

## Review

# The most stirring technology in future: Cellulase enzyme and biomass utilization

Xing-hua Li<sup>1</sup>, Hua-jun Yang<sup>1</sup>, Bhaskar Roy<sup>1</sup>, Dan Wang<sup>1</sup>, Wan-fu Yue<sup>1</sup>, Li-jun Jiang<sup>1</sup>, Enoch Y. Park<sup>2</sup> and Yun-gen Miao<sup>1\*</sup>

<sup>1</sup>Key Laboratory of Animal Epidemic Etiology and Immunological Prevention of Ministry of Agriculture, College of Animal Sciences, Zhejiang University, Hangzhou 310029, P. R. China.

<sup>2</sup>Laboratory of Biotechnology, Department of Applied Biological Chemistry, Faculty of Agriculture, Shizuoka University, Shizuoka 422-8529, Japan.

Accepted 3 March, 2009

**In recent years, fundamental and applied researches on cellulase enzyme have not only generated significant scientific knowledge but also have revealed their enormous potential in biotechnology. Growing attention has been devoted to its bioconversion of biomass into fuel ethanol, considered the cleanest liquid fuel alternative to fossil fuels. Significant advances have been made towards the production and alteration technology of cellulase enzyme. This review simply introduces cellulose and cellulase enzyme, gives a broad overview of the current research status of cellulase enzyme, briefly refers to its applied fields, and lastly summarizes its promising prospects.**

**Key words:** Cellulose, cellulase enzyme, fuel ethanol, biomass.

## INTRODUCTION

Currently, the problems of energy and environment are large obstacles to the development of human civilization. The world's energy demand is increasing steadily as the human population grows and economic development. However, the current predominant energy resource, which is the fossil fuel supply, is limited. In addition, since an accelerated release of fossil entombed CO<sub>2</sub> due to human activities, the global greenhouse effect is more and more serious. Therefore, continuous efforts have to be emphasized towards the solution of the energy supply depletion problem and the environmental impacts caused by the human activities. There has been also an increasing worldwide interest in alternative sources of energy (Aristidou and Penttila, 2000; Jeffries and Jin, 2000; Zaldivar et al., 2001), such as agricultural biomass, to substitute fossil-fuel-based energy resources. The use of biomass for energy can complement solar, wind, and other intermittent energy resources in the renewable energy mix of the future, reduce fossil fuel greenhouse

gas emissions, and contribute to social sustainable development.

One of the most immediate and important applications of biomass energy systems could be in the fermentation of ethanol from biomass (Lin and Tanaka, 2006). Each year, photosynthetic fixation of CO<sub>2</sub> yields more than 10<sup>11</sup> tons of dry plant material worldwide, and almost half of this material consists of cellulose (Leschine, 1995). In addition, agronomic residues arising from human activities, such as corn stover (corn cobs and stalks), sugarcane waste, wheat or rice straw, forestry, and paper mill discards, the paper portion of municipal waste and dedicated energy crops, also have plentiful cellulose, which can be converted into fuel ethanol. Nowadays, nearly all fuel ethanol is produced by fermentation of corn glucose in America or sucrose in Brazil, and any countries advanced in agriculture can use current technology for fuel ethanol fermentation (Lin and Tanaka, 2006). But this technology consumes a lot of food materials, costs too much, and under the pressure of world's food crisis at present, there is even a serious competition between human food and fuel ethanol. Thus, there is need to find another better way to solve this problem, and now it is mostly concentrated in the field of cellulase enzymes,

\*Corresponding author. E-mail: [miaoyg@zju.edu.cn](mailto:miaoyg@zju.edu.cn). Tel.: +86 571 86971659.

which can hydrolyze cellulose in biomass with high efficiency.

This review will introduce some background knowledge of cellulase enzyme and its hydrolyzation mechanism, focus on its current status in research and application, and give insight into some promising prospects for its future.

## CELLULOSE AND CELLULASE

Cellulose is a fibrous, insoluble, crystalline polysaccharide. It is a major polysaccharide constituent of plant cell walls, composed of repeating D-glucose units linked by  $\beta$ -1,4-glucosidic bonds (Jagtap and Rao, 2005) and being the most abundant carbohydrate polymer on earth (Guo et al., 2008).

