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ANTIFUNGAL PROPERTIES OF METHANOLIC EXTRACTS OF SOME MEDICAL PLANTS IN ENUGU, SOUTH EAST NIGERIA

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ABSTRACT

Objective: The current study evaluated the anti-fungal activities of methanolic extracts of the leaves and fruits of some medicinal plants of health value in Enugu, southeast Nigeria.

Materials and methods: Volumes of the different extracts into molten Sabouraud Dextrose Agar to make up for concentrations of 25 mg/ml, 50 mg/ml and 100 mg/ml respectively. Concentrations of 125 µg/ml, 250 µg/ml and 500 µg/ml of Miconazole were incorporated to serve as a positive control while 0.5 ml of DMSO served as a negative control. The plants extracts employed were from *Azadirachta indica* (Neem), *Anacardium occidentale* (Cashew), *Euphorbia hirta* (Asthma weed), *Jatropha curcas* (Physic nut), *Acantus montanus* (Mountain thistle) and *Picalima nitida* (ósúgwe in Igbo). A 2 mm agar disc cut out from the margin of actively growing cultures of each dermatophyte (*Trichophyton soudanense*, *T.mentagrophytes*, *Cladosporium* sp, *T.rubrum* and *Fusarium* sp) were inoculated on the agar plates containing varying concentrations of the different plant extracts, the standard antifungal agent and negative control in triplicates and incubated at 28 °C.

Results: All the extracts exhibited antifungal activities of varying degrees with radial growth inhibitions (RGIs) ranging from 0-0.45 mm radius. Fifty percent (50 %) of the plants extracts comprising of *Picalima nitida*, *Euphorbia hirta* and *Acantus montanus* exhibited complete inhibition at 100 mg/ml against all the clinical isolates under investigation (p < 0.05). *Picalima nitida* seed was the only plant extract with complete inhibition at 25 mg/ml against *T.soudanense* and *T.rubrum*. *Picalima nitida* was the closest in activity to miconazole. *Azadirachta indica* (Neem) did not exhibit complete inhibition against any of the clinical isolates at 100 mg/ml yet exhibited a significantly lower RGI at the lowest concentration of 25 mg/ml better than others.

Conclusion: Antifungal activities showed that leaves extract of fruits plants against the clinical isolates are good source of medicinal applications

Key words: Antifungal agents, medical plants, Enugu, Nigeria.

PROPRIÉTÉS ANTIFONGIQUES D'EXTRAITS MÉTHANOLIQUES DE CERTAINES PLANTES MÉDICALES À ENUGU, AU SUD-EST DE NIGERIA

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RÉSUMÉ

Objectif : La présente étude a évalué les activités anti-fongiques des extraits méthanoliques des feuilles et fruits de certaines plantes médicinales de valeur santé à Enugu, au sud-est du Nigeria. **Matériel et méthodes :** Volumes des différents extraits dans molten Sabouraud Dextrose Agar pour compenser pour les concentrations de 25 mg/ml, 50 mg/ml et 100 mg/ml respectivement. Les concentrations de 125 µg/ml, 250 µg/ml et 500 µg/ml de miconazole ont été intégrés pour servir comme contrôle positif alors que 0,5 ml de DMSO a servi de témoin négatif. Les extraits de plantes utilisés étaient d'*Azadirachta indica* (neem), noix de cajou (*Anacardium occidentale*), *Euphorbia hirta* (asthme), *Jatropha curcas* (pourghère), *Acantus montanus* (chardon de montagne) et de *Picalima nitida* (ósúgwe en Igbo).

