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ISOLATION OF AEROMANAS SPECIES FROM CHILDREN WITH AND

WITHOUT DIARRHOEA IN JOS, NIGERIA.

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**ABSTRACT** 

An investigation on the prevalence and antibiogram of Aeromonas species

among children in Jos was conducted. The samples analysed included a total of

104 (52 diarrhoeal and 52 non - diarrhoea) stool samples collected from Vom

Christian and Plateau Specialists Hospital in Jos. Aeromonas isolates were

identified using standard biochemical tests. Of the total number examined, 6

(5.7%) were positive for Aeromonas species, 2 (3.9%) from diarrhoeal and 4 (7.7%)

from diarrhoeal samples non

(P>0.05). All isolates were identified as Aeromonas hydrophilia. The

highest number of isolates 3 (10.7%) were recovered from the group 7-12 months.

No isolates were recovered from exclusively breast fed children while the highest

number 4 (9.8%) was found in children fed with breast milk and formula. The

isolates were found to be very sensitive to ciprofloxacin, but resistant to

penicillin.

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### INTRODUCTION

Diarrhoeal diseases constitute major childhood mortality and morbidity world wide especially in developing countries (1). Estimates show that diarrhoeal diseases cause nearly 5 million deaths annually in children under 5 years old in developing countries. Traditional aetiologic agents diarrhoea include Entamoeba histolytica, Giardia lamblia, Salmonella species, Shigella species and Vibrio cholerae (2). However, other agents as Campylobacter, Yersinia, Aeromonas, Plesiomonas and Crytosporidium have also been implicated in gastrointestinal diseases and are often referred to as new agents of diarrhoea (3,4).

Of growing importance in recent times is Aeromonas which affects all age groups but is said to be most common in children under 5 years, the elderly and the immunocompromised (5).

Aeromonas species are gramnegative bacilli of the Aeromonadaceae family. These motile bacteria are involved in both intestinal and extraintestinal human infections (6) with clinical manifestations ranging from skin and soft tissue infection, bacteremia, to gastroenteritis (7). However, acute watery diarrhoea with a short duration is the most common clinical feature (8).

The first reported association of Aeromonas with gastrointestinal disease was in 1958 in Jamaica(9), since then numerous reports have appeared from several countries including Italy, England, Australia and the United States regarding the isolation of Aeromonas from faeces of patients with diarrhoea (10,11).

In Nigeria Obi et al (12) identified Aeromonas species and Plesiomonas shigelloides as bacterial agents of diarrhoea in urban and rural areas. Aeromonas have also been found in cases of acute diarrhoea and asysmptomatic infections in Nigerian school children (13).

Reported frequency of isolation from symptomatic (diarrhoeic) as compared with asymptomatic (non-diarrhoeic) cases varies considerably, with some studies showing no significant difference in isolation rates (14, 15).

This study was therefore undertaken to examine the prevalence of Aeromonas species among children with and without diarrhoea and to identify the antibiogram of recovered isolates.

### **MATERIALS AND METHODS**

## Samples

The samples analysed in this study included a total of one hundred and four (52 diarrhoeal and 52 non-diarrhoeal) stool specimens collected from Vom Christian and Plateau Specialist Hospital in Jos.

Stool samples were collected from patients in clean, transparent wide-mouthed bottles. Information was also obtained from each subject regarding age, sex, major symptoms (diarrhoea, vomiting and fever) duration of disease, source of water and feeding pattern.

### **Processing of Specimens**

The specimens were processed according to guidelines provided by Cheesbrough(16) for the laboratory diagnosis of enteric pathogens. These include, macroscopy, microscropy, gram stain, motility testing, culture, biochemical testing and antimicrobial sensitivity testing.

Specimens were inoculated into the medium of Agger et al (5) for the isolation of Aeromonas species (5% sheep blood agar containing 30µg/ml ampicillin). The inoculated plates were then incubated aerobically at 37°C for 24 hours. Resultant colonies were identified using biochemical tests.

### Biochemical testing

Isolates that were beta haemolytic on sheep blood agar and gram -negative bacilli were identified as Aeromonas species using the following standard tests; oxidase test, indole test, urease test, citrate utilization test and test to determine motility after distilled water and peptone water subcultures. All tests were done using the methods

described by Collee and Miles (17) and Porter and Duguid (18).

# Characterization of Species

Isolates were characterized to the species level based on seven biochemical tests as described by Carnahan et al (19). These included aesculin hydrolysis, gas from glucose, acid from arabinose, indole production, acid from sucrose, Voges-Proskauer reaction and resistance to cephalothin (30µg).

