

Review Article

Value of immunohistochemical IMP3 expression with endoscopic ultrasound-guided-fine needle aspiration (EUS-FNA) in diagnosing pancreatic lesions

Hayam E. Rashed, MD ¹, Awatef, N. Nasr MD ¹, Nora T. Wasfi¹, Ramy ElHendawy MD ², Nelly M. Said, MD ¹

¹ Pathology Department, Faculty of Medicine, Zagazig University, Egypt.

² Tropical Medicine Department, Faculty of Medicine, Zagazig University, Egypt.

The corresponding authors' address: Pathology Department, Faculty of Medicine, Zagazig University, Zagazig, Egypt.

E-mail address of the corresponding author: hayam_rashed@yahoo.com

Mobile No: +201022932069.

DOI: [10.21608/ajgh.2022.112624.1000](https://doi.org/10.21608/ajgh.2022.112624.1000)

Type of manuscript: a narrative review

Conflict of interest: N / A

Funding source: This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Date of submission: 22 December 2021.

Revised: 11, January 2022.

Accepted: 24, January 2022.

First online: 24, January 2022.

Abstract:

Pancreatic cancer (PC), a lethal condition with a poor prognosis, ranks fourth among the most common causes of cancer-related mortality as early diagnosis of PC is so tricky. Consequently, most cases at the time of initial diagnosis already harbor metastasis. PC cases' early detection and survival depend mainly on improving diagnostic approaches. This review sheds light on the role of endoscopic ultrasound-guided fine-needle aspiration (EUS-FNA) as a minimally invasive method in early PC diagnosis and differentiation

Review Article

between different pancreatic lesions. The discovery of new diagnostic and prognostic markers for PC will raise the accuracy of proper diagnosis, and in turn, patients will gain better survival and prognosis. Insulin-like growth factor II mRNA binding protein3 (IMP3) is overexpressed in several malignant tumors, including pancreatic cancer, which may raise its role in diagnosis and prognosis as well as its therapeutic benefit for PC.

Keywords: Pancreatic cancer; IMP3; EUS-FNA; Immunohistochemistry

Introduction

Cytology in diagnosis of pancreatic cancer:

Cytological sampling from the pancreas is achieved mainly by either percutaneous FNA under ultrasound guidance, EUS-FNA transduodenal for head or neck lesions, or transgastric for body and tail lesions. Percutaneous and EUS FNA have the same complication rate from 0 -5%, and the percutaneous technique is an accurate and safe method for diagnosing pancreatic lesions [1]. Both procedures have the same accuracy, and percutaneous FNA can be used as an alternative for EUS FNA as it is cheaper [2]. However, EUS-FNA replaced percutaneous FNA. It is superior in detecting smaller lesions even if they are less than 2 cm, correctly detects vascular involvement, and can stage pancreatic cancer cases, affecting the prognosis and therapeutic decisions [3]. Also, the peritoneal metastasis rate is higher in patients undergoing percutaneous FNA for PC diagnosis due to tumor seeding into the peritoneal cavity during this technique [4].

Indications and contraindications of EUS-FNA:

EUS-FNA has been used more frequently to improve the diagnosis of pancreatic lesions using cytopathological evaluation and constitutes a handy tool for the loco-regional staging of PC [5]. The indications of EUS-FNA include the presence of solid or cystic mass, enlarged lymph node, intrapleural/abdominal fluid, differentiation between benign and malignant lesions, staging of cancers, and providing histopathological evidence for starting therapy.

All solid pancreatic lesions should be aspirated before surgery to rule out autoimmune/focal pancreatitis lymphoma and assess for different types of cancer other than adenocarcinoma. Contraindications that limit its use include the inability to visualize the lesion site and the presence of vessels along the target's pathway. In addition, the FNA result cannot affect the management and pseudocysts aspiration due to high complications rate unless in case of therapeutic drainage of the cyst after its aspiration [6], [7].

Review Article**Technique:**

The diagnostic ability of EUS-FNA depends on several factors such as size, site, and properties of target tissues and procedural and technical elements (type of needle, material processing, and biopsy technique) [8].

- The size of the needle used in EUS-FNA is so essential. Smaller gauge needles as 25 gauge are safer and more effective than large gauge needles like 19-gauge. They reduce the bleeding risk, especially in highly vascular tumors such as neuroendocrine neoplasms, improving diagnosis [9]. Also, the failure of a 19-gauge needle in sampling pancreatic lesions presenting in the head or uncinate process [10], 22 gauge is influential in both aspiration and core biopsy [8]. Also, a 22-gauge needle is the most commonly used, but this needle was unsuccessful in about 33% of cases with uncinate lesions. However, 25-gauge needle was more flexible and successful in EUS-FNA of head and uncinate process lesions [11]. Gimenco-Garcia stated no significant difference between 22- and 25-gauge needles. [12]. 22- or 25-gauge needles can be used in any FNA approach; however, a 25-gauge needle is the best choice for transdoudenal FNA [13]. There is no significant difference between using 19- or 25-gauge needles, and the needle choice depends only on endoscopist desire. The number of aspirates in pancreatic lesions is five or six passes. For a highly accurate diagnosis, seven passes are recommended, which is so high compared to other organs, requiring only two or three passes[14]. Nevertheless, this will require less frequent needle passes, such as specific cytologic diagnoses, prolonged procedure time, higher risk, and additional needles [15].

Stains:

-There are multiple types of stains used in FNA staining. Romanowsky stain, although rapid, defines cell size and stromal components so well, but its nuclear morphology is so limited. Rapid Papanicolaou stain shows a high ability to focus through overlapping cell clusters and thicker smears. Toluidine blue stain is an ultra-fast stain, but it requires constant stain for destaining and restaining. Finally, hematoxylin and eosin stain is more time-consuming, but most pathologists prefer it [16].

