

PRELIMINARY STUDIES ON THE PROTECTIVE EFFECT OF SHEA OIL ON THE GONADS OF MALE NORWEGIAN RATS INTOXICATED WITH UNTREATED REFINERY EFFLUENTS

¹EKAYE, Sese-Owei, ²UWAGIE-ERO, Edwin Aihanuwa, ³ODIGIE, Eugene Amienwanlen and ¹AGHAYEDO, Cosmos Oghogho

¹Department of Animal and Environmental Biology, University of Benin, Benin City, Nigeria.

²Department of Surgery, Faculty of Veterinary Medicine, University of Benin, Benin City, Nigeria.

³Department of Public Health, Faculty of Veterinary Medicine, University of Benin, Benin City, Nigeria.

Corresponding Author: Uwagie-Ero, E. A. Department of Surgery, Faculty of Veterinary Medicine, University of Benin, Benin City, Nigeria. **Email:** edwin.uwagie-ero@uniben.edu **Phone:** +234 8033977590

Received: September 14, 2018 **Revised:** February 4, 2019 **Accepted:** February 11, 2019

ABSTRACT

This study reports the possible ameliorative effect of Shea oil on the testes of Norwegian rats intoxicated with untreated refinery effluent. 3 groups of 10 animals each were treated with Effluent only, Effluent with Shea oil and an untreated control daily for 9 weeks. Rats were euthanised at 3, 6 and 9 weeks interval. After 9 week, rats were left for 21 days post-exposure to recuperate. Body weight, testicular weight and morphology, heavy metal concentration in testes and histopathology of testes were estimated across the groups. There was a significant increase ($p > 0.05$) in body and organ weights of treated rats compared to control. Gross organ morphology also varied widely between treated and control but this was not statistically significant. Lead and chromium concentrations in the testes were high in rats treated with effluent only. Histopathology of gonads also showed evidence of pathological effects in rats treated with effluent only as compared with control and ameliorated group. Rats treated and ameliorated with Shea oil showed normal histological architecture in their organs. The study concluded that Shea oil was effective in ameliorating the effects of untreated refinery effluent toxicity on the gonads of male rats.

Keywords: Shea oil, Refinery effluents, Testis, Amelioration

INTRODUCTION

A myriad of industrial activities cause environmental pollution, industries such as refinery and agrochemical industries contribute to the degradation of the environment as they release effluent that are toxic into the environment. According to the United State Environmental Protection Agency (USEPA), effluent (treated or untreated) generated from industrial plants are discharged to water bodies; with effluent from petroleum refinery been reported to be a major source of aquatic pollution (Wake, 2005). Refinery effluent majorly consist of several organic and inorganic compounds

including phenolic groups, organochlorides, grease and oil and also heavy metals such as chromium, lead, mercury and cadmium (Diya'uddeen *et al.*, 2011). Thus, the effect of petroleum refinery effluent on the environment is severe as; they are widely distributed and pose environmental and occupational exposure risks, which may result in adverse health effects to man (Permenter *et al.*, 2011). Exposure to effluent that contains heavy metals may occur through direct exposure to contaminated substrate such as soil, air, water and food or via consumption of aquatic lives through uptake and biomagnification (Marouani *et al.*, 2011).

Heavy metals found in petroleum refinery effluent such as chromium, lead, mercury, copper and zinc have received special attention in ecotoxicology (Guertin, 2005). Although, the biological functions of organisms require some of these metals in trace amount, exposure to high concentrations might be lethal or cause damages to cells and tissues in the body (Cohen *et al.*, 1993).

It has been reported that exposure to chromium (VI) causes reproductive toxicity in human and laboratory animals (Danadevi *et al.*, 2003; Subramanian *et al.*, 2006). Furthermore, a decreased concentration of sperm cells and increase in abnormal spermatozoa were observed in Swiss mice after chromic acid exposure (Acharya *et al.*, 2006), in rats (Marouani *et al.*, 2017), in rabbits exposed to chromium (Yousef *et al.*, 2006) and bonnet monkeys that were equally exposed to chromium (Subramanian *et al.*, 2006).

The therapeutic role of medicinal plants in ameliorating the toxicity effect of effluent and other contaminants has been widely reported (Chatterjee *et al.*, 2012; Adikwu *et al.*, 2013; Ugwu *et al.*, 2013). Essential oils are used as therapeutic supplements because they are rich in biologically active compounds. Shea butter oil, extracted from Shea tree (*Vitellaria paradoxa*) reportedly possesses antimicrobial, antifungal, antiviral and insecticidal properties (Kordali *et al.*, 2005). About 85 to 90 % of the fatty acid composition is stearic and oleic acids. A study had characterized and quantified the most important phenolic compounds in Shea oil (Maranz *et al.*, 2004). Phenolic compounds that are a major constituent of Shea oil are known to have antioxidant properties against tissue damage. Assessment of the toxic effect of effluent have been carried out using animal models such as the Norwegian rat (*Rattus norvegicus*) in order to extrapolate the toxic effect of these harmful substances and compounds in humans (Isselhard and Kushe, 1998; Giridharan *et al.*, 2000; Ihedioha *et al.*, 2004).

