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Anti-Salmonella activity of metabolites from African soldier termites, Macrotermes bellicosus

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Abstract:

Background: The global emergence and rapid dissemination of multidrug resistant *Salmonella* strains necessitate research to find new antimicrobials that will effectively be used against these pathogens. In the present study, anti-*Salmonella* activity of metabolites from African Soldier Termites, *Macrotermes bellicosus* was demonstrated and subsequently compared with a potent antibiotic, ciprofloxacin.

Materials and Methods: N-hexane, ethylacetate, methanol and aqueous extracts of metabolites from the *M. bellicosus* were assayed for anti-*Salmonella* activity using the agar dilution method in the determination of the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC). The inhibitory activities of the extracts were compared to ciprofloxacin (256µg/ml). Also, the bioactive components of the extracts were determined using standard techniques.

Results: At 4000 µg/ml, N-hexane extract inhibited the growth of *Salmonella* Typhi, *S.* Paratyphi A, B and C while ethylacetate extract was able to inhibit *S.* Paratyphi A and C. Methanolic and aqueous extracts at the same concentration were unable to inhibit these strains of *Salmonella*. Furthermore, our findings revealed that the MIC of ethylacetate extract was 2000µg/ml for *S.* Paratyphi A and B, 250µg/ml for *S.* Typhi, and 125µg/ml for *S.* Paratyphi C. Also, the MIC of hexane extract was 4000µg/ml for *S.* Paratyphi B, 2000 µg/ml for *S.* Paratyphi C, 500µg/ml for *S.* Typhi and 250µg/ml for *S.* Paratyphi A respectively. The screening of bioactive components revealed the presence of cardiac glycosides and alkaloids.

Conclusion: Our results provide evidence of anti-Salmonella action of metabolites from African Soldier Termites, *M. bellicosus*. N-hexane and ethylacetate extracts of *M. bellicosus* may be explored as novel antimicrobials for the treatment of typhoid and paratyphoid fevers thereby reducing the pressure exerted on available antibiotics.

Keywords: Salmonella, antimicrobials, insects, extracts

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Activité anti-Salmonella des métabolites de termites soldats africains, Macrotermes bellicosus

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Abstrait:

Contexte: L'émergence et la dissémination rapide de souches de *Salmonella* multirésistantes nécessitent des recherches pour trouver de nouveaux antimicrobiens qui seront utilisés efficacement contre ces agents pathogènes. Dans la présente étude, l'activité anti-*Salmonella* de métabolites de *Macrotermes bellicosus*, African Soldier Termites, a été démontrée et comparée par la suite à un antibiotique puissant, la ciprofloxacine.

Matériels et méthodes: L'activité anti-Salmonella a été dosée avec du N-hexane, de l'acétate d'éthyle, du méthanol et des extraits aqueux de métabolites provenant de *M. bellicosus* en utilisant la méthode de dilution en gélose dans la détermination de la concentration minimale inhibitrice (CMI) et de la concentration bactéricide (MBC). Les activités inhibitrices des extraits ont été comparées à la ciprofloxacine (256µg/ml). En outre, les composants bioactifs des extraits ont été déterminés à l'aide de techniques classiques.

Résultats: À 4000μg/ml, l'extrait de N-hexane inhibe la croissance de *Salmonella* Typhi, *S.* Paratyphi A, B et C alors que l'extrait d'acétate d'éthyle est capable d'inhiber *S.* Paratyphi A et C. Les extraits méthaniques et aqueux à la même concentration ne peuvent inhiber ces souches de *Salmonella*. En outre, nos résultats ont révélé que la CMI de l'extrait d'acétate d'éthyle était de 2000μg/ml pour *S.* Paratyphi A et B, de 250 μg/ml pour *S.* Typhi et de 125μg/ml pour *S.* Paratyphi C. De plus, la CMI de l'extrait d'hexane était de 4000μg/ml pour *S.* Paratyphi B, 2000μg/ml pour *S.* Paratyphi C, 500μg/ml pour *S.* Typhi et 250μg/ml pour *S.* Paratyphi A respectivement. Le dépistage des composants bioactifs a révélé la présence de glucosides et d'alcaloïdes cardiaques.