- Rapid on-site evaluation (ROSE) means the evaluation of cytological smears at the endoscopic suite point of care (Point of care in the endoscopy ward). This process is done by a pathologist using the light microscope to provide rapid feedback to the endosonographer [17]. In addition, ROSE raises the diagnostic ability of EUS-FNA to reduce the number of needles passed, reducing the time of procedure and allowing proper earlier therapeutic decisions [9].

Review Article**Factors affecting diagnosis:****Difficulties:**

-Although EUS-FNA has many advantages, it also has some complications such as bleeding, infection, perforation, and malignant seeding. [16] complications happen only in 2.2% of cases in the form of pancreatitis, retroperitoneal bleeding, and bradycardia (may be caused by mechanical factors).[18]. Besides, many challenges may face the EUS-FNA, such as the fibrotic or inflammatory nature of tumors and cystic tumor aspirations being hypocellular, which may yield false results [16], so the number of aspirates from tumor should be at least five to seven for optimal results. Also, the small lesions and those far away from GIT lumen are very challenging targets [19].

Among the disadvantages of EUS-FNA is the defective diagnostic material in cases of cystic lesions compared to solid and solid cystic lesions. Radiological findings and cell block preparation with the help of immunohistochemical markers can give better diagnostic results and a more accurate diagnosis of cystic lesions [20].

Interpretation of EUS-FNA results:

-The interpretation of pancreatic cytology requires proper knowledge of the following normal cells such as acinar, ductal, and islet cells to avoid pitfalls. [16] Some contaminants during the FNA procedure may appear, such as benign hepatocytes, duodenal epithelium, gastric mucosa, and mesothelial cells. The pancreatic FNA includes the background pattern, which may be mucinous, bloody, clean, inflammatory, or necrotic, the type of parenchymal epithelium that may be ductal, acinar, or islet cells, the stromal elements, which may be spindle cells, fibrovascular cores, or fibrous stromal elements [16].

Solid cellular neoplasms:

-Highly cellular smears characterize specific solid pancreatic lesions such as pancreatic endocrine neoplasm, acinar cell carcinoma, solid-pseudopapillary neoplasm, pancreatoblastoma [21]. Comparing these lesions, solid pseudopapillary neoplasm (SPN) shows a cribriform pattern of cells as they have cytoplasmic extensions with or without cytoplasmic vacuoles and hyaline globules [16], [21]. Meanwhile, neuroendocrine neoplasms are characterized by plasmacytoid nuclei, salt, pepper chromatin, and cytoplasmic neurosecretory granules usually surrounded by a bloody background. Acinar cell carcinoma (ACC) is characterized by acinar or grape-like clusters, granular cytoplasm, and minimal anisonucleosis.

Review Article

SPN, ACC, and neuroendocrine neoplasms share close features of cytological smears, so depending on immunohistochemical markers could help solve this, as neuroendocrine neoplasms exhibit synaptophysin chromogranin and CD56. At the same time, ACC shows a positive expression of trypsin, chymotrypsin, and phospholipase A2. Vimentin and beta-catenin are positive in SPN [22],[23].

Pancreatoblastoma (PB) is characterized by round blast-like cells twice the size of RBCs, squamoid morules, epithelioid cells with eosinophilic cytoplasm, and syncytial arrangements can be detected on both smears and cell blocks [24].

Mucinous Cystic Lesions:

Pancreatic lesions characterized by their mucinous background in FNA are intraductal papillary mucinous neoplasm (IPMN) and mucinous cystic neoplasms (MCN). Although they share some features such as classification into low grade, usually with hypocellular smear and abundant mucin, and a high degree of variable cellularity of atypical ductal cells and less prominent mucin, they differ in their incidence, gross and microscopic features, presence of papillary structures in IPMN and cystic fluid analysis of amylase (clinicopathological criteria) [21].

The criteria that favor the diagnosis of IPMN are the occurrence in the head of the pancreas. Male gender [27]. besides that IPMNs, are cystic pancreatic lesions that show two characteristic features: papillary projections bulging into the pancreatic duct and mucin production and microscopically IPMN offers four distinctive morphologic types of papillae (1) intestinal pattern, which has the same appearance of colonic villous adenomas (2) pancreatobiliary pattern, their papillae are lined by cuboidal cells with prominent nucleoli (3) gastric pattern, rarely some papillae have a gastric foveolar appearance. (4) Oncocytic pattern is characterized by abundant granular eosinophilic neoplastic cells and also contains intracellular mucin [28],[29].

Determining the degree of dysplasia is a crucial target in FNA interpretation. Several cytological features can identify the grade of dysplasia of IPMN as the hypercellularity of the specimens in the form of crowded epithelial clusters, the presence of necrosis, the presence of papillary fragments, parachromatin clearing, open chromatin, irregular nuclear membranes and nucleoli, and background of acute inflammation, all of these features support IPMN carcinoma diagnosis or at least IPMN-CIS [21].

Review Article

Notwithstanding, MCN is more common in women in 95% of cases and distal pancreas in 97% of patients [30]. It is predominant in premenopausal females in the body and tail. [24].

Grossly known as multilocular large cysts surrounded by thick fibrotic walls, while at the microscopic view, the cysts are lined by tall, columnar mucin-producing epithelium. The stroma of MCN is characterized by being similar to the ovarian stroma, which is an essential key in defining these neoplasms [28]. Usually, it is associated with an elevated cystic fluid of CEA, while the occurrence of elevation of CEA, CA19-9 levels in cystic fluid of IPMN means the presence of invasive carcinoma [24]. The examination of cystic fluid is beneficial, as elevated amylase levels connect with the duct and are characteristically high in IPMNs while low in other cystic lesions [31]. Specific mutated genes are identified in the cystic fluid of cystic pancreatic lesions—K-ras gene mutation in cystic fluid diagnoses mucinous cysts [32]. GNAS gene mutation is detected in more than half of cases of IPMN [33].