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Un disque de 2 mm d'agar coupé à partir de la marge de croissance active, les cultures de chaque (dermatophytes *Trichophyton soudanense*, *T. mentagrophytes*, *Cladosporium* sp, *T. rubrum* et *Fusarium* sp) ont été inoculés sur les géloses contenant des concentrations variables des différents extraits de plantes, la norme agent antifongique et contrôle négatif en triplicats et incubées à 28 °C. Résultats : Tous les extraits de l'exposé des activités antifongiques avec divers degrés d'inhibitions de la croissance radiale (RGIs) allant de 0-0,45 mm de rayon. Cinquante pour cent (50 %) des extraits de plantes comprenant de *Picalima nitida*, *Euphorbia hirta* et *Acantus montanus* présentaient une inhibition complète à 100 mg/ml contre tous les isolats cliniques soumis à l'enquête ($p < 0,05$). *Picalima nitida* semence a été le seul extrait de plantes avec une inhibition complète à 25 mg/ml contre *T. soudanense* et *T. rubrum*. *Picalima nitida* était la plus proche de l'activité pour le miconazole. *Azadirachta indica* (neem) n'ont pas d'inhibition complète à l'égard d'aucun des isolats cliniques à 100 mg/ml mais montrent un souci nettement inférieur à la plus faible concentration de 25 mg/ml mieux que d'autres. Conclusion: activités antifongique a montré que l'extrait de feuilles de fruits plantes contre les isolats cliniques sont une bonne source d'applications médicinales

Mots clés: antifongiques, plantes médicinales, Enugu, Nigéria.

INTRODUCTION

The increasing incidence of growing resistance to antifungal agents despite the intensive use of antifungal drugs in the treatment of fungal infection [1], has become a great health challenge. However, there has been some claims by the traditional healers that some medicinal plants are more efficient in the treatment of infectious diseases than synthetic antibiotics. Medicinal plants have been practiced for centuries as remedies for human diseases as they incorporate elements of therapeutic value. At that place are numerous plant natural products which have antifungal, antibacterial and antiprotozoal activities that could be used either systemically or locally [2]. Medicinal plants are considered to be an important source of new chemical substances with possible therapeutic effects [3]. The World Health Organization (WHO) estimates that up to 85% of people still rely primarily on traditional remedies such as herbaceous plants for their medicine [4]. Traditional healing plays an integral role in black African culture as it provides health care needs for a large majority of the society [5]. Presently, there is growing awareness of scientific and medicinal plants in the health care system of many developing countries [6]. In Nigeria, there is a rich tradition in the use of herbal plant products for the treatment of several ailments. Plants such as *Euphorbia hirta* (*Asthma weed*), *Anacardium occidentale* (*Cashew*), *Picalima nitida* (*Ösúigwe*), *Jatropha curcas* (*Physic nut*), *Azadirachta indica* (*Neem*) and *Acantus montanus* (*Mountain thistle*) have been used by the people within the Enugu metropolis for the local treatment of skin diseases because of their effectiveness, availability and cost effectiveness.

Furthermore, nature has bestowed a very rich botanical wealth of vegetation, and a large number of diverse types of plants grow in different parts of the country. They constitute the richest source of drugs for traditional systems of medicine, modern medicines, food supplements [7]. The increasing prevalence of serious mycoses coupled with the frequent use of the available antifungal drugs has resulted in rising resistance of fungal pathogens to

antifungal agents. However, these topical drugs are generally ineffective against fungal infections of the nails due to their inability to penetrate the entire nail unit and eradicate the infection. The increasing prevalence of serious mycoses coupled with the frequent use of antifungal drugs currently available has resulted in rising resistance of fungal pathogens to antifungal agents. There is, therefore, a need for new broad-spectrum antifungal agents that can be used empirically in immune compromised patients, organ transplant patient, and other challenging situations. Use of herbal medicine in the treatment of infection with microorganisms predates the introduction of antibiotics [8]. Herbs are widely exploited in the traditional medicine and their curative potentials are well documented too, as well as representing a rich source of antimicrobial agents [9], with many of them readily available in rural areas at a relatively cheaper price. Medicinal plants extracts are promising as alternative or complementary control means because of their antimicrobial activity, non-phytotoxicity, as well as biodegradability properties [10].

Considering the vast potential of plants materials in the health care and the challenges in the management of fungal infections, this current study was designed to evaluate the antifungal properties of methanolic extracts of some traditional medicinal plants in Enugu, South East, Nigeria and their phytochemistry properties in order to reformulate the existing antifungals that are essential for improving patient management.

