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Research Article

## Anti-inflammatory and Antipyretic Activities of *Panicum Maximum*

\***Jude E. Okokon<sup>1</sup>, Anwanga E. Udoh<sup>1</sup>, Samuel G. Frank<sup>2</sup>, Nsikak M. Udo<sup>1</sup>**

<sup>1</sup>Department of Pharmacology and Toxicology, Faculty of Pharmacy, University Of Uyo, Uyo, Nigeria.

<sup>2</sup>Department of Science Technology, Akwa Ibom State Polytechnic, Ikot Osurua, Akwa Ibom State.

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**ABSTRACT:** Anti-inflammatory and antipyretic activities of leaf extract of *Panicum maximum* were evaluated to ascertain the folkloric claim of its use in fevers and inflammatory conditions. The crude leaf extract (48 – 144 mg/kg) of *Panicum maximum* was investigated for anti-inflammatory and antipyretic activities using various experimental models. The extract caused a significant ( $p < 0.05 - 0.001$ ) dose-dependent reduction in inflammation and fever induced by different agents used. The anti-inflammatory and antipyretic effects of this plant may in part be mediated through the secondary metabolites present in the plant.

**Keywords:** *Panicum maximum*, Anti-inflammatory, antipyretic

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### INTRODUCTION

*Panicum maximum*. Jacq ( Poacace) is a perennial, tuft grass with a short, creeping rhizome. The stem of this robust grass can reach a height of up to 2m; the leaf sheath are found at the bases of the stems and are covered in fine hairs. It is widely distributed in Africa where it originates and almost all tropical parts of the world (Van Oudtshoorn, 1999). The plant (leaf) is used traditionally by the Ibibios of Akwa Ibom State, Nigeria in the treatment of various ailments such as malaria, infections, rheumatism pain, inflammation and diabetes. Antia *et al.*, (2010) reported the antidiabetic activity of the leaf extract. Antiplasmodial and analgesic activities of the leaf extract have also been reported (Andrew, 2011). Ethnopharmacological and scientific reports on this plant is scarce. To confirm its

ethnobotanical uses in the treatment of these ailments, we investigated the anti-inflammatory and antipyretic properties of an ethanolic leaf extract of *Panicum maximum* in experimentally-induced inflammation and fever in rodents.

### MATERIALS AND METHODS

#### *Plant materials*

Fresh leaves of *Panicum maximum* were collected in July, 2010 at a farmland in University of Uyo, Uyo, Nigeria. The plant was identified and authenticated by Dr. Margaret Bassey, a taxonomist in the Department of Botany, University of Uyo, Uyo, Nigeria. Herbarium specimen was deposited at Faculty of Pharmacy Herbarium (FPH 76c). The fresh leaves of the plant were shade-dried for 2 weeks and reduced to powder. The powdered leaves (1kg) was macerated in 97% ethanol (3L) for 72 hours to give the crude ethanolic extract. The liquid filtrate was concentrated and evaporated to dryness in vacuo 40°C using rotary evaporator. The dry extract was stored in a refrigerator at -4°C until used for experiments reported in this study.

#### *Phytochemical Screening*

Phytochemical screening of the crude leaf extract was carried out employing standard procedures and tests (Trease and Evans, 1989, Sofowora, 1993), to reveal

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\*Address for correspondence: [judeefiom@yahoo.com](mailto:judeefiom@yahoo.com)

Tel. +234802-345-3678

the presence of chemical constituents such as alkaloids, flavonoids, tannins, terpenes, saponins, anthraquinones, reducing sugars, cardiac glycosides among others.

#### *Animals*

Albino rats (120-155g) and mice (18 – 23g) of either sex were obtained from the University of Uyo animal house. They were maintained on standard animal pellets and water *ad libitum*. Permission and approval for animal studies were obtained from the College of Health Sciences Animal Ethics committee, University of Uyo.

