

Full Length Research Paper

Disease occurrence and fruit quality of pre-harvest calcium treated red flesh dragon fruit (*Hylocereus polyrhizus*)

Muhd Azlan Abd Ghani, Yahya Awang* and Kamaruzaman Sijam

Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM, Serdang, Selangor, Malaysia.

Accepted 16 November, 2010

A study aiming at increasing Ca content in red flesh dragon fruit (*Hylocereus polyrhizus*) via pre-harvest fruit CaCl₂ spray in relations to postharvest disease occurrence and fruit quality parameters was conducted. From day 7 after anthesis, red flesh dragon fruits (*H. polyrhizus*) were sprayed at weekly interval for four times with different concentration of CaCl₂ (0, 1, 2, 3, and 4 g l⁻¹). Fruit Ca content in fruit peel (DW basis) were markedly increased with the increasing Ca concentration in the applied solution but the Ca application did not affect Ca content in flesh. The compositions of N, P, K and Mg were not affected by the treatment. The severity of anthracnose and brown rot, caused by *Colletotrichum gloeosporioides* and *Monilinia fructicola*, respectively, of artificially wounded fruits was reduced in pre-harvest CaCl₂-treated fruits. The concentrations of soluble solids and titratable acidity in fruit were not affected by the treatment. The firmness of fresh cut fruit was markedly enhanced at higher concentration of CaCl₂. Beneficial effects of increasing Ca in fruit can be seen in the increase of fruit firmness although this did not contribute in enhancement of fruit quality-related parameters. Increased Ca content in treated fruits, together with no effects of treatment on other mineral nutrients, increased the ratio of Ca to other elements and this may contribute directly to the reduction of anthracnose and brown rot severity in CaCl₂-treated fruits.

Key words: Pitaya, anthracnose, brown rot, calcium chloride, fruit quality.

INTRODUCTION

Calcium is a vital macronutrient in plant cycle including fruit development and securing of good fruit quality. Lack of Ca might cause an abnormal growth in fruit and its low mobility into fruit make Ca concentration in fruit decreasing as the fruit grows (Saure, 2005). Fruits with low Ca are generally poor in its quality (Serrano et al., 2002) and become more sensitive to physiological disorders and disease infection (Fallahi et al., 1997; Biggs et al.,

1997; Biggs, 1999; Chardonnet et al., 1999; Elmer et al., 2007). Many studies showed a positive relationship between Ca and fruit shelf life and quality retention (Luna-Guzman and Barret, 2000; Alcaraz et al., 2003).

The ability to retain quality or at least to slowdown the quality degradation process probably become the ultimate goal for any professional that is involve in the postharvest operations and handling of fresh fruits. Quality of fruit may associate with many attributes but in the present study we are only concern on a few aspects of them, that is, disease infection and selected physico-chemical attributes in relations to pre-harvest fruit Ca treatment. Our earlier reports clearly indicated that, dragon fruit is highly susceptible to many diseases (Masyahit et al., 2009) and such diseases have caused significant losses to the growers. Besides diseases,

*Corresponding author. E-mail: yahya_awg@putra.upm.edu.my. Tel: 603-89466917. Fax: 603-89435973.

Abbreviations: SSC, Soluble solids contents; TA, titratable acidity.

degradation of quality is also link to many other quality parameters as seen in the reduction of tissue firmness and moisture content, changes in color and in some other nutritional values.

Fruit tissue softening is associated with many factors, one of them is calcium content of the fruits (Luna-Guzman and Barret, 2000; Aguayo et al., 2007). The role of Ca in developing the resistant of fruit tissue to softening is attributed to the stabilization of membrane systems and the formation of Ca-pectates, which increases rigidity of the middle lamella. Ca also makes the tissue become more resistant to cell wall degradation enzymes such as polygalacturonase (PG) and pectine-methylesterase (PME) (Siti Hajar et al., 2010; Manganaris et al., 2005). Positive relationships between fruit Ca and firmness retention were observed in the existence of many type of fruits (Saftner et al., 2003; Omaira and Karima, 2007).

Loss of calcium from cell wall and middle lamella of fruit occurs during the maturing process (Cutting et al., 1992) and causes fruit softening (Stow, 1993), while increasing fruit Ca content through Ca application which reduces Ca loss. Thus, it increases the fruit firmness (Gerasopoulos et al., 1996; Singh et al., 2007), possibly by several mechanisms including an increase in Ca-pectin bond in middle lamella (Grant et al., 1973), a reducing fruit respiration (Eaks, 1985) and maintenance of cell turgor potentials (Mignani et al., 1995).

Since the mobility of calcium in plants is low, Ca root uptake from soil-applied fertilizer is less effective in increasing Ca content in fruit. Direct application of liquid source of Ca on leaf and fruit may offer an alternative solution. In this study, we were aiming to increase Ca content in red flesh dragon fruit (*H. polyrhizus*) via pre-harvest fruit CaCl_2 spray and to examine its effects on postharvest disease occurrence and fruit quality parameters. As the absorption of Ca into fruit may interact with the uptake and translocation of other nutritional elements, thus affecting their balance which could have impacts on fruit quality, the fruit contents of N, P, K and Mg were also considered in the study.

