

*Full Length Research Paper*

# The effects of valproic acid on renal corpuscle of pregnant rats and protective role of folic acid and vitamin E

Ayfer Aktaş\*, Yusuf Nergiz, Murat Akkuş and Yasemin Nasır

Medical Faculty, Departments of Histology and Embryology, Dicle University, 21280 Diyarbakır, Turkey.

Accepted 18 June, 2010

We aimed to investigate the potential harmful effects of valproic acid (VPA), a widely used anticonvulsant in child delivery, and the protective effects of vitamin E (Vit E) and folic acid (FA) on kidney. Sodium valproate (400 mg/kg), folic acid (400 mg/kg) and vitamin E (250 mg/kg) were administered to rats on each of gestation days, 8<sup>th</sup>, 9<sup>th</sup>, and 10<sup>th</sup>. The rats were sacrificed on the 20<sup>th</sup> day of pregnancy. With thin sections of kidney biopsies, they were stained with uranyl acetate-lead citrate and examined under transmission electron microscope. The animals were divided into four groups randomly: control, VPA, VPA+FA and VPA+Vit E groups. In each group, drug procedure, surgical procedure and histological methods were performed. The histopathological findings of control group was normal. In VPA group, it showed degenerative changes especially in renal glomerular basal membrane and foot process. Both VPA+FA and VPA+Vit E groups exhibited similar ultrastructural changes and had almost the normal structure. Administration of single doses of SV (400 mg/kg) resulted in degenerative changes on kidney at ultrastructural level. Administration of FA and Vit E had a protective effect by preventing the degenerative changes to a certain degree. The aim of the present study is to examine histopathologic changes which may occur in a high risk experimental model after the administration of valproic acid. In addition, protective roles of the administration of folic acid and vitamin E are assessed.

**Key words:** Folic acid, kidney, rat, valproic acid, vitamin E.

## INTRODUCTION

Valproic acid (VPA, 2-propylpentanoic acid or dipropylacetic acid) is a broad spectrum anticonvulsant which has proved useful especially in the treatment of primarily generalized epilepsies (Loscher and Nau, 1983). This drug is applied in the form of sodium valproate (SV). VPA and its derivatives are widely used in treatment of various seizure disorders and some psychiatric conditions. The majority of childbearing women exposed to VPA deliver healthy infants, however, the risk of congenital malformation in the developing fetus is 2 or 3-fold higher (4-6%) than the risk in general population (2%)

(2%) (Jamsheer et al., 2008; Yerby, 2003). First reports on possible teratogenic activity of VPA appeared in early 1980s (Dalens et al., 1980; Gomez, 1981).

Many animal studies were carried out in order to mimic the effects of VPA on the human embryo and elucidate the mechanism of its teratogenic action (Wagner et al., 2006; Arndt et al., 2005; Holmes et al., 2005; Emmanouil-Nikoloussi et al., 2004). In most animals the drug was teratogenic but the effective teratogenic doses differed widely. VPA induced malformations of multiple organs in mice, rats, and gerbils, renal and skeletal defects in rabbits, neural tube defects in mice and hamsters, craniofacial and appendicular skeletal defects in primates and behavioral deficits resembling autism in mice and rats (Ingram et al., 2000; Emmanouil-Nikoloussi et al., 2004; Rodier et al., 1996; Binkerd et al., 1988; Hendrickx et al., 1988).

In this study, the effects of sodium valproate in rat renal

\*Corresponding author E-mail: [aaktas@dicle.edu.tr](mailto:aaktas@dicle.edu.tr).

corpuscle and the possible protective roles of folic acid and vitamin E in the ultrastructural examination were investigated.

## MATERIALS AND METHODS

### Animals

The study was conducted in accordance with the National Institutes of Health guidelines for the use of experimental animals. Twenty-four (24) adult female Wistar-Albino rats weighing 200 to 250 g, obtained from Experimental Research Institute of Dicle University (DÜSAM) were used. Rats were maintained on a 12 h light/dark cycle at  $21 \pm 1^\circ\text{C}$  and  $50 \pm 10\%$  humidity. The pregnant rats ( $n = 24$ ) were randomly divided into four groups (18 pregnant rats in treatment groups, and 6 pregnant rats in control group). Each pregnant rat was put into an individual cage.

### Experimental protocol

Control group ( $n=6$ ): The first group of rats was used as control. This group was not given the drug. These rats were only fed with standard laboratory chow and tap water.