For a clearer insight into the effects of exposure to untreated refinery effluent and its wider effect on the environment, an assessment on the toxicity response of *Rattus norvegicus*

exposed to untreated refinery effluent with possible abatement using Shea oil was carried out. The specific aims of this study therefore were to; determine the level of heavy metal concentration on the testicles of the rats and observe the histopathological alteration in the testes of the treated rats. The study also evaluated the possible ameliorative effect of Shea oil on intoxicated rats and determined if rats can recuperate over a short period of time post exposure to effluent.

MATERIALS AND METHODS

Experimental Design: Thirty adult male albino rats were used for the study. Rats were grouped randomly into three groups of 10 animals each. Rats in Group 1 were given feed and drinking water *ad-libitum* all through the experiment. Group 2 rats were given feed and drinking water *ad-libitum* and 2 ml of 100 % untreated refinery effluent daily for 9 weeks *per os*. Group 3 rats were given feed and drinking water *ad-libitum*, 2 ml of 100 % of refinery effluent and 2 ml of Shea oil 1 hour after, daily for 9 weeks. Every three weeks, two rats were euthanized from each group (1 – 3). Blood and testicular tissue samples were collected and analysed. Treatment was discontinued after nine weeks. The remaining rats in groups 2 and 3 were designated groups 4 and 5. They were untreated for additional three weeks (representing weeks 10 – 12 recovery period), but given food and drinking water *ad-libitum*. Blood and testicular tissue samples from groups 4 and 5 were collected and analysed after this post exposure phase as well.

Refinery Effluent Collection: Refinery effluent was collected from a crude oil refinery this included both the tank farm drainage water and the spent caustic and MEA (monoethanolamine) transferred to the laboratory in pre-cleaned 1.5 litre plastic containers and stored at room temperature until use. This was the stock effluent (100 %).

Physical and Chemical Analysis of Refinery Effluent: Physical and chemical components of the untreated refinery effluent were analysed

and parameters such as pH, temperature, free oil, sulphides, phenols, total nitrogen, total alkalinity, chemical oxygen demand (COD), total suspended solids (TSS), turbidity, and dissolved solids, were determined. The concentrations of heavy metals namely dissolved iron (Fe), lead (Pb), copper (Cu), manganese (Mn), chromium (Cr), mercury (Hg), arsenic (As) and tetrahydrocannabinol THC were also determined using APHA (2005).

Phytochemical Analysis of Shea Oil:

Phytochemical analysis was carried out to determine the presence of flavonoids, tannins, cardiac glycosides, saponin, steroids, terpenoids, alkaloids and reducing sugar in Shea oil according to the methods in Harborne (1989).

Anti-Microbial Analysis of the Effluent and Shea Oil

Preparation of culture media: All media were prepared according to manufacturer instruction. The media used in this study include nutrient agar and MacConkey agar.

Isolation of bacteria: Serial dilution of each sample was made to 10^{-1} , 10^{-2} and 10^{-3} dilutions. Total viable heterotrophic bacterial counts were determined using pour plate technique (Taylor *et al.*, 1983). Colony counts were taken and recorded in colony forming unit per millilitre.

Enumeration of microorganisms: Total viable counts of bacterial and fungal isolates were enumerated using the standard methods for estimating bacterial and fungal counts (Collins *et al.*, 1989).

Characterization and identification of bacterial isolates: The bacterial isolates were identified based on microbiological methods and cultural characteristics and the fungal isolates were identified based on their macroscopic and microscopic characteristics with reference to standard identification keys and atlas (Cowan, 1985).

Physical Observations, Body and Organ Weight Measurement:

Each rat in each of the treatment groups was observed twice daily (before and after exposure) for signs of clinical toxicity in the skin and fur, eyes and mucous membrane, behavioural pattern, respiratory rates, morbidity and mortality. The body weights of each animal in the control and treatment groups were measured at the beginning of the experiment and at the end of exposure period using a digital weighing balance (OHAUS® Scout™ Pro, Model: SPU202). The testes of the animals were surgically removed, weighed to measure the absolute organ weight, and fixed in Bouin's fluid for histological analysis using the method of Waheed and Ansari (2012).

Heavy Metal Concentration Analysis: Metal concentrations in the testicles was determined using atomic absorption Spectrophotometer (Buck Scientific 210 VGP) as described by Brzoska *et al.* (2002).

Histopathology

Testicular histopathology: Testes of each rat fixed in Bouin's fluid, were passed through ascending series of ethanol and then through xylene and embedded to paraffin wax. The tissues were sectioned at 15 μ , stained with Haematoxylin and Eosin, and mounted. All sections were examined under light microscope (Nikon Eclipse E400) in $\times 100$ and $\times 400$ magnifications. Photomicrographs of the lesions were taken for observation and documentation of histopathologic lesions. All alterations from the normal structure were registered.

Statistical Analysis: All data were analysed using the statistical software, Sigma Plot version 12.0. Significant difference between treatment(s) and control were analysed using analysis of variance (ANOVA). Variant means were separated using Duncan's New Multiple Range Test. Significant differences at $p \leq 0.05$ were accepted.

RESULTS

Phytochemical Analysis of Shea Oil: Shea oil was found to be rich in essential constituent that could serve as anti-oxidants and useful in nutrient enrichment. Flavonoids, terpenoids, steroids, anthraquinones, saponin, reducing sugar, alkaloids, tannin and cardiac glycosides.

Body Weight: There was a significant variation ($p < 0.05$) in the body weight of rats treated for 9 weeks (Figures 1 – 4) across the three groups. From week 10 to 12 recovery period, there was a significant decrease ($p < 0.05$) in body weight gain of rats treated with effluent only and in rats treated with effluent and Shea oil (Figure 1).