MATERIALS AND METHODS

Pre-harvest calcium application

The study was conducted at a two-year old commercial dragon fruit farm at Pajam in Negeri Sembilan, Malaysia. Prior to pre-harvest Ca treatment, four well developed flowers on four different plants were tagged at one day after anthesis and their development were closely monitored. The pre-harvest fruit CaCl_2 treatment began at 7 DAA. The fruits were sprayed till drip (approximately 20 s) at 0900 to 1000 h with the respective treatment of five different concentrations of CaCl_2 : 0-distilled water, 1, 2, 3 and 4 g l^{-1} Ca (4 fruits/plot). 5% (v/v) of a non-ionic wetting agent was added into the CaCl_2 solution to increase Ca retention on the fruit skin. During the course of experiment, the fruits were sprayed four times, at day 7, 14, 21 and 28 after anthesis. The fruits were wrapped in clear perforated plastic bags after every spray and re-opened again before each CaCl_2 application. The fruits were harvested at fully ripened stage

(33 ± 2 DAA) and stored overnight at 13°C before being used for the study.

Disease occurrence

Sample fruits (4 fruits/plot x five treatments) were artificially wounded with a cork borer (0.5 cm diameter) and inoculated with 1×10^6 spores ml^{-1} of *C. gloeosporioides* and *M. fructicola* (two fruits for each fungus/plot). The controlled fruits were 'inoculated' with distilled water and placed in moisturized plastic trays, covered with cling-films and incubated for three days at 22°C after which the disease incidence (% of fruits infected) and severity of the infections (size of lesions of the infected fruits) were evaluated.

Fruit N, P, K, Ca and Mg contents

The inoculated fruits were divided into flesh and peel portions, cut into small pieces and dried at 60°C in an air-circulating oven and finely ground once dried. 0.25 g of the fruit peel and flesh were digested in 5 ml H_2SO_4 on hot plate at 450°C in a fume chamber for 7 min. 10 ml H_2O_2 was added into the mixtures and the heating was continued for another 4 min. The solution mixtures were made-up to 100 ml with distilled water. N and P contents in the samples were determined using an auto-analyzer (LACHART Instruments, Model Quikchem IC + FIA 8000 Series, Milwaukee, USA) while K, Ca and Mg were measured using an atomic absorption spectrophotometer (Perkin Elmer, Model AAS 3110, Palo Alto, California, USA).

Fruit quality

To examine the impact of CaCl_2 treatment and disease infection on fruit quality, fruit firmness, soluble solids contents (SSC) and titratable acidity (TA) of the inoculated fruits were measured. The firmness of the whole fruit was determined using a texture analyzer (Instron Universal Testing machine, Model 5543, Instron Corporation, USA) by measuring the maximum penetration force required during peel tissue breakage, using a 5 mm diameter flat probe. The measurement of firmness was done at two locations for each fruit at 2.0 cm away from the point of fungus inoculation. SSC of the inoculated fruits was determined using a digital refractometer meter (Model N- α , Atago, Japan) by squeezing the fruit flesh onto the meter's prism. TA was measured using diluted fruit juice (1 juice to 4 distilled water) prepared using the same fruits as for the SSC measurement. 10 ml of the juice for each treatment (with three determinations of each) were titrated with 0.1N NaOH to pH 8.1. The TA was calculated and expressed as percentage of citric acid. The pH of the same fruit juice was also measured using a glass electrode pH meter.

The experiment was conducted in a completely randomized design with four replications. Data were subjected to analysis of variance (ANOVA) and comparison of means was performed using Tukey HSD with SPSS (Version 13).

RESULTS

Effects on disease occurrence

Regardless of CaCl_2 concentration in the applied solution, fruit sprays did not reduce the incidence percentage of disease infection. After three days of incubation period, all fruits were inoculated with 10^6 spores ml^{-1} of both fungi exhibited 100% infections. Increasing CaCl_2 concentra-

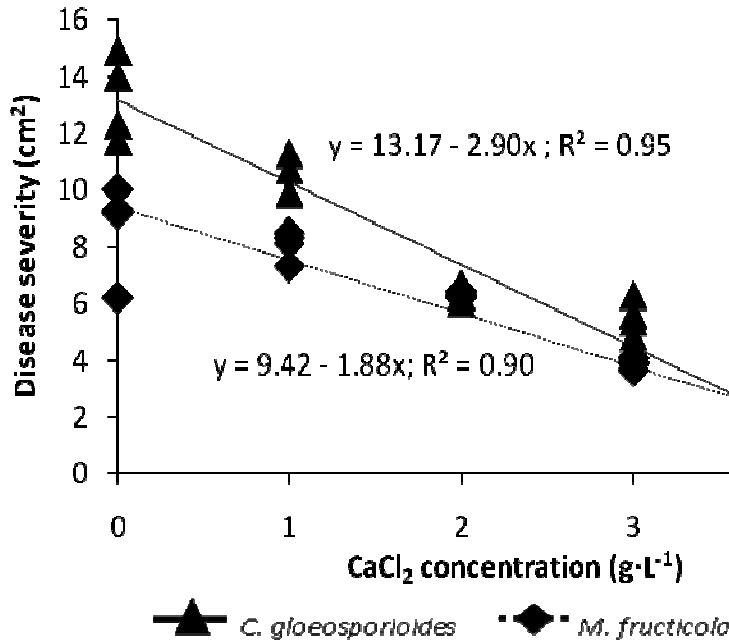


Figure 1. Relationship between CaCl₂ concentration and disease severity of pre-harvest CaCl₂ treated fruit at three days after inoculated with *C. gloeosporioides* and *M. fructicola* spores (10⁶ spores per ml⁻¹).

tion in the spray solution linearly reduced the severity of disease symptom, as seen in the reduction of lesion size at high CaCl₂ concentration (Figure 1). The diameter of lesion was reduced from 8.3 to 2.0 cm² in fruits inoculated with *C. gloeosporioides* as the CaCl₂ concentration increased from 0 to 4 g l⁻¹. The corresponding values for *M. fructicola* were 13.4 and 1.8 cm². Disease severity caused by *C. gloeosporioides* and *M. fructicola* was negatively correlated with Ca in peel ($r = 0.91$, $p \leq 0.01$) but it has no correlation with Ca concentration ($r = 0.23$) in flesh (Figure 2).