VPA group ( $n=6$ ): Once per day on days 8<sup>th</sup>, 9<sup>th</sup> and 10<sup>th</sup> of gestation, sodium valproate (Sodium valproate, Sigma P 4543) was administered subcutaneously into a loose fold of skin on the back leg at the dose of 400 mg/kg.

VPA + FA group ( $n=6$ ): Once per day on days 8<sup>th</sup>, 9<sup>th</sup> and 10<sup>th</sup> of gestation, sodium valproate was administered subcutaneously into a loose fold of skin on the back leg at the dose of 400 mg/kg and FA (folic acid, pteryoglutamic acid vit M, Sigma F 8798) was given (400 µg) ordinarily in drinking water per day during pregnancy.

VPA + Vit E group ( $n=6$ ): Once per day on days 8<sup>th</sup>, 9<sup>th</sup> and 10<sup>th</sup> of gestation, sodium valproate was administered subcutaneously into a loose fold of skin on the back leg at the dose of 400 mg/kg. One hour before the injection of VPA, vitamin E ( $\alpha$ -tocopherol, Sigma T-3251) was administered by gastric intubation to dams at a dose of 250 mg/kg (Vorhees, 1987).

### Surgical procedure and histopathological examination

The rats were sacrificed using ketamine (25 mg/kg) anesthesia, on the 20<sup>th</sup> day of pregnancy. Kidney biopsies were fixed in 2.5% phosphate buffered glutaraldehyde. After postfixation with 1% osmic acid, they were dehydrated within acetone and semi-thin sections of tissue samples embedded in araldite were stained with toluidin blue. Then, the thin sections were stained with uranyl nitrate-lead and evaluated under Jeol 1010 transmission electron microscope and microphotographs were taken.

## RESULTS AND DISCUSSION

No pathology was observed in the control group of rats kidneys. The basal lamina, foot processes and infiltration slits were observed in normal view (Figure 1). In the kidneys of the group of valproic acid, the irregularities on the basal lamina and deletion on the foot processes were observed (Figure 2). In the group of valproic acid + folic

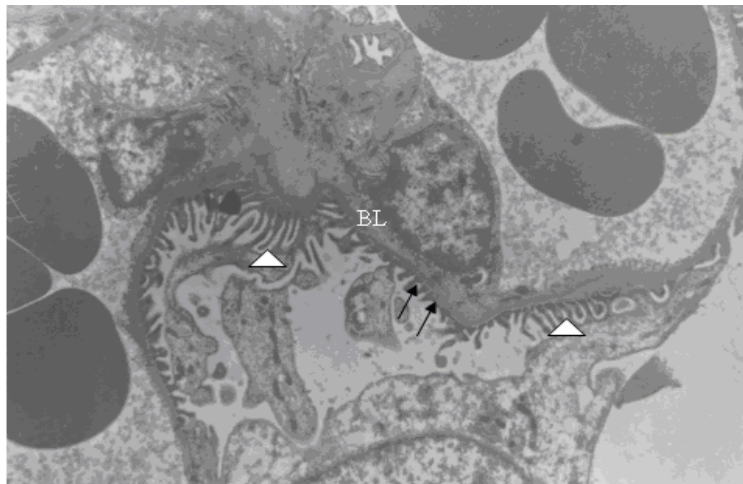
acid, sections belonging to this group showed relatively normal ultrastructure when compared to VPA group. Glomerular basal lamina, foot processes and infiltration slits were observed in normal view (Figure 3). And in the group of valproic acid + vitamin E, glomerular basal lamina, foot processes and infiltration slits view were found close to the control group (Figure 4).

VPA is in the market as an anticonvulsant since 1974, and is used in many countries because of its efficiency against several types of epilepsy and as a mood stabilizer. One of its main actions is the increase in the level of gamma amino butyric acid (GABA) in the brain. GABA is an important inhibitor of seizures, and reduction of GABA levels may potentiate seizures. For seizure control, the daily doses range between 300 mg to 2 g, aiming to achieve therapeutic plasma levels of 50 - 100 µg/mL. Lower doses are usually administered in the treatment of bipolar disorder-for manic patients, and against migraine (Ornoy, 2009).