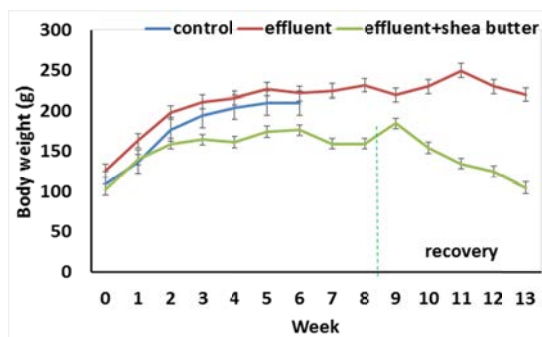


Figure 1: Changes in mean body weight of control, effluent treated and Shea oil ameliorated male rats from week 0 – 13 respectively

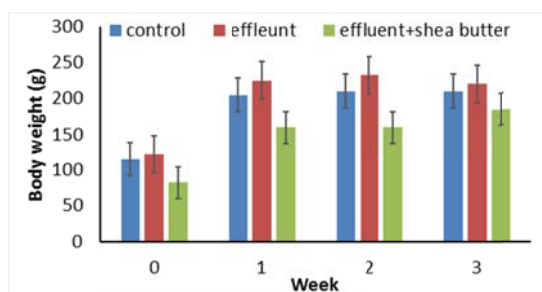


Figure 2: Changes in mean body weight of control, effluent treated and Shea oil ameliorated male rats at week 0 – 3 respectively

Testicular Weight and Morphology: There was no significant difference ($p > 0.05$) in the weight of testes across all the treatment groups at the end of the 9th week of test duration (Figure 5).

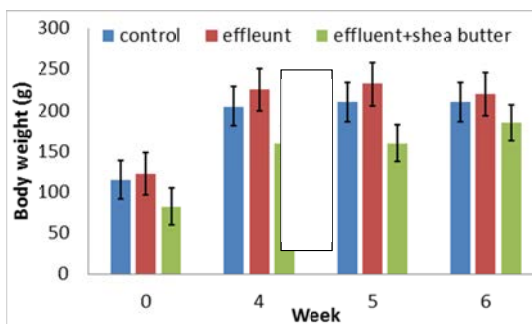


Figure 3: Changes in mean body weight of control, effluent treated and Shea oil ameliorated male rats at week 4 – 6 respectively

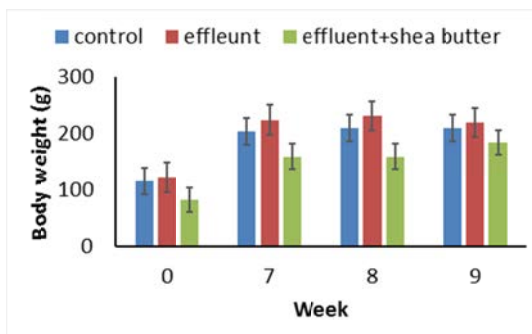


Figure 4: Changes in mean body weight of control, effluent treated and Shea oil ameliorated male rats at week 7 – 9 respectively

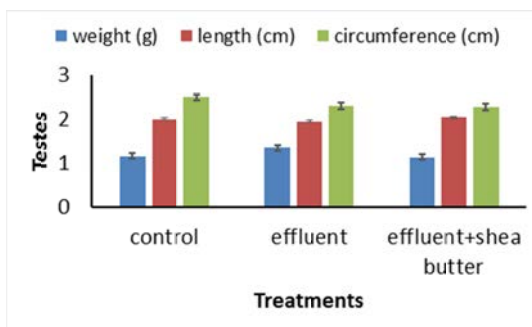


Figure 5: Changes in mean weight, length and circumference of testes of control, effluent treated and Shea oil ameliorated male rats

Rats that were left for week 10 – 12 recovery period showed significance differences between treated and ameliorated rats (Figure 5). Morphology of testes showed significant variation between treated groups. The circumference of testes of treated rats significantly increases at the 3rd week of treatment compared to weeks 3 – 6, 6 – 9 and during week 10 – 12 recovery period.

Testes length of control rats was constant at 6 weeks but reduced in length from the 9th week to the recovery period. Testes length of rats given Shea oil as abatement reduced at the 3rd week of treatment but increased from the 6th week to 9th week (Figure 5). There was also reduction in testes length during the recovery period. Rats treated with effluent alone without abatement showed no significant difference ($p>0.05$) in testes length during the recovery period.

Tissue Concentration of Heavy Metals: The testes were found to contain detectable concentrations of Chromium (Cr) and Lead (Pb). Tissue Cr and Pb concentrations were found to be significantly different ($p<0.05$) in rats treated with effluent only (Figure 6).

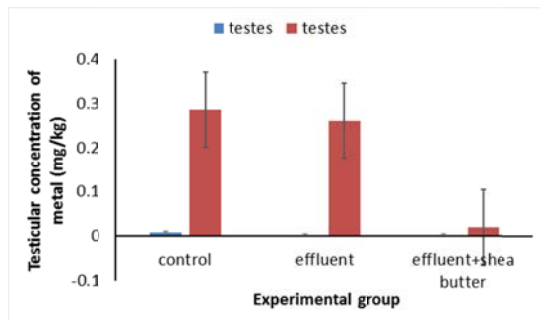


Figure 6: Tissue concentration of chromium (Cr) and lead (Pb) in the testes of control, effluent treated and Shea oil ameliorated male rats respectively

The maximum Cr and Pb concentration in gonads were in effluent treated rats. In the testes, the highest concentration of Cr (0.018 mg/kg) was observed at week 3 in the effluent treated rats. While the highest concentration of Pb (0.9 mg/kg) was observed at the recovery period.