Effects on fruit mineral contents

Fruit Ca content increased markedly with the increasing concentration of CaCl₂ applied (Figure 3). However, the effects of treatment on Ca content in fruit tissues were more apparent in peel compared to the flesh. The peel of the fruits treated with Ca concentration at 4 g l⁻¹ had the highest concentration of Ca (83.56 mg per 100 g⁻¹), followed by 3 g l⁻¹ (66.79 mg per 100 g⁻¹), 2 g l⁻¹ (28.62 mg·100 g⁻¹), 1 g l⁻¹ (20.62 mg per 100 g⁻¹) and 0 g l⁻¹ (22.77 mg per 100 g⁻¹). Although not significant, Ca content in the flesh was also enhanced at higher CaCl₂ concentration which is ranged from 6.68 to 10.53 mg per 100 g⁻¹. There was no significant correlation existence between Ca in fruit flesh and in fruit peel (Figure 4). Results in Table 1 show that, the contents of N, P, K and Mg in peel and flesh were not significantly affected with CaCl₂

concentration.

Effects on fruit quality

Fruit firmness was progressively increased at higher CaCl₂ regardless of fungal treatments (Figure 5). The firmness level of the non-treated fruit ranged from 22.12 to 24.81 N but the firmness of the fruits inoculated with *C. gloeosporioides* and *M. fructicola* ranged from 14.95 to 23.88 N and 14.95 to 23.81 N, respectively. The firmness of the non-inoculated fruit increased as the CaCl₂ concentration increased from 0 to 4 g l⁻¹, but the effects were not significantly differed among them. Overall, fruit firmness was positively correlated to Ca concentration in peel, regardless of inoculation types ($r = 0.93$). There was no correlation existed between firmness and Ca concentration in flesh (data not shown).

Results in Table 2 show that the pH of fruit was not significantly affected by CaCl₂ treatment. In contrast, pH of fruit inoculated with *C. gloeosporioides* and *M. fructicola* decreased significantly compared to the controlled fruit. There was no interaction effect between CaCl₂ concentration and inoculums type on pH. The pH of fruit was correlated positively with severity of infection ($r = 0.80$). The negative correlation was also observed between pH and Ca concentration in peel ($r = -0.87$).

Soluble solids contents of fruit inoculated remains constant after experiencing CaCl₂ treatment and SSC was significantly lower ($p \leq 0.05$) in fruit inoculated with

