



www.ajbrui.org

Afr. J. Biomed. Res. Vol. 26 (May 2023); 239 - 247

Research Article

Efficacy, Safety and Pharmacokinetics of a Triple Combination of Artemether-Lumefantrine and Amodiaquine in Laboratory Rodents

***Nwaiwu O.¹, Okafor S.¹, Amao O.¹, Ojobor P.², Akinyede A.¹ and Dike S.³**

¹*Department of Pharmacology, Therapeutics and Toxicology, Faculty of Basic Medical Sciences, College of Medicine, University of Lagos Nigeria.*

²*Central Research Laboratories, College of Medicine, University of Lagos.*

³*Department of Physiology, Faculty of Basic Medical Sciences, College of Medicine, University of Lagos.*

ABSTRACT

Presently artemisinin-based combination therapies (ACT) are now widely recommended as first-line treatment of uncomplicated malaria, however there are some reports and evidence of treatment failure despite adequate drug concentrations. Addition of a second long-acting partner drug to the existing single partner artemisinin-based combination therapy may delay the development of resistance. The objective of the present study is to determine the efficacy, of a triple combination of artemether-lumefantrine and amodiaquine in laboratory rodents. The blood schizonticidal activity of the proposed triple combination of artemether-lumefantrine (AL) and amodiaquine (AQ) was evaluated in a rodent model of *Plasmodium berghei*. Animals were treated orally with standard doses of artemether-lumefantrine (AL), amodiaquine (AQ) or the triple combination (ALAQ). Parasitological activity and survival of the animals were assessed over 24 days. Safety and plasma concentrations of artemether, amodiaquine and lumefantrine were determined both in the standalone and in the triple combination treatment groups using uninfected but treated albino rats. There was a progressive significant decline in parasitemia in all therapeutic groups with the triple combination (ALAQ) achieving a 100% suppression of parasites by day 16. ALAQ resulted in significant elevations in total white blood cell counts, platelet counts, alanine transaminase and urea levels. There were significant reductions in blood pressure and heart rate. Compared to artemether-lumefantrine administered alone the triple combination (ALAQ) showed lower plasma artemether levels and area under the curve (AUC) values at 72hours with low day 7 lumefantrine plasma levels in the triple combination (ALAQ). These preliminary results showed that the triple therapy's efficacy, safety and pharmacokinetics are quite encouraging. Human studies are required to confirm the efficacy, safety and pharmacokinetic findings in this study.

Keywords: *Uncomplicated malaria, Artemisinin-Based Combination Therapy, Amodiaquine, Artemether-Lumefantrine Plasmodium berghei.*

*Author for correspondence: Email: onwaiwu@unilag.edu.ng; Tel: + 2348037790925

Received: August 2020; Accepted: March 2021

DOI: 10.4314/ajbr.v26i2.13

INTRODUCTION

Despite the scale up of antimalarial interventions and the resultant reduction in malaria transmission in some areas, the morbidity and mortality due to malaria disease is still high in other endemic areas of sub-Saharan Africa (Prudhomme *et al.*, 2010). The emergence of antimalarial drug resistance has led to many countries in sub-Saharan Africa to adopt the World Health Organization (WHO) recommended artemisinin-based combination therapy for treatment of uncomplicated *P. falciparum* malaria (World Health Organization, 2006). Artemisinin-based combination therapies (ACT) have a fast-acting artemisinin derivative with rapid effects on parasite clearance, and a long-acting drug to prevent recrudescence

and development of resistance (World Health Organization, 2006; World Health Organization, 2010). Artemether-Lumefantrine and artesunate-amodiaquine are both recommended by the WHO and have shown good efficacy and safety outcomes which has resulted in a reduction of malaria morbidity and mortality in different populations (Van Vugt *et al.*, 2000 ; Mutabingwa *et al.*, 2005 ; Makanga *et al.*, 2006; Pecoul *et al.*, 2008).

Delayed clearance times suggest emergence of resistance and there are reports of artemisinin resistance in *P. falciparum* malaria in western Cambodia, Thai-Burmese border and the Greater Mekong region (Noedl *et al.*, 2008; Mizuno *et al.*, 2009 ; Dondorp *et al.*, 2009; Phyo *et al.*, 2012; Carrara *et al.*,

2013; Arley *et al.*, 2014). Mutations associated with artemisinin resistance on chromosome 13 have been identified and treatment failures with artemisinin-based combination therapies have also been reported in non-immune Europeans and travelers after visiting endemic regions (Jackson *et al.*, 2006 ; Repetto *et al.*, 2011; Färnert *et al.*, 2012). Failure of artemisinin efficacy would render partner drugs susceptible to greater selection pressure for the development of resistance, compromising the value of the combination (White and Pongtavornpinyo , 2003; Kay and Hastings ,2015).

In endemic regions with high malaria transmission the use of one long acting partner drug with a very short acting artemisinin may expose parasites to the emergence of resistance because the single partner drug exposes remaining and new parasites to sub therapeutic drug levels over a long time. A perfect drug combination is one with an excellent overlap in the exposure of the drug components. It has been postulated that addition of a second long acting partner drug to the present artemisinin-based combination therapy will strengthen the combination and prevent the parasites from developing resistance to artemisinins. Addition of a third partner drug with a longer half-life than lumefantrine such as amodiaquine (2 weeks), mefloquine (2–3 weeks) or piperazine (4-5weeks) will strengthen the present combination., and delay development of resistance. A third partner drug with a different mechanism of action and longer half-lives will further reduce the time window in which remaining and new parasites can be selected by offering better therapeutic drug levels over a long time(White ,1998 ; Dondorp *et al.*, 2010). There are unpublished reports of the use of triple combination therapy for the treatment of uncomplicated *P. falciparum* malaria in Nigeria. In such situations, a third schizonticidal drug usually a monotherapy is administered to a patient either together or after a presumed treatment failure of an artemisinin-based combination therapy. The present study evaluated the efficacy, safety and pharmacokinetics of a triple combination of artemether-lumefantrine and amodiaquine administered together in laboratory rodents. A major objective is to assess if the simultaneous administration of artemether-lumefantrine with amodiaquine will have better cure rates, be safe and will not affect the bioavailability of the individual drugs when compared with existing one partner artemisinin-based combination.