Histopathology: The results of the histopathology of the testes of rats revealed that after 3 weeks of exposure the testis appeared normal in shape and morphology with no visible signs of degeneration composing of seminiferous tubules and interstitial space in the control group (Figure 7). Animals in Group 2 showed mild interstitial congestion and oedema

(Figure 8). While the animals in Group 3 showed normal spermatogenic series (Figure 9).

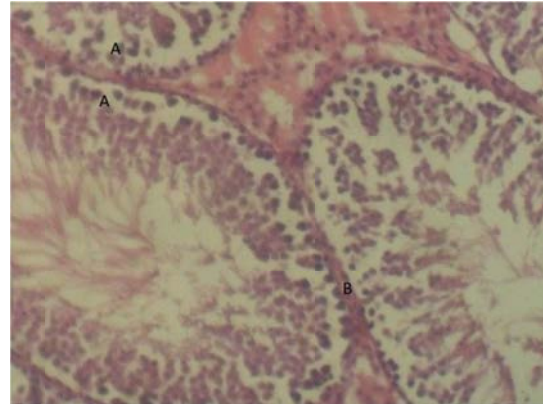


Figure 7: Testis of control rat showing (A) seminiferous tubules and (B) interstitial space (H & E x100)

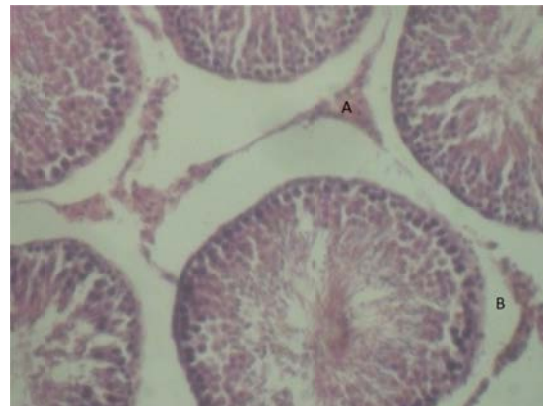


Figure 8: Testis of refinery effluent treated rat showing (A) mild interstitial congestion and (B) interstitial oedema (H & E x100)

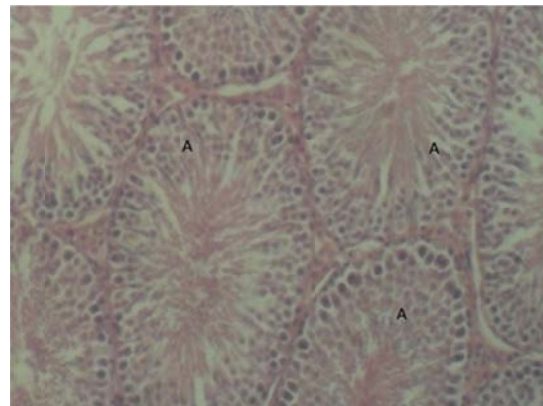


Figure 9: Testis of refinery effluent treated rat ameliorated with Shea oil showing (A) normal spermatogenic series (H & E x100)

At week 6, the testes of rats in the control group appeared normal with no visible signs of degeneration, composing of normal seminiferous tubules and normal interstitial space (Figure 10). The testes of rats given effluent only showing patchy spermatogenic arrest (Figure 11), while the testes of the rats given effluent and Shea oil showed normal tubules (Figure 12).

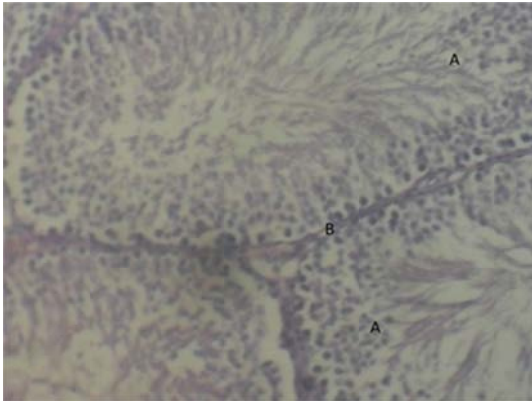


Figure 10: Testis of control rat showing (A) seminiferous tubules and (B) interstitial space (H & E x100)

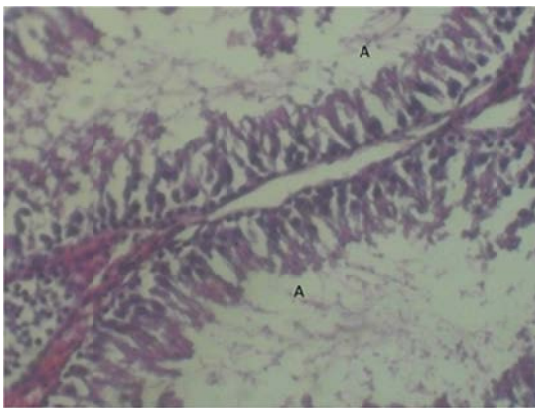


Figure 11: Testis of refinery effluent treated rat showing (A) patchy spermatogenic arrest (H & E x100)

At week 9, the testes of the rats given effluent only showed mild patchy spermatogenic arrest (Figure 13) while the testes of the rat given effluent ameliorated with Shea oil showed normal sequential maturation (Figure 14). At 21 days post exposure period the testes of the male rat in which effluent only was discontinued showed patchy spermatogenic arrest (Figure 15).