MATERIALS AND METHODS

Preparation of Plant Materials

Fresh leaves of the plants, *Azadirachta indica* (Neem), *Anacardium occidentale* (cashew), *Euphorbia hirta* (Asthma weed), *Jatropha curcas* (Physic nut), *Acantus montanus* (Mountain thistle) and the *Picalima nitida* (Ösúigwe) Seeds & Rind were collected within the Enugu metropolis and authenticated in the Botany Department of the University of Nigeria, Nsukka. They were washed

under running tap water and air dried in room for 5 days (for the leaves) and 12 days (for the seeds and rind). The materials were ground into fine powder and stored in labelled air tight containers.

Preparation of Crude Extracts

100 g of each of the powdered form of the plant materials was exhaustively extracted by Soxhlet extraction method [11], using absolute methanol at 30 °C.

Preparation of fungal isolates: Microscopically identified isolates of *Trichophyton soudanense*, *T. mentagrophytes*, *T. rubrum*, *Cladosporium* sp and *Fusarium* sp, were obtained from the Mycology Laboratory of the University of Nigeria Teaching Hospital, Ituku-Ozalla, Enugu. The isolates were subcultured in Sabouraud Dextrose Agar (SDA) to get pure cultures. Pure cultures were prepared in slanted cultures, stored in McCartney bottles and kept at 2-8 °C for further experimental purposes.

Determination of Antifungal Activities

Reconstitution of plant extracts: A stock concentration was reconstituted by weighing 2 g of the individual plant residue by adding 1 ml of DMSO to it in a test tube to make a concentration of 2 g/ml (2000 mg/ml).

Preparation of molten SDA plates containing the extract: The agar plate method was adopted as described [12].

Culture Process: The Culture medium of the different concentrations was inoculated with 2 mm agar disc cut out from the margin of the actively growing culture of the dermatophyte. This was carried out in triplicates and incubated at 28 °C, with controls run concurrently. The radial growth was measured daily for 4 days and the average reading taken for the test, positive and negative controls.

Percentage radial growth inhibition was calculated by employing the following formula:

Percent inhibition = $C - T \div C \times 100$ [13]. Where, C = Radial growth of negative control; T = Radial growth of the test

Column Chromatography

This was done using n-hexane, chloroform and ethyl acetate based on increasing polarity [14].

Phytochemical Analysis

The phytochemical analysis of fractionated *Picralima nitida* seed extract was carried out using standard methods [15].

Other test analysis for alkaloids, flavonoids, steroids and terpenoids, saponins, tannins carbohydrate, glycosides, resins (Precipitation test), proteins, fats and oil were carried using standard methods.

Data analysis: All generated data were subjected to statistical analysis using a one-way analysis of variance (ANOVA), followed by Dunnett's test (multiple comparison post-test) at $p < 0.05$.

RESULTS

Antifungal activities of the methanolic extracts of six medicinal plants were determined against five clinical isolates of fungi species. Miconazole nitrate was used as the standard for comparing the plant's extract. In Table 1, Miconazole nitrate at 500 µg/ml had the highest antifungal effect on the growth of *Cladosporium* sp by inhibiting it completely with a percentage radial growth inhibition (PRGI) of 100 %. *Picralima nitida* seed extract exhibited the highest antifungal effect with all the three concentrations (Tables 2 and 3).

TABLE 1: PERCENTAGE RADIAL GROWTH INHIBITION OF POSITIVE CONTROL ANTIFUNGAL AGENT (MICONAZOLE) AGAINST FUNGAL ISOLATES

Fungal isolates	Concentration of Miconazole nitrate		
	125 µg/ml	250 µg/ml	500 µg/ml
<i>T.soudanense</i>	89.2 %	90 %	93.8 %
<i>T.mentagrophytes</i>	75 %	78 %	85 %
<i>Cladosporium</i> sp	86.7 %	95 %	100 %
<i>T.rubrum</i>	94 %	95 %	98 %
<i>Fusarium</i> sp	82.9 %	75.3 %	95.3 %