MATERIALS AND METHODS

Animals: Adult albino mice of weight range 20-25g (average age 7 weeks) were obtained from the laboratory animal center, College of Medicine, University of Lagos, Lagos Nigeria. The animals were housed in well ventilated cages in 4 groups of 5 animals per cage at a regular 12 h light-dark cycle. The animals were acclimatized for a minimum period of three days prior to the experiment and allowed free access to drinking water and standard pellets diet (Pfizer standard rodent pellet diet). All experimental protocols maintenance and care of experimental animals complied with research guidelines for the use and care of laboratory animals (American Physiological Society, 2002; National Research Council, 1985; National Academy of Science, 2011).

Administration of drugs: Treatment was carried out twice daily for artemether-lumefantrine and once daily for amodiaquine for a period of 3 days. Artemether-lumefantrine 20mg/120mg (AL) was sourced from Novartis Pharma, Lagos Nigeria while Amodiaquine syrup 60ml (AQ) was sourced from Pfizer, Lagos Nigeria. The amodiaquine syrup and the AL tablets dissolved in distilled water were administered to the mice orally using an oral cannula as follows;

Group 1: Control group. Each mouse received distilled water for 7 days

Group 2: This group was treated with amodiaquine (0.02ml) 10g/kg orally (p.o.) once daily, for 3 days starting from the fourth day post infection

Group 3: This group was treated with artemether and lumefantrine.(0.025ml) [artemether 2mg/kg and lumefantrine 12mg/kg orally (p.o.) twice daily, for 3 days starting from the fourth day post infection

Group 4: This group was treated with artemether and lumefantrine(AL) twice daily plus amodiaquine. Each mouse was administered 0.025ml of AL and 0.02 ml of amodiaquine orally once daily for 3 days starting from the fourth day post infection

Inoculation of mice with malaria parasites. : Evaluation of schizonticidal activity on established infection was done by the curative test (Madara *et al.*, 2010; Akinyede *et al.*, 2013). The rodent parasite chloroquine sensitive *Plasmodium berghei* NK 65 was sourced from National Institute for Medical Research (NIMR), Lagos, Nigeria. Animals were infected with parasitized blood obtained from the tail veins of infected experimental mice. Approximately 0.1 mL of infected blood (3–4 drops) was diluted in freshly prepared phosphate buffer solution (PBS). Mice were inoculated intraperitoneally with 0.1 mL of parasitized saline suspension containing approximately 1×10^7 parasites on the first day. On the 4th day the mice were divided into four groups of five mice each and a smear was taken to confirm presence of *P.berghei* in the mice before the start of treatment (White, 1997; Shida *et al.*, 1989).

Parasite counts in infected mice and assessment of efficacy: Blood smears were collected daily and examined microscopically to monitor the parasitaemia level until the parasites were cleared. Peripheral blood smears were prepared by using blood obtained from the tail veins of infected experimental mice. The thin films were fixed in methanol (3 min) and then stained with May-Grunewald Giemsa. Blood smears were examined at a magnification of X 100 by oil immersion light microscopy. Parasitemia was determined by counting 30 or 100 fields of view for $>0.5\%$ and $<0.5\%$ infected erythrocytes, respectively ensuring an acceptable standard error of 22% at 0.1% parasitemia and a limit of detection on the order of 0.002% parasitemia. The total number of erythrocyte and the parasitized erythrocyte in the four experimental groups were counted and recorded. Infected erythrocytes were counted by using the formula;

Malaria parasite density/mL of blood

$= \text{No. parasites counted} + 8,000 / 200 \text{ leukocytes.}$

(Shida *et al.*, 1989; World Health Organization, 2009).

Mean parasite counts mice in each treatment group was determined and compared on days 4, 8, 12, 16, 20 and 24.

Mice were euthanized by sodium pentobarbitone injection (50 to 100 mg/kg i.p.) (Fish *et al.*, 2008).

Assessment of safety: We assessed the haematological, biochemical and cardiovascular safety of the proposed triple artemisinin-based combination therapy. Normotensive albino rats, of either sex weighing 220-250g not infected with malaria parasites were used for the assessment. The rats were divided into four groups of control, amodiaquine alone, artemether-lumefantrine alone and artemether-lumefantrine co-administered with amodiaquine for 3 days. Blood samples were collected from the ocular veins with capillary tubes and put into EDTA and heparinized sample bottles to assess the haematological (haemoglobin level, leukocyte count), and biochemical parameters (serum alanine aminotransferase, aspartate aminotransferase, total bilirubin, glycaemia, urea and creatinine) respectively (Osonuga *et al.*, 2012; Olayemi *et al.*, 2012; Kotepui *et al.*, 2014). The cardiovascular safety of the triple combination therapy was assessed by determination of blood pressure and heart rate (Feng, *et al.*, 2015). The rats were anaesthetized with intra-peritoneal injection of 0.5 ml of 25% urethane (1000mg/kg) (Fish *et al.*, 2008). The anaesthetized rat was fixed in a supine position on a dissecting table and the temperature of the animal was maintained at 37°C by the use of an overhead lamp. A longitudinal midtracheal incision approximately 2 cm long was made in order to expose the trachea, the right jugular vein and left carotid artery. After tracheotomy, the trachea was cannulated with polyethylene tube 92.75 mm diameter to maintain a free airway for spontaneous respiration. The right jugular vein was cannulated with polyethylene tube for flushing with 0.9% NaCl (normal saline). The cannulation of the carotid artery was performed in the same manner as the cannulation of jugular vein and the polyethylene tube (1 mm diameter) filled with heparin sodium in saline solution was used. The arterial blood pressure was measured from the left carotid arterial cannula connected to a research grade blood pressure transducer (Harvard, 60-3003) which was connected to an oscillograph (Harvard) for recording. The animal was allowed to equilibrate for at least 30 min. before recording. The mean arterial Blood pressure was calculated using the following formula $MABP = DP + 1/3 (SP-DP)$; Where, DP= diastolic pressure; SP= systolic pressure. The response of mean arterial blood pressure (MABP) was expressed as percent change from the control group measurement (Adeboye *et al.*, 1999; Lahlou *et al.*, 2002; Feng *et al.*, 2015).

Pharmacokinetic study: Simultaneous determination of artemether, lumefantrine and amodiaquine, plasma concentrations was done in treated non-infected albino rats by high-performance liquid chromatography with UV detector (HPLC-UV) at the Central Research Laboratories, College of Medicine, University of Lagos. Artemether, lumefantrine, amodiaquine concentrations and the internal standard (Halofantrine) were measured using high performance liquid chromatography methodologies adapted from previous studies (Virendra *et al.*, 2004; Huang *et al.*, (2010); Maddela *et al.*, 2015). The area under the blood concentration-time curve (AUC) of artemether, amodiaquine and lumefantrine were determined using a non-compartmental model in WinNonLin

Professional pharmacokinetic software version 2.1 (Pharsight Corporation, Mountain View, USA). The concentrations and the AUCs were then compared between treatment arms.

