Local Government Area of Benue State, Nigeria (61%–71%).

Further, all the samples that were positive using Standard BIOLINE Onchocerciasis test strips were subjected to PCR amplification of *O. volvulus* Mt DNA COX1 gene subunit and microsatellites. This showed marked amplification across the samples which ascertain that onchocerciasis infection remains a serious neglected tropical disease of public health concern in



Figure 5: A representative agarose gel picture showing the resolution of the amplified products of (GT) GC (GT) 10 microsatellite using GACGCATACCGAGTCCTTGT-F TACGCACACATTTTTCTATTTC-R in 5'-3' direction



**Figure 8:** A representative agarose gel picture showing the resolution of the amplified products of (GT) AT (GT) AT (GT) 10 microsatellite using CCCATTTGCCAGTTGAGGTGA-F CCCGTCAACATTGTGGCTACG-R in 5'-3' direction

Benue and Cross River state of Nigeria. This established a nexus between the persistent ongoing transmission of onchocerciasis infection in these states in Nigeria and the continuous exposure to black flies in these localities as confirmed by marked amplification of *O. volvulus* microsatellites and Mt DNA from infected patients. This shows that though the burden of onchocerciasis is reduced in some region of the country due to community-based ivermectin distribution, it is still of health concern in riverine areas of Benue and Cross River states of Nigeria. This is in agreement with a study carried out in Ghana which showed that despite the community-directed ivermectin treatment, the prevalence of *O. volvulus* infection still remains a health concern in different parts of the tropics.<sup>[2,28,34]</sup>

## CONCLUSION

Although the burden of onchocerciasis is reduced in different parts of the world, this study clearly showed that active



**Figure 6:** A representative agarose gel picture showing the resolution of the amplified products of (CAG)  $_{10}$ (CAA)  $_{10}$ (CAG) microsatellite



**Figure 7:** A representative agarose gel picture showing the resolution of the amplified products of (GT) <sub>11</sub>TT (GT) microsatellite

transmission of human onchocerciasis infection is still on going in Wanikade and Igede communities of Benue and Cross River States as evident by the skin lesions and depigmentation presented by a 55-year-old woman and the molecular parasitological evidence of the incidence *O. volvulus* using the parasites' genetic materials. Therefore, we recommend intensifying community-directed ivermectin intervention in these states.

#### **Ethical approval**

Ethical approval for this study was obtained from Research and Ethics Committee of College of Medicine, University of Nigeria, with ethical clearance notification. Protocol No: 082/12/2019 and Government of Benue State of Nigeria with Ref. MOH/OFF340/VOL/4P8912.

#### Acknowledgments

I want to use this medium to appreciate the Head of Department of Medical Biochemistry, Prof. (Mrs.) J. E. Ikekpazu for her motherly love and counsel during the course of this research and the staffs of the department of Medical Biochemistry, College of Medicine, University of Nigeria, Enugu Campus, Enugu, especially Prof. I. E. Ezeagu (of Blessed Memory), Prof. F. E. Ejezie, Prof. Cambell, Dr. M. D. Ibegbu and Dr. (Mrs.). I. Nwigwe, for the various roles they played in ensuring the completion of this research.

My thanks also go to the management of Bioinformatics Services and Staffs of International Institute of Tropical Agriculture, Ibadan for their efforts and assistance throughout the period of this research.

### **Declaration of patient consent**

The authors certify that they have obtained all appropriate patient consent forms. In the form the patient(s) has/have given his/her/their consent for his/her/their images and other clinical information to be reported in the journal. The patients understand that their names and initials will not be published and due efforts will be made to conceal their identity, but anonymity cannot be guaranteed.

## Financial support and sponsorship Nil.

**Conflicts of interest** 

There are no conflicts of interest.

## REFERENCES

- Akinbo FO, Okaka CE. Prevalence and socio-economic effects of onchocerciasis in Ovia North East Local Government Area of Edo State, Nigeria. Riv Parasitol 2005;22:215-21.
- Cotton JA, Bennuru S, Grote A, Harsha B, Tracy A, Beech R, *et al.* The genome of *Onchocerca volvulus*, the agent of river blindness. Nat Microbiol 2016;2:16216.
- 3. Choi Y, Tyagi R, McNaulty NS, Rosa AB, Ozersky P, Mafrtin J, *et al.* Genomic diversity in *Ocnhocerca volvulus* and its *Wolbachia endosymbiont*. Nat Microbiol 2017;2:16207.
- Dunn C, Callahan K, Katabarwa M, Richards F, Hopkins D, Withers PC Jr., *et al.* The contributions of onchocerciasis control and elimination programs toward the achievement of the millennium development goals. PLoS Negl Trop Dis 2015;9:e0003703.
- 5. Crump A, Morel CM, Omura S. The onchocerciasis chronicle: From the beginning to the end? Trends Parasitol 2012;28:280-8.
- Brady MA, Hooper PJ, Ottesen EA. Projected benefits from integrating NTD programs in sub-Saharan Africa. Trends Parasitol 2006;22:285-91.
- World Health Organization (WHO); 2018. Available from: http://www. who.int/news-room/factsheets/detail/onchocerciasis. [Last accessed on 2018 Aug 28].
- Centre for Disease Control (CDC). Onchocerciasis (also known as River Blindness) Prevention and Control. Parasite. CDC; 2013. Achieved from the Original on April 2014. [Last accessed on 2018 Aug 28].
- Cupp EW, Sauerbrey M, Richards F. Elimination of human onchocerciasis: History of progress and current feasibility using ivermectin (Mectizan(®)) monotherapy. Acta Trop 2011;120 Suppl 1:S100-8.
- Akagun BO. Onchocerciasis in Taraba state, Nigeria: Clinical epidemiology study of at-risk males in Bakundi district. Zentralbl Bakteriol 1999;289:371-9.
- Darvin SS, Muzzammil IM, Francisco T, John WK. Onchocerciasis (River blindness), treatment and management, Medline. 2018. Available from https://emedicine.mrdscape.com/article/224309-treatment. [Last accessed on 2018 Aug 28].
- Stewart D. Everything you should know about onchocerciasis (River blindness). 2017. Retrieved from https://www.healthline.com/health/ onchocerciasis-river-blindness #1. [Last accessed 2018 on 28].
- Ogbonna EC, Ikani OR. Systematic review on onchocerciasis infection in Nigeria in the past five decades. Int J Med Public Health 2020;10:1-7.