Conclusion: Nos résultats fournissent des preuves de l'action anti-*Salmonella* des métabolites de termites de soldat africains, *M. bellicosus*. Des extraits d'N-hexane et d'acétate d'éthyle de *M. bellicosus* peuvent être explorés comme nouveaux antimicrobiens pour le traitement des fièvres typhoïde et paratyphoïde, réduisant ainsi la pression exercée sur les antibiotiques disponibles

Mots-clés: Salmonella, antimicrobiens, insectes, extraits

Introduction:

Typhoid and paratyphoid fevers are a severe, life-threatening disease caused primarily by serovars Typhi and Paratyphi A, B, or C of Salmonella enterica subspecies enterica. Typhoid accounts for approximately 90% of enteric fevers in Africa, Asia, and Latin America, mostly regarded as endemic, with morbidities and mortalities (1). It is transmitted by the faecal-oral route via contaminated food and water and is therefore common where sanitary conditions are inadequate and access to clean water is limited (2).

Although, antibiotics have been used for the treatment and control of *Salmonella* infections in human and animals, recent studies show increase resistance to conventional drugs, including fluoroguinolones and third generation

cephalosporins which are the drugs of choice for infections caused by Salmonella other members of Enterobacteriaceae family (3, 4). The emergence of Salmonella strains with multidrug-resistant (MDR) genes has constituted а serious public health problem, resulting in higher treatment cost, longer stay in the hospital, and increase morbidity and mortality especially in developing countries (5). Therefore, an urgent search for novel anti-Salmonella agents with high potency which will be used as alternatives to conventional drugs is imperative.

Insects have been recognized recently to possess highly potent immune defenses that synthesize constitutive and inducible antimicrobial compounds capable of combating a wide spectrum of pathogens (6). Consequently, insects are now the major target as the abundant

potential source of different antimicrobial compounds (7, 8). Although insects make up 90% of the total number of animals on earth, insect-derived antimicrobial peptides (AMPs) only account for approximately 10% of more than 2,830 AMPs listed in the Antimicrobial Peptide Database; thus there are more AMPs with activity just waiting to be discovered (9).

African Soldier Termites, Macrotermes bellicosus are well distributed in Nigeria and other African countries. These termites can survive and in diverse ecological propagate environments due to their ability to develop resistance mechanisms against different diseases (10). This study, therefore, investigated the In vitro anti-Salmonella activity of different extracts from African soldier termite, M. bellicous against strains of Salmonella.

Materials and Methods:

Zoological Method

Termites were sampled form the mound using a hoe in Minna, North-Central Nigeria by methods previously described (11). The samples were put in plastic bags with mound soil and immediately transported to the laboratory. The termites were further picked using forceps and transferred to a bottle containing methanol, and were identified as African soldier termites, *M. bellicous*, by the Entomology section of the Department of Biological Sciences, Federal University of Technology, Minna, Nigeria.

Preparation and Screening of Extracts for Bioactive Components

The head of the termite which is distinct from the body was carefully removed using a sterile scalpel, which was air-dried for two weeks and pounded using clean mortar and pestle. Successive reflux extraction of the powdered termites' heads (80g) was carried out using solvents ranging from non-polar to polar to obtain various soluble portions. The resulting mixtures were filtered using Whatman filter paper No. 1 while the soluble portion was evaporated to dryness

using a steam bath. At the end of the extraction protocol, four extracts of N-hexane, ethylacetate, methanol, and aqueous extracts were obtained and stored in sterile universal bottles at 4°C until further processing. The extracts obtained were screened for the presence of bioactive components as previously described (12). Different bioactive components including saponins, tannins, phlobatannin, cardiac glycosides, alkaloids and reducing sugars were sought for.

Bacterial Strains

Pure clinical strains of Salmonella Typhi and S. Paratyphi A, B and C were collected from the Vaccine Laboratory, Department of Microbiology, Federal University of Technology Minna, Nigeria. The strains were further confirmed by biotyping using the MicrobactTM 12A (12 E) kit.

Anti-Salmonella Activity

The extracts were reconstituted in DMSO and distilled water as 0.4g of the different extract was dissolved in 1ml of DMSO plus 4ml of distilled water. anti-Salmonella activities of all extracts were determined using agar dilution method (13). Under aseptic conditions, 1 ml of each reconstituted extract was dispensed into sterile Petri plates, and 19 ml of sterilized nutrient agar was added to make a final concentration of 2000 µg/ml, after which the plates were swirled for homogeneity. The plates were prepared in duplicates and control plates consisted of organism viability control (OVC), extract sterility control (ESC), medium sterility control DMSO control plate was prepared. The activities of extracts were compared with that of ciprofloxacin (256µg/ml). Ciprofloxacin, which is a fluoroguinolone known to be potent against Salmonella, was obtained from General Hospital, Minna, Nigeria and dissolved in sterile water before use.