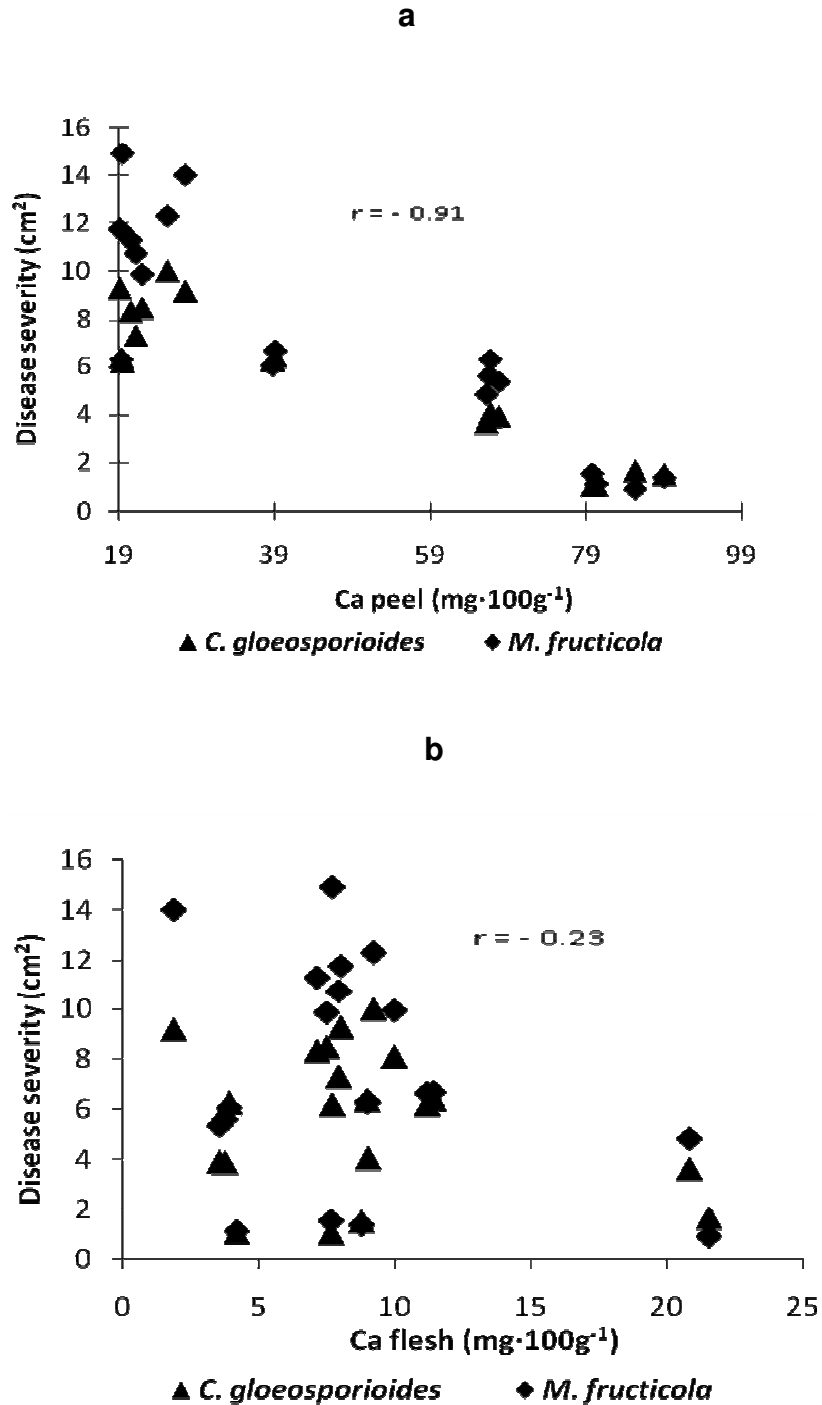


Figure 2. Correlation between disease severity and Ca concentration in peel (a) and disease severity and Ca concentration in flesh (b) of pre-harvest CaCl_2 treated fruit at three days after inoculated with *C. gloeosporioides* and *M. fructicola* spores (10^6 spores/ml).

either *C. gloeosporioides* or *M. fructicola* compared to the non-inoculated fruit (Table 2). Results of correlation analysis show that SSC was negatively correlated to severity of infection ($r = -0.84$) and positively correlated

with Ca concentration in peel ($r = 0.88$), TA of the inoculated fruit was not influenced by the treatment. In fruit inoculated with *C. gloeosporioides*, *M. fructicola* and TA increased significantly with CaCl_2 concentration

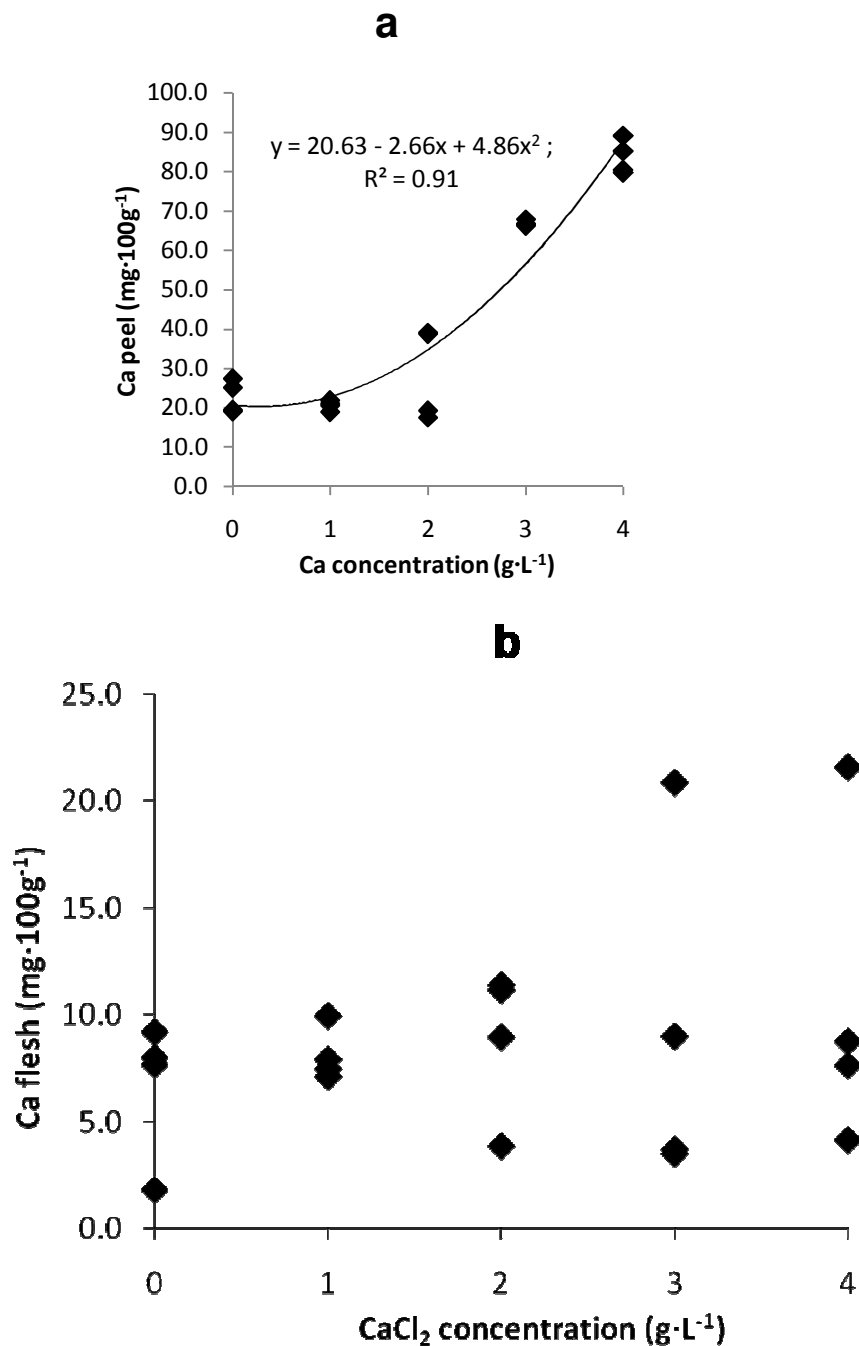


Figure 3. Relationship between CaCl₂ concentrations of the applied solution with Ca content of peel (a) and flesh (b) of dragon fruit.

compared to the non-inoculated fruits but TA was negatively correlated with severity of infection ($r = -0.76$).