#### *Determination of Median Lethal dose (LD<sub>50</sub>)*

The median lethal dose (LD<sub>50</sub>) of the extract was estimated using albino mice by intraperitoneal (i.p) route using the method of Miller and Tainter (1944). The animals were observed for manifestation of physical signs of toxicity such as writhing, decreased motor activity, decreased body/limb tone, decreased respiration and death. The number of deaths in each group within 24 hours was recorded.

#### *Evaluation of anti-inflammatory activity of the extract Carrageenin – induced mice hind paw oedema*

Increase in the mice hind paw linear circumference induced by planar injection of the phlogistic agent was used as the measure of acute inflammation (Winter *et al.*, 1962). Adult albino mice of either sex were used after 24 hours fast and deprived of water only during experiment. Inflammation of the hind paw was induced by injection of 0.1ml of freshly prepared carrageenin suspension in normal saline into the sub planar surface of the hind paw. The linear circumference of the injected paw was measured before and 0.5, 1, 2, 3, 4 and 5 hrs after administration of phlogistic agent. For routine drug testing, the increase in paw circumference at 0.5, 1, 2, 3, 4 and 5 hrs after administration of phlogistic agent was adopted as the parameter for measuring inflammation (Winter, *et al.*, 1962; Akah and Nwambie, 1994; Ekpendu *et al.*, 1994, Besra *et al.*, 1996). Edema (inflammation) was assessed as difference in paw circumference between the control and 0.5, 1, 2, 3, 4 and 5 hrs after administration of phlogistic agent [Hess and Milonig, 1992]. The extract (48, 96 and 144 mg/kg i.p) was administered to various groups of mice, 1 hr before inducing inflammation. Control mice received carrageenin only, while reference group received ASA (100 mg/kg) before induction of inflammation with carrageenin. The average (mean) oedema was assessed by measuring with vernier calipers.

#### *Egg-albumin induced inflammation*

Inflammation was induced in mice by the injection of egg albumin (0.1ml, 1% in normal saline) into the sub planar tissue of the right hind paw (Akah and Nwambie, 1994). The linear circumference of the injected paw was measured before and 0.5, 1, 2, 3, 4 and 5hrs after the administration of the phlogistic agent. The leaf extract (48, 96 and 144 mg/kg i.p) and ASA (100 mg/kg orally) were administered to 24 hrs fasted mice 1 hr before the induction of inflammation. Control group received 10 ml/kg of distilled water orally. Edema (inflammation) was assessed as the difference in paw circumference between the control and 0.5, 1, 2, 3, 4 and 5 hrs after the administration of the phlogistic agent (Hess and Milonig, 1972). The average (mean) edema was assessed by measuring with vernier calipers.

#### *Evaluation of antipyretic activity of the extract*

##### *2, 4 – Dinitrophenol (DNP) induced pyrexia*

Adult albino rats (132 – 175 g) of both sexes fasted for 24 hours but allowed water *ad libitum* were used for the experiment. They were randomized into groups of 6 rats each. DNP (10 mg/kg, i.p) was administered to the rats after obtaining the basal rectal temperatures. Hyperthermia developed within 30 min of DNP administration. Different doses of extract (48, 96, and 144 mg/kg i.p), aspirin (100 mg/kg) and distilled water (10 ml/kg, orally) were administered respectively to the treatment and control groups of animals. Rectal temperatures of the animals were obtained at an hour interval for 5 hrs (Backhouse *et al.*, 1994; Winter *et al.*, 1962; Mbagwu *et al.*, 2007).

##### *Yeast-induced pyrexia*

Adult albino rats (124 – 180 g) of both sexes fasted for 24 hours but allowed water *ad libitum* were used for the experiment. They were randomized into groups of 6 rats each. At zero hour, the basal temperature of the rats was taken using digital clinical thermometer. Thereafter, each animal was administered subcutaneously with 20% W/V aqueous suspension of yeast at a volume of 10 ml/kg (Gural *et al.*, 1955, Okokon and Nwafor, 2010). At suitable intervals beginning one hour after yeast injection, rectal temperature of animals were taken, animals with increase of 1°C were selected and grouped for the study. The extract under study was administered i.p. after the pyrogen at doses of 48, 96 and 144 mg/kg to respective groups of rats. The control group received distilled water (10 ml/kg) and the reference group administered with ASA (100 mg/kg) both intraperitoneally. The rectal temperature of the groups was taken at 1hr interval for 5hrs.