TABLE 2: PERCENTAGE RADIAL GROWTH INHIBITION OF *PICRALIMA NITIDA* SEED EXTRACT AGAINST FUNGAL ISOLATES

Fungal Isolates	Concentrations of extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial Growth inhibition		
<i>T.soudanense</i>	100 %	100 %	100 %
<i>T.mentagrophytes</i>	90 %	100 %	100 %
<i>Cladosporium</i> sp	91.7 %	95 %	100 %
<i>T.rubrum</i>	100 %	100 %	100 %
<i>Fusarium</i> sp	94.1 %	100 %	100 %

TABLE 3: PERCENTAGE RADIAL GROWTH INHIBITION OF *PICRALIMA NITIDA* SEED FRACTIONS AGAINST FUNGAL ISOLATES

Fungal Isolates	Hexane fraction (100 mg/ml)	Chloroform fraction (100 mg/ml)	Ethylether fraction (100 mg/ml)
	Radial growth	Inhibition	
<i>T.soudanense</i>	55 %	77.1 %	26.7 %
<i>T.mentagrophytes</i>	-33.3 %	25 %	-66.7 %
<i>Cladosporium sp</i>	25 %	50 %	-50 %
<i>T.rubrum</i>	35.7 %	88.6 %	57.1 %
<i>Fusarium sp</i>	-14.3 %	83.3 %	0%

At 100 mg/ml the extract inhibited completely all the investigated isolates with PRGI of 100 %. At 50 mg/ml, there was complete inhibition against all isolates except against *Cladosporium spp.* and at 25 mg/ml it inhibited completely *T.soudanense* and *T.rubrum*. The inhibitions exhibited by *T.mentagrophytes*, *Cladosporium spp.* and *Fusarium spp.* were, however, significant. *T.soudanense* and *T.rubrum* appeared to be the most sensitive of the fungi under investigation. This was followed by *T.mentagrophytes* and *Fusarium spp.* with a PRGI of 90 - 100 %. *Cladosporium spp.* with a PRGI of 91.7 -100 % appeared to be the least sensitive. The inhibition range of *Picalima nitida* seed extract for all the isolates was 90 -100 %. There was no inhibition observed in the negative control. The PRGIs for the antifungal activities of *Picalima nitida* seed extract at 25 mg/ml, 50 mg/ml and 100 mg/ml, respectively were significant ($P < 0.05$) compared with the standard.

The phytochemistry of the chloroform fraction of *Picalima nitida* seed revealed the presence of flavonoids, alkaloids, and terpenoids (Table 3). The highest antifungal effect was exhibited by Chloroform fraction with a PRGI of 88.6 % against *T.rubrum*. All the fungi under investigation appeared sensitive to the chloroform extract. Ethyl ether fraction had a lesser antifungal effect on the experimental isolates. There was a significant difference of ($p < 0.05$) in the treatment of chloroform fraction against control.

From Table 4, *Picalima nitida* rind extract had its highest antifungal effect at 100 mg/ml by inhibiting completely all the investigated isolates with a PRGI of 100 %. RGs of 0.03 ± 0.03 mm to 0.1 ± 0.05 mm at 25 mg/ml and 0.03 ± 0.03 mm to 0.05 ± 0.05 mm at 50 mg/ml were observed for all the isolates. The activities of *Picalima nitida* rind at these concentrations show a significant difference ($p < 0.05$) compared to the activities of the standard antifungal agent. Table 5 shows that *Azadirachta*

indica (Neem) seed extract did not inhibit any of the fungi completely at its highest concentration of 100 mg/ml. The inhibition range of Neem seed extract for all the isolates was 54.2-95.4 %. Neem seed was statistically significant ($p < 0.05$) only at 100mg/ml when compared with Miconazole nitrate.