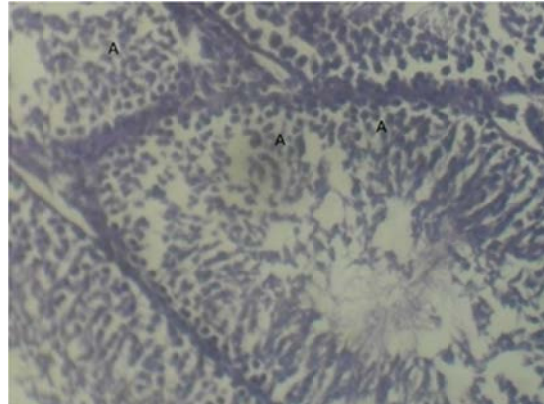


Figure 12: Testis of refinery effluent treated rat ameliorated with Shea oil showing (A) normal tubules (H & E x100)

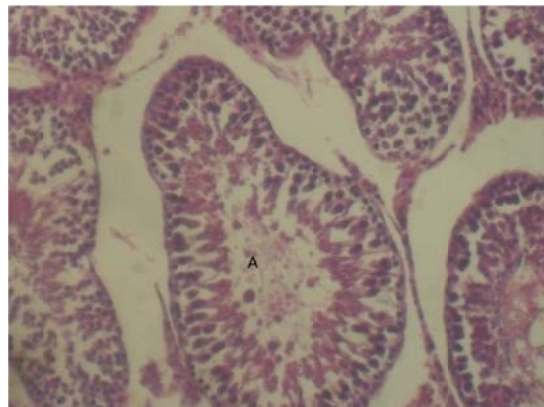


Figure 13: Testis of refinery effluent treated rat showing (A) patchy spermatogenic arrest (H & E x100)

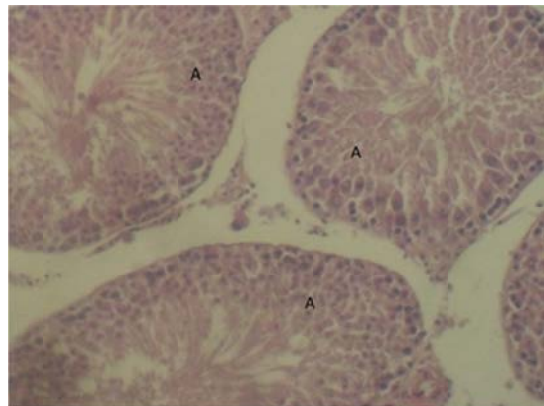


Figure 14: Testis of refinery effluent treated rat ameliorated with Shea oil showing (A) normal sequential maturation (H & E X100)

While the testes of the rats in which effluent and Shea oil was discontinued showed normal sequential maturation (Figure 16).

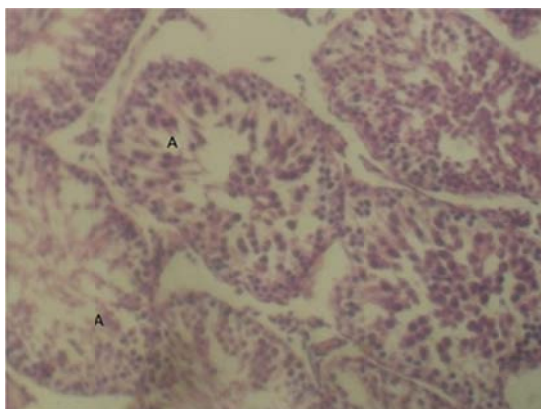


Figure 15: Testis of refinery effluent treated rat ameliorated with Shearbutter oil after 21 days recovery period showing (A) patchy spermatogenic arrest (H & E X100)

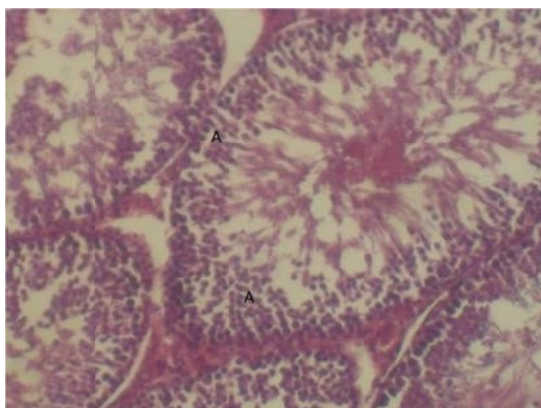


Figure 16: Testis of refinery effluent treated rat after 21 days recovery period showing (A) normal sequential maturation (H & E x100)

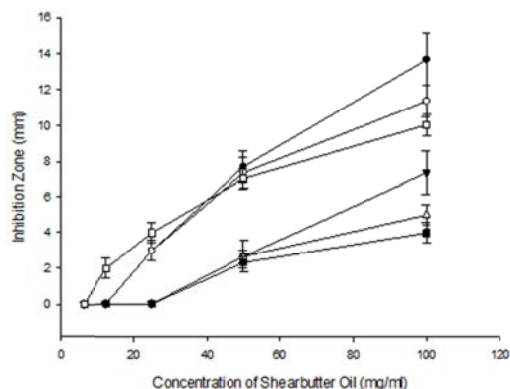


Figure 17: Antimicrobial inhibition of Shearbutter oil on microbial culture of untreated refinery effluents at various concentrations

Antimicrobial Activities of Shea Oil: There was an increase anti-microbial activity with Shea oil (Figure 17). All the isolates were inhibited at concentration of 100 mg/ml of Shea oil with *Micrococcus* spp. and *Klebsiella* spp. having the highest and lowest zones of inhibition, respectively.