*Statistical analysis and data evaluation*

Data obtained from this work were analyzed statistically using Students' t-test and ANOVA (One- or Two- way) followed by a post test (Tukey-Kramer multiple comparison test). Differences between means will be considered significant at 1 % and 5 % level of significance i.e  $P \leq 0.01$  and  $0.05$ .

**RESULTS**

*Phytochemical screening*

The phytochemical screening of the ethanolic extract of the leaf extract of *Panicum maximum* revealed the presence of alkaloids, cardiac glycosides, tannins, saponins, terpenes and flavonoids.

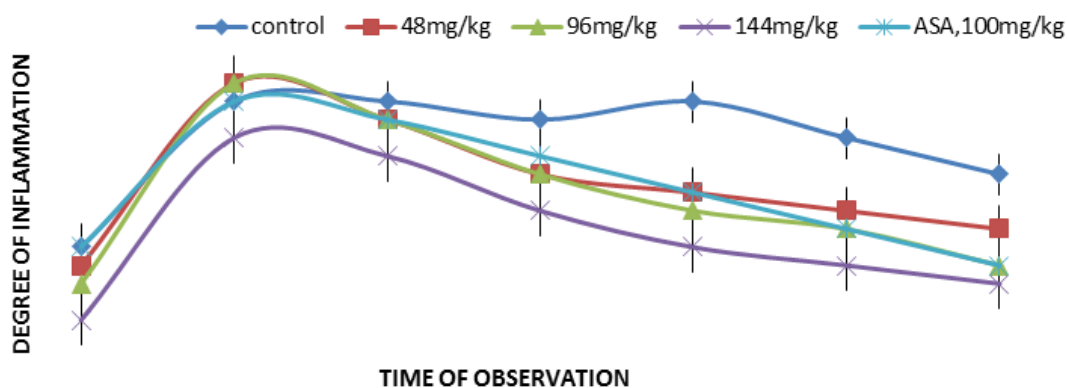
*Determination of Median lethal dose (LD<sub>50</sub>)*

The median lethal dose (LD<sub>50</sub>) was calculated to be  $480.0 \pm 4.91$  mg/kg. The physical signs of toxicity included excitation, paw licking, increased respiratory rate, decreased motor activity, gasping and coma which was followed by death.

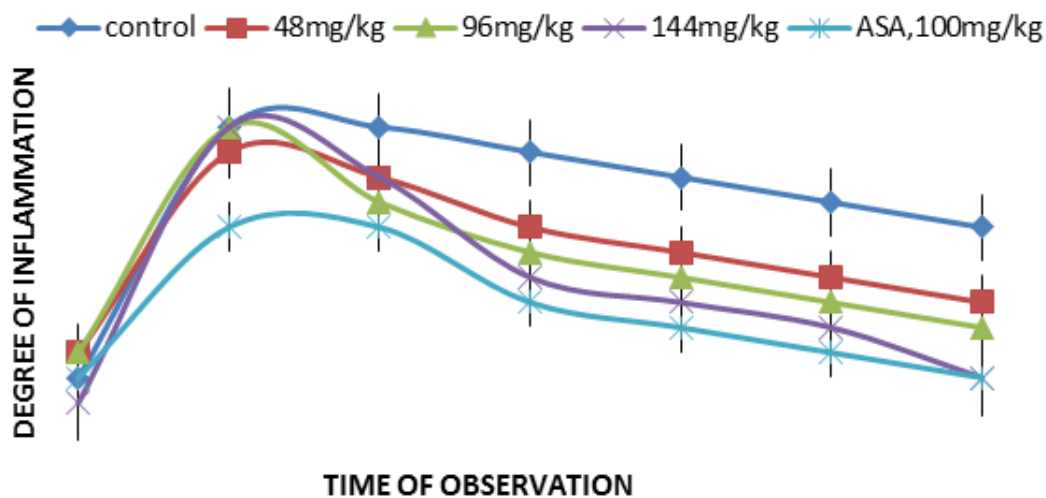
*Anti-inflammatory activity*

*Carragenin-induced oedema in mice*

The effect of ethanolic leaf extract of *Panicum maximum* on carragenin-induced oedema is shown in figure 1. The extract exerted a significant ( $P < 0.05 - 0.001$ ) anti-inflammatory effect in a dose –dependent manner which was comparable to the standard drug, ASA, 100mg/kg.