# Antimicrobial Susceptibility Testing

Sensitivity of isolates to antimicrobial agents was determined on Mueller-Hinton agar plates using the disc diffusion method of Scott (20). From a pure culture of the isolate to be tested a uniform streak was made on the agar plate. The antibiotic (Antec Diagnostics, UK) discs were placed on the plates and incubated at 37°C overnight. Interpretation of results was done using the zone sizes. Zones of inhibition of ≥ 18mm were considered

while sensitive 13-17mm were considered intermediate and <13mm were considered resistant. All isolates were tested for sensitivity to the following antibiotics, ciprofloxacin (5mcg) cotrimoxazole (25mcg) streptomycin gentamycin (10mcg), erythromycin (10mcg), (5mcg), tetracycline (10mcg) penicillin (5mcg) peflacine (10mcg) and tarivid (10mcg).

# Statistical Analysis

The data obtained were subjected to the chi-squared test using a probability of P=0.05 as the level of significance.

### RESULTS

A total of 104 (54 diarrhoeal and 52 non-diarrhoeal) stool samples were examined. The age range of the patients was 0-72 months. Of the total number of specimen examined, 6 (5.7%) were positive for Aeromonas spp. 2 (3.9%) of Aeromonas spp were recovered from specimens while diarrhoeal stool 4(7.7%) from non-diarrhoeal samples 1). difference is (Table The not significant statistically 1

P>0.05). All the isolates were found to be Aeromonas hydrophila.

The highest numbers of isolates 3(10.7%) were recovered form the age group 7-12 months. The age brackets 13-18 months, 19-24 months and 67-72 months had 1 isolate each. No isolates were recovered from age group 0-6 months and from 25-66 months (Table 2). The difference is not statistically significance (P>0.05).

Macroscopic examination of the specimens showed that 36 were watery was found in children fed with breast milk and formular 4 (9.8%) followed by formular and family diet 2(4.7%). No

13 mucoid, 3 blood stained, 40 soft-formed and 12 hard-formed. The soft-formed specimens yielded the highest number of isolates 3 (7.5%), watery samples 2(5.6%) and, hard-formed 1(8.3%). The blood stained and mucoid specimens yielded no isolates (Table 3). This difference is not statistically significant (P>0.05).

Table 4 shows the prevalence of Aeromonas spp in relation to the feeding pattern of the children. The highest number of isolates isolates were recovered from exclusively breast fed children. This result is not statistically significant.

Table 1: Prevalence of Aeromonas species among symptomatic and asymptomatic patients.

Patients	No. of Examined	Specimens	No. (%) Positive
Symptomatic (with diarrhoea)	52		2(3.9)
A symptomatic (without diarrhoea)	52		4(7.7)
Total	104		6(5.8)

 $X^2 = 1.2$ 

df = 1 P > 0.05

Table 2: Prevalence of Aeromonas species isolated in relation to age and sex:

Age Group (Months)		No of Specimens Collected		No (%) Positive	
Male	Female	Male	Female	Total	
0-6	5	14	0(0.0)	0(0.0)	0(0.0)
7-12	17	11	2(7.1)	1(3.6)	3(10.7)
13-18	10	5	1(6.7)	0(0.0)	1(6.7)
19-24	8	5	1(7.7)	0(0.0)	1(7.7)
25-30	5	2	0(0.0)	0(0.0)	0(0.0)
31-36	3	1	0(0.0)	0(0.0)	0(0.0)
37-42	3	2	0(0.0)	0(0.0)	0(0.0)
43-48	2	1	0(0.0)	0(0.0)	0(0.0)
49-54	1	0	0(0.0)	0(0.0)	0(0.0)
55-60	2	1	0(0.0)	0(0.0)	0(0.0)
61-66	2	1	0(0.0)	0(0.0)	0(0.0)
67-72	1	2	0(0.0)	1(33.3)	1(33.3)
Total	59	45	4(3.9)	2(1.9)	6(5.8)

 $X^2 = 21.35, df = 11 P>0.05$ 

Table 3: Types of samples treated and the number (%) of Aeromonas species isolated.

Types of Stool	No. Examined	No. (%) Positive
Watery	36	2(5.6)
Mucoid	13	0(0.0)
Blood stained	3	0(0.0)
Soft formed	40	3(7.5)
Hard formed	12	1(8.3)
Total	104	6(5.8)
$X^2 = 132,  \alpha$	If =4 P>0.05	

Table 4: Prevalence of Aeromonas species in Relation to the type of Feeding

Type of Feeding	No. of Patient Tested	No. (% positive)	
Breast milk	20	0(0.0)	
Breast milk & formular	41	4(9.8)	
Formular & family diet	43	2(4.7)	
Total	104	5(5.8)	

 $X^2 = 60.17 \text{ df} = 2 \text{ P} > 0.05$ 

Table 5 shows the in-vitro susceptibility pattern of the isolates. Six (100.0%) of the isolates were sentive to ciprofloxacin, 5 (83.33%) to gentamycin, peflacine and tarivid, 4 (66.67%) to erythromycin and streptomycin, the highest isolation rates were found in infants 7-12 months. This result correlates with the findings of Abraham et al (21) and Regua et al., (22). They both observed that the highest incidence of gastroenteritis in children was found within the age range of 7-12

The protective role of breast milk against diarrhoeal bacterial aetiologic is well documented (23, 24).

the had their breast feeding interrupted with mixed feeding or stopped completely.