DISCUSSION

Shea oil extracted from Shea tree (*Vitellaria paradoxa*) is one of the essential oils, known to possess antimicrobial, antifungal, antiviral, insecticidal and antioxidant properties (Kordali *et al.*, 2005). Shea oil is composed of five principal fatty acids: palmitic, stearic, oleic, linoleic and arachidonic acids. About 85 to 90 % of the fatty acid composition is stearic and oleic acids. Phenolic compounds are known to have antioxidant properties. A study characterized and quantified the most important phenolic compounds in Shea butter (Marouani *et al.*, 2012). Shea oil used in this study demonstrated antimicrobial activities against all bacteria isolates from refinery effluent, antimicrobial activity by Shea oil against *Staphylococcus aureus* has been reported earlier (Wahedi and David, 2014).

Results of this study showed that fifteen days exposure of adult male rats to refinery effluent induced reproductive toxicity. The relative weight of testes was reduced in effluent treated rats. Significant increase in body weight gain across the duration of exposure was seen in rats treated with refinery effluent and those given Shea oil for abatements. In the period when rats were withdrawn from treatment i.e. recovery period, there was significance reduction in body weight across the treated groups. Although, rats during exposure showed no obvious signs of loss of appetite as rats feed and consumed water that was added *ad-libitum*, the increased body weight might be an indication of toxicity via accumulation of fluids in organs or other body tissues and heavy metal accumulation in tissues and organs of treated rats. Studies have reported weight gain in some organs of rats upon exposure to contaminants such as leachates and the authors attributed it to the heavy metal constituents in the

contaminants this may be responsible for the change noticed in organ weight in this study as these organs are involved with sequestration of metals (Barbier *et al.*, 2005). A similar study reported reduction in body and organ weight of rats exposed to textile effluent at concentrations reported toxic (Suryavathi *et al.*, 2005). Similar to previously published reports, testicular Cr and Pb levels increased in a dose-dependent manner in treated rats, confirming the absorption of the metals after administration (Marouani *et al.*, 2012).

Moreover, histological changes observed in the gonads of treated animals, included enlarged intracellular space; which may have resulted from the destruction of Sertoli cells by the intoxication with heavy metals; as well as a dramatic loss of gametes in the lumen of the seminiferous tubule. This might also be a consequence of the disruption of the blood-testis barrier with the heavy metal accumulation in the tissues. Changes in the permeability of the blood-testis barrier and alterations in testicular and epididymal histo-architecture was also demonstrated in rats exposed to Cr (Marouani *et al.*, 2012). Many reports exist on the effects of heavy metal accumulation in humans, but only a few studies have verified these effects in experimental animals (Flora *et al.*, 2012). Oral exposure of both male and female rabbits to sodium dichromate (0.1% solution, 0.2 – 5 mg/kg body weight for up to 545 days) resulted in significant morphological changes in the gonads, including atrophy of the epithelium and dystrophic alterations of the Sertoli and Leydig cells in the testes (Nigam *et al.*, 2014). Large doses of hexavalent Cr are highly toxic and may cause death especially when injected subcutaneously (Nigam *et al.*, 2014). A study in which sodium dichromate (VI) was administered by gastric intubations to groups of 10 mature male rats at levels of 20, 40 and 60 mg Cr(VI)/kg/day for 90 days also reported reproductive effects in rats and mice orally exposed to Cr(VI) (Chandra *et al.*, 2007).

In this study, the highest concentration of Cr in the testes (slightly > 0.018 mg/kg) was at 3 weeks in the effluent treated rats while for

Pb, the highest concentration (0.9 mg/kg) was at the recovery period.

The testis of the rats in the refinery effluent treated and Shea oil ameliorated group showed mild inflammatory changes (interstitial congestion and oedema) similar to that seen in Pb intoxication; the ameliorated group did not show these pathologies at 3 weeks. There was some degree of spermatogenic arrest after six weeks in the effluent intoxicated group, which was indicated by alterations of the normal testicular architecture. There have been previous report of damage to testicular architecture in rabbits given a dose of 200 mg of Pb this study reported deformities in the architecture with serious damage within the seminiferous tubules (Ibrahim *et al.*, 2012). However, these pathologies were abated in the group treated with Shea oil in this study. Some degree of spermatogenic arrest also occurred at the 9th week, which abated in the Shea oil treated group also. At the end of the three weeks recovery period, the spermatogenic arrest reduced with visible signs of return of the testes to normal testicular architecture.