DISCUSSION

Regardless of its concentration in the applied solution, CaCl₂ fruit sprays did not reduce the incidence percen-

tage of disease infection. The results recorded here were paralleled with studies involving other fruit species (For example, Kiwi - Gerasopoulos et al., 1996; Strawberry-Hernandez-munoz et al., 2006; Nectarine-Vasilakakis et al., 2006; Peach-Elmer et al., 2007). Increasing CaCl₂ concentration in the spray solution however has reduced the severity of symptoms, as seen in the reduction of lesion area. The roles of Ca in reducing disease severity

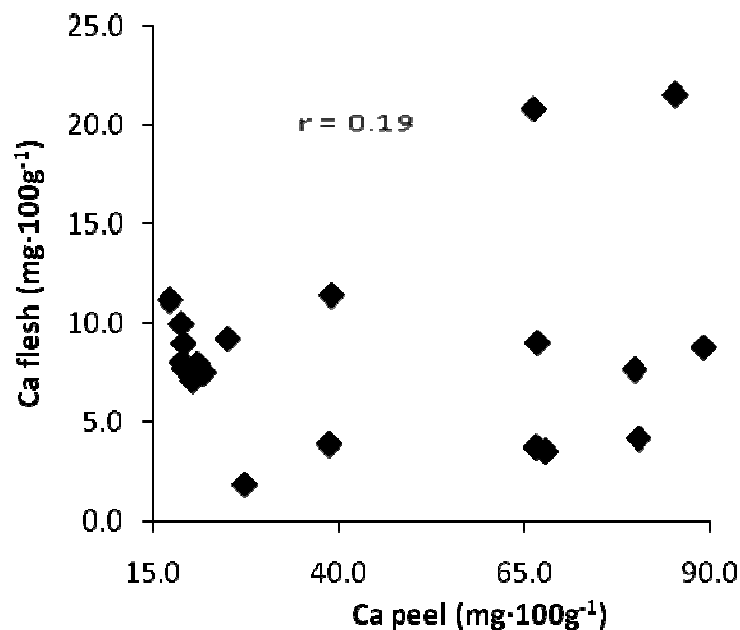


Figure 4. Correlation between Ca peel and Ca flesh of CaCl_2 treated fruit.

Table 1. Effects of pre-harvest CaCl_2 treatment on mineral concentrations in flesh and peel of *H. polyrhizus*

Part of fruit	CaCl_2 treatment (g l^{-1})	Nutrient concentration ($\text{mg } 100\text{g}^{-1}$)			
		N	P	K	Mg
Peel	0 (control)	126.90	10.28	243.30	25.10
	1	100.80	10.81	251.17	25.36
	2	97.10	12.37	250.83	25.36
	3	95.50	10.76	249.97	25.09
	4	100.70	10.99	243.91	26.16
Flesh	0 (control)	208.90	19.53	148.68	20.19
	1	234.40	21.76	144.74	21.19
	2	238.40	21.49	147.47	20.68
	3	173.50	17.38	173.43	18.59
	4	208.90	18.80	157.33	20.57
Tukey HSD _{0.05}		55.40	4.50	25.30	1.20

HSD, Honestly significant difference.

can be discussed in two ways; by direct effect of Ca to fungal itself or by its indirect effect. The direct effect of Ca in the reduction of disease severity could begin at the spore germination stage. Droby et al. (1997) reported that, calcium reduced the germination of *Penicillium digitatum* spores on Ca treated grapefruit thus reducing the severity of infection. Maintenance of low basal concentrations of internal calcium is essential for normal cell functions of organisms and the inability of cells to regulate calcium may affect the organism's normal growth (Biggs, 1999). Increasing CaCl_2 concentration might have increased free Ca in fruit tissues thus ele-

vates the free calcium in the cytosol of fungus and hence inhibit the growth of germ tube and mycelial growth of *C. gloeosporioides* (Biggs et al., 1997; Biggs, 1999). The same mechanism might apply in Ca treated fruit inoculated with *M. fructicola*. Ca also was reported to have the ability to inhibit the activity of pectolytic enzymes secreted by fungus (Droby et al., 1997), thus reduce disease severity in fruit treated with high Ca concentration.

Indirect effect of Ca in reducing disease severity could also relate to the role of Ca on fruit cell wall integrity. Calcium in the cell wall has a role in fruit texture (Quiles