The risk features of mucinous pancreatic cystic neoplasms (MCN and IPMN) are moderate and high-risk. Moderate risk features include more than 3 cm cystic size, sudden change in diameter of the main pancreatic duct, regional lymphadenopathy, size of main pancreatic duct ranges from 5-9 mm, mural nodules, and cystic wall thickening. High-risk features include pancreatic head lesions associated with the common bile duct obstruction, the primary pancreatic duct size of more than 10 mm, and enhanced solid components within the cyst. The detection of just one of the hazardous features and two of the moderate features of a lesion is highly dysplastic or invasive cancer [34].

-Neoplastic mucin should be differentiated from contaminating mucin by its quality and quantity, and the presence of degenerated inflammatory cells and histiocytes confirm the neoplastic changes. The presence of neoplastic mucin, thick and colloid inconsistency, is enough for diagnosing the neoplastic mucinous cyst [21].

Intraductal tubulopapillary neoplasm (ITPN) is differentiated from IPMN by lacking mucin and papillary structures, but the cytological smears are highly cellular and arranged in tubular patterns with no mitosis. The cytological features of ITPN reported that it resembles IPMN by being ductal in origin and ACC by some morphology features [35].

Serous cystic lesion:

-FNA hardly detects serous cystic neoplasms as serous epithelium is seen only in 20% of cases, and cytology is usually non-diagnostic due to their scarcity of cellularity even after re-aspiration [21]. In serous cystadenoma FNA, the sparsely cellular smear is present with

Review Article

cuboidal cells, arranged in small sheets, harboring rounded central or eccentric nuclei and scanty cytoplasm in a clean background without features of malignancy such as mitosis, nuclear enlargement, or necrosis [16], [21].

Solid mass lesions:

-The solid lesions of ductal origin such as chronic pancreatitis and pancreatic ductal adenocarcinoma (PDA) remain the most crucial obstacle in EUS-FNA, as the false-negative cases are very high (23%) [36], (15%) [37]. Until now, FNA could not correctly differentiate between pseudo-tumorous pancreatitis and adenocarcinoma.

- PDA is characterized by ductal cells with overlapping nuclei, chromatin clearing, mitosis, and necrosis. Nevertheless, the components of a cytological smear of pancreatitis vary according to the stage. The smear mainly contains ductal, acinar cells, and inflammatory cells early. In contrast, later stage, mostly ductal cells are due to atrophy of acinar cells with no or rare mitosis [16], [38]. Several important cytological features can diagnose Well-differentiated PDA: 1) chromatin clearing, 2) anisonucleosis, 3) nuclear overlapping 4) nuclear membrane irregularity, 5) nuclear enlargement, 6) macronuclei 7) hyperchromasia 8) necrosis, 9) mitosis, 10) gap versus confluent cell space [38]. Supporting the diagnosis of PDA, CA19-9 is not only the most important serum marker, but it is also a valuable prognostic marker that can detect the survival and response of cases to chemotherapy with a cut-off value above 200u\mL as reported by Ballehaninna UK (2013) [39].

Regarding autoimmune pancreatitis, FNA is characterized by mainly plasma cells and lymphocytes, fibrous tissue fragments, a population of ductal or acinar cells, and occasionally fibroblasts [40]. In addition, the serum level of IgG4 is highly elevated in autoimmune pancreatitis, which helps differentiate it from other causes of pancreatitis [41].

Nonneoplastic cystic lesions:

-Pseudocysts represent the most common benign pancreatic cystic lesions, which happen mainly because of acute pancreatitis and auto-digestion of pancreatic tissue by the release of pancreatic enzymes. According to FNA, Pseudocysts are classified into complicated forms with mucinous background and numerous inflammatory cells or uncomplicated forms with transparent non-mucinous background, few inflammatory cells, bile, and histiocytes. No mitosis or nuclear features of malignancy occur [21]. Very high amylase

Review Article

levels are characteristically demonstrated [31]. Also, Martínez-Ordaz (2016) reported elevated serum levels in 79% of cases [42].

- Lymphoepithelial cyst (LEC) is a rare lesion, but its incidence begins to rise and mimics pseudocyst clinically and radiologically. The cytological findings include anucleated squamous cells, keratin, amorphous debris, a few nucleated squamous cells, and lymphocytes [43].

IMP3 (Insulin-like growth factor II mRNA binding protein 3):

The structure of IMP3 is composed of six RNA-binding domains: four K-homology (KH) domains in the C-terminal region and two RNA recognition motifs (RRMs) in the N-terminal area. The six parts are arranged in three pairs (RRM1 with 2, KH1 with 2, and KH3 with 4) and separated by flexible linkers [44], [45]. It binds to RNA through the C-terminal of KH domains. [46]

IMP3, its other name is IGF2BP3 or KOC, which means K homology domain-containing protein overexpressed in cancer, is a gene detected on the chromosome 7p11.5 by fluorescence in situ hybridization (FISH) [46].

It plays a vital role during embryogenesis in cell migration [47]. In addition, IMP3-RNA binding protein participates in post-transcriptional gene regulation [48].

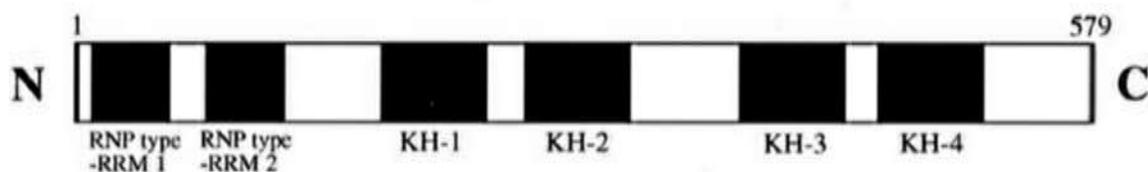


Fig1: Schematic diagram IMP3 structure, N, N-terminal, C, C-terminal, RRM, RNA recognition motifs, KH, K-homology [49].