Minimum Inhibitory Concentration Determination

The broth dilution method of Cheesbrough (13) was employed for MIC

determination. The hexane and ethylacetate extracts were diluted with nutrient broth to obtain 4000µg/ml, 2000 µg/ml, 500µg/ml, 250µg/ml and 150µg/ml respectively. A loopful of each *Salmonella* strain was added to the diluents and incubated at 37°C for 24hr. The tubes with the least concentration of the extracts that showed no turbidity were recorded as the MIC.

Minimum Bactericidal Concentration Determination

The MBC of the extracts were determined as earlier described (13). Briefly, the MIC tubes were sub-cultured on sterile nutrient agar and incubated at 37°C for 24hr. The plates with the least concentration of the extracts that showed no growth after sub-culturing was recorded as the MBC.

Results:

The anti-Salmonella activity of crude extracts of *M. bellicous* are

presented in Table 1. The N-hexane exracts showed activity against the four isolates, *S.* Typhi and *S.* Paratyphi A, B and C screened while activity was only recorded against *S.* Paratyphi A and C from the Ethylacetate extract respectively. There were no anti-*Salmonella* activities recorded for both methanol and aqueous extracts.

minimum inhibitory concentration (MIC) of N-hexane extract showed activity ranged between 4000 to $250\mu g/ml$ only for S. Typhi and S. Paratyphi A, B and C while that of Ethylacetate extract ranged between 2000 to $125\mu g/ml$ only for S. Typhi and S. Paratyphi A, B and C (Tables 2 and 3). The presence of bioactive components as N-hexane obtained from the and Ethylacetate extracts indicated the presence of cardiac alycosides and alkaloids (Table 4).

Table 1: Anti-Salmonella activity of crude extracts of Macrotermes bellicous

| Test organisms | Crude Extract (4000 μg/ml) | | | |
|----------------|----------------------------|------------------|------------------|------------------|
| | ST ^{SH} | ST ^{SE} | ST SM | ST ^{SW} |
| S. Typhi | + | - | - | - |
| S. Paratyphi A | + | + | - | - |
| S. Paratyphi B | + | - | - | - |
| S. Paratyphi C | + | + | - | - |

^{+ =} activity; - = no activity; ST^{SH} = N-hexane extract; ST^{SE} = Ethylacetate extract; STSM = Methanol extract; ST^{SW} = Aqueous extract

Table 2: Minimum Inhibitory Concentration (MIC) of N-Hexane Extract Crude Extract of $Macrotermes\ bellicous$

| Test organisms | | N-hexane (μg/ml) | | | | | |
|--------------------------|------|------------------|------|-----|-----|-----|--|
| _ | 4000 | 2000 | 1000 | 500 | 250 | 125 | |
| S. Typhi | - | - | - | + | - | - | |
| <i>S.</i> Paratyphi A | - | - | - | - | + | - | |
| <i>S.</i> Paratyphi B | + | - | - | - | - | - | |
| <i>S.</i> Paratyphi C | - | + | - | - | - | - | |

Activity = +; - = No activity

Table 3: Minimum Inhibitory Concentration (MIC) of Ethylacetate Extract Crude Extract of Macrotermes bellicous

| Test organisms | Ethylacetate (μg/ml) | | | | | |
|--------------------------|----------------------|------|------|-----|-----|-----|
| <u> </u> | 4000 | 2000 | 1000 | 500 | 250 | 125 |
| S. Typhi | - | - | - | - | + | - |
| <i>S.</i> Paratyphi A | - | + | - | - | - | - |
| <i>S.</i> Paratyphi B | - | + | - | - | - | - |
| <i>S.</i> Paratyphi C | - | - | - | - | - | + |

+ = activity: - = no activity

Table 4: Bioactive components of the N-hexane and Ethylacetate extracts

| Bioactive components | Extract | |
|----------------------|----------|--------------|
| | N-hexane | Ethylacetate |
| Tannin | - | - |
| Phlobatannin | - | - |
| Reducing sugar | - | - |
| Alkaloids | - | + |
| Steroids | - | - |
| Saponin | - | - |
| Cardiac glycosides | + | + |

+ = Present; - = Absent

Discussion:

The study demonstrates metabolites from the head of African termites have antibacterial activities that are effective against some human pathogens. The pathogens tested were found to show different degree of susceptibility to various extracts obtained from the head of African soldier termites. In this study, N-hexane extract inhibited the growth of typhoid and paratyphoid bacilli while the Ethylacetate extract inhibited S. Paratyphi A and C only. Termites have developed the ability to deal with a rich microbial community inhabiting their nests and feeding sites. One of the ways which they do this is by the synthesis of antimicrobial peptides, among other defense mechanisms (14). These antimicrobial peptides may explain

the reason for the anti-Salmonella activity observed in this study. This claim was supported by the observation of Zeng (15), who reported the antibacterial activity of some peptides, spinigerin and termicin obtained from termites against pathogenic organisms. Furthermore, spinigerin and termicin peptides that are rich in cysteine and contained α-helical properties have high ability to permeate microbial cytoplasmic membranes (16). This was further corroborated by Lee (17), who studied the antimicrobial properties of spinigerin.