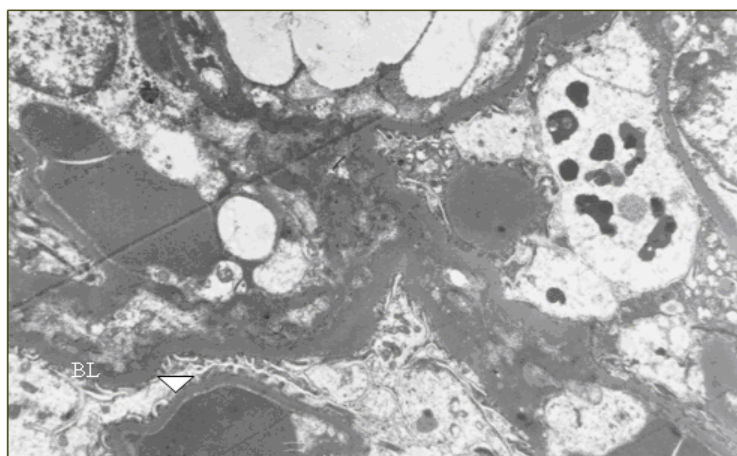
The use of VPA during pregnancy is associated with a 1 - 2% incidence of neural tube defects. Although this term refers to all types of neural tube defects, including anencephaly/exencephaly, VPA is associated mainly with lumbosacral meningocele (spina bifida aperta), the latter being 10–20 times the rate in the general population (Fried et al., 2004; Omtzigt et al., 1992; Lindhout et al., 1992).

In our study pathology was not observed in the first control group of rats kidneys (Figure 1). In the kidneys of the group of valproic acid, the irregularities on the renal glomerular basal membranes and deletion on the foot processes were observed (Figure 2).

SV is an antiepileptic drug known to induce hyperammonemia in humans. This hyperammonemia might result from a reduced detoxification of ammonium in the liver and/or from an accelerated renal ammoniogenesis (Rengel-Aranda et al., 1988). SV treatment has been indicated to produce hyperglycaemia and hyperglycinuria. Hyperammonaemia may occur asymptotically (Takeuchi et al., 1988). The main source of production is deamination of aminoacids taking place in different tissues (kidney, skeletal muscle and the colon) (Kvamme, 1983). Lenoir et al. (Lenoir et al., 1981) reported a case of severe myopathic syndrome and biochemical evidence suggestive of proximal tubular defect. Renal biopsy indicated giant mitochondria in the proximal tubular cells and abnormal round granular inclusions in the cytosol of tubular cells, podiocytes and interstitial cells which were recovered on withdrawal of valproic acid. Gossrau and Graf (Gossrau and Graf, 1989) reported a very severe damage to the tubules of the kidney in pregnant mice at a single dose of valproic acid (500 mg kg<sup>-1</sup> i.p.) which persisted even after 48 h of drug administration. Warter et al. (1983) showed that an important source of ammonia production is located in renal tubule cells. Hulsman (1989) demonstrated that ammonia is metabolised in the ornithine cycle, beginning in the liver mitochondria. The initiating step ( $\text{NH}_3 + \text{CO}_2 + \text{ATP}$ ) is



**Figure 1.** Electronmicrograph of control group rat kidney. Basal lamina (BL), foot processes (arrowheads) and infiltration slits (arrow) observed normal on rat kidney (uranyl acetate-lead citrate, X 3000).



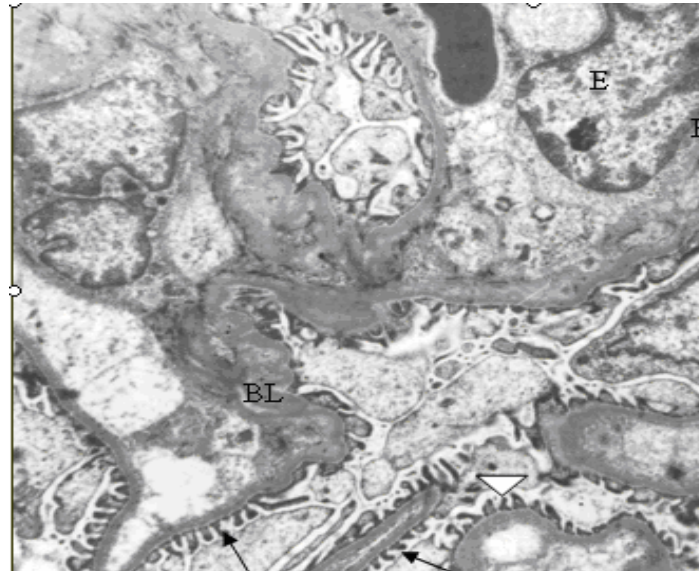
**Figure 2.** Electronmicrograph of valproic acid group rat kidney. The irregularities on the basal lamina (BL) and food deletion on the foot processes (arrowheads) observed (uranyl acetate-lead citrate, X 3000).