IMP3 normal function:

IMP3 expression in normal tissues:

High IMP3 is found in pancreatic tissue during embryogenesis and ductal PC and not in adult exocrine pancreatic tissue [50]. Typically, IMP3 is absent in various tissues, e.g.,

Review Article

pancreas, esophagus, stomach, heart, lung, kidney, and other tissues. However, it could be seen in a few tissues and particular cell types, for example, syncytiotrophoblast, cytotrophoblast, decidua, lymph follicles in lymph nodes and tonsils, absorptive cells of the ileum, crypt cells of the rectal mucosa, mucus cells of submandibular and sublingual glands, spermatogonia, ciliated cells of the bronchial mucosa and the fallopian tube, secretory cells of the endocervix, and cells of the adenohypophysis of the anterior lobe of the pituitary gland [47].

IMP3's role in modulating tumor cell fate:

- Heterogeneous nuclear ribonucleoprotein M (HNRNPM) manage the nuclear stabilization and transport of IMP3, as some studies confirmed its presence as nuclear not only cytoplasmic distribution relevant to its role in cyclins regulation and cancer cells proliferation, so IMP3 cytoplasmic to nuclear ratio could be used to determine the rate of cancer cells growth and might also be used as a therapeutic target [51].

IMP3 binds to CD44 mRNA, which acts as an adhesion molecule with extracellular matrix proteins including collagen, hyaluronan, laminin, and fibronectin, promoting tumor invasiveness [52].

Through a pro-metastatic behavior of pancreatic cancer cells, IMP3 shows enhanced aggressiveness of PDA by promoting the dissemination of cancer cells [53]. In addition, IMP3 has been detected to increase the levels of nerve growth factor β (NGF β) and facilitate the translation of IGF-2 mRNA, which enhances the angiogenesis and lymphangiogenesis of the tumor [50]. Also, the regulation of KIF11 mRNA, a mitotic kinesin, has been suggested to promote cancer cell proliferation tumor formation and play a vital role in coordinating cell movement [54].

IMP3 in malignant vs. normal pancreas:

IMP3 marker shows cytoplasmic distribution with evidence of malignant pancreatic tumors and high-grade dysplastic lesions, somehow in low-grade dysplastic lesions. Nevertheless, it is scarcely found in normal pancreatic tissue or benign pancreatic lesions. IMP3 was positive in 78.7% of PAC, 91.7% of MCN high-grade dysplasia, 100% of IPMN high-grade dysplasia. While negative nearly in all benign cases (95.8%) [36]. Also, it was detected in 80.8%, 92% of malignant lesions Senoo (2018) and Yantiss (2008) respectively, and 0% of benign lesions in both studies [55], [56]. Evaluation of IMP3 on different pancreatic lesions and specimens (core needle biopsies and resection) showed that, in contrast to normal or inflamed pancreatic tissue, which was negative in 47 of 65 (72.3%) cases and weakly

Review Article

positive in 15 of 65 (23.1%) cases, strong IMP3 expression was found in 99 of 112 (88.4%) PDA. So, IMP3 expression sensitivity and specificity in PDA differentiation from chronic sclerosing pancreatitis on core needle biopsies were 88.4% and 94.6%, respectively [57].

IMP3 with high grades and stage pancreatic cancer:

Expression of IMP3 usually accompanies poor prognostic factors. Higher expression was detected in the advanced TNM stage and poorer prognosis. [56], [58]. In urinary bladder cancers and esophageal adenocarcinomas, overexpression was correlated with high stage and grade. Also, a significant correlation with shortened survival in gastric and lung adenocarcinomas was observed [47].

-IMP3 expression is positive mainly in malignant cases such as (PDA MCN with high-grade dysplasia) and harmful in benign cases (pseudocyst, serous cystadenoma, and pancreatitis). The score is directly proportional to the grade. Higher expression of IMP3 detected in advanced TNM stages and poorer prognosis. Malignant lesions with over-expression of IMP3 often suggest a poorer prognosis [56], [58]. Higher expression was detected with perineural, vascular, deeper, and metastasis to lymph nodes [59].

IMP3 diagnostic and prognostic factors in different organs

IMP3 expression in various cancers:

Expression of IMP3 was widely demonstrated in several human cancers. For example, it was seen in neuroblastoma (88%), Hodgkin's lymphoma (90%), and squamous cell carcinoma in distinct organs [47].

IMP3 was detected in nearly 67% of the cases in hepatocellular carcinoma and gastric cancer. Over-expression in both cancers was associated with poor outcomes [59], [60]. Also, high expression was demonstrated in 41% of lung adenocarcinomas [61]. IMP3 expression in colon cancer is correlated with cancer metastasis and has a high recurrence rate [62]. IMP3 is a diagnostic and prognostic marker in renal cell carcinoma [63].

IMP3 has been studied in different organs to assess its diagnostic and prognostic value. For diagnosis of endometrial cancers and their premalignant lesions, IMP3 showed a marked and diffuse expression, mainly in endometrial serous and clear cell carcinomas, including their precursor lesions [64]. On the other hand, high expression was considered a poor prognostic predictor for duodenal papillary carcinoma. Also, an objective diagnosis based

Review Article

on IMP3 evaluation can be offered for patients with papillary tumors to determine if endoscopic papillectomy can be employed [65].