**Figure 1:**  
Effect of *Panicum maximum* leaf extract on carragenin- induced oedema in mice



**Figure 2:**  
Effect of *Panicum maximum* leaf extract on egg- albumin induced oedema in mice.

**Egg albumin- induced oedema:** Administration of leaf extract of *Panicum maximum* on egg albumin - induced oedema in mice caused a significant ( $p < 0.05 - 0.001$ ) dose-dependent anti-inflammatory effect against oedema caused by egg albumin. The effect was comparable to that of standard drug, ASA (100 mg/kg) (Figure 2).

#### Antipyretic test

**Dinitrophenol induced pyrexia:** The antipyretic effect of the extract on DNP induced pyrexia is shown in figure 3. Administration of the leaf extract of *Panicum maximum* (48, 96 and 144 mg/kg) in the presence of the pyrogen caused a significant ( $P < 0.05 - 0.001$ ) reduction in the temperatures of the extract treated rats when compared with the control. The antipyretic effect was dose-dependent and comparable to that of the standard drug, ASA (100 mg/kg).

**Yeast-induced pyrexia:** Figure 4 shows the effect of the extract against yeast-induced pyrexia. There was a dose-dependent reduction in the temperature of rats treated with the leaf extract. The reductions caused by the extract was significant ( $P < 0.005 - 0.001$ ) when compared to control and comparable to that of the standard drug, ASA (100 mg/kg).

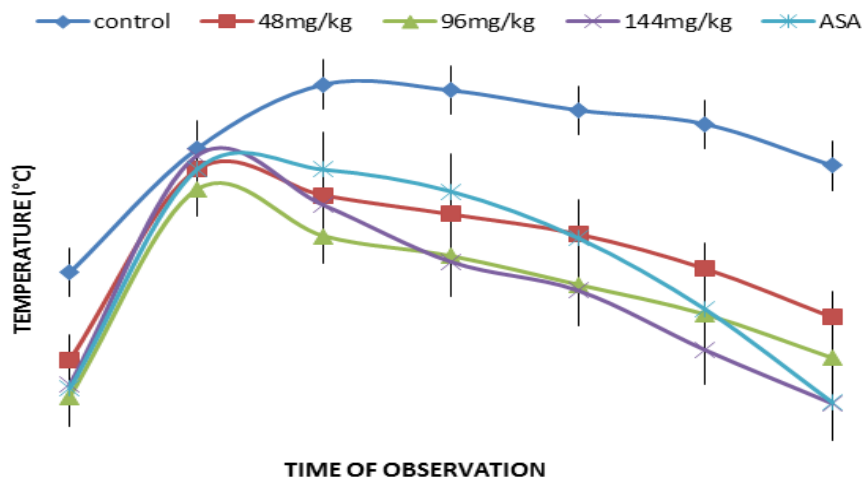
## DISCUSSION

The major folkloric uses of *P. maximum* have been in the treatment of malaria, infections, rheumatism pain, inflammation and diabetes (Antia *et al.*, 2010). The Ibibios of Niger Delta region of Nigeria have been