complex and is catalysed by carbamoyl phosphate transferase (CPT). When this enzyme is partially deficient hyperammonaemia occurs (Hjelm et al., 1986; Tripp et al., 1981). Furthermore, CPT may be inhibited with propionate, a degradation moiety of valproate. Although the exact biochemical mechanisms of SV toxicity to liver and kidney have not been well defined, several hypothesis have been proposed. The recent hypothesis suggests an involvement of lipid peroxidation (Olson et al., 1986; Buchi et al., 1984). The involvement of peroxidative injury in SV induced renal tubular disorder and hepatotoxicity is based on several lines of evidence. Valproate treatment decreased the rate of oxidized glutathione released into the bile (Olson et al., 1986); free radical scavengers like vitamin E and/or N-N'-diphenyl-p-phenylene-diamine

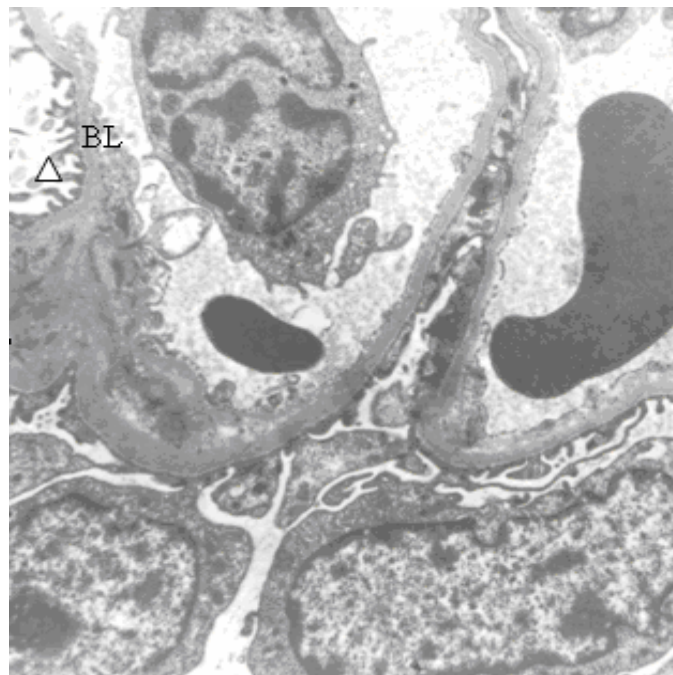
supplements provided adequate protection against SV toxicity in hepatocyte cultures (Buchi et al., 1984).

Although these studies suggest that lipid peroxidation may play a role in SV toxicity, a detailed time course of biochemical changes in liver and kidney following subacute exposure has not been reported and may be critical to understanding the mechanism of SV toxicity. These experiments were designed to demonstrate the initiation of biochemical events and the role of lipid peroxidation in SV toxicity.

SV may undergo omega oxidation and delta-dehydrogenation (Dreifuss et al., 1987; Kassahun et al., 1994) producing unsaturated derivatives (2-n-propyl-4-pentenoic acid; 2-n-propyl-2 (E) pentenoic acid; 2 hydroxy, 3 hydroxy, and 4 hydroxy valproic acid) by the



**Figure 3.** Electronmicrograph of valproic acid + folic acid group rat kidney. Regularity in basal lamina (BL), foot processes (arrowheads), infiltration slits (arrow) and endothelial cells (E) observed normal structure (uranyl acetate-lead citrate, X 3000).



**Figure 4.** Electronmicrograph of valproic acid + vitamin E group rat kidney. Note basal lamina (BL) and foot processes (arrow heads) in normal view (uranyl acetate-lead citrate, X 3000).

inhibition of either mitochondrial or microsomal P-450 (Rittle et al., 1987). This may generate reactive oxygen species, to combine with polyunsaturated lipids and generate lipid hydroperoxides. SV, its omega oxidation products and delta dehydrogenation moieties are

reported to be depressants of gluconeogenesis (Rogiers et al., 1985; Turnbull et al., 1986), and may inhibit the mitochondrial or microsomal reactions, a rate limiting factor.