Another probable reason for the increase incidence of gastroenteritis around 7-12 of age months might be due to faulty weaning practices and poor hygiene in preparing food.

The low isolation rate in asymptomatic children older than age 12 months might be attributed to immunity (50.0%) to tetracycline and cotrimoxazole. All isolates were resistant to penicillin.

#### DISCUSSION

months where weaning practices begin ...in many parts of the world (Nigeria inclusive). The finding indicates that breast milk confers considerable protection to children as positive cases were not reported in children below 7 months whose mothers practice exclusive breast feeding.

Disease developed by the older children who may have come in contact with the agent through exposure. Aeromonas spp was found to be higher in males (6.8%) than in females (4.4%). This finding may be related to the number of male and female children from who samples were collected. I.e. more samples were collected from males than females. However, this result is not statistically significant and no sex preference has been reported.

Aeraomonas spp. were isolated more frequently from loose and watery stools.

The result of in-vitro antibiotic sensitive test showed 100% sensitivity ciprofloxacin and more than 80% sensitivity to peflacine, tarvid and gentamicin. Cirpoflaxacin therefore is the drug of choice, when treating Aeromonas infections from this study. This presents cause for concern since it is expensive. Conventional and cheaper drugs like (cotrimoxazole, tetracycline, streptomycin and erythromycin) showed marked reduced in vitro susceptibility. This may be due to indiscriminate usage or an antibiotic (drug abuse) which has resulted in multiple drug resistance of many microorganisms in Nigeria (25). In addition, other enteric Bacteria isolated in patyients with diarrhea in Jos are resistant to common antibiotics (26, 27).

rate from children withot diarrhoea (7.7%) compare to those with diarrhoea (3.9%). However this result is not statistically significant This agrees with findings from some researchers (Pitaragis et al., (14) and Figura et al., (15). Aeromonas spp was isolated from infant below 6 months.

Other common enteric pathogens like Salmonella, Shegella and Escherichia coli were not sought for in this study therefore it can not be concluded that the Aeromonas spp isolated were the actual cause or the diarrhoea in this study.

A total of 104 stool samples were analysed in this study in which the prevalence rate for Aeromonas spp was 5.8%. This result is similar to the 5% prevalence rate documented by Obi et al., (12) for urban population in Edo, Lagos and Cross River States of Nigeria. All isolates identified were found to be Aeromonas hydrophilia. This Aeromonas spp has been associated with many cases of diarrhoea (5).

### References

- 1. Odugbemi T, Addoyin MA,
  Okoro E, Agbede O.
  Study of a new
  formulation of Diapec
  without antibiotics in
  acute diarrhoea
  diseases. Current
  Therapeutic Research.
  1986; 39: 106-111.
- 2. Odugbemi T, Research
  Priorities on bacterial
  infections in Nigeria.
  In: Essien EM, Idigbe
  EO, Olukoya DK

- (eds.) International Conference on Health Research Priorities for Nigeria in 1990's and strategies for their achievement, 1992: 66-73.
- 3. Idigbe EO, Bacteria infections. In: Essien EM, Idigbe EO, Olukoya DK (eds). International Conference on Health Research **Priorities** for Nigeria in 1990's and Strategies for their achievement. 1992: 66-73.
- 4. Loughon BE, Druckman DA,
  Vermon A, Quin TC,
  Polk BF, Modlin JF,
  Prevalence of enteric
  pathogens in homosexual men without
  AIDS Hasterology,
  1998; 94: 983-993.
- 5. Agger WA, McCormick JD, Gurwith MJ. Clinical and Microbiological Features o Aeromonas associated diarrhoea, J. Clin Microbiol. 1985; 21:909-913.
- Duffey 6. Janda JM, PS. Mesophilic aeromonads in human diseases: Current taxonomy, laboratory identification, and infectious diseases spectrum, Rev. Infect. Dis. 1988; 10: 980.
- 7. Jones BL, Wilcox M.H.

  Aeromonas infections
  treatment. J.

  Antimicrobial
  Chemotherapy 1995;
  35 (4): 453-461.