Blood samples for pharmacokinetic analysis were collected from the ocular veins with capillary tubes and put into EDTA and heparinized sample bottles for assessment of artemether, amodiaquine, and lumefantrine levels. 1 ml of blood was drawn at pre-determined time peaks from each treated albino rat at the following times 24hrs, 48hrs, 72hrs and on day7 for lumefantrine levels. The Blood samples were centrifuged at a speed of 4000rpm for 10mins with a centrifuge (5702 by Eppendorf). The plasma was stored at -70 degrees centigrade.

All chemicals (Methanol (MeOH), KH₂PO₄ (potassium dihydrogen orthophosphate, acetonitrile) were of HPLC grade and purchased from Merck (Darmstadt, Germany). Reference standards of artemether, amodiaquine, lumefantrine, and halofantrine (Internal Standard) were sourced from Ipcalaboratories Ltd. (Mumbai, India). Blank plasma was collected from untreated rats. Water was distilled water. The HPLC system consisted of Agilent 1100 series with UV detector. Separation was achieved on a Zorbax Eclipse XDB RP C8 HPLC column (150 X 4.6mm, 5 µm, Agilent). All chromatographic experiments were carried out in the isocratic mode at room temperature. The mobile phase was vacuum degassed before use. The mobile phase consisted of a mixture of Acetonitrile: Potassium dihydrogen orthophosphate (H₂5mMKH₂PO₄) ; (70:30) %. Injection volume was 20µl at a flow rate of 1.0ml/min and wavelength 216 nm at ambient temp and pH 3.9.

The powdered drugs were weighed and dissolved in in MeOH (methanol) and water (1:1) at room temperature to prepare working stock solutions of 1.0 mg/ml and working solutions. Serial dilutions of the stock solutions were appropriately diluted to lower concentration for spiking the calibration standards. Spiked mixed standards of Amodiaquine (1.25) + lumefantrine (46.875)+Artemether 7.812+ halofantrine (Internal Standard) (10) µg/mL were added to drug-free human plasma and mixed properly by vortexing. Working solutions were made by diluting with appropriate volume of mobile phase. Calibration standards and QC samples were prepared from separately weighted stock solutions. The stock solutions, standards, QC samples, and the I.S. working solution (100 g/mL) were stored at -70 °C between uses.

A Liquid-liquid extraction method was followed for extraction of artemether, lumefantrine and amodiaquine from rat plasma. An aliquot of 1 mL of plasma spiked with 0.05ml of internal standard (3µg/ml) and extraction solvent (2 ml of chilled acetonitrile) was dispensed into 5 mL plain polypropylene sample bottle, vortex mixed for 1 minute and sonicated for 10 minutes using ultrasonic bath. The resultant precipitant was centrifuged at 4000 rpm for 10 minutes. The supernatant was aspirated using disposable Pasteur pipette and filtered using 0.45 µm syringe filter and this was subjected to HPLC analysis. Extraction recoveries of artemether, lumefantrine and amodiaquine were satisfactory (75-80%) .

A complete validation was carried out as per US FDA (US DHHS, FDA and CDER.(2001) and EMEA guidelines (European Medicines Agency (2011). The validation

parameters tested include specificity, linearity, precision and accuracy, limit of detection (LOD) and limit of quantification (LOQ), recovery and stability

Statistical Analysis: Results were expressed as mean \pm standard error of mean. Descriptive summary statistics was presented for values and concentrations of artemether, amodiaquine and lumefantrine. The results of the effects of artemether-lumefantrine alone, amodiaquine alone, was compared to the effects of the triple combination of the three drugs administered together. The two way ANOVA test Bonferonni Post Test was used to analyse and compare the results at a 95% confidence level. Values of $p < 0.05$ and < 0.001 were considered significant. Graph pad prism version 5.1 was used for statistical analysis.

RESULTS

Efficacy (Parasite clearance): There was a progressive significant decline in parasite counts in all three therapeutic groups. In the group that received amodiaquine or artemether-lumefantrine as separate medications there were residual parasitaemia up till the 20th day of observation. In the animal group that received a combination of artemether lumefantrine and amodiaquine administered together (ALAQ), there were no parasites detected after day 12. In this group there was 100% clearance of parasites on days 16 and 20. In the control group that received distilled water there was a progressive increase in parasite counts with all mice dying after the 16th day. Table 1 below shows the effect of amodiaquine alone, artemether-lumefantrine alone and the combination of

artemether-lumefantrine and amodiaquine on parasite counts in *P. berghei* infection in albino mice.

Safety:

The effect of artemether-lumefantrine alone (AL), amodiaquine alone (AQ), and the combination of artemether-lumefantrine and amodiaquine (ALAQ) on haematological, hepatic, renal and cardiovascular parameters were evaluated and shown in Tables 2, 3 and 4 respectively. The triple combination of artemether-lumefantrine and amodiaquine (ALAQ) showed significant elevations in total white blood cell counts, red blood cells platelet counts, alanine transaminase and urea levels compared to the control, amodiaquine alone and artemether-lumefantrine alone groups. On the cardiovascular function, the triple combination showed significant reductions in both systolic and diastolic blood pressure, mean arterial blood pressure and heart rate compared to the control and artemether-lumefantrine alone groups. However, the amodiaquine alone group showed lower values in these cardiovascular parameters compared to the triple combination therapy.