Our findings in the present study, the group of SV+FA

(Figure 3), show that renal glomerular basal membranes and foot process were under normal view. Folic acid was studied in animals of one class of antimetabolites, those of pteroylglutamic acid, or folic acid as it is most commonly called, that led to interest in the role of this vitamin in human abnormal development, an interest that has extended over 50 years. In the first of such experimental work, the antimetabolites, when administered to female mice and rats at the time of uterine implantation or earlier, caused early prenatal death, but not malformation; suggesting an all-or-none action (Kalter, 2003). Many antiepileptic drugs interfere with folic acid absorption (phenytoin, barbiturates, carbamazepine and lamotrigine) or metabolism (VPA) (Yerby, 2003). It is therefore recommended to treat women on antiepileptic drugs at preconception and in the first 2–3 months of pregnancy with folic acid that protects human and animals from neural tube defect (NTD). Although the use of folic acid supplementation has been shown to generally decrease the incidence of NTD in humans, there is disagreement as to the benefit of folic acid in reducing the rate of NTD or other anomalies following VPA or other antiepileptic drugs exposure, as several studies and case reports were unable to demonstrate any significant reduction of these anomalies by folic acid (Yerby, 1994; Yerby, 2003; Genton et al., 2006). In spite of the uncertainty of effectiveness, it is recommended to take 4-5 mg/day of folic acid prior to any planned pregnancy. Indeed, in a recent study, Pittschieler et al. (2008) found that periconceptual folic acid significantly reduced the rate of spontaneous abortions and premature delivery in women treated with VPA and carbamazepine. They did not study the rate of congenital anomalies. Other studies were inconclusive as to the beneficial effects of folic acid, even in reducing the rate of spontaneous abortions

Similar disagreement exists in experimental animal studies. It seems that the protective effect of folic acid on VPA induced NTD is strain dependent, as different results were obtained in different strains of mice. For example, administration of high doses of folic acid to valproic acid-treated pregnant mice did not alter the production of exencephaly (Wegner and Nau, 1991; Wegner and Nau, 1992). On the other hand, pretreatment of pregnant CD-1 mice with high doses of folic acid and/or of pantothenic acid prior to the administration of valproic acid decreased the rate of NTD and almost normalized the levels of proteins involved in neurulation (Dawson et al., 2006). These conflicting results of human and animal studies imply that folic acid deficiency is not the major pathogenetic mechanism of VPA-induced teratogenicity.

Vitamin E is a beneficial antioxidant in pharmacological treatment of mitochondrial disorders (Bandyopadhyay and Dutta, 2005). In our findings in the group of SV+ Vit E, there was an appearance nearest to the control group (Figure 4). The antioxidant, vitamin E, has been shown to decrease the frequency of valproic acid-induced neural tube defects in mice, suggesting that ROS may play a role

in the failure of the neural tube to develop properly (Al Deeb et al., 2002). In addition, vitamin E has provided significant protection against hydrophobic bile acid toxicity in an *in vivo* rat model (Sokol et al., 1998). Vitamin E decreased the rate of VPA induced anomalies and embryonic damage in Balb mice (Ehlers et al., 1996), pointing to the possibility that oxidative stress is involved in VPA induced embryonic damage.

Valproic acid seems to be a highly teratogenic antiepileptic drug. VPA treated women at childbearing age should use contraceptives and stop the medication before any planned pregnancy (Ornoy, 2009).

## CONCLUSION

In conclusion, our findings show that the usage of SV during gestational period might clinically create risk, but this risk can be reduced by FA administration. In addition, Vit E was found to have a similar protective effect as FA. This effect of Vit E may be due to its antioxidant property. We consider that administration of FA and Vit E in case of maternal SV administration during pregnancy may reduce degenerative changes seen in kidney.