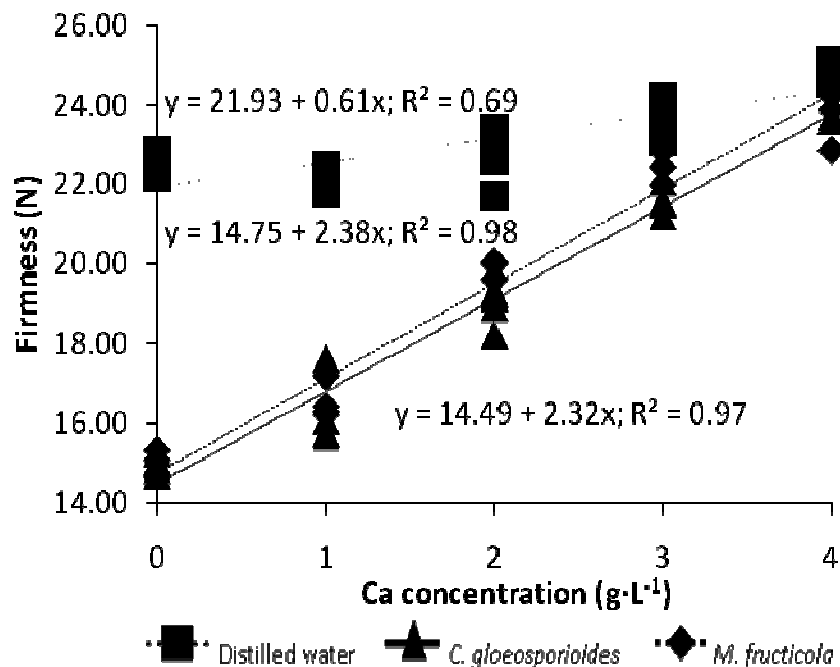


Figure 5. Relationship between CaCl_2 treatment and fruit firmness after three days of incubation.

Table 2. Effects of pre-harvest CaCl_2 treatment on fruit pH, soluble solid content and titratable acidity of red flesh dragon fruits infected by *C. gloeosporioides* and *M. fructicola* after three days of incubation.

Inoculum (10^6 spores. ml^{-1})	CaCl_2 treatment (gl^{-1})	Quality parameter		
		pH	SSC (%)	TA (%)
Sterilized water	0 (control)	5.43	9.26	0.10
	1	5.24	9.80	0.13
	2	5.28	10.12	0.12
	3	5.31	9.88	0.11
	4	5.31	9.77	0.14
<i>C. gloeosporioides</i>	0 (control)	5.76	6.16	0.05
	1	5.68	7.05	0.06
	2	5.60	8.21	0.07
	3	5.47	9.45	0.09
	4	5.36	9.97	0.12
<i>M. fructicola</i>	0 (control)	5.76	6.99	0.06
	1	5.68	7.21	0.06
	2	5.55	8.94	0.09
	3	5.51	9.64	0.09
	4	5.45	9.93	0.12
F-test				
Inoculum		**	**	**
Calcium concentration		ns	ns	ns
Interaction		ns	ns	ns
Tukey HSD _{pooled (0.05)}		0.26	0.98	0.04

SSC, Soluble solids contents; TA, titratable acidity; ns, * and ** indicate non-significant, significant at $P < 0.05$ and $P < 0.01$, respectively.

et al., 2007) but Ca content in the fruit could have reduced through the maturing process and resulting in the lack of Ca at the end of fruit maturity period. Deficiency of Ca would increase cell membrane permeability thus permits ions to escape and lead to the breakdown of intercellular compartmentalization, as well as the escapes of enzymes, such as polygalacturonase and pectin methylesterase which accelerates fruit ripening and softening processes (Deytieux-Belleau et al., 2008). This predisposes fruit to fungal attack. Reduction in fungal infection in Ca treated fruits as observed here could also be attributed to the increase of cell wall-bound Ca (Chardonnet et al., 1999) which stabilize cell wall structure and protect it from pectolytic enzymes produced by the fungi (Conway et al., 1994). The roles of Ca treatment in increasing cell wall integrity thus protect the wall from fungal pectolytic enzymes that have been proposed and discussed in many studies (Fallahi et al., 1997; Elmer et al., 2007; Singh et al., 2007). In another perspective, plants are known to produce phytoalexin and phenolics compound as a self-defense mechanism and Ca application increases the synthesis of these compounds thus inhibit the activity of fungal pectolytic enzymes (Kohle et al., 1985).

Increased Ca content in fruit and specifically in fruit peel following pre-harvest CaCl₂ application recorded here is in agreement with the results of studies reported by Dris (1998) for apples, Elmer et al. (2007) for peaches and Singh et al. (2007) for strawberry. As fruit is an organ with high metabolic rate and dependent on continuous supply of Ca, it is highly demanded during fruit development and rapid fruit growth causes a dilution of Ca in fruit tissues (Saure, 2005). Fruit Ca spray creates a concentration gradient of calcium between exogenous and endogenous portion of fruit, resulted in passive uptake of Ca into fruit (Alcaraz et al., 2003). In its early development, Ca is evenly distributed but when the fruit is becoming more mature, Ca will be transported to peel of fruit (Dris, 1998), making the concentration of Ca in peel higher than that in the flesh.

The concentration of N, P, K and Mg contents in fruit were not affected by the treatment. Such results would give a good indication on the positive effects of increasing Ca in the fruit tissue as this did not affect the balance of nutrient composition. This leads to higher ratios of Ca to other elements, thus elevating the possible benefit of the CaCl₂ treatment. Higher Ca:N ratio in fruit tissue for example could reduce fruit physiological disorders (Ferguson and Boyd, 2002).

Lack of interaction between the content Ca and other mineral nutrients in fruit could be attributed by differences in the regulation of uptake and absorption process for Ca when calcium is directly applied on fruits, whereby calcium enters the fruit tissue mainly via stomata, lenticels and fruit cleavage (Harker and Ferguson, 1988; Eichert and Burkhardt, 2001; Schlegel and Schonherr, 2002; Paul and Srivastava, 2006). In contrast to soil

applied calcium, the element is absorbed and moved radially across the roots before being transported upward via the xylem. In the process, Ca²⁺ may have to compete with other ions such as K⁺ and NH₄⁺ at the site of absorption and with Mn⁺ in upward translocation (Mengel et al., 2001). However, such competition does not occur when Ca is directly sprayed on fruit surfaces.