Cellulose has attracted worldwide attention as a renewable resource that can be converted into biobased products and bioenergy. But nowadays, enormous amounts of agricultural, industrial and municipal cellulose wastes have been accumulating or used inefficiently due to the high cost of their utilization processes (Kim et al., 2003). Therefore, it has become of considerable economic interest to develop processes for the effective treatment and utilization of cellulosic wastes as cheap carbon sources. Cellulose is used as a food source by a wide variety of organisms including fungi, bacteria, plants and protists, as well as a wide range of invertebrate animals, such as insects, crustaceans, annelids, molluscs and nematodes (Watanabe and Tokuda, 2001; Davison and Blaxter, 2005). These organisms possess cellulases and the complete enzymatic system of them include three different types, that is, exo- $\beta$ -1,4-glucanases (EC 3.2.1.91), endo- $\beta$ -1,4-glucanases (EC 3.2.1.4), and  $\beta$ -1,4-glucosidase (EC 3.2.1.21) (Wilson and Irwin, 1999). These enzymatic components act sequentially in a synergistic system to facilitate the breakdown of cellulose and the subsequent biological conversion to an utilizable energy source, glucose (Beguin and Aubert, 1994). The endo- $\beta$ -1,4-glucanases randomly hydrolyzes the  $\beta$ -1,4 bonds in the cellulose molecule, and the exo- $\beta$ -1,4-glucanases in most cases release a cellobiose unit showing a recurrent reaction from chain extremity. Lastly, the cellobiose is converted to glucose by  $\beta$ -1,4-glucosidase (Bhat and Bhat, 1997). This whole enzymatic process to hydrolyze cellulosic materials could be accomplished through a complex synergistically reaction of these various enzymatic components in an optimum proportion (Tomme et al., 1995).

Cellulases provide a key opportunity for achieving tremendous benefits of biomass utilization (Wen et al., 2005). But currently, two significant points of these enzyme-based bioconversion technologies are reaction conditions and the production cost of the related enzyme system. Therefore, there has been much research aimed at obtaining new microorganisms producing cellulase enzymes with higher specific activities and greater effi-

ciency (Subramaniyan and Prema, 2000).

## RESEARCH STATUS OF CELLULASE ENZYME

Cellulase is an important and essential kind of enzyme for carrying out the depolymerization of cellulose into fermentable sugars. As a major resource for renewable energy and raw materials, it is widely used in the bioconversion of renewable cellulosic biomass. Glucose, from appropriate hydrolysis of this cellulosic biomass under the treatment of advanced biotechnology can be used in different applications such as production of fuel ethanol, single cell protein, feed stock, industrially important chemicals and so on (Gawande and Kamat, 1999; Fujita et al., 2002; Lynd et al., 2002). Given the importance of this enzyme to these so many industries, extensive interest and considerable research efforts are focused on the understanding of reaction mechanism and industrial application of cellulase, which began in the early 1950s. Nature has evolved a number of cellulases for the hydrolysis of cellulose, including exoglucanase enzymes that depolymerize cellulose from the reducing and non-reducing ends processively, and endoglucanase enzymes that cleave along the cellulose chains randomly (Warren, 1996; Coughlan, 1985; Wilson and Irwin, 1999).

Prior studies for natural cellulose hydrolysis have revealed many cellulolytic microorganisms and their complex cellulases (Lowe et al., 1987; Lynd et al., 2005). A number of fungi and bacteria capable of utilizing cellulose as a carbon source have been identified (Kim et al., 2003). Among the cellulolytic fungi, *Trichoderma reesei* has the strongest cellulose-degrading activity, and its cellulase has been widely investigated (Penttila et al., 1986; Tomme et al., 1988). Meanwhile, cellulases produced by other fungi such as the *Aspergillus* and *Rhizopus* species have also been extensively studied by several researchers (Murashima et al., 2002; Saito et al., 2003). Interestingly, recent works confirmed the production of cellulases from insect for themselves (Watanabe et al., 1998; Girard and Jouanin, 1999; Tokuda et al., 1999) along with reports on the production from symbiotic organisms harboring in the insect gut and both (Ohtoko et al., 2000; Scharf et al., 2003). These new findings challenged the traditional view of cellulase activity that cellulose digestion in insects was mediated by microbial cellulase activity in their gut.

Current understanding of the modes of catalysis and the role of various structural domains has been gained from protein engineering, X-ray crystallography and fluorescence studies, which evaluate cellulase-cellulose interactions in bulk (Pilz et al., 1990; Chapon et al., 2001; Violot et al., 2005; Pinto et al., 2007). To date, a battery of cellulase genes have been found and their gene structures and functions have also been studied (Mae et al., 1995; Murray et al., 2003; Lee et al., 2005). The purification and properties of cellulases have been described in many papers (Whitaker, 1951; Kanda et al., 1976;