Pharmacokinetics: The plasma concentrations of artemether lumefantrine and amodiaquine were measured up to 72 hours with day 7 concentrations for lumefantrine after oral administration. Total-assay coefficients of variation (CVs) for artemether during analysis was $< 5\%$ at all quality-control levels. The lower limit of quantification (LLOQ) was set to 1.4 ng/mL for lumefantrine with an LLOQ of 50 ng/ml, a calibration range of 50 to 10,000 ng/ml, and a CV range of 1.1 to 6.7%

Table 1:

Effect of artemether-lumefantrine alone, amodiaquine alone and the combination of artemether-lumefantrine with amodiaquine on parasite counts in *Plasmodium berghei* infection in albino mice

Experimental animal groups	Day 4 parasite count/ μ L	Day 8 parasite count/ μ L	Day 12 parasite count/ μ L	Day 16 Parasite count/ μ L	Day 20 parasite count/ μ L	Day 24 parasite count/ μ L
Control	24.64 \pm 0.49	30.04 \pm 0.19	60.64 \pm 2.46	85.97 \pm 2.46	All animals died	All animals died
Artemether-lumefantrine (AL)	15.50 \pm 0.32	12.82 \pm 0.30 ^c	10.44 \pm 0.19	8.31 \pm 0.27 ^{cY***}	1.83 \pm 0.13 ^{cY***}	0
Amodiaquine(AQ)	16.26 \pm 0.31	11.52 \pm 0.57 ^c	7.52 \pm 0.16 ^{cY}	6.26 \pm 0.24 ^{cY*}	0.13 \pm 0.12 ^{cY***}	0
Artemether-lumefantrine + amodiaquine (ALAQ)	14.60 \pm 0.08	10.06 \pm 0.09 ^c	5.89 \pm 0.18 ^{cY}	0.00 \pm 0.00	0.00 \pm 0.00 ^{cY***}	0

Values are Mean \pm SEM; (n=5); ^c $P < 0.001$ vs Day 0, ^Y $P < 0.001$ vs Day 1; ^{*} $P < 0.01$, ^{*} $P < 0.05$, ^{***} $P < 0.001$ vs Day 2 (Two-Way NOVA followed by Bonferonni Post Test). Day 0 refers to 1st day of therapy.

Table 2:

Effect of artemether-lumefantrine alone (AL), amodiaquine alone (AQ), and the combination of artemether-lumefantrine and amodiaquine (ALAQ) on haematological values in albino rat

Parameter	Control	Artemether-lumefantrine (AL)	Amodiaquine (AQ)	Artemether-lumefantrine + Amodiaquine (ALAQ)
White blood cell ($\times 10^3/\mu$ L)	8.70 \pm 1.06	11.45 \pm 0.40*	11.02 \pm 0.92*	13.77 \pm 0.59* \neq α
Haemoglobin (g/dl)	10.66 \pm 0.72	12.35 \pm 0.36*	11.42 \pm 0.79	11.32 \pm 0.22 \neq
Red Blood Cell ($\times 10^6/\mu$ L)	5.20 \pm 0.22	6.06 \pm 0.18*	6.13 \pm 0.18*	6.02 \pm 0.10*
Mean cell volume (MCV /fl)	61.27 \pm 0.75	56.47 \pm 1.48*	55.92 \pm 1.66*	55.05 \pm 1.51*
Mean cell haemoglobin (MCH)(pg/cell)	20.00 \pm 0.47	20.77 \pm 0.52	17.65 \pm 1.13* \neq	18.72 \pm 0.31* \neq
Mean cell haemoglobin conc.(MCHC) (g/dl)	33.22 \pm 0.46	31.70 \pm 1.51	31.92 \pm 1.00*	32.60 \pm 0.98
Platelet count ($\times 10^3/\mu$ L)	255.28 \pm 118.8	605.25 \pm 2.17*	530.25 \pm 48.58* \neq	682.50 \pm 42.73* \neq α

* $p < 0.05$ when triple combination group is compared with the control group, * \neq $p < 0.05$ when compared with the artemether-lumefantrine group and * \neq α $p < 0.05$ when compared with the amodiaquine group

The within-assay and between-assay coefficients of variations for amodiaquine were always <10% at the limits of quantification. In the three treatment groups there were no significant changes to amodiaquine levels. Compared to artemether-lumefantrine administered alone the triple combination (ALAQ) showed significant lower artemether levels and a lower area under the curve at 72hours. The

lumefantrine levels were similar in all treatment groups with a lower area under the curve for day 7 lumefantrine levels. (Tables 5, 6 and 7). Figure 1 shows a representative chromatogram for amodiaquine, lumefantrine, artemether and halofantrine (Internal standard) at 72 hours with 1.451, 4.646, 5.542 and 2.756 minutes retention time respectively

Table3:

Effect of amodiaquine alone (AQ), artemether-lumefantrine alone (AL) and the tripple combination of artemether-lumefantrine and amodiaquine (AL+ AQ) on hepatic and renal parameters in albino rat.

Parameter	Control	Artemether-lumefantrine (AL)	Amodiaquine (AQ)	AL + AQ
Aspartate transferase (AST) U/L	95.25±5.96	235.72±5.84* \neq	191.40±6.56*	214.75±3.59* $\neq\alpha$
Alanine transaminase (ALT) U/L	32.37±0.44	22.02 ± 4.41 * \neq	94.80±4.57*	69.07±6.72* $\neq\alpha$
Alkaline phosphatase (U/L)	164.40±4.80	260.75 ± 36.77*	221.92±13.67*	295.55±17.83* \neq
Total Protein (g/dl)	0.82±0.02	1.57 ± 0.52 *	1.62±0.35*	0.62±0.20 $\neq\alpha$
Urea (mmol/L)	1.62 ±0.21	2.92 ± 1.47	4.27±0.21*	5.27±0.16* $\neq\alpha$
Creatinine (µmol/l)	31.60±1.16	42.09 ± 5.35	27.83±0.95	33.82±3.58

*=p<0.05 when combination group is compared with the control group, * \neq =p<0.05 when compared with the artemether-lumefantrine group and * $\neq\alpha$ =p<0.05 when compared with the amodiaquine group

Table 4:

Effect of amodiaquine alone, artemether-lumefantrine alone and the combination of artemether- lumefantrine plus amodiaquine on cardiovascular parameters in rat

Cardiovascular Parameters	Control	Artemether lumefantrine	Amodiaquine (AQ)	ALAQ
Systolic blood pressure (mmHg)	99.32±2.12	83.55±3.09*	59.00±1.77* \neq	66.72±1.79* $\neq\alpha$
Diastolic blood pressure (mmHg)	79.00±0.57	63.62±3.13*	38.20±1.95* \neq	45.77±4.56* $\neq\alpha$
Pulse pressure (mmHg)	20.32±1.76	19.92±1.80	20.80±1.51	19.95±3.15
The mean arterial blood pressure (mmHg)	85.77± 1.01	70.26 ±3.00*	45.13±1.75* \neq	52.42±3.57* $\neq\alpha$
HR (beats /min)	495.00±45.00	450.00±17.32	330.00 ±17.32* \neq	375.00±28.72* $\neq\alpha$

*=p<0.05 when triple combination group is compared with the control group, * \neq =p<0.05 when compared with the artemether lumefantrine group and * $\neq\alpha$ =p<0.05 when compared with the amodiaquine group

Table 5:

Plasma concentrations and area under the curve of artemether in albino rat administered with artemether-lumefantrine (AL) alone and the triple combination of artemether-lumefantrine plus amodiaquine (AL+AQ)

Parameter	Artemether conc. in AL Alone	Artemether conc. in AL +AQ	P-value
Cmax (ng/ml)	1.38 (0.68 – 2.74)	0.26 (0.14 – 0.51)	0.12
24 hour conc. (ng/ml)	0.255 (0.218 -0.280)	0.230 (0.150 -0.264)	0.287
48hour conc. (ng/ml)	0.270 (0.255 – 0.289)	0.250 (0.238-0.273)	0.086
72 hour conc. (ng/ml)	0.289 (0.272 -0.315)	0.260 (0.249 -0.271)	0.006*
AUC ^{0-∞} µgh/ml	17.558 (11.259 -24.263)	11.778 (6.021 – 17.897)	0.003**

*P<0.05

Table 6:

Plasma concentrations and area under the curve of amodiaquine in albino rat administered with amodiaquine (AQ), and the combination of artemether- lumefantrine and amodiaquine (AL+AQ)

Parameter	Amodiaquine conc. in AQ alone	Amodiaquine conc. in AL +AQ	P-value
Cmax (ng/ml)	59.0(29.0 -118.0)	62.6 (31.1 – 131.5)	0.215
24 hour conc. (ng/ml)	58.66 (38.50 -62.79)	55.83 (43.15 - 59.87)	0.902
48hour conc. (ng/ml)	59.01 (28.55 -64.08)	57.40 (46.66 - 62.23)	0.496
72 hour conc. (ng/ml)	57.80 (38.50 – 60.17)	62.61 (51.39 - 69.85)	0.127
AUC ^{0-∞} µgh/ml	2703.995 (1291.995-4105.664)	2606.700 (1355.393 – 4046.900)	0.052

*P<0.05

Table 7:

Plasma concentrations and area under the curve of lumefantrine in albino rat administered with artemether-lumefantrine (AL) and the combination of artemether- lumefantrine and amodiaquine . (AL+AQ)

Parameter	Lumefantrine conc. in AL	Lumefantrine conc. in ALAQ	P-value
Cmax (ng/ml)	1924 (984 -3865)	2189 (1094 -4339)	0.115
24 hour conc. (ng/ml)	1928 (1653 -2493)	1951 (1786 -2106)	0.64
48hour conc. (ng/ml)	1768 (1587 -1955)	1603 (1564 -1894)	0.331
72 hour conc. (ng/ml)	1912 (1776- 2294)	2189 (1973 – 2383)	0.212
Day 7 conc. (ng/ml)	1670 (1153 -21187)	1683 (774-2591)	0.992
AUC ^{0-∞} µgh/ml	84921.17 (41830.25-127814.60)	88347.50 (45699.50 – 133851.50)	0.031*

*P<0.05

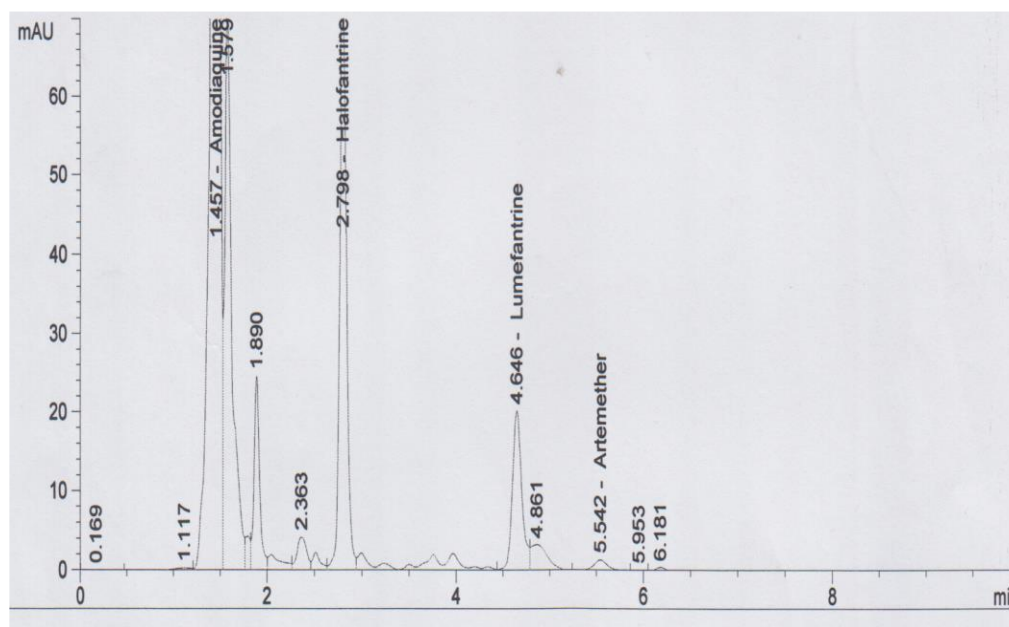


Figure 1: Representative chromatograms for amodiaquine and artemether, lumefantrine and halofantrine (internal standard) at 72 hours.

DISCUSSION

Deployment of a triple artemisinin based combinations as a standard regimen is one of the options to delay the development of resistance to artemisinin and its derivatives (Dondorp *et al.*, 2010). In the present study, the proposed triple combination of artemether-lumefantrine plus amodiaquine was compared to the conventional artemisinin-based combination therapy. The triple combination therapy was very effective with a rapid blood schizonticidal action. Strengthening the conventional ACT with a second long acting partner drug will make the emergence of mutant parasites unlikely as the parasites will be exposed to three drugs instead of two and the drugs are all schizonticidal and have independent mechanisms and sites of action. This complete parasite clearance is expected to have positive implications for prevention of recrudescence because the combination of artemether, which rapidly reduces parasite biomass, with longer-acting lumefantrine that is capable of eliminating residual parasites and amodiaquine with its active metabolites that have a long half-life will achieve parasitologic cure and offer protection against recrudescence infection after therapy (White, 1999a). A rapid and complete clearance time will reduce the probability of the parasite developing resistance because delayed clearance times are

associated with development of resistance (Noedl *et al.*, 2008). The parasites would need to develop mutations at three resistance loci at the same time and this appears very unlikely and may further delay development of resistance (Meshnick, 1998)

Both artemether lumefantrine and amodiaquine are safe and well tolerated (Makanga *et al.*, 2006; Olliaro and Mussano, 2003; Faye *et al.*, 2007). Haematological changes during malaria infection, such as thrombocytopenia and leucocytosis or leucopenia have been extensively documented (Maina *et al.*, 2010; Kotepui *et al.*, 2014). Amodiaquine prophylaxis has been associated with agranulocytosis, neutropenia and hepatitis (Thomas *et al.*, 2004). In this study the triple combination of amodiaquine and artemether-lumefantrine showed significant elevations in total white blood cell counts, red blood cell and platelet counts. These findings suggest that the triple combination may have a good effect on post-treatment haematological recovery. Previous studies have documented anaemia of persistent parasitaemia and showed that repeated exposure to artemether-lumefantrine improved anaemia and thrombocytopenia (Okafor and Nwaiwu, 2001; Bakshi *et al.*, 2000). The observation of significant elevations in alanine aminotransferase and urea calls for more human safety evaluations.