The infected fruits were observed to have high pH. Upon infection, the production and activity of cell wall degrading enzymes secreted by fungus could be influenced by host pH (Prusky et al., 2001). Fungus secretes NH₃ in order to activate the enzymes thus increase the pH of host tissue (Drori et al., 2003). Although not significant, CaCl₂ application seems to reduce to fruit pH, possibly related to the reduction of fungal activity in the treated fruits.

Generally, the fruit SSC and TA are not directly influenced by CaCl₂ application. TA naturally decreases during ripening (Park et al., 2006) but fungal infection accelerates the decrease. Working with strawberry, Wang and Galletta (2002) reported that anthracnose reduced the fruit SSC and TA, which may link to the utilization of the carbon (C) skeleton in sugars and organic acids by the fungus (Steinberg et al., 1999). For this reason, infected fruit would eventually have lower sugar and acid content, as recorded in this study.

Conclusion

CaCl₂ application as liquid spray at pre-harvest stage elevated Ca content in fruit. Beneficial effects of increasing calcium in fruit can be seen in the increase of fruit firmness although this did not contribute in enhancement of fruit quality-related parameters. Except for fruit Ca content, the composition of other mineral nutrients measured were not affected by the treatment, thus increasing the ratio of Ca to other elements. This may contribute directly to the reduction of anthracnose and brown rot severity observed in Ca-treated fruits.

REFERENCES

- Aguayo E, Jansasithorn R, Kader, AA (2007). Combined effects of 1-methylcyclopropene, calcium chloride dip, and/or atmospheric modification on quality changes in fresh-cut strawberries. *Postharvest Biol. Technol.* 40: 269-278.
- Alcaraz CF, Alcaraz-Lopez C, Botia M, Riquelme F (2003). Effects of foliar sprays containing calcium, magnesium, and titanium on plum (*Prunus domestica* L.) fruit quality. *J. Plant Physiol.* 160: 1442-1446.
- Biggs AR (1999). Effects of calcium salts on apple bitter rot caused by two *Colletotrichum* spp. *Plant Dis.* 83: 1001-1005.
- Biggs AR, El-Kholi MM, El-Neshawy S, Nickerson R (1997). Effects of calcium salts on growth, polygalacturonase activity and infection of peach fruit by *Monilinia fructicola*. *Plant Dis.* 81: 399-403.
- Chardonnet CO, Sams CE, Conway WS (1999). Calcium effect on the mycelia cell walls of *Botrytis cinerea*. *Phytochemistry*, 52: 967-973.
- Conway WS, Sams CE, Kelman A (1994). Enhancing the natural resistance of plant tissues to postharvest disease through calcium applications. *Hort. Sci.* 29: 751-754.