TABLE 4: PERCENTAGE RADIAL GROWTH INHIBITION OF *PICRALIMA NITIDA* RIND EXTRACT AGAINST FUNGAL ISOLATES

Fungal isolates	Concentration of Extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial growth inhibition		
<i>T.soudanense</i>	92.3 %	95.4 %	100 %
<i>T.mentagrophytes</i>	80 %	94 %	100 %
<i>Cladosporium sp</i>	86.7 %	91.7 %	100 %
<i>T.rubrum</i>	94 %	94 %	100 %
<i>Fusarium sp</i>	90.6 %	94.1 %	100 %

Anacardium occidentale (cashew) leaf extract as shown in Table 5 had its highest antifungal effect of 100 % RGI at 100 mg/ml. All investigated isolates were completely inhibited at 100 mg/ml by the extract except for *Cladosporium sp* with a PRGI of 91.7 %. *T.rubrum* had a PRGI of 88-100 % and appeared to be the most sensitive followed by *T.soudanense* with a PRGI of 76.9-100 %. The percentage inhibition range of *A.occidentale* extract for all the isolates was between 23.5-100 % giving a significant difference ($p < 0.05$) on all the concentrations with respect to the standard control (Table 6),

TABLE 5: PERCENTAGE RADIAL GROWTH INHIBITION OF *AZADIRACHTA INDICA* SEED EXTRACT AGAINST FUNGAL ISOLATES

Fungal isolates	Concentration of Extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial growth inhibition		
<i>T.soudanense</i>	86.2 %	95.4 %	89.2 %
<i>T.mentagrophytes</i>	64 %	94 %	94 %
<i>Cladosporium sp</i>	54.2 %	66.7 %	91.7 %
<i>T.rubrum</i>	94 %	94 %	95 %
<i>Fusarium sp</i>	88.2 %	79.4 %	89.4 %

In table 7, *Euphorbia hirta* exhibited its highest antifungal effect at 100 mg/ml by completely inhibiting all the fungal isolates. No inhibition was observed in most of the isolates under investigation at 25 mg/ml. *T.rubrum* appeared to be the most sensitive fungi with a PRGI of 60 % at 50 mg/ml and 100 % at 100 mg/ml. However, the concentrations of 25 mg/ml, 50 mg/ml and 100 mg/ml had a significant difference of ($p < 0.05$)

when compared with the Standard. In Table 8, *Jatropha curcas* exhibited its highest antifungal effect of 100 % at 100 mg/ml by inhibiting all tested isolates except for *Fusarium* sp. Mild inhibition of 58.3 % was observed at 25 mg/ml and 50 mg/ml for *Cladosporium* sp. Others appeared to be resistant at these concentrations.

TABLE 6: PERCENTAGE RADIAL GROWTH INHIBITION OF ANACARDIUM OCCIDENTALE LEAF (CASHEW) SEED EXTRACT AGAINST FUNGAL ISOLATES

Fungal isolates	Concentration of Extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial growth inhibition		
<i>T.soudanense</i>	76.9 %	84.6 %	100 %
<i>T.mentagrophytes</i>	68 %	82 %	100 %
<i>Cladosporium</i> sp	43.3 %	80 %	91.7 %
<i>T.rubrum</i>	88 %	92 %	100 %
<i>Fusarium</i> sp	23.5 %	52.9 %	100 %

TABLE 7: PERCENTAGE RADIAL GROWTH INHIBITION OF EUPHORBIA HIRTA LEAF EXTRACT AGAINST FUNGAL ISOLATES

Fungal isolates	Concentration of Extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial growth inhibition		
<i>T.soudanense</i>	30.8 %	46.2 %	100 %
<i>T.mentagrophytes</i>	0 %	40 %	100 %
<i>Cladosporium</i> sp	0 %	33.3 %	100 %
<i>T.rubrum</i>	0 %	60 %	100 %
<i>Fusarium</i> sp	-5.9 %	35.3 %	100 %