IMP3 was considered crucial in predicting mucoepidermoid carcinoma of salivary glands outcome as positive expression was related to age above 60 years, tumors of the submandibular gland, size more than 4 cm, higher grade, lymph node involvement, perineural invasion, surgical margins involvement, distant spread, higher stages, tumor relapse, and death. Also, as a diagnostic marker, IMP3 could distinguish between benign and malignant lesions of salivary glands, as it was negative in pleomorphic adenoma and normal salivary gland tissues and positive in 51.4% mucoepidermoid carcinoma. Increased expression in submandibular gland tumors and lymph node involvement are independent prognostic factors of free survival [66]. In addition to pilocytic and pilomyxoid astrocytomas, overexpression was considered a poor prognostic predictor [67].

By using EUS-FNA, the sensitivity and, specificity were 80.8%, 100%, and 80.3%, 92.3%, respectively [55], [68]. While combination of EUS-FNA and IMP3 expression raised the sensitivity reaching 89% [57], 87.9% [54], and 92%, [56] [69]. The four studies showed specificity of 100% for this combined diagnostic tool. Also, Rashed (2021) and Ezzat et al 2016 have reported sensitivity, and specificity of 78.2%, 95.8%, and 91.2%, 86.7%, respectively [36][70].

Conclusion:

EUS-FNA is considered a flexible, safe procedure, particularly with the ROSE technique to have sufficient material for a proper diagnostic approach of pancreatic lesions. In addition, immunohistochemical expression of IMP3 on cytological smears or cell blocks obtained by EUS-FNA will add an excellent diagnostic and prognostic value if added to the diagnosis panel.

Footnotes.

Peer- Reviewers: Amr Shaban Hanafy (professor of internal medicine), Aziza E Abdelrahman (Assistant professor of pathology), Abdalla Hussein (Military hospitals, department of pathology).

E- Editor: Salem Y Mohamed.

Copyright ©. This open-access article is distributed under the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution, or reproduction in other forums

Review Article

is permitted, provided the original author(s) and the copyright owner(s) are credited. The original publication in this journal is cited by accepted academic practice. No use, distribution, or reproduction is permitted, complying with these terms.

Disclaimer: All claims expressed in this article are solely those of the authors and do not necessarily represent their affiliated organizations or those of the publisher, the editors, and the reviewers. Any product evaluated in this article or claim that its manufacturer may make is not guaranteed or endorsed by the publisher.

References:

1. D'Onofrio M, De Robertis R, Barbi E, Martone E, Manfrin E, Gobbo S, Puntel G, Bonetti F, Mucelli RP. Ultrasound-guided percutaneous fine-needle aspiration of solid pancreatic neoplasms: 10-year experience with more than 2,000 cases and literature review. *European radiology*. 2016 Jun 1;26(6):1801-7.
2. Okasha HH, Naga MI, Serag Esmat MN, Hassanein M, Hassani M, El-Kassas M, Mahdy RE, El-Gemeie E, Farag AH, Foda AM. Endoscopic ultrasound-guided fine-needle aspiration versus percutaneous ultrasound-guided fine-needle aspiration in the diagnosis of focal pancreatic masses. *Endoscopic ultrasound*. 2013 Oct;2(4):190.
3. Jani BS, Rzouq F, Saligram S, Lim D, Rastogi A, Bonino J, Olyae M. Endoscopic ultrasound-guided fine-needle aspiration of pancreatic lesions: a systematic review of technical and procedural variables. *North American journal of medical sciences*. 2016 Jan;8(1):1.
4. Micames C, Jowell PS, White R, Paulson E, Nelson R, Morse M, Hurwitz H, Pappas T, Tyler D, McGrath K. Lower frequency of peritoneal carcinomatosis in patients with pancreatic cancer diagnosed by EUS-guided FNA vs. percutaneous FNA. *Gastrointestinal endoscopy*. 2003 Nov 1;58(5):690-5.
5. Ahmad NA, Lewis JD, Ginsberg GG, Rosato EF, Morris JB, Kochman ML. EUS in preoperative staging of pancreatic cancer. *Gastrointestinal endoscopy*. 2000 Oct 1;52(4):463-8.
6. Hawes RH. Indications for EUS-directed FNA. *Endoscopy*. 1998 Aug;30(S 1):A155-7.
7. Yamao K, Sawaki A, Mizuno N, Shimizu Y, Yatabe Y, Koshikawa T. Endoscopic ultrasound-guided fine-needle aspiration biopsy (EUS-FNAB): past, present, and future. *Journal of gastroenterology*. 2005 Nov 1;40(11):1013.