## REFERENCES

- Al Deeb S, Al Moutaery K, Arshaduddin M, Tariq M (2002). Vitamin E decreases valproic acid induced neural tube defects in mice. *Neurosci. Lett.* 292(3): 179-182.
- Arndt TL, Stodgell CJ, Rodier PM (2005). The teratology of autism. *Int. J. Dev. Neurosci.* 23: 189-199.
- Bandyopadhyay SK, Dutta AA (2005). Mitochondrial Hepatopathies. *JAPI*, p. 53.
- Binkerd PE, Rowland JM, Nau H, Hendrickx AG (1988). Evaluation of valproic acid (VPA) developmental toxicity and pharmacokinetics in Sprague-Dawley rats. *Fundam. Appl. Toxicol.* 11: 485-493.
- Buchi KN, Gray PD, Rollins DE, Tollman KG (1984). Protection against sodium valproate injury in isolated hepatocytes by alphas-tocopherol and N-N'-diphenyl-p-phenylenediamine. *J. Clin. Pharmacol.* 24(4): 148-154.
- Dalens B, Raynaud EJ, J Gaulme (1980). Teratogenicity of valproic acid. *J. Pediatr.* 97: 332-333.
- Dawson JE, Raymond AM, Winn LM (2006). Folic acid and pantothenic acid protection against valproic acid-induced neural tube defect in CD 1 mice. *Toxicol. Appl. Pharmacol.* 211(124): 132.
- Dreifuss FE, Santilli N, Langer DH, Sweeney KP, Moline KA, Manander KB (1987). Valproic acid hepatic fatalities: a retrospective review. *Neurol.* 37:379-385.
- Ehlers K, Elmazar MM, Nau H (1996). Methionine reduces the valproic acid-induced spina bifida in mice without altering valproic acid kinetics. *J. Nutr.* 126: 67-75.
- Emmanouil-Nikoloussi EN, Foroglou NG, Kerameos-foroglou CH, Thliveris JA (2004). Effects of valproic acid on fetal and maternal organs in the mouse: a morphological study. *Morphologie*, 88: 41-45.
- Fried S, Kozer E, Nulman I, Einarson TR, Koren G (2004). Malformation rates in children of women with untreated epilepsy. *Drug Saf.* 27: 197-202.
- Genton P, Semach F, Trinka E (2006). Valproic acid in epilepsy: pregnancy related issues. *Drug Saf.* 29(1): p. 21.
- Gomez MR (1981). Possible teratogenicity of valproic acid. *J. Pediatr.* 98: 508-509.
- Gossrau R, Graf R (1989). Lesions and repair of cells of maternal mice after valproic acid treatment on day 8 pregnancy: an enzyme histochemical analysis. *Acta Histochemica*, 86(1): 23-32.