Shea oil had inhibitory action against the six bacteria isolates (*Staphylococcus aureus*, *Micrococcus*, *Bacillus subtilis*, *Klebsiella* spp., *Escherichia coli* and *Pseudomonas aeruginosa*) from the refinery effluent. This agrees with previous reports (Coulibaly *et al.*, 2009; Wahedi and David, 2014). This study has provided adequate information on the toxic effects of untreated refinery effluent on the testes of rats. The histological examinations revealed varying pathological changes in the testis. These include spermatogenic arrest. However, the use of Shea oil for amelioration of the toxicity induced by the untreated refinery effluent proved effective.

Conclusion: The study concluded that; the level of heavy metal concentration on the testicles of the rats exposed to refinery effluent was high, there was histopathological alteration in the testes of the effluent treated rats, Shea oil proved effective against the toxic effects of the untreated refinery effluent on the testes of treated rats. Finally untreated refinery waste should be treated and disposed of properly to avoid contaminating the environment and

causing deleterious effects on organs and tissues of unsuspecting animals and possibly humans.

ACKNOWLEDGMENTS

The authors wish to acknowledge the assistance of Dr Ekeolu, O. K. of the Department of Veterinary Anatomy, University of Benin, Benin City, Nigeria during the histopathology study.

REFERENCES

- ACHARYA, U. R., MISHRA, M., TRIPATHY, R. R. and MISHRA, I. (2006). Testicular dysfunction and antioxidative defense system of Swiss mice after chromic acid exposure. *Reproductive Toxicology*, 22(1): 87 – 91.
- ADIKWU, E., OPUTIRI, D., ORU-BO, P. G. and ENIMEYA, D. A. (2013). Lead organ and tissue toxicity: roles of mitigating agents (Part 1). *British Journal of Pharmacology and Toxicology*, 4(6): 232 – 240.
- APHA (2005). *Standard Methods for the Examination of Water and Wastewater*. 21st Edition, American Public Health Association/American Water Works Association/Water Environment Federation, Washington DC, USA.
- BARBIER, O., JACQUILLET, G., TAUC, M., COUGNON, M. and POUJEOL, P. (2005). Effect of heavy metals on, and handling by the kidney. *Nephron Physiology*, 99(4): 105 – 110.
- BRZÓSKA, M. M., MONIUSZKO-JAKONIUK, J., JURCZUK, M. and GAŁAŻYN-SIDORCZUK, M. (2002). Cadmium turnover and changes of zinc and copper body status of rats continuously exposed to cadmium and ethanol. *Alcohol and Alcoholism*, 37(3): 213 – 221.
- CHANDRA, A. K., CHATTERJEE, A., GHOSH, R., SARKAR, M. and CHAUBE, S. K. (2007). Chromium induced testicular impairment in relation to adrenocortical activities in adult albino rats. *Reproductive Toxicology*, 24(3-4): 388 – 396.
- CHATTERJEE, S., SINGH, L., CHATTOPADHYAY, B., DATTA, S. and MUKHOPADHYAY, S. K. (2012). A study on the waste metal remediation using floriculture at East Calcutta Wetlands, a Ramsar site in India. *Environmental Monitoring and Assessment*, 184(8): 5139 – 5150.
- COHEN, M. D., KARGACIN, B., KLEIN, C. B., COSTA, M. (1993). Mechanisms of chromium carcinogenicity and toxicity. *Critical Reviews in Toxicology*, 3(23): 255 – 281.
- COLLINS, C. H., LYNE, P. M. and GRANGE, G. M. (1989). *Collins and Lyne Microbiological Methods*. Sixth Edition, Butterworth, London.
- COULIBALY, Y., OUÉDRAOGO, S. and NICULESCU, N. (2009). Experimental study of Shea butter extraction efficiency using a centrifugal process. *ARPN Journal of Engineering and Applied Sciences*, 4: 14 – 19.
- COWAN, S. T. (1985). *Cowan and Steel's Manual for Identification of Medical Bacteria*. Cambridge University Press, Cambridge, United Kingdom.
- DANADEVII, K., ROZATI, R., REDDY, P. P. and GROVER, P. (2003). Semen quality of Indian welders occupationally exposed to nickel and chromium. *Reproductive Toxicology*, 17(4): 451 – 456.
- DIYA'UDDEEN, B. H., DAUD, W. M. A. W. and AZIZ, A. A. (2011). Treatment technologies for petroleum refinery effluents: A review. *Process Safety and Environmental Protection*, 89(2): 95 – 105.
- FLORA, G., GUPTA, D. and TIWARI, A. (2012). Toxicity of lead: a review with recent updates. *Interdisciplinary Toxicology*, 5(2): 47 – 58.
- GIRIDHARAN, N. V., KUMAR, V. and MUTHUSWAMY, V. (2000). *Use of Animals in Scientific Research*. Indian Council of Medical Research, Ministry of Health and Family Welfare, New Delhi, India. <http://icmr.nic.in/bioethics/Animalsbiomedicalresearch.pdf> Retrieved May 28, 2013.