Review Article

8. Jenssen C, Dietrich CF. Endoscopic ultrasound-guided fine-needle aspiration biopsy and trucut biopsy in gastroenterology—An overview. *Best practice & research Clinical gastroenterology*. 2009 Oct 1;23(5):743-59.
9. Forcione DG. On-site cytopathology for endoscopic ultrasound-guided fine-needle aspiration of solid pancreatic masses: Is it Time to Make it Standard of Care?. *Cancer cytopathology*. 2013 Sep;121(9):471-2.
10. Pioppo L, Tyberg A. Indications and techniques of fine-needle aspiration of the pancreas. In *Pancreas and Biliary Tract Cytopathology 2019* (pp. 1-20). Springer, Cham.
11. Jani BS, Rzouq F, Saligram S, Lim D, Rastogi A, Bonino J, Olyae M. Endoscopic ultrasound-guided fine-needle aspiration of pancreatic lesions: a systematic review of technical and procedural variables. *North American journal of medical sciences*. 2016 Jan;8(1):1.
12. Gimeno-García AZ, Elwassief A, Paquin SC, Gariépy G, Sahai AV. Randomized controlled trial comparing stylet-free endoscopic ultrasound-guided fine-needle aspiration with 22-G and 25-G needles. *Digestive Endoscopy*. 2014 May;26(3):467-73.
13. Bang JY, Ramesh J, Trevino J, Eloubeidi MA, Varadarajulu S. Objective assessment of an algorithmic approach to EUS-guided FNA and interventions. *Gastrointestinal endoscopy*. 2013 May 1;77(5):739-44.
14. Ramesh J, Bang JY, Hebert-Magee S, Trevino J, Eltoun I, Frost A, Hasan MK, Logue A, Hawes R, Varadarajulu S. Randomized trial comparing the flexible 19G and 25G needles for endoscopic ultrasound-guided fine needle aspiration of solid pancreatic mass lesions. *Pancreas*. 2015 Jan;44(1):128.
15. Erickson RA, Sayage-Rabie L, Beissner RS. Factors predicting the number of EUS-guided fine-needle passes for diagnosis of pancreatic malignancies. *Gastrointestinal endoscopy*. 2000 Feb 1;51(2):184-90.
16. Conrad R, Castelino-Prabhu S, Cobb C, Raza A. Cytopathology of the pancreatobiliary tract—the agony, and sometimes, its ease. *Journal of gastrointestinal oncology*. 2013 Jun;4(2):210.
17. Petrone MC, Arcidiacono PG. Basic technique in endoscopic ultrasound-guided fine-needle aspiration for solid lesions: How many passes?. *Endoscopic ultrasound*. 2014 Jan;3(1):22.
18. Lee LS, Saltzman JR, Bounds BC, Poneris JM, Brugge WR, Thompson CC. EUS-guided fine-needle aspiration of pancreatic cysts: a retrospective analysis of complications and their predictors. *Clinical Gastroenterology and Hepatology*. 2005 Mar 1;3(3):231-6.
19. Ylagan LR, Edmundowicz S, Kasal K, Walsh D, Lu DW. Endoscopic ultrasound-guided fine-needle aspiration cytology of pancreatic carcinoma: a 3-year

Review Article

20. experience and review of the literature. *Cancer Cytopathology: Interdisciplinary International Journal of the American Cancer Society*. 2002 Dec 25;96(6):362-9.
21. Ieni A, Barresi V, Todaro P, Caruso RA, Tuccari G. Cell-block procedure in endoscopic ultrasound-guided-fine-needle-aspiration of gastrointestinal solid neoplastic lesions. *World journal of gastrointestinal endoscopy*. 2015 Aug 25;7(11):1014.
22. Pitman MB, Deshpande V. Endoscopic ultrasound-guided fine needle aspiration cytology of the pancreas: a morphological and multimodal approach to the diagnosis of solid and cystic mass lesions. *Cytopathology*. 2007 Dec;18(6):331-47.
23. Ohara Y, Oda T, Hashimoto S, Akashi Y, Miyamoto R, Enomoto T, Satomi K, Morishita Y, Ohkohchi N. Pancreatic neuroendocrine tumor and solid-pseudopapillary neoplasm: Key immunohistochemical profiles for differential diagnosis. *World journal of gastroenterology*. 2016 Oct 14;22(38):8596.
24. Skacel M, Ormsby AH, Petras RE, McMahon JT, Henricks WH. Immunohistochemistry in the differential diagnosis of acinar and endocrine pancreatic neoplasms. *Applied Immunohistochemistry & Molecular Morphology*. 2000 Sep 1;8(3):203-9.
25. Reid MD, Bhattarai S, Graham RP, Pehlivanoglu B, Sigel CS, Shi J, Saqi A, Shirazi M, Xue Y, Basturk O, Adsay V. Pancreatoblastoma: Cytologic and histologic analysis of 12 adult cases reveals helpful criteria in their diagnosis and distinction from common mimics. *Cancer cytopathology*. 2019 Nov;127(11):708-19.
26. Sigel CS, Klimstra DS. Cytomorphologic and immunophenotypical features of acinar cell neoplasms of the pancreas. *Cancer cytopathology*. 2013 Aug;121(8):459-70.
27. Chatzipantelis P, Salla C, Konstantinou P, Karoumpalis I, Sakellariou S, Doumani I. Endoscopic ultrasound-guided fine-needle aspiration cytology of pancreatic neuroendocrine tumors: a study of 48 cases. *Cancer Cytopathology: Interdisciplinary International Journal of the American Cancer Society*. 2008 Aug 25;114(4):255-62.
28. Kamisawa T, Tu Y, Egawa N, Nakajima H, Tsuruta K, Okamoto A. Malignancies associated with intraductal papillary mucinous neoplasm of the pancreas. *World journal of gastroenterology: WJG*. 2005 Sep 28;11(36):5688.
29. Adsay NV. Cystic lesions of the pancreas. *Modern Pathology*. 2007 Feb;20(1):S71-93.
30. Yonezawa S, Nakamura A, Horinouchi M, Sato E. The expression of several types of mucin is related to the biological behavior of pancreatic neoplasms. *Journal of hepato-biliary-pancreatic surgery*. 2002 Sep 1;9(3):328-41.