Although extracts used in this study were not obtained from plants, it has been suggested that for plants to be used for medicinal purposes, their extracts should possess antimicrobial activity with MIC<1000µg/mL (18). With reference to

their report, results of the MIC obtained from this study indicated that N-hexane extracts possess strong antimicrobial activity for S. Typhi and S. Paratyphi A Ethylacetate extracts possess antimicrobial activity against S. Typhi and S. Paratyphi C since their MICs were less than 1000µg/mL. The evaluation of the MBC to ascertain the bactericidal effect of the N-hexane and Ethylacetate extracts on the Salmonella strains examined at various concentrations did not showed any bactericidal activity (results presented). This further shows that a higher concentration above the MIC is required for bactericidal activity.

Phytochemicals are known complementary and overlapping have mechanisms of action in the body modulation including of hormone metabolism and enzyme detoxification, antioxidant effects, stimulation of the immune system and antibacterial activity (19, 20). The qualitative screening of bioactive components from the head of soldier termites shows African the presence of some plant secondary alkaloids and cardiac metabolites; glycosides. The presence of these components in termites could undoubtedly arise from their host plants. This is because this species of termites is mainly xylophage. They are herbivorous whose diets consist primarily of wood. As such, Phyto-compounds present in the plants consumed may still be in circulation within the insects in relatively high amounts (21, 22). The presence of these bioactive components may also be the reason for the antimicrobial activity recorded in this study.

It was observed that alkaloids and cardiac glycosides also significantly decreased the growth and proliferation of pathogenic Klebsiella pneumoniae and Staphylococcus aureus (23). Similarly, Fernandez-Melendez (24) observed that the alkaloids in fire ant, Solenopsis invicta inhibited the growth of Gram-positive and Gram-negative bacteria acting as a broadspectrum antimicrobial agent. findings are in consonance with the results obtained from this current study.

The extracts of *M. bellicosus* at 4000µg/ml displayed similar bacteriostatic effects on the test organisms when compared with ciprofloxacin (265µg/ml). It is noteworthy to further state that such results as obtained from this study can be attributed to the crude state of the extract used. Furthermore, upon purification and quantification of the extracts used in this study, their activities may be more potent and biocidal even at lower concentrations.

Conclusion:

The findings of the current study show that metabolites from the head of solider termites can inhibit the growth and survival of some pathogenic bacteria. However, further studies are needed to standardize potential use of *M. bellicosus* as complementary agent for eliminating pathogenic bacteria. Also, synergistic effect of the use of metabolites of *M. bellicosus* and conventional antibiotics can also be examined.

References:

- Laxminarayan, R., Duse, A., Watta, C., et al. Antibiotic resistance-the need for global solutions. Lancet Infect Dis. 2013; 13: 1057–1098
- Donald, S. T., Donatien, G., Siméon, P. C. F., Charles, F., Fabrice, K., and Merline, N. D. *In vivo* anti-salmonella activity of aqueous extract of *Euphorbia prostrata* Aiton (Euphorbiaceae) and its toxicological evaluation. Asian Pac J Trop Biomed. 2015; 5 (4): 310-318
- Aggarwal, A., Vij, A. S., and Oberoi, A. A three-year retrospective study on the prevalence, drug susceptibility pattern, and phage types of Salmonella enterica subspecies Typhi and Paratyphi in Christian Medical College and Hospital, Ludhiana, Punjab. JIACM. 2007; 8: 32-35.
- Medalla, F., Sjölund-Karlsson, M., Shin, S., Harvey, E., Joyce, K., and Theobald, L. Ciprofloxacin-resistant Salmonella enterica serotype typhi, United States, 1999-2008. Emerg Infect Dis. 2011; 17 (6): 1095-1098.
- Tsobou, R., Mapongmetsem, P. M., Voukeng, K. I., and Vandamme, P. Phytochemical screening and antibacterial activity of medicinal plants used to treat typhoid fever in Bamboutos division, West Cameroon. J Appl Pharma Sci. 2015; 5 (6): 34-49
- 6. Haine, E. R., Moret, Y., Siva-Jothy, M. T., and Rolff, J. Antimicrobial defense and