TABLE 8: PERCENTAGE RADIAL GROWTH INHIBITION OF JATROPHA CURCAS LEAF EXTRACT AGAINST FUNGAL ISOLATES

Fungal isolates	Concentration of Extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial Growth Inhibition		
<i>T.soudanense</i>	38.5 %	53.8 %	100 %
<i>T.mentagrophytes</i>	10 %	20 %	100 %
<i>Cladosporium</i> sp	58.3 %	58.3 %	100 %
<i>T.rubrum</i>	30 %	40 %	100 %
<i>Fusarium</i> sp	29.4 %	47.1 %	97.1 %

TABLE 9: PERCENTAGE RADIAL GROWTH INHIBITION OF ACANTUS MONTANUS LEAVES EXTRACT AGAINST FUNGAL ISOLATES

Fungal Isolates	Concentration of extracts		
	25 mg/ml	50 mg/ml	100 mg/ml
	Radial Growth inhibition		
<i>T.soudanense</i>	84.6 %	92.3 %	100 %
<i>T.mentagrophytes</i>	52 %	64 %	100 %
<i>Cladosporium</i> sp	50 %	66.7 %	100 %
<i>T.rubrum</i>	-40 %	0 %	100 %
<i>Fusarium</i> sp	41.2 %	52.9 %	100 %

Cladosporium sp appeared to be the most sensitive fungi with a PRGI of 58.3-100 %. The extract was significantly higher ($p < 0.05$) at the three concentrations with respect to the standard. In Table 9 shows that *Acanthus montanus* at 100 mg/ml exhibited its highest antifungal effect by completely inhibiting all the tested isolates at a 100 % RGI. *T.rubrum* and *Fusarium* sp were not inhibited at 25 mg/ml and 50 mg/ml of extract. *T.soudanense* appeared to be the most sensitive with a PRGI of 84.6-100 % followed by *T.mentagrophytes* with PRGI of 52-100 %. However, there was a significant difference ($p < 0.05$) in activities of the plant extract to the standard. The phytochemical studies of these plants revealed the presence of secondary metabolites such as tannins, terpenoids, alkaloids, flavonoids, phenols, steroids, glycosides and volatile oils (Table 10).

DISCUSSION AND CONCLUSION

The methanolic extracts of six different plant species were assayed for antifungal activities and compared with Miconazole nitrate using the agar plate method. Miconazole belongs to the azole group of antifungal agents used clinically against fungal infections. They owe their antifungal activity by inhibiting the enzyme lanosterol 1,4- α -demethylase; the enzyme necessary to convert lanosterol to ergosterol, which is the predominant sterol in fungal cell membranes responsible for maintaining cell integrity, viability, function and normal growth [16]. Results from this study showed that the methanolic extract of *Picralima nitida* (seed and rind), *Azadirachta indica* (Neem), *Anacardium occidentale* (Cashew), *Euphorbia hirta*, *Acanthus montanus* and *Jatropha curcas* exhibited antifungal activity against *T. soudanense*, *T.mentagrophyte*, *T.*

rubrum, *Cladosporium* sp and *Fusarium* sp. These plants were considered to possess biochemically related substances with similar inhibitory properties with miconazole with the antifungal activities varying differently at the same concentrations of each of the extracts against the respective fungal isolate. It generally increases with increase in concentration of the extracts. This finding agrees with other study [17], which showed higher concentrations of antimicrobial substances led to appreciable growth inhibition.

TABLE 10: PHYTOCHEMICAL CONSTITUENTS OF CHLOROFORM EXTRACT

S/No	Phytoconstituent	CE
1	Alkaloids	+
2	Carbohydrates	-
3	Flavonoids	+
4	Glycosides	-
5	Fats and oil	-
6	Tannins	-
7	Saponins	-
8	Resins	-
9	Proteins	-
10	Terpenoids	+
11	Steroids	-