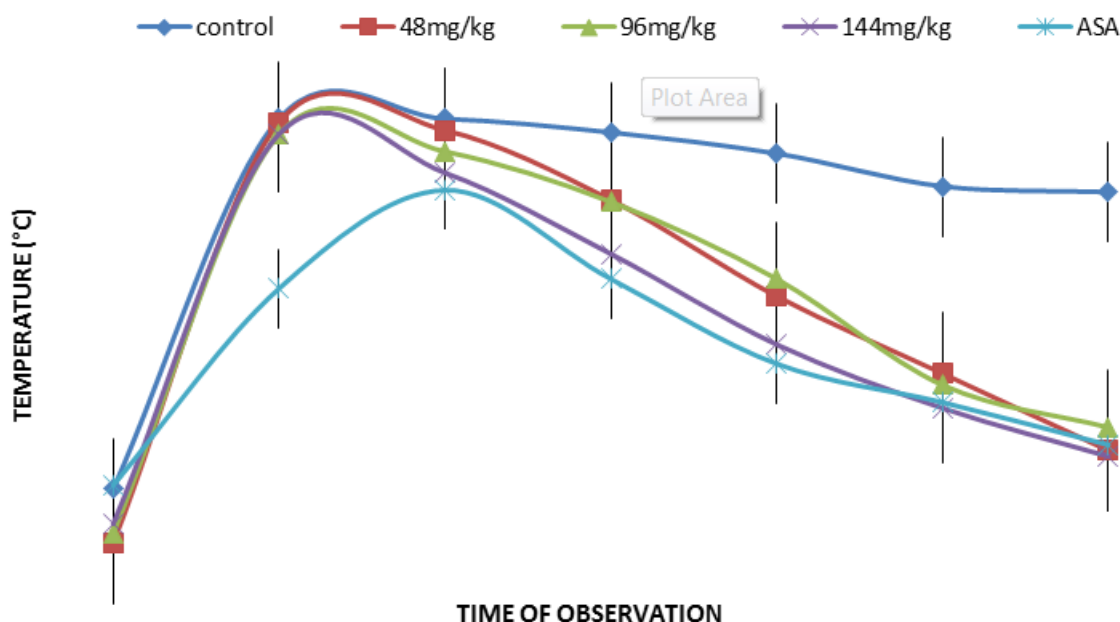
using it to treat malaria and other febrile illnesses. These prompted the need to evaluate the anti-inflammatory and antipyretic potentials of the crude extract of the leaves of *P. Maximum* to ascertain the folkloric claims.

In this work, median lethal dose ( $LD_{50}$ ) was determined to be  $480.0 \pm 4.91$  mg/kg, and the extract was moderately safe (Homburger, 1989). In the carragenin induced oedema, the extract (48 – 144 mg/kg) exerted pronounced effect at the early stage of inflammation (1-2hr) indicating effect probably on histamine, serotonin and kinins that are involved in the early stage of carragenin induced oedema (Vane and Booting, 1987). The extract also reduced later stage of the oedema maybe due to its ability to inhibit prostaglandin which is known to mediate the second phase of carragenin induced inflammation (Vane and Booting, 1987). However, ASA (100 mg/kg) a prototype NSAID, a cyclooxygenase inhibitor whose mechanism of action involves inhibition of prostaglandin, inhibited significantly the paw swelling due to carragenin injection.

The extract also inhibited egg albumin-induced oedema demonstrating that it can inhibit inflammation by blocking the release of histamine and 5-HT, two mediators that are released by egg albumin (Nwafor *et al.*, 2007). However, ASA, a cyclooxygenase inhibitor reduced significantly oedema produced by egg albumin. Flavonoids are reported to be involved in anti-inflammatory activity of plants (Parmer and Gosh, 1978). These have been found to be present in the extract.



**Figure 3:** Effect of *Panicum maximum* leaf extract on DNP induced pyrexia in rat



**Figure 4:**  
Effect of *Panicum maximum* leaf extract on yeast induced pyrexia in rat

In this study, the extract was observed to inhibit greatly DNP- and yeast -induced pyrexia. The extract is likely to reduce pyrexia by reducing brain concentration of prostaglandin E<sub>2</sub> especially in the hypothalamus through its action on COX-3 (Botting and Ayoub,2005) or by enhancement of the production of the body's own antipyretic substances like vasopressin and arginine (Chandrasekharan,2002). This effect is due to the presence of the phytochemical compounds in the extract.