Assessment of cardiovascular safety is important because delay in ventricular depolarisation resulting in prolongation of the QRS complex and worsening malaria-associated orthostatic hypotension have been documented with some antimalarial drugs as well as malaria disease itself (White, 2007; von Seidlein *et al.*, 1998). In this study we observed reductions in heart rate, and blood pressure and this should be explored with ECG studies in humans.

In the present study we evaluated the effect of the triple combination on the plasma concentrations of artemether, lumefantrine and amodiaquine. Co-administration of amodiaquine had no significant effect on plasma concentration of artemether and lumefantrine in the first 48 hours. However, the proposed triple combination showed significant lower artemether levels and the area under the curve at 72hours. The area under the concentration-time curve (AUC) is a measure of drug exposure and the routine measurement of drug concentration at day 7 as part of anti-malarial treatment is recommended (White *et al.*, 2008). The lumefantrine levels in this study were similar in all treatment groups, however we observed a lower area under the curve at day 7. The observed low AUC of lumefantrine could raise concerns that the triple therapy may put the patients at risk of treatment failure because day 7 lumefantrine concentration is the principal determinant of artemether-lumefantrine anti-malarial activity which drives the 28-day cure rate (Singh *et al.*, 2011 ; Ezzet *et al.*, 2000). Previous studies show lumefantrine displays similar pharmacokinetics in the rat as in humans with a long terminal elimination half-life, and therefore a higher post-treatment prophylactic effect (Singh *et al.*, 2011). Lumefantrine, is mainly metabolized by cytochrome P450 3A4 (CYP3A4), to desbutyl lumefantrine (Ezzet *et al.*, 2000) which has a higher in-vitro antiparasitic effect and could reduce the impact of low day 7 plasma concentrations observed.

There were some limitations in this study and the results should be interpreted within the limits of the study design and context which is to determine preliminary data on efficacy, safety and kinetics of the proposed triple artemisinin combination therapy. The efficacy, safety and pharmacokinetics of antimalarials have been documented in infected and uninfected rodents (Olayemi *et al.*, 2012; Osonuga *et al.*, 2012) and humans (Van Vugt *et al.*, 2000; Makanga *et al.*, 2006; White *et al.*, 1999b; Jullien *et al.* 2010). We looked at the hematological profile in uninfected rats to limit blood sampling and improve survival rates in the infected mice. The duration of pharmacokinetic sampling was too short (72hrs). However we determined up to day 7 lumefantrine concentration which is the principal determinant of artemether-lumefantrine anti-malarial activity and drives the 28-day cure rate (Singh *et al.*, 2011; Ezzet *et al.*, 2000). Furthermore chromatographic separation and simultaneous quantification of artemether and lumefantrine is better with reversed-phase HPLC with MS/MS detection. Artemether and the biologically active metabolite dihydroartemisinin are difficult to measure in body fluids. These compounds lack UV or fluorescent chromophores, are thermally labile, and lack functional groups for reliable derivatisation. Post-column derivatisation methods in which artemether and metabolite are converted to a UV-detectable product have yielded variable

results, often with poor reproducibility (Melendez *et al.*, 1991; White, 1994; Navaratnam *et al.*, 1995). HPLC with MS/MS detection was not available and the use of HPLC with UV detector could explain the low artemether levels observed (Sandhya *et al.*, 2015).

In conclusion, the present study in rodents, suggest that the proposed triple artemisinin-based combination will improve efficacy and further delay the emergence of resistance to artemisinins as there was a complete clearance of parasites with the combination of artemether-lumefantrine and amodiaquine after day 12. The lower area under the curve at 72hours with lower artemether and lumefantrine levels observed with the proposed triple artemisinin-based combination (ALAQ) will need further evaluation. We recommend randomized controlled human studies to further evaluate efficacy and safety and pharmacokinetics.

Acknowledgements

We are grateful thanks to Mr Chijioko Micah, and Duncan Ota of the departments of pharmacology and physiology, CMUL for technical assistance.

REFERENCES

- Adeboye JO, Fajonyomi MO Makinde JM and Taiwo OB (1999).** A preliminary study on the Hypotensive activity of *Persea americana* leaf extracts in anaesthetized normotensive rats. *Fitoterapia*. 70:15-20.
- Akinyede A, Akintonwa A, Awodele O, Olayemi S, Oreagba I, Okany C, Aina O, and Akindele S (2013)** Antimalaria action of antiretroviral drugs on *Plasmodium berghei* in mice *American Journal Tropical. Medicine and Hygeine* 88(1):14–19
- American Physiological Society (2002).** Guiding principles for research involving animals and human beings. *American Journal of Physiology* 283:281–283.
- Ariey F, Witkowski B, Amaratunga C, Beghain J, Langlois AC, Khim N, Kim S, Duru V, Bouchier C, Ma L, Lim P, Leang R, Duong S, Sreng S, Suon S, Chuor CM, Bout DM, Menard S, Rogers WO, Genton B, Fandeur T, Miotto O, Ringwald P, Le Bras J, Berry A, Barale JC, Fairhurst RM, Benoit-Vical F, Mercereau-Puijalon O, Menard D (2014,)** A molecular marker of artemisinin resistant *Plasmodium falciparum* malaria. *Nature* 505:50–55.
- Bakshi R, Hermeling-Fritz I, Gathmann I, Alteri E(2000):** An integrated assessment of the clinical safety of artemether-lumefantrine: a new oral fixed-dose combination antimalarial drug. *Tropical medicine and hygiene* 94: 419-424.
- Carrara VI, Lwin KM, Phyo AP, Ashley E, Wiladphaingern J, Sriprawat K, Rijken M, Boel M, McGready R, Proux S, Chu C, Singhasivanon P, White N, Nosten F(2013).** Malaria burden and artemisinin resistance in the mobile and migrant population on the thai-myanmar border, 1999–2011: an observational study. *PLoS Med* 10:e1001398.
- Dondorp AM, Nosten F, Yi P, Das D, Phyo AP, Tarning J, Lwin KM, Ariey F, Hanpithakpong W, Lee SJ, Ringwald P, Silamut K, Imwong M, Chotivanich K, Lim P, Herdman T, An SS, Yeung S, Singhasivanon P, Day NPJ, Lindegardh N, Socheat D, White NJ(2009)** .Reduced in-vivo susceptibility of *Plasmodium falciparum* to artesunate in Western Cambodia. *New England Journal of Medicine* 361:455–467.
- Dondorp A M, Yeung S, White L, Nguon C, Day N PJ, Socheat D & von Seidlein L(2010).** Artemisinin resistance:

- current status and scenarios for containment *Nature Reviews Microbiology*.8: 272-280
- European Medicines Agency (2011)**. Guideline on bioanalytical method validation, Science and Medicinal Health, EMEA/CHMP/EWP/192217/2009.
- Ezzet F, vav Vugt M, Nosten F, Looareesuwan S, White NJ. (2000)**. Pharmacokinetics and pharmacodynamics of lumefantrine (benflumetol) in acute falciparum malaria. *Antimicrob Agents Chemother*.44:697-704.
- Färnert A, Ursing J, Tolfvenstam T, Rono J, Karlsson L, Sparrelid E and Lindegårdh N (2012)**. Artemether-lumefantrine treatment failure despite adequate lumefantrine day 7 concentration in a traveller with Plasmodium falciparum malaria after returning from Tanzania *Malaria Journal* 11:176
- Faye B, Ndiaye JL, Ndiaye D, Dieng Y, Faye O, Gaye O (2007)** Efficacy and tolerability of four antimalarial combinations in the treatment of uncomplicated Plasmodium falciparum malaria in Senegal. *Malaria Journal*. 6:80
- Feng, J., Fitz, Y., Li, Y., Fernandez, M., Cortes Puch, I., Wang, D., Pazniokas, S., Bucher, B., Cui, X., Solomon, S.B. (2015)**. Catheterization of the Carotid Artery and Jugular Vein to Perform Hemodynamic Measures, Infusions and Blood Sampling in a Conscious Rat Model. *Journal of Visualized Experiments*. (95),e51881, doi:10.3791/51881
- Fish E., Brown, M.J , Danneman, P.J and Karas, A.Z . Eds. (2008)**. Anesthesia and Analgesia in Laboratory Animals (Second Edition. American College of Laboratory Animal Medicine Series, Academic Press, San Diego.
- Huang L, Lizak PS , Jayewardene AL , Marzan F, Lee MT , Aweeka FT (2010)**. A Modified Method for Determination of Lumefantrine in Human Plasma by HPLC-UV and Combination of Protein Precipitation and Solid-Phase Extraction: Application to a Pharmacokinetic Study *Analytical Chemistry Insights* .5 :15–23
- Jackson Y, Chappuis F, Loutan L Taylor W (2006)**. Malaria treatment failures after artemisinin-based therapy in three expatriates: could improve manufacturer information help to decrease the risk of treatment failure? *Malaria Journal*, 5:81
- Jullien V, Ogotu B, Juma E, Carn G, Obyono C, Kiechel JR.(2010)**. Population Pharmacokinetics and Pharmacodynamic considerations of Amodiaquine and Desethylamodiaquine in Kenyan Adults with uncomplicated malaria receiving the artesunate- Amodiaquine combination therapy. *Antimicrobial Agents and Chemotherapy* 54(6):2611-2717
- Kay K, Hastings IM (2015)**. Measuring windows of selection for antimalarial drug treatments *Malaria Journal* .14:292 -302.
- Kotepui M, Phunphuech B, Phiwklam N, Chupeerach C and Duangmano S (2014)** Effect of malarial infection on haematological parameters in population near Thailand-Myanmar border. *Malaria Journal* .13:218.
- Lahlou S, Galindo CAB, Leal-Cadoso JH, Fonteles MC, and Durate GP (2002)**. Cardiovascular effects of the essential oil of *Alpinia zerumbet* leaves and its main constituents, Terpinen-4-ol, in rats: Role of the autonomic nervous system. *Planta Medica*, 68: 1097-1102.
- Madara, A. A, Ajayi, J.A, Salawu O. A., and Tijani. A.Y (2010)**. Anti-malarial activity of ethanolic leaf extract of *Piliostigma thonningii* Schum. (Caesalpinaceae) in mice infected with Plasmodium berghei . *African Journal of Biotechnology* 9(23):3475-3480
- Maddela R, Pilli NR, Ravi VB, Adireddy V, Makula A (2015)**. Simultaneous determination of artesunate and amodiaquine in human plasma using lc-ms/ms and its application to a pharmacokinetic study *International Journal of Pharmacy and Pharmaceutical Sciences* ISSN- 0975-1491;7 : 12.
- Makanga M, Premji Z, Falade C, Karbwang J, Mueller EA, Andriano K, Hunt P, De Palacios P (2006)** Efficacy and safety of the six-dose regimen of artemether-lumefantrine in pediatrics with uncomplicated Plasmodium falciparum malaria: a pooled analysis of individual patient data. *American Journal of Tropical Medicine and Hygiene*. 74:991-998.
- Melendez V, Peggins JO, Brewer TG, Theoharides AD (1991)**. Determination of the antimalarial arteether and its deethylated metabolite dihydroartemisinin in plasma by high performance liquid chromatography with reductive electrochemical detection. *Journal of Pharmaceutical Sciences*. 80:132-138.
- Meshnick SR (1998)** Artemisinin antimalarials ;Mechanisms of action and resistance *Tropical Medicine* 58: 13-17
- Mizuno Y, Kato Y, Kudo K, Kano S. (2009)**. First case of treatment failure of artemether-lumefantrine in a Japanese traveler with imported falciparum malaria. *Jpn J Infect Dis* 62:139–141.
- Mutabingwa T, Anthony D, Heller A, Hallett R, Ahmed J, Drakeley C, Greenwood BM, Whitty CJ(2005)** Amodiaquine alone, amodiaquine + sulfadoxine-pyrimethamine, amodiaquine + artesunate, and artemetherlumefantrine for outpatient treatment of malaria in Tanzanian children:a four-arm randomised effectiveness trial. *Lancet*, 365:1474-1480.
- National Academy of Science (2011)**. Guide for the care and use of laboratory animals, 8th ed, Washington:National Academic Press; p 246.
- National Research Council (1985)**. A Guide for the Care and Use of Laboratory Animals. A report of the Institute of Laboratory Animal Resources Committee on Care and Use of Laboratory Animals. National Institutes of Health Publication No. 85-23. Washington D.C: National Academy Press.
- Navaratnam V, Mansor SM, Chin LK, Mordi MN, Asokan M, Nair NK (1995)** Determination of artemether and dihydroartemisinin in blood plasma by highperformance liquid chromatography for application in clinical pharmacological studies. *Journal of Chromatography (B) Biomed Appl* . 669: 289-294
- Noedl H, Se Y, Schaefer K, Smith BL, Socheat D, Fukuda MM(2008)**. Artemisinin Resistance in Cambodia 1 (ARC1) study consortium. Evidence of artemisinin-resistant malaria in western Cambodia. *New England Journal of Medicine* .359:2619-2620.
- Olayemi SO, Arikawe AP, Akinyede A, Oreagba AI , Awodele O, (2012)**. Effect of malarial treatments on Biochemical parameters and plasma pH of mice infected with Plasmodium berghei. *International Journal of Pharmacology*. ISSN; 1811-7775/DOI:10.3923/ijp.2012
- Okafor H.U., Nwaiwu O. (2001)** Anaemia of Persistent Malarial Parasitemia in Nigeria Children. *Journal of Tropical Paediatrics* 47, 271-275.
- Olliaro P, Mussano P(2003)** Amodiaquine for treating malaria .Cochrane database-systematic review . CDOO016
- Osonuga IO, Osonuga OA, Osonuga A, Onadeko AA, Osonuga AA (2012)**. Effect of artemether on hematological parameters of healthy and uninfected adult Wistar rats *Asian Pacific Journal of Tropical Biomedicine*. 2(6): 493-495
- Pecoul B, Sevcik A, Amuasi J, Diap G, Kiechel J (2008)**. The story of ASAQ: the first antimalarial product development partnership success. *Health Partnerships Review*.
- Phyo AP, Nkhoma S, Stepniewska K, Ashley EA, Nair S, McGready R, Ler Moo C, Al-Saai S, Dondorp AM, Lwin KM, Singhasivanon P, Day NP, White NJ, Anderson TJ, Nosten F:**