Churilova et al., 1980; Anzai et al., 1984; Mori, 1992; Ye et al., 2001; Kim et al., 2005; Li et al., 2005; Ogura et al., 2006; Lee et al., 2008; Thongekkaew et al., 2008). These studies have allowed scientists to further understand the cellulase molecular mechanisms at the fundamental scale of cellulose, expand their understanding of microscale heterogeneous kinetics and identify essential amino acids in the catalytic site of various enzymes (Wilson and Irwin, 1999; Tomme et al., 1995; Henrissat et al., 1998; Jung et al., 2002; Jung et al., 2003; Zhang and Lynd, 2004). By the way, although there have been many papers dealing with more efficient cellulose degrading enzyme from various organisms such as *Trichoderma reesei*, *Trichoderma viride*, *Trichoderma lignorum*, *Chrysosporium lignorum*, *Chrysosporium pruinatum* and *Fusarium solani* (Selby and Maitland, 1967; Toyama and Ogawa, 1975; Tong et al., 1980), only limited research has identified the yeast as cellulase producer Oikawa et al., 1998; Hong et al., 2007). In addition, in the last decades, the high production cost and low yields of this enzyme are the major problems for industrial application. Therefore, investigations on ability of microbial strains to utilize inexpensive substrate (Griffin, 1973; Hurst et al., 1978; Liaw and Penner, 1990; Ju and Afolabi, 1999; Stenberg et al., 2000) and improvement of enzyme productivity (Kumakura et al., 1984; Chadha and Garcha, 1992; Hayward et al., 2000; Bailey and Tahhtiharju, 2003; Villena and Gutierrez-Correa, 2006) have been done. However, by far, although the cellulase enzyme cost has dropped due to improvements in expression vectors and on-site production (Barros and Thomson, 1987; Din et al., 1990; Sahasrabudhe and Ranjekar, 1990; Harkki et al., 1991; Okamoto et al., 1994; Kobayashi et al., 2003; Kashima and Udaka, 2004), there is still a necessity of engineering a new generation of cellulase cocktails that would further reduce cellulase cost.

## APPLICATION OF CELLULASE ENZYME

Along with the extensive fundamental researches of cellulases for decades of years, their application studies have also developed at a tremendous speed. Biotechnology of cellulases began in early 1980s, first in animal feed followed by food applications (Voragen et al., 1980). Subsequently, they were used in the textile, laundry as well as in the pulp and paper industries (Godfrey et al., 1996; Ito, 1997; Bajpai, 1999). Through developing for several decades, the use of cellulases has increased considerably, demonstrating their huge biotechnological potential in various industries, including above-mentioned industrial fields, brewery and wine, agriculture, as well as in research and development (Bhat and Bhat, 1997; Mandels, 1985; Bayer et al., 1994; Ohmiya et al., 1997; Bhat, 2000). These have been reviewed in many papers (Mandels, 1985; Coughlan, 1985a, 1985b; Godfrey and West, 1996; Harman and Kubicek, 1998; Uhlig, 1998).

## PROSPECTS

Enhancing the activity of cellulase enzyme and reducing its production cost are two key issues in the enzymatic hydrolysis of cellulosic materials. Genetic techniques will be used to clone the cellulase coding sequences into bacteria, yeasts, fungi, plants and animals to create new cellulase production systems with possible improvement of enzyme production and activity. In addition, using cellulosic materials such as agricultural residues, grasses, forestry wastes, and other low-cost biomass can significantly reduce the cost of raw materials for ethanol production compared to corn. It is also predicted that the use of genetically engineered raw materials with higher carbohydrate content combined with the improvement of conversion technology could reduce the cost of ethanol a lot. All those will give a great help for solving the problems of energy and food in the world. In a word, the cellulase enzymes will be commonly used in many industrial applications, and the demand for more stable, highly active and specific enzymes will be also growing rapidly. So, cellulase enzyme will be as the most stirring technology in 2009, gaining the whole world attention.

## ACKNOWLEDGEMENTS

The work was supported by a key project of Zhejiang Government No. 2008C24011 and by the Hi-Tech Research and Development Program of China (No. 2008AA10Z132 and No. 2006AA10A119).