In conclusion, the results of this study support the ethnobotanical use of the plant in the treatment of febrile illnesses and inflammatory conditions. Further investigation is being advocated especially in elucidating cellular mechanisms and establishing structural components of the active ingredients with a view of standardizing them.

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#### REFERENCES

**Andrew, U. (2011).** Antimalarial and analgesic activities of ethanolic crude leaf extract and fractions of *Panicum*

*maximum*. M.Sc. Dissertation. University of Uyo, Uyo, Nigeria.

**Antia, BS., Okokon JE, Umoh E E., Udobang JA .(2010).** Antidiabetic activity of *Panicum maximum*. Int. Journal of Drug Dev. and Research 2(3): 488 - 492.

**Akah, P. A. and Nwanbie, A. (1994).** Evaluation of Nigerian traditional medicines plants used for rheumatic (inflammatory) disorder. *Journal of Ethnopharmacology* 42: 179 – 182.

**Amico-Roxas, M., Caruso, A., Trombadore, S., Scifo, R., Scapagnime, U. (1984).** Gangliosides antinociceptive effects in rodents. *Archives of International Pharmacodynamics and Therapeutics*. 272:103-117.

**Bamgbose, S. O, Noamesi, B. K., (1981).** Studies on crytolepine II: inhibition of carrageenan-induced oedema by Crytolepine. *Planta Medica*. 42:392-396.

**Bentley, G. A., Newton, S.H., Starr, J. (1983).** Studies on the antinociceptive action of agonist drugs and their interaction with opoid mechanisms. *British Journal of Pharmacy*. 79,125 - 134.

**Berken, T., Ostunes, L., Lermioglu, F., Ozer, A. (1991).** Antiinflammatory analgesic and antipyretic effect of an aqueous extract of *Erythraea ceulaurum*. *Planta Medica*. 57,34 -37.

**Besra, S. E., Sharma, R. M., Gomes., A. (1996).** Anti-inflammatory effect of petroleum ether extract of leaves of *Litchi Chinensis*. Caertn (sapindaceae). *Journal of Ethnopharmacology*. 54:1-6.