- 14. Kayembe DL, Kasonga DL, Kayembe PK, Mwanza JC, Boussinesq M. Profile of eye lesions and vision loss: A cross-sectional study in Lusambo, a forest-savanna area hyperendemic for onchocerciasis in the Democratic Republic of Congo. Trop Med Int Health 2003;8:83-9.
- Reddy M, Gill SS, Kalkar SR, Wu W, Anderson PJ, Rochon PA. Oral drug therapy for multiple neglected tropical diseases: A systematic review. JAMA 2007;298:1911-24.
- Onchocerciasis. WHO MediaCenter. 2017. Available from http://www. who.int/mediacentre/factsheets/fs374/en/. [Last accessed on 2018 Aug 28].
- Kelly-Hope LA, Cano J, Stanton MC, Bockarie MJ, Molyneux DH. Innovative tools for assessing risks for severe adverse events in areas of overlapping loaloa and other filarial distributions: The application of microstratification mapping. Parasit Vectors 2014;7:307.
- Sara L, James PM. Ivermectine resistance in onchocera volvulus: Toward a genetic basis. Negl Trop Dis 2007;1:e76.
- Africa: Nigeria. World Factbook of Central Intelligence Agency. 2011. Available from https://www.cia.gov/ library/publications/ the- world factbook/geos/ni. htmml. [Last accessed on 2020 Mar 14].
- Andem AB, Udofie UU, Okorafor KA, George UU. Bioaccumulation of some heavy metal and total hydrocarbon (THC) in the tissue of Periwinkle (*Tympanotonus fuscatus* Var Radula) in the intertidal regions of Qua Iboe River Basin, Ibero Akwa Ibom State, Nigeria. Gen J Biol Sci 2013;3:258-68.
- List of tribes in Cross River State, Nigeria. Available from https:// allnigeriainfo.ng/tribes-cross-river/. [Last accessed on 2021 Aug 15].
- Onekutu A, Ayom IF, Iboyi OM. Prevalence and distribution of human onchocerciasis and dermatological features in Kwande local government area of Benue State, Nigeria. Br J Med Med Res 2018;27:1-10.
- 23. Opera KN, Fegbemi BO, Atting IA, Oyene UE. State of forest onchocerciasis in lower cross river basin, Nigeria: Changes in clinical and parasitological indices after 6 years of Ivermectin intervention. J R Inst Public Health 2006;121;202-7.
- SD Bioline Onchocerciasis IgG4; 2020. Available at https://www. globalpointofcare .abbott/en/product-details/bioline-onchocerciasisigg4-elisa.html. [Last accessed on 2020 Sep 20].
- DNeasy Blood & Tissue Handbook 07/20064; 2020. Available from https://www.qiagen.com/ca/resources/resourcedetail?id=68f29296-5a9f-40fa-8b3d-1c14d0b3030&lang=en. [Last accessed on 2021 Mar 15].
- Saiki R, Gelfand D, Stoffel S, Scharf S, Higuchi R, Horn G, *et al.* Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. Science 1988;239:487-91.
- Ho S. The molecular clock and estimating species divergence. Nat Educ 2008;1:168.
- Crainey JL, Medeiros JF, Pessoa FA, Luz SL. Onchocerciasis. In: Marcondes CB, editor. Arthropod Borne Disease. Heidelberg: Springer; 2016.
- 29. Ta Tang TH, López-Vélez R, Lanza M, Shelley AJ, Rubio JM, Luz SL. Nested PCR to detect and distinguish the sympatric filarial species *Onchocerca volvulus, Mansonella ozzardi*, and *Mansonella perstans* in the Amazon Region. Mem Inst Oswaldo Cruz 2010;105:823-8.
- Dumas V, Herder S, Bebba K, Cadoux-Barnabé C, Christian B, Géarq K. Polymorphic microsatellites in *Simulium damnosum* S.I. and their use for differentiating two savannah populations: Implications for epidemiological studies. Genome 1998;41:154-61.
- Wogu MD, Okaka CE. Prevalence and socio-economic effects of onchocerciasis in Okpuje, Owan West Local Govt. Area, Edo State, Nigeria. Int J Biomed Health Sci 2008;4:113-9.
- Adesina FP, Awosolu OB, Olusi TA. Incidence of onchocerciasis and vector knowledge among residents of some part of Ondo state, Nigeria. Sci Acad Publ 2017;7:107-11.
- Adesina FP, Awosolu OB, Olusi TA. Incidence of onchocerciasis and vector knowledge among residents of some part of Ondo state, Nigeria. Scientific & academic publishing 2017;7:107-111.
- Hotez PJ. Control of onchocerciasis The next generation. Lancet 2007;369:1979-80.