- persistent infection in insects. Science. 2008; 322 (5905): 1257–1259. doi: 10.1126/science.1165265 PMID: 19023083.
- 7. Slocinska, M., Marciniak, P., and Rosinski, G. Insects antiviral and anticancer peptides: New leads for the future? Protein Peptide Lett. 2008; 15 (6): 578–585.
- 8. Dossey, A. T. Insects and their chemical weaponry: New potential for drug discovery. Nat Prod Rep. 2010; 27 (12):1737–1757. doi: 10.1039/c005319h PMID: 20957283
- 9. Yi, H. Y., Chowdhury, M., Huang, Y. D., and Yu, X. Q. Insect antimicrobial peptides and their applications. Appl Microbiol Biotechnol. 2014; 98: 5807–5822.
- 10. Chovenc, T., Efstathion, C. A., Elliott, M. L., and Su, N. Y. Extended disease resistance emerging from the faecal nest of a subterranean termite. Proc Royal Society Biol. 2013; 18: doi: 10.1098/rspb.2013.1885
- 11. Imhasly, P., and Leuthold, R. H. Intraspecific colony recognition in the termites *Macrotermes subhyalinus* and *Macrotermes bellicosus* (Isoptera: Termitidae). Insects Sociaux. 1999; 46 (2): 35-38.
- 12. Trease, G. E., and Evans, W. C. Pharmacognosy. 15th edition London; Saunders Publishers 2002; 24-27.
- 13. Cheesbrough, M. District Laboratory Practice in Tropical Countries. Part 2, 2nd Edition, Cambridge University Press Publication, South Africa, 2006; 1-434.
- Schmid-Hempel, P. Evolutionary ecology of insect immune defenses. Ann Rev Entomol. 2005; 50: 529–551.
- 15. Zeng, Y., Hu, X. P., and Suh, S. J. Multiple antibacterial activities of proteinaeceous compounds in crude extracts from the eastern Subterranean termite, Reticulitermes flavipes Kollar (Blattodea: Isoptera:Rhinotermitidae). Adv Res. 2014; 2 (8): 455-461.
- 16. Dimarcq, J. L., Bulet, P., Hetru, C., and Hoffmann, J. Cysteine-rich antimicrobial peptides in invertebrates. Peptide Sci. 1998; 47: 465–477

- Lee, K. H., Shin, S. Y., Hong, J. E., Yang, S. T., Kim, J. I, Hahm, K. S., and Kim, Y. Solution structure of termite-derived antimicrobial peptide, spinigerin, as determined in SDS micelle by NMR spectroscopy. Biochem Biophy Res. 2003; 309: 591–597
- 18. Rı'os, J. L., and Recio, M. C. Medicinal plants and antimicrobial activity. J Ethnopharmacol. 2005; 100, 80–84.
- 19. Tula, M. Y., Azih, A. V., Iruolaje, F. O., Okojie, R. O., Elimian, K. O., and Toy, B. D. Systematic study on comparing phytochemicals and the antimicrobial activities from different parts of V. amygdalina. Afr J Microbiol Res. 2012; 6 7089 (43): 7093. 10.5897/AJMR12.1474
- 20. Farombi, E. O. African Indigenous plants with chemotherapeutic potentials and biotechnological approach to the production of bioactive prophylactic agents. Afr J Biotechnol. 2003; 2 (12): 662-671.
- 21. Noirot, C., and Darlington, J. Termite nests: architecture, regulation and defense. In: Abe, T., Higashi, M., and Bignell, D. E. (eds.). Termites: Evolution, Sociality, Symbiosis, Ecology. Kluwer Academic Publications, Dordrecht, 2002.
- 22. Thorne, B. L., and Haverty, M. I. Nest Growth and Survivorship in Three Species of Neotropical Nasutitermes (Isoptera: Termitidae). J Environ Ecol. 2000; 29, 256–264.
- 23. Ibraheem, O., and Maimako, R. F. Evaluation of alkaloids and cardiac glycosides contents of *Ricinus communis* Linn. (Castor) whole plant parts and determination of their biological properties. Int J Toxicol Pharmacol Res. 2014; 6 (3): 34-42.
- 24. Fernandez-Melendez, S., Miranda, A., and Garcia-Gonzalez, J. Anaphylaxis caused by imported red fire ant sting in Malaga, Spain. J Invest Allergol Clin Immunol. 2007; 17: 48-49.