**Blackhouse, N., Delporte, C., Negrete, R., Munoz, O., Ruiz, R. (1994).** Anti inflammatory and antipyretic activities

- of *Maytenus boaria*. International Journal of Pharmacognosy. 32: 239-244.
- Botting, R., Ayoub, S. S. (2005).** COX-3 and the mechanism of action of paracetamol/acetaminophen. Prostaglandins, Leukotrienes and Essential fatty Acids. 72(2):85 – 87.
- Chandrasekharan, N.V. (2002).** COX-3, a cyclooxygenase-1 variant inhibited by acetaminophen and other analgesic/antipyretic drugs:cloning, structure and expression. Proceeding of National Academy of Science. 99,13926 – 13931.
- Correa, C. R., Calixto, J. B. (1993).** Evidence of participation of  $\beta_1$  and  $\beta_2$  receptors, in formalin induced nociceptive response in mouse. British Journal of Pharmacology 110,193-198.
- Correa, C. R., Kyle, D. J., Chakravarty, S., Calixto, J.B. (1996).** Antinociceptive profile of the pseudopeptide  $\beta_2$  bradykinin receptors antagonist NPC 18688 in mice. British Journal of Pharmacology. 117,552-556.
- Deraedt, R., Jougnay, S., Falhout, M. (1980).** Release of Prostaglandin E and F in an allogenic reaction and its inhibition. European Journal of Pharmacy. 51,17-24.
- Ekipendu, T.O., Akah, P. A., Adesomoju, A. A., Okogun, J. I. (1994).** Antinflammatory and antimicrobial activities of *Mitracarpus scaber* extracts. International Journal of Pharmacology. 32:191-195.
- Gundidza, M., Chiyangaya, F., Chagonda, L., Depooter, H. L., Mavi, S. (1994).** Phytoconstituents and antimicrobial activity of the leaf essential oil of *Clausena anisata* (Wild) Hook. f. ex Benth. Flavour and Fragrance Journal. 9(6), 299 – 303.
- Gural, M. L., Kohli, P.P., Saxena, P.H. (1955).** Antipyretic activity of some indigenous drugs. Indian Journal of Medical Research. 6:89-92.
- Hess, S. M., Milonig, R. C. (1972).** Inflammation In: Lepow, L. H., Ward, P.S. (Eds). Inflammation, Mechanism and control. Academic Press, New-york, USA. pp.1-2.
- Homburger, F., (1989).** In vivo testing in the study of toxicity and safety evaluation. In: A guide to general toxicology. Marquis J. K. (Ed). 2nd Edn Karger, New York.
- Lembeck, F., Holzer, P. (1979).** Substance P as neurogenic mediator of antidromic, vasodilatation and neurogenic plasma extravasation. Naunyn-Schmiedeberg's Archives of Pharmacy. 310, 175-183.
- Liang, Y. C., Huang, Y. T., Tsau, S. H., Lin-Shiau, S. Y., Chen, C. F., Lin, J. K. (1999).** Suppression of inducible cyclo-oxygenase and inducible nitric acid synthase by apigenin and related flavonoid in mouse macrophages. Carcinogenesis. 20, 1945-52.
- Lin, L. L., Lin, A. Y. Knoop, J. L. (1992).** Cytosolic phospholipase A<sub>2</sub> is coupled to hormonally regulated release of arachidonic acid. *Proceeding of National Academy of Science, U. S. A.* 89:6147-6157.
- Mbagwu HO, Anene RA, Adeyemi OO. (2007).** Analgesic, antipyretic and antiinflammatory properties of *Mezoneuron benthamianum* Baill Caesalpiniaceae. Nigerian Quarterly Journal of Hospital Medicine. 17(1), 35 – 41.
- Miller, L. C. Tainter, M. L. (1944).** Estimation of ED<sub>50</sub> or LD<sub>50</sub> values and their error using logarithmic-probit graph paper. Proc. Soc. Exp. Biol and Medicine. 57, 261 – 264.
- Ngadjui, B. T., Ayafor, J. F., Sodengam, B. L., Collony, J.D. (1989).** Limonoids from *Clausena anisata*. Journal of Natural Products. 52, 243 – 247.
- Nwafor P. A., Jacks T. W., Ekanem A. U. (2007).** Analgesic and anti-inflammatory effects of methanolic extract of *Pausinystalia mecroceras* stem bark in rodents. Journal of Pharmacology; 3,86-90.
- Okokon JE, Nwafor PA. (2010).** Antiinflammatory, analgesic and antipyretic activities of ethanolic root extract of *Croton zambesicus*. *Pakistan Journal of Pharmaceutical sciences.* 23(4): 383 - 390.
- Parmer, N. S., Ghosh M. N. (1978).** Anti-inflammatory activity of gossypin a biflavonoid isolated from *Hibiscus vitifolicus* Linn. Indian Journal of Pharmacology. 10: 277-293.
- Sofowora, A. (1993).** Medicinal Plants and Traditional Medicine in Africa. 2nd edn, Spectrum Book Ltd, Ibadan, Nigeria. pp. 2 – 23.
- Tjolsen, A., Berge, O. G., Hunnskaar, S., Rosland, J. H., Hole K. (1992).** The formalin test: An evaluation of the method. Pain 51,5-17.
- Trease, G.E., Evans, W. C. (1989).** Pharmacognosy, 13<sup>th</sup> ed. Bailliere Tindal, London. pp. 683 - 684.
- Van Oudtshoorn, F. (1999).** Guide to grasses of Southern Africa. Briza Publications, Pretoria.
- Vane T., Booting R. (1987).** Inflammation and Mechanism of action of antiinflammatory drugs. *FASSEB J.* 1:89-96.
- Winter, C. A., Risley, E. A., and Nuss, G. W. (1962).** Carrageenin-induced oedema in hind paw of the rats as an assay of anti-inflammatory drugs. *Proc. Soc. Expt. Biol. Med.* 111,544-547.