- 8. Albert ML, Ansaruzzaman, Talukder M. KA. Prevalence of entrotoxin in genes in Aeromonas spp isolated from children with diarrhoea healthy controls and environment. J. Clin. Microbiol. 2000; 38 (10): 3785-3790.
- 9. Caselitz F.H. Zur Frage Von.

  Pseudomonas
  aeruginosa und Ver
  Wandten
  Mikrooganismen as
  enteritiserregen.
  Topen Med. Parasitol.
  1958; 9: 269-275.
- 10. Gracey M, Burke V,
  Robinson J.
  Aeromonas associated
  gastroenteritis.
  Lancet 1982; 2:
  1304-1306.
- 11. Challapalli M, Tess BR.

  Aeromonas
  associated diarrhoea
  in children Pediatr.
  Infect. Dis. J. 1988; 7:
  693-698.
- Obi CL, Coker AO, Epoke J, 12. Ndip RN. Enteric bacterial pathogens in stools of residents of urban and rural regions in Nigeria: a comparison of patients with and without diarrhoea and controls without diarrhoea. Diarrhoeal. Dis. Res. 1997; 15(4): 241-247.
- 13. Utsalo SY, Eko FO, Antia-Obong OE, Nwaigwe CU. Aeromonas in acute diarrhoea and asymptomatic

infection in Nigerian children European J of Epidemiol. 1995; 11 (2): 271-275.

- 14. Pitarangsi CP, Echeverria P, Whitmire R, Tirapat C, Formal s, Dammin GJ, Tingtalapong M. Enteropathogenicity **Aeromonas** hydrophilia and Plesiomonas shigelloides. Prevalence among individuals with and without diarrhoeal in Thailand. Infect Immun. 1982; 35: 666-673.
- 15. Figura N, Marri L, Verdiani Ceccherini Barberi A. Prevalence species differentiation, and toxigenicity of Aeromonas strains in cases of childhood gastroenteritis and in J. controls. Clin Microbiol. 1986; 23: 595-599.
- 16. Cheesbrough M. Medical laboratory manual for Tropical Countries. Press Syndicate of Cambridge University. 1985, 2: 192-193.
- 17. Collee JG, Miles RS. Tests for identification of bacteria. In: Collee JG, Dugid JP, Fraser AG, Marmion BP (eds.) Mackie and McCartney Practical Medical Microbiology, 1989: 141-160.
- 18. Porter IA, Duguid JP. Vibrio, *Aeromonas*Plesiomonas,

Spirillum, Campylobacter. In: Collee JG, Duguid JP, Fraser AG, Marmion BP (eds) Mackie and McCartney practical Medical Microbiology, 1989: 505-524.

- 19. Carnahan AM, Behram S,
  Joseph SW. Aerokey
  II: a flexible key for
  identifying clinical
  Aeromonas species. J.
  Clin. Microbiol 1991;
  29: 2843-2849.
- 20. Scott AC. Laboratory control of antimicrobial therapy. In Collee JG, Duguid JP, Fraser AG, Marmion BP (eds) Mackie and McCartney Practical Medical Microbiology, 1989: 161-181.
- 21. Abraham AA, Cahill Y,
  Davies T, Kawaguche
  LF, Miller JD,
  Norllway L, Damen
  GJ. Studies on
  Infantile diarrhoea in
  Cairo, Egypt J.
  Tropical Paediatric
  and Environmental
  Child Tealsts 1978;
  33: 187-193.
  - Rega AH, Barvo VLP, Lead
    MG, Lobe MEL.
    Epidemiology Survey
    of the
    enteropathogenic
    Escherichia coli
    isolated from children
    with diarrhoea. J.
    Tropical Paediatric
    1990; 36: 176-178.
- 23. Cameron M, Hofvander Y.

  Manual on Feeding
  Infants and Young
  Children. Oxford

22.

University Press, New York 1983: 87.

24. Kebede D, Ketsela, T, Astaw W, Patterns of Breast feeding in Western Ethiopia and their relationship to acute diarrhoea in infants.

J. of Paediatrics 1990; 36: 180-183.

25. monigho SE, Nwokoji AE,
Ophori EA.
Antimicrobial
Susceptibility of
Staphylococcus
aureus isolates from
operating theaters. J.
of Medical Lab
Science 1999; 8: 1822.

Nigeria. Afr. J Clin Exper Microbiol 2004; 5(3): 267-271.

27. Kandakai-Olukemi YT,
Okewu MS, Mawak
JD, Olukemi MA,
Zumbes HJ.

28.

Prevalence of Yersina
enterocolitica among
patients in Jos and
environs. J. Pharmacy
Bioresources 2004;
1(1): 46-50.

26. Opojabi SO, KandakaiOlukemi YT, Mawak
JD, Olukemi MA,
Bello CSS. Vibrio.
Cholerae of
infections in Jos,