- Cutting JGM, Wolstenholme BN, Hardy J (1992). Increasing relative maturity alters the base mineral composition and phenolic concentration of avocado fruit. *J. Hort. Sci.* 67(6): 761-768.
- Dris R (1998). Effect of preharvest calcium treatments on postharvest quality of apples grown in Finland. Helsinki, Finland, University of Helsinki, Publication 34.
- Droby S, Wisniewski ME, Cohen L, Weiss B, Touitou D, Eilam Y, Chalutz E (1997). Influence of CaCl_2 on *Penicillium digitatum*, grapefruit peel tissue, and biocontrol activity of *Pichis quilliermondii*. *Phytopathol.* 87: 310-315.
- Drori N, Kramer-Haimovich H, Rollins J, Dinooor A, Okon Y, Pines O, Prusky D (2003). External pH and Nitrogen source affect secretion of pectatae lyase by *Colletotrichum gloeosporioides*. *Appl. Environ. Microbiol.* 69: 3258-3262.
- Deytioux-Belleau C, Vallet A, Doneche B, Geny L (2008). Pectin methylesterase and polygalacturonase in the developing grape skin. *Plant Physiol. Biochem.* 46: 638-646.
- Eaks IL (1985). Effect of calcium on ripening, respiratory rate. Ethylene production, and quality of avocado fruit. *J. Am. Soc. Hort. Sci.* 110(2): 145-148.
- Eichert T, Burkhardt J (2001). Quantification of stomatal uptake of ionic solutes using a new model system. *J. Exp. Bot.* 52: 771-781.
- Elmer PAG, Spiers TM, Wood PN (2007). Effects of pre-harvest foliar calcium sprays on fruit calcium levels and brown rot of peaches. *Crop Prot.* 26: 11-18.
- Fallahi E, Conway WS, Hickey KD, Sams CE (1997). The role of calcium and nitrogen in post harvest quality and disease resistance of apples. *Hort. Sci.* 32: 831-835.
- Ferguson IB, Boyd, LM (2002). Inorganic nutrients and fruit quality. In: *Fruit Quality and its Biological Basis* (Knee M ed.). Sheffield Academic Press, Sheffield, UK. pp. 17-45.
- Gerasopoulos D, Chouliaras V, Lionakis S (1996). Effects of preharvest calcium chloride sprays on maturity and storability of Hayward kiwifruit. *Postharvest Biol. Technol.* 7: 65-72.
- Grant GT, Morris ER, Rees DA, Smith PJC, Thom D (1973). Biological interactions between polysaccharides and divalent cations: The egg-box model. *FEBS Lett.* 32: 195-198.
- Harker FR, Ferguson IB (1988). Transport of calcium across cuticles isolated from apple fruit. *Scientia Hort.* 36: 205-217.
- Hernandez-munoz P, Almenar E, Ocio MJ, Gavara R (2006). Effects of calcium dips and chitosan coatings on postharvest life of strawberries (*Fragaria-ananassa*). *Postharvest Biol. Technol.* 39: 247-253.
- Kohle H, Jeblick W, Poten F, Blaschek W, Kauss H (1985). Chitosan-elicited callose synthesis in soybean cells as a Ca^{2+} -dependent process. *Plant Physiol.* 77: 544-551.
- Luna-Guzman I, Barrett DM (2000). Comparison of calcium chloride and calcium lactate effectiveness in maintaining shelf stability and quality of fresh-cut cantaloupes. *Postharvest Biol. Technol.* 19: 61-72.
- Manganaris GA, Vasilakakis M, Diamantidis G, Mignani I (2005). The effect of postharvest calcium application on tissue calcium concentration, quality attributes, incidence of flesh browning and cell wall physicochemical aspects of peach fruits. *J. Food Chem.* 100: 1385-1392.
- Masyahit M, Kamaruzaman S, Yahya A, Ghazali M (2009). First report of the occurrence of the anthracnose disease caused by *Colletotrichum gloeosporioides* (Penz.) Penz. & Sacc. on dragon fruit (*Hylocereus spp.*) in Peninsular Malaysia. *Am. J. Appl. Sci.* 6: 902-912.
- Mengel K, Kirkby EA, Kosegarten H, Appel T (2001). *Principles of Plant Nutrition* (5th ed.). Dordrecht, Netherlands: Kluwer Academic Publishers.
- Mignani I, Grane LC, Benane R, Stotz HU, Li CY, Shackel LA, Labaritch JM (1995). The effects of GA_3 and divalent cations on aspects of pectin metabolism and tissue softening in ripening tomato pericarp. *Physiologia Planta*, 93: 108-115.
- Omaira MH, Karima HEH (2007). Quality improvement and storability of apple cv. Anna by pre-harvest application of boric acid and calcium chloride. *J. Agric. Biol. Sci.* 3: 176-183.
- Park YS, Jung ST, Gorinstein S (2006). Ethylene treatment of 'Hayward' kiwifruits (*Actinidia deliciosa*) during ripening and its influence on ethylene biosynthesis and antioxidant activity. *Scientia Hort.* 108: 22-28.
- Paul V, Srivastava GC (2006). Role of surface morphology in determining the ripening behaviour of tomato (*Lycopersicon esculentum* Mill.) fruits. *Scientia Hort.* 110: 84-92.
- Prusky D, McEvoy JL, Levrentz B, Conway WS (2001). Local modulation of host pH by *Colletotrichum* species as a mechanism to increase virulence. *Mol. Plant-Microbe Interactions*, 14: 1105-1113.
- Quiles A, Hernando I, Perez-Munuera I, Llorca E, Larrea V, Lluch MA (2007). The effect of calcium and cellular permeabilization on the structure of the parenchyma of the osmotic dehydrated 'Granny Smith' apple. *J. Sci. Food Agric.* 84: 1765-1770.
- Saftner RA, Bai J, Abbott JA, Lee YS (2003). Sanitary dips with calcium propionate, calcium chloride or a calcium amino acid chelate maintain quality and shelf stability of fresh-cut honeydew chunks. *Postharvest Biol. Technol.* 29: 257-269.
- Saure MC (2005). Calcium translocation to fleshy fruit: its mechanism and endogenous control. *Scientia Hort.* 105: 65-89.
- Schlegel TK, Schonherr J (2002). Selective permeability of cuticles over stomata and trichomes to calcium chloride. *Acta Hort.* 549: 91-96.
- Serrano M, Amoros A, Pretel MT, Martinez-Madrid MC, Madrid R, Romojaro F (2002). Effect of calcium deficiency on melon (*Cucumis melo* L.) texture and glassiness incidence during ripening. *Food Sci. Technol. Int.* 8: 147-154.
- Siti Hajar C, Yahya A, Mahmud TMM (2010). Cell wall enzymes activities and quality of calcium treated fresh-cut red flesh dragon fruit. *Int. J. Agric. Biol.* 12: 713-718.
- Singh R, Sharma RR, Tyagi SK (2007). Pre harvest foliar application of calcium and boron influences physiological disorders, fruit yield and quality of strawberry (*Fragaria x ananassa* Duch.). *Scientia Hort.* 112: 215-220.
- Steinberg C, Whipps JM, Wood DA, Fenlon J, Alabouvette C (1999). Effects of nutritional sources on growth of one non-pathogenic strain and four strains of *Fusarium oxysporium* pathogenic on tomato. *Mycol. Res.* 103: 1210-1216.
- Stow J (1993). Effect of calcium ions on apple fruit softening during storage and ripening. *Postharvest Biol. and Technol.* 3: 1-9.
- Vasilakakis M, Manganaris GA, Diamantidis G, Mignani I (2006). Effect of in-season calcium applications on cell wall physicochemical properties of nectarine fruit (*Prunus persica* var. nectarine Ait. Maxim) after harvest or cold storage. *J. Sci. Food Agric.* 86: 2597-2602.
- Wang SY, Galletta GJ (2002). Compositional change in *Colletotrichum* (anthracnose) infected strawberry fruit. *Acta Hort.* 567: 815-819.