## REFERENCES

- Anzai H, Nisizawa K, Matsuda K (1984). Purification and characterization of a cellulase from *Dolabella auricularia*. *J. Biochem.* 96(5): 1381-1390.
- Aristidou A, Penttila M (2000). Metabolic engineering applications to renewable resource utilization. *Curr. Opin. Biotechnol.* 11(2): 187-198.
- Bajpai P (1999). Application of enzymes in the pulp and paper industry. *Biotechnol. Prog.* 15(2): 147-157.
- Bailey MJ, Tahhtiharju J (2003). Efficient cellulase production by *Trichoderma reesei* in continuous cultivation on lactose medium with a computer-controlled feeding strategy. *Appl. Microbiol. Biotechnol.* 62(2-3): 156-162.
- Barros ME, Thomson JA (1987). Cloning and expression in *Escherichia coli* of a cellulase gene from *Ruminococcus flavefaciens*. *J. Bacteriol.* 169(4): 1760-1762.
- Bayer EA, Morag E, Lamed R (1994). The cellulosome--a treasure-trove for biotechnology. *Trends Biotechnol.* 12(9): 379-386.
- Beguín, P, Aubert JP (1994). The biological degradation of cellulose. *FEMS Microbiol. Rev.* 13(1): 25-58.
- Bhat MK, Bhat S (1997). Cellulose degrading enzymes and their potential industrial applications. *Biotechnol. Adv.* 15(3-4): 583-620.
- Bhat MK (2000). Cellulases and related enzymes in biotechnology. *Biotechnol. Adv.* 18(5): 355-383.
- Chapon V, Czjzek MEI, Hassouni M, Py B, Juy M, Barras F (2001). Type II protein secretion in gram-negative pathogenic bacteria: the study of the structure/secretion relationships of the cellulase Cel5 (formerly EGZ) from *Erwinia chrysanthemi*. *J. Mol. Biol.* 310(5): 1055-1066.
- Chadha BS, Garcha HS (1992). Mixed cultivation of *Trichoderma reesei* and *Aspergillus ochraceus* for improved cellulase production. *Acta Microbiol. Hung.* 39(1): 61-67.

- Churilova IV, Maksimov VI, Klesov AA (1980). Purification and properties of low molecular weight endoglucanase of the cellulase complex from *Trichoderma koningii*. *Biokhimiia*, 45(4): 669-678.
- Coughlan MP (1985a). Cellulases: Production properties and applications. *Biochem. Soc. Trans.* 13: 405-406.
- Coughlan MP (1985b). The properties of fungal and bacterial cellulases with comment on their production and application. In: *Biotechnol. Genet. Eng. Rev.* 3: 39-109.
- Davison A, Blaxter M (2005). Ancient origin of glycosyl hydrolase family 9 cellulase genes. *Mol. Biol. Evol.* 22(5): 1273-1284.
- Din N, Beck CF, Miller Jr RC, Kilburn DG, Warren RA (1990). Expression of the *Cellulomonas fimi* cellulase genes *cex* and *cenA* from the divergent tet promoters of transposon Tn10. *Arch Microbiol.* 153(2): 129-133.
- Fujita Y, Takahashi S, Ueda M, Tanaka A, Okada H, Morikawa Y, Kawauchi T, Arai M, Fukuda H, Kondo A (2002). Direct and efficient production of ethanol from cellulosic material with a yeast strain displaying cellulolytic enzymes. *Appl. Environ. Microbiol.* 68(10): 5136-5141.
- Gawande PV, Kamat MY (1999). Production of *Aspergillus niger* by lignocellulosic waste fermentation and its application. *J. Appl. Microbiol.* 87(4): 511-519.
- Godfrey T, West S (1996). *Industrial Enzymology*. London: Macmillan Press, p. 2.
- Godfrey T, West S, (1996). *Textiles In: 2<sup>nd</sup> ed. Industrial enzymology*, London, Macmillan Press, p. 360-371.
- Girard C, Jouanin L (1999). Molecular cloning of cDNAs encoding a range of digestive enzymes from a phytophagous beetle, *Phaedon cochleariae*. *Insect Biochem. Mol. Biol.* 29(12): 1129-1142.
- Griffin HL (1973). Filter paper assay--effect of time and substrate concentration on cellulase activity. *Anal. Biochem.* 56(2): 621-625.
- Guo R, Ding M, Zhang SL, Xu GJ, Zhao FK (2008). Molecular cloning and characterization of two novel cellulase genes from the mollusc *Ampullaria crosseana*. *J. Comp. Physiol. [B]*, 178(2): 209-215.
- Harkki A, Mantyla A, Penttila M, Muttillainen S, Buhler R, Suominen P, Knowles J, Nevalainen H (1991). Genetic engineering of *Trichoderma* to produce strains with novel cellulase profiles. *Enzyme Microb. Technol.* 13(3): 227-233.
- Harman GE, Kubicek CP (1998). *Trichoderma and Gliocladium: Enzymes, biological control and commercial applications*. London: Taylor & Francis Ltd, 2: p. 393.
- Hayward TK, Hamilton J, Tholudur A, McMillan JD (2000). Improvements in titer, productivity, and yield using Solka-floc for cellulase production. *Appl. Biochem. Biotechnol.* 84-86: 859-874.
- Henrissat B, Teeri TT, Warren RA (1998). A scheme for designating enzymes that hydrolyse the polysaccharides in the cell walls of plants. *FEBS Lett.* 425(2): 352-354.