- (2012.) Emergence of artemisinin-resistant malaria on the western border of Thailand: a longitudinal study. *Lancet* 379:1960–1966.
- Prudhomme O'Meara W, Nekesa-Mangeni J, Steketee R, Greenwood B. (2010).** Changes in the burden of malaria in sub-Saharan Africa. *Lancet Infectious Disease*. 10:545–555.
- Repetto EC, Traverso A, Giacomazzi CG.(2011)** Possible clinical failure of artemether-lumefantrine in an Italian traveler with uncomplicated falciparum malaria. *Mediterr J Hematology Infectious Disease* . 3:e2011041.
- Sandhya SM , Kumar PS , Meena S (2015).** A sensitive liquid chromatographic assay for the simultaneous determination of lumefantrine and artemether in human plasma *Indo American Journal of Pharmaceutical Research*. 2231-6876 : 720
- Shida KK, Lewchalermvongse B, Pang LW (1989).** Plasmodium berghei malaria infection causes increased cardiac output in rats, Rattus rattus. *Exp Parasitol* 68: 253–259.
- Singh WSP, Raju KSR, Nafis A, Puri SK and Jain GK (2011)** Intravenous pharmacokinetics, oral bioavailability, dose proportionality and in situ permeability of anti-malarial lumefantrine in rats, *Malaria Journal*. 10:293
- US DHHS, FDA and CDER.(2001)** Guidance for Industry: Bioanalytical Method Validation. US Department of Health and Human Services, Food and Drug Administration, Center for Drug Evaluation and Research and Center for Veterinary Medicine. Available from: <http://www.fda.gov/cder/guidance/index.htm>.
- Van Vugt M, Looareesuwan S, Wilairatana P, McGready R, Villegas L, Gathmann I, Mull R, Brockman A, White NJ, Nosten F(2000). Artemether-lumefantrine for the treatment of multidrug resistant falciparum malaria. *Transactions of Royal Society of Tropical Medicine and Hygiene*. 94:545-548.
- Virendra K, Dua NC , Gupta VP, Sharma SK, Subbarao W (2004).** Liquid chromatographic determination of amodiaquine in human plasma. *Journal of chromatography* 803, (2) ;25 : 371-374.
- Von Seidlein I, Jaffar S, Greenwood B.(1998)** Prolongation of the QTC interval in African children treated for falciparum malaria. *American journal of Tropical, medicine and hygiene* .56; 494-497
- White N (1994)** Clinical pharmacokinetics and pharmacodynamics of artemisinin and derivatives. *Transactions of the Royal Society of Tropical Medicine and Hygiene* .88 (Suppl. 1): 41-43.
- White NJ (1997).** Assessment of the pharmacodynamic properties of antimalarial drugs in vivo. *Antimicrob Agents Chemother* 41: 1413–1422.
- White NJ (1998).** Preventing antimalarial drug resistance through combinations. *Drug Resistance Updates* .1:3-9.
- White N. (1999a)** Antimalarial drug resistance and combination chemotherapy. *Transactions of Royal Society . London Biological Science* 354:739-749
- White NJ, van Vugt M, Ezzet F (1999b)** Clinical pharmacokinetics and pharmacodynamics of artemether-lumefantrine. *Clinical pharmacokinetics* 37:105-125.
- White NJ and Pongtavornpinyo W (2003).** The de-novo selection of drug resistance in malaria parasites. *Transactions of Royal Society London B* 270:545–554.
- White NJ, Stepniewska K, Barnes K, Price RN, Simpson J(2008).** Simplified antimalarial therapeutic monitoring using the day-7 drug level? *Trends in Parasitology* 24:159-163.
- World Health Organization (2006).** WHO Guidelines for the treatment of malaria. World Health Organization. Geneva, Switzerland
- White NJ (2007).** Cardiotoxicity of antimalarial drugs. *Lancet Infectious Disease*. 7: 549-558.
- World Health Organization,(2009).** Malaria Microscopy Quality Assurance Manual Version 1. Malaria Light Microscopy: Creating a Culture of Quality. Geneva: *World Health Organization*.
- World Health Organization (2010)** .Guidelines for the treatment of malaria (second edition) World Health Organization, Geneva.