Review Article

31. Naveed S, Qari H, Banday T, Altaf A, Para M. Mucinous cystic neoplasms of pancreas. *Gastroenterology research*. 2014 Apr;7(2):44.
32. Buerlein RC, Shami VM. Management of pancreatic cysts and guidelines: what the gastroenterologist needs to know. *Therapeutic Advances in Gastrointestinal Endoscopy*. 2021 Sep;14:26317745211045769.
33. Khalid A, Zahid M, Finkelstein SD, LeBlanc JK, Kaushik N, Ahmad N, Brugge WR, Edmundowicz SA, Hawes RH, McGrath KM. Pancreatic cyst fluid DNA analysis in evaluating pancreatic cysts: a report of the PANDA study. *Gastrointestinal endoscopy*. 2009 May 1;69(6):1095-102.
34. Wu J, Matthaei H, Maitra A, Dal Molin M, Wood LD, Eshleman JR, Goggins M, Canto MI, Schulick RD, Edil BH, Wolfgang CL. Recurrent GNAS mutations define an unexpected pathway for pancreatic cyst development. *Science translational medicine*. 2011 Jul 20;3(92):92ra66-.
35. Clores MJ, Thosani A, Buscaglia JM. Multidisciplinary diagnostic and therapeutic approaches to pancreatic cystic lesions. *Journal of multidisciplinary healthcare*. 2014;7:81.
36. Aslan DL, Jessurun J, Gulbahce HE, Pambuccian SE, Adsay V, Mallery JS. Endoscopic ultrasound-guided fine needle aspiration features of a pancreatic neoplasm with predominantly intraductal growth and prominent tubular cytomorphology: Intraductal tubular carcinoma of the pancreas?. *Diagnostic cytopathology*. 2008 Nov;36(11):833-9.
37. Rashed H, Wasfi N, Nasr A, ElHendawy R, Said NM. A Promising Diagnostic Role of Immunohistochemical Expression of Insulin-Like Growth Factor II mRNA Binding Protein3 (IMP3) in Pancreatic Lesions Using Endoscopic Ultrasound–Guided–Fine Needle Aspiration (EUS-FNA) Cytology. 2021 Nov 5.
38. Woolf KM, Liang H, Sletten ZJ, Russell DK, Bonfiglio TA, Zhou Z. False-negative rate of endoscopic ultrasound-guided fine-needle aspiration for pancreatic solid and cystic lesions with matched surgical resections as the gold standard: one institution's experience. *Cancer cytopathology*. 2013 Aug;121(8):449-58.
39. Lin F, Staerkel G. Cytologic criteria for well differentiated adenocarcinoma of the pancreas in fine-needle aspiration biopsy specimens. *Cancer Cytopathology: Interdisciplinary International Journal of the American Cancer Society*. 2003 Feb 25;99(1):44-50.
40. Ballehaninna UK, Chamberlain RS. Biomarkers for pancreatic cancer: promising new markers and options beyond CA 19-9. *Tumor Biology*. 2013 Dec;34(6):3279-92.
41. Salla C, Chatzipantelis P, Konstantinou P, Karoumpalis I, Pantazopoulou A, Tsiotos G. EUS-FNA contribution in the identification of autoimmune pancreatitis: a case report. *JOP*. 2007 Sep 7;8(5):598-604.

Review Article

42. Kim KP, Kim MH, Song MH, Lee SS, Seo DW, Lee SK. Autoimmune chronic pancreatitis. *Official journal of the American College of Gastroenterology| ACG*. 2004 Aug 1;99(8):1605-16.
43. Martínez-Ordaz JL, Toledo-Toral C, Franco-Guerrero N, Tun-Abraham M, Souza-Gallardo LM. Surgical treatment of pancreatic pseudocysts. *Cirugía y Cirujanos (English Edition)*. 2016 Jul 1;84(4):288-92.
44. Liu J, Shin HJ, Rubenchik I, Lang E, Lahoti S, Staerckel GA. Cytologic features of lymphoepithelial cyst of the pancreas: two preoperatively diagnosed cases based on fine-needle aspiration. *Diagnostic cytopathology*. 1999 Nov;21(5):346-50.
45. Nielsen J, Kristensen MA, Willemoes M, Nielsen FC, Christiansen J. Sequential dimerization of human zipcode-binding protein IMP1 on RNA: a cooperative mechanism providing RNP stability. *Nucleic acids research*. 2004 Jan 1;32(14):4368-76.
46. Jia M, Gut H, Chao JA. Structural basis of IMP3 RRM12 recognition of RNA. *RNA*. 2018 Dec 1;24(12):1659-66.
47. Wächter K, Köhn M, Stöhr N, Hüttelmaier S. Subcellular localization and RNP formation of IGF2BPs (IGF2 mRNA-binding proteins) is modulated by distinct RNA-binding domains. *Biological chemistry*. 2013 Aug 1;394(8):1077-90.
48. Burdelski C, Jakani-Karimi N, Jacobsen F, Möller-Koop C, Minner S, Simon R, Sauter G, Steurer S, Clauditz TS, Wilczak W. IMP3 overexpression occurs in various important cancer types and is linked to aggressive tumor features: a tissue microarray study on 8,877 human cancers and normal tissues. *Oncology reports*. 2018 Jan 1;39(1):3-12.
49. Schneider T, Hung LH, Schreiner S, Starke S, Eckhof H, Roszbach O, Reich S, Medenbach J, Bindereif A. CircRNA-protein complexes: IMP3 protein component defines subfamily of circRNPs. *Scientific reports*. 2016 Aug 11;6(1):1-1
50. Mori H, Sakakibara SI, Imai T, Nakamura Y, Iijima T, Suzuki A, Yuasa Y, Takeda M, Okano H. Expression of mouse igf2 mRNA-binding protein 3 and its implications for the developing central nervous system. *Journal of neuroscience research*. 2001 Apr 15;64(2):132-43.
51. Wagner M, Kunsch S, Duerschmied D, Beil M, Adler G, Mueller F, Gress TM. Transgenic overexpression of the oncofetal RNA binding protein KOC leads to remodeling of the exocrine pancreas. *Gastroenterology*. 2003 Jun 1;124(7):1901-14.
52. Vargas TR, Boudoukha S, Simon A, Souidi M, Cuvellier S, Pinna G, Polesskaya A. Post-transcriptional regulation of cyclins D1, D3 and G1 and proliferation of human cancer cells depend on IMP-3 nuclear localization. *Oncogene*. 2014 May;33(22):2866-75.