Key: + = Present, - = Absent, CE = Chloroform extract

The phytochemical studies of these plants also reveal the presence of secondary metabolites such as tannins, terpernoids, alkaloids, flavonoids, phenols, steroids, glycosides and volatile oils (Table 10). The antifungal activities of these plants may probably be due to the presence of these rich secondary metabolites in plants which is tandem with other studies [18, 19, 20]. The patterns of radial growth inhibition of these plant extracts in this study were similar to those of miconazole nitrate. This could suggest the presence of similarly active ingredients present in Miconazole nitrate (the control antifungal agent) which is used predominantly as a drug of choice against superficial fungal infection especially *Trichophyton* spp, *Epidermophyton* spp and *Micosporium* spp. The activities of *Picalima nitida* seed extract appeared to be better than the other plant extracts investigated. This could be based on the concentration of active antifungal ingredients such as the phenolics (Tannins and flavonoids) present in that part of the plant material. The activities of *Picalima nitida* (seed

and rind) in this study justifies its use in the treatment of skin conditions including of *Tinea corporis* (ringworm of the skin), *Tinea capitis* (ringworm of the head) as reported by other works [21, 22]. *Azadirachta indica* (Neem) seed oil has been used in the treatment of various skin infections by alternative system of medicine [23]. Its activities in this study are in agreement with the work done [24], where neem oil was shown to have different inhibitory effects on different fungi including *T. rubrum*, *T. mentagrophytes* and *Fusarium* sp. However, though there was no complete inhibition observed in this study with any of the fungi, which could be due to neem oil from different localities may possess different rates of the antifungal activities with significant effect on all the fungal species tested. This variation could be due to the difference in the quality of the active ingredients in the oil sample. The leaves of *Anacardium occidentale* (cashew) in this study exhibited poor inhibitions at lower concentrations, though Rajesh *et al* [25] reported that, the nuts have proven antifungal properties of more than 94 % inhibition against *Aspergillus fumigatus*, *A niger*, *Curvularia* sp and *Fusarium* sp. This is an indication of uneven distribution of the active ingredients of the plant. *Euphorbia hirta* exhibited resistance at lower concentrations on some of the isolates, but complete inhibition at a higher concentration. This finding agrees with the other findings [9], as sensitivity increases with increase in concentration. *Jatropha curcas* in this study showed a poor inhibition of 10 % and 30 % at 25 mg/ml for *T. mentagrophytes* and *T.rubrum* respectively. However, Adejumo *et al.* 2009 who worked on *Fusarium* sp observed it to be the most resistant. The difference in their susceptibilities could be attributed to an inherent resistance factor of the test organisms among other factors, though other studies [26, 27] observed that, the leaves of *Jatropha curcas* had no inhibitory effect against *T. mentagrophytes* rather it was observed in the seed extract. This discrepancy is likely due to the fact that no two plants of the same species may have experienced the same environmental challenges considering the fact that plants phytoanticipins are uniformly distributed within the plants while phytoalexins are restricted to the tissue colonized by the fungus and the cells surrounding the infection site [28]. Some antifungal compounds may be present constitutively in one part of a plant, but induced as phytoalexins in other organs. *Acanthus montanus* had not really been widely used in folk medicine for the treatment of skin diseases, rather for pain, inflammation and other ailments [29, 30]. Its antifungal activities in this study support its usage in folklore treatment. *T.rubrum* and *T.soudanense* appear to be the most sensitive fungi at lower concentrations of the extracts. This could be due to the difference in the concentrations of the bioactive components in the sample or synergistic reactions of the various phytochemicals in the extract.

Euphorbia hirta (Asthma weed), *Anacardium occidentale* L (Cashew), *Picralima nitida* (Akuamma plant), *Jatropha curcas* (Barbados nut), *Azadirachta indica* A (Neem plant), and *Acanthus montanus* (Mountain thistle) have antifungal activities. Their antifungal activities increase with increase in concentrations. Crude extract of *Picralima nitida* seed possesses the highest antifungal activity. The phytochemistry of its chloroform fraction reveals

the presence of flavonoids, alkaloids and terpenoids. *Trichophyton rubrum* was the most sensitive fungal under investigation. Further investigation of the purified components of the seed extracts of *Picralima nitida* to determine the metabolites responsible for their activities will make it serve as a good base for consideration in the pharmaceutical industries for the production and packaging of antifungal products.

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