- Hendrickx AG, Nau H, Binkerd P, Rowland JM, Rowland JR, Cukierski MJ (1988). Valproic acid developmental toxicity and pharmacokinetics in the rhesus monkey: an interspecies comparison. *Teratology*, 38(329): 45.
- Hjelm M, Oberholzer V, Seakins J, Thomas S, Kay TDS (1986). Valproate induced inhibition of urea synthesis and hyperammonaemia in healthy subjects. *Lancet II*, p. 859.
- Holmes LB, Coull BA, Dorfman J, Rosenberger PB (2005). The correlation of deficits in IQ with midface and digit hypoplasia in children exposed in utero to anticonvulsant drugs. *J. Pediatr.* 146: 118-122.
- Hulsman J (1989). Hyperammonaemia and the use of antiepileptic drugs, including valproate. In: Chadwick D (Ed) Fourth international symposium on sodium valproate and epilepsy. London: R. Soc. Med. Services, pp. 163-168.
- Ingram JL, Peckham SM, Tisdale B, Rodier PM (2000). Prenatal exposure of rats to valproic acid reproduces the cerebellar anomalies associated with autism. *Neurotoxicol. Teratol.* 22: 319-324.
- Jamsheer A, Materna-Kiryluk A, Latos-Bieleńska A (2008). Valproic acid and pregnancy: clinical presentation of 3 cases with valproate embryopathy. *Arch. Perinatal Med.* 14(1): 57-60.
- Kalter H (2003). Teratology in the 20th century. Environmental causes of congenital malformations in humans and how they were established. *Neurotoxicol. Teratol.* 26(1): 1-12.
- Kassahun K, Hu P, Grillo MP, Davis MR, Jin L, Baillie TA (1994). Metabolic activation of unsaturated derivatives of valproic acid. Identification of novel glutathione adducts formed through co-enzyme. A dependent and independent processes. *Chem. Biol. Interact.* 90(3): 253-275.
- Kvamme E (1983). Ammonia metabolism in the CNS. *Prog. Neurobiol.* 20: 109-132.
- Lenoir GR, Perignon JL, Gubler MC, Broyer M (1981). Valproic acid: a possible cause of proximal tubular renal syndrome. *J. Pediatr.* 98: 503-504.
- Lindhout D, Omtzigt JG, Cornel MC (1992). Spectrum of neural-tube defects in 34 infants prenatally exposed to antiepileptic drugs. *Neurol.* 42(5): 111-118.
- Loscher W, Nau H (1983). Distribution of valproic acid and its metabolites in various brain areas of dogs and rats after acute and prolonged treatment. *J. Pharmacol. Exp. Ther.* 226(3): 845-854.
- Olson MJ, Handler JA, Thurman RG (1986). Mechanism of zone specific hepatic steatosis caused by valproate: inhibition of ketogenesis in periportal regions of the liver lobule. *Mol. Pharmacol.* 30(6): 520-525.
- Omtzigt JG, Los FJ, Grobbee DE, Pijpers L, Jahoda MG, Brandenburg H (1992). The risk of spina bifida aperta after first-trimester exposure to valproate in a prenatal cohort. *Neurology*, 42: 119-125.
- Ornoy A (2009). Valproic acid in pregnancy: How much are we endangering the embryo and fetus? *Reprod. Toxicol.* 28: 1-10.
- Pittschieler S, Brezinka C, Jahn B, Trinkka E, Unterberger I, Dobesberger J (2008). Spontaneous abortions and the prophylactic effect of folic acid supplementation in epileptic women undergoing antiepileptic therapy. *J. Neurol.* 255: 1926-1931.
- Rengel-Aranda M, Gougoux A, Vinay P, Lopez-Novoa JM (1988). Effect of valproate on renal metabolism in the intact dog. *Kidney Int.* 34: 645-654.
- Rittle AE, Rettewmeier AW, Howard WN, Baillie TA (1987). Cytochrome P-450 catalysed formation of VPA, a toxic metabolite of valproic acid. *Sci.* 235: 890-893.
- Rodier PM, Ingram JL, Tisdale B, Nelson S, Romano J (1996). Embryological origin for autism: developmental anomalies of the cranial nerve motor nuclei. *J. Comp. Neurol.* 370: 247-261.
- Rogiers V, Vandenberghe Y, Vereruyse A (1985). Inhibition of gluconeogenesis by sodium valproate and its metabolites in isolated rat hepatocytes. *Xenobiotica*, 15(8-9): 759-765.
- Sokol RJ, McKim JM Jr, M0 Goff, Ruyle SZ, Devereaux MV, Han O, Packer I, Everson G (1998). Vitamin E reduces oxidant injury to mitochondria and the hepatotoxicity of taurochenodeoxy-cholic acid in the rat. *Gastroenterol.* 114: 164-174.
- Takeuchi T, Sugimoto T, Nishida N, Kobayashi Y (1988). Protective effect of D,L-carnitine on valproate induced hyperammonaemia and hypoketonaemia in primary cultured hepatocytes. *Biochem. Pharmacol.* 37(11): 2255-2258.
- Tripp JH, Hargreaves T, Anthony PP, Searle JF, Miller P, Leonard JV, Patrick AD, Oberholzer VG (1981). Sodium valproate and ornithine carbamoyl-transferase deficiency. *Lancet.* I: 1165-1166.
- Turnbull DM, Dick DJ, Wilson L, Sherratt HS, Alberti KG (1986). Valproate causes metabolic disturbances in normal man. *J. Neurol. Neurosurg. Psychiat.* 49(4): 405-410.
- Wagner GC, Reuhl KR, Chen M, McRae P, Halladay AK (2006). A new neurobehavioral model of autism in mice: pre and postnatal exposure to sodium valproate. *Autism. Dev. Disord.* 36: 779-793.
- Warter JM, Brandt C, Marescaux C, Rumach L, Michelletti G, Chabrier G, Krieger J, Imler M (1983). The renal origin of sodium valproate induced hyperammonaemia in fasting humans. *Neurobiology*, 33: 1136-1140.
- Wegner C, Nau H (1992). Alteration of embryonic folate metabolism by valproic acid during organogenesis: implication for mechanism of teratogenesis. *Neurol.* 42(5): 17-24.
- Wegner C, Nau H (1991). Diurnal variation of folate concentrations in mouse embryo and plasma: the protective effect of folic acid on valproic acid induced teratogenicity is time dependent. *Reprod. Toxicol.* 5: 465-471.
- Yerby MS (2003). Management issues for women with epilepsy. Neural tube defects and folic acid supplementation. *Neurol.* 61(2): 23-26.
- Yerby MS (1994). Pregnancy, teratogenesis and epilepsy. *Neurol. Clin.* 12: 749-771.