Review Article

53. Vikesaa J, Hansen TV, Jønson L, Borup R, Wewer UM, Christiansen J, Nielsen FC. RNA-binding IMPs promote cell adhesion and invadopodia formation. *The EMBO journal*. 2006 Apr 5;25(7):1456-68.
54. Pasiliao CC, Chang CW, Sutherland BW, Valdez SM, Schaeffer D, Yapp DT, Ng SS. The involvement of insulin-like growth factor 2 binding protein 3 (IMP3) in pancreatic cancer cell migration, invasion, and adhesion. *BMC cancer*. 2015 Dec;15(1):1-9.
55. Sun XD, Shi XJ, Sun XO, Luo YG, Wu XJ, Yao CF, Yu HY, Li DW, Liu M, Zhou J. Dimethylenastron suppresses human pancreatic cancer cell migration and
56. invasion in vitro via allosteric inhibition of mitotic kinesin Eg5. *Acta pharmacologica Sinica*. 2011 Dec;32(12):1543-8.
57. Senoo J, Mikata R, Kishimoto T, Hayashi M, Kusakabe Y, Yasui S, Yamato M, Ohyama H, Sugiyama H, Tsuyuguchi T, Yoshitomi H. Immunohistochemical analysis of IMP3 and p53 expression in endoscopic ultrasound-guided fine needle aspiration and resected specimens of pancreatic diseases. *Pancreatology*. 2018 Mar 1;18(2):176-83.
58. Yantiss RK, Cosar E, Fischer AH. Use of IMP3 in identification of carcinoma in fine needle aspiration biopsies of pancreas. *Acta cytologica*. 2008;52(2):133-8.
59. Wachter DL, Schlabrakowski A, Hoegel J, Kristiansen G, Hartmann A, Riener MO. Diagnostic value of immunohistochemical IMP3 expression in core needle biopsies of pancreatic ductal adenocarcinoma. *The American journal of surgical pathology*. 2011 Jun 1;35(6):873-7.
60. Tadic M, Stoos-Veic T, Kujundzic M, Turcic P, Aralica G, Boskoski I. Insulin-like growth factor 2 binding protein 3 expression on endoscopic ultrasound guided fine needle aspiration specimens in pancreatic ductal adenocarcinoma. *European journal of gastroenterology & hepatology*. 2020 Apr 1;32(4):496-500.
61. Damasceno EA, Carneiro FP, de Magalhães AV, de Vasconcelos Carneiro M, Takano GH, de Sousa Vianna LM, Seidler HB, de Castro TM, Muniz-Junqueira MI, Amorim RF, Ferreira VM. IMP3 expression in gastric cancer: association with clinicopathological features and HER2 status. *Journal of cancer research and clinical oncology*. 2014 Dec;140(12):2163-8.
62. Jeng YM, Chang CC, Hu FC, Chou HY, Kao HL, Wang TH, Hsu HC. RNA-binding protein insulin-like growth factor II mRNA-binding protein 3 expression promotes tumor invasion and predicts early recurrence and poor prognosis in hepatocellular carcinoma. *Hepatology*. 2008 Oct;48(4):1118-27.
63. Yan J, Wei Q, Jian W, Qiu B, Wen J, Liu J, Fu B, Zhou X, Zhao T. IMP3 predicts invasion and prognosis in human lung adenocarcinoma. *Lung*. 2016 Feb 1;194(1):137-46.

Review Article

64. Li D, Yan D, Tang H, Zhou C, Fan J, Li S, Wang X, Xia J, Huang F, Qiu G, Peng Z. IMP3 is a novel prognostic marker that correlates with colon cancer progression and pathogenesis. *Annals of surgical oncology*. 2009 Dec;16(12):3499-506.
65. Jiang Z, Chu PG, Woda BA, Rock KL, Liu Q, Hsieh CC, Li C, Chen W, Duan HO, McDougal S, Wu CL. Analysis of RNA-binding protein IMP3 to predict metastasis and prognosis of renal-cell carcinoma: a retrospective study. *The lancet oncology*. 2006 Jul 1;7(7):556-64.
66. Zheng W, Yi X, Fadare O, Liang SX, Martel M, Schwartz PE, Jiang Z. The oncofetal protein IMP3: a novel biomarker for endometrial serous carcinoma. *The American journal of surgical pathology*. 2008 Feb 1;32(2):304-15.
67. Tanaka H, Kawashima H, Ohno E, Ishikawa T, Iida T, Ishikawa E, Furukawa K, Nakamura M, Honda T, Shimoyama Y, Miyahara R. Immunohistochemical staining for IMP3 in patients with duodenal papilla tumors: assessment of the potential for diagnosing endoscopic resectability and predicting prognosis. *BMC gastroenterology*. 2021 Dec;21(1):1-0.
68. Elshafey MR, Ahmed RA, Mourad MI, Gaballah ET. The oncofetal protein IMP3 is an indicator of early recurrence and poor outcome in mucoepidermoid carcinoma of salivary glands. *Cancer Biol Med*. 2016;13(2):286-295.
69. Barton VN, Donson AM, Birks DK, et al. Insulin-like growth factor 2 mRNA binding protein 3 expression is an independent prognostic factor in pediatric pilocytic and pilomyxoid astrocytoma. *J Neuropathol Exp Neurol*. 2013;72(5):442-449.
70. Alizadeh AH, Shahrokh S, Hadizadeh M, Padashi M, Zali MR. Diagnostic potency of EUS-guided FNA for the evaluation of pancreatic mass lesions. *Endoscopic ultrasound*. 2016 Jan;5(1):30.
71. Han L, Patel C. Utility of IMP3 Immunohistochemistry in the Distinction of Pancreatic Adenocarcinoma and Chronic Pancreatitis. *American Journal of Clinical Pathology*. 2014 Oct 1;142(suppl_1):A225-.
72. Ezzat NE, Tahoun NS, Ismail YM. The role of S100P and IMP3 in the cytologic diagnosis of pancreatic adenocarcinoma. *J Egypt Natl Canc Inst*. 2016 Dec;28(4):229-234.