- GUERTIN, J. (2005). Toxicity and health effects of chromium (all oxidation states). Pages 216 – 234. *In*: GUERTIN, J., JACOBS, J. A. and AVAKIAN, C. P. (Editors). *Chromium (VI) Handbook*. CRC Press, Boca Raton, Florida, USA.
- HARBORNE, J. B. (1973). *Phytochemical Methods*. Chapman and Hall Limited, London.
- IBRAHIM, N. M., EWEIS, E. A., EL-BELTAGI, H. S. and ABDEL-MOBDY, Y. E. (2012). Effect of lead acetate toxicity on experimental male albino rat. *Asian Pacific Journal of Tropical Biomedicine*, 2(1): 41 – 46.
- IHEDIOHA, J. I., OKAFOR, C. and IHEDIOHA, T. E. (2004). The haematological profile of the Sprague-Dawley outbred albino rat in Nsukka, Nigeria. *Animal Research International*, 1(2): 125 – 132.
- ISSELHARD, W. H. and KUSHE, J. (1998). Animal experimentation. Pages 419 – 434. *In*: TROIDL, H., SPITZER, W. O., MCPPEEK, B., MULDER, D. S., MCKNEALLY, M. F., WECHSLER, A. and BALCH, C. M. (Eds.). *Principles and Practice of Research: Strategies for Surgical Investigators*. 3rd Edition, Springer-Verlag, New York.
- KORDALI, S., CAKIR, A., MAVI, A., KILIC, H. and YILDIRIM, A. (2005). Screening of chemical composition and antifungal and antioxidant activities of the essential oils from three Turkish Artemisia species. *Journal of Agricultural and Food Chemistry*, 53(5): 1408 – 1416.
- MARANZ, S., WIESMAN, Z., BISGAARD, J. and BIANCHI, G. (2004). Germplasm resources of *Vitellaria paradoxa* based on variations in fat composition across the species distribution range. *Agroforestry Systems*, 60(1): 71 – 76.
- MAROUANI, N., TEBOURBI, O., HALLEGUE, D., MOKNI, M., YACOUBI, M. T., SAKLY, M., BENKHALIFA, M., RHOUMA, K. B. (2017). Mechanisms of chromium hexavalent-induced apoptosis in testis rats. *Toxicology and Industrial Health*, 33(2): 97 – 106.
- MAROUANI, N., TEBOURBI, O., MAHJOUR, S., YACOUBI, M. T., SAKLY, M., BENKHALIFA, M. and RHOUMA, K. B. (2012). Effects of hexavalent chromium on reproductive functions of male adult rats. *Reproductive Biology*, 12(2): 119 – 133.
- MAROUANI, S., CHAËBA, R., KADRI, H., SAIDI, B., BOUAIN, A., MALTAGLIATI, F., LAST, P., SÉRET, B. and BRADAI, M. N. (2011). Taxonomic research on *Squalus megalops* (Macleay, 1881) and *Squalus blainvillei* (Risso, 1827) (Chondrichthyes: Squalidae) in Tunisian waters (central Mediterranean Sea). *Scientia Marina*, 76(1): 97 – 109.
- NIGAM, A., PRIYA, S., BAJPAI, P. and KUMAR, S. (2014). Cytogenomics of hexavalent chromium (Cr6+) exposed cells: a comprehensive review. *Indian Journal of Medical Research*, 139(3): 349 – 370.
- PERMENTER, M. G., LEWIS, J. A. and JACKSON, D. A. (2011). Exposure to nickel, chromium, or cadmium causes distinct changes in the gene expression patterns of a rat liver derived cell line. *PLoS One*, 6(11): e27730. <https://doi.org/10.1371/journal.pone.0027730>
- SUBRAMANIAN, S., RAJENDIRAN, G., SEKHAR, P., GOWRI, C., GOVINDARAJULU, P. and ARULDHAS, M. M. (2006). Reproductive toxicity of chromium in adult bonnet monkeys (*Macaca radiata* Geoffrey). Reversible oxidative stress in the semen. *Toxicology and Applied Pharmacology*, 215(3): 237 – 249.
- SURYAVATHI, V., SHARMA, S., SHARMA, S., SAXENA, P., PANDEY, S., GROVER, R., KUMAR, S. and SHARMA, K. P. (2005). Acute toxicity of textile dye wastewaters (untreated and treated) of Sangner on male reproductive systems of albino rats and mice. *Reproductive Toxicology*, 19(4): 547 – 556.
- TAYLOR, R., ALLEN, M. and GELDREICH, E. (1983). Standard plate count: a comparison of pour plate and spread plate methods. *Journal of American Water Works Association*, 75(1): 35 – 37.

- UGWU, O. P. C., NWODO, O. F. C., JOSHUA, P. E., BAWA, A., OSSAI, E. C. and ODO, C. E. (2013). Phytochemical and acute toxicity studies of *Moringa oleifera* ethanol leaf extract. *International Journal of Life Science Biotechnology and Pharma Research*, 2(2): 65 – 71.
- WAHEDI, J. A. and DAVID, L. D. (2014). Anti-microbial activity of essential oil extracted from Shea tree seed (*Butyrospermum parkii*) in Mubi, north-eastern Nigeria. *International Journal of Pure and Applied Sciences and Technology*, 23(1): 1 – 7.
- WAHEED, U. and ANSARI, A. (2012). *Histotechniques: Laboratory Techniques in Histopathology: A Handbook for Medical Technologists*. Lap Lambert Academic Publishing, Saarbrucken, Germany.
- WAKE, H. (2005). Oil refineries: a review of their ecological impacts on the aquatic environment. *Estuarine, Coastal and Shelf Science*, 62(1-2): 131 – 140.
- YOUSEF, M. I., EL-DEMERDASH, F. M., KAMIL, K. I. and ELASWAD, F. A. (2006). Ameliorating effect of folic acid on chromium (VI)-induced changes in reproductive performance and seminal plasma biochemistry in male rabbits. *Reproductive Toxicology*, 21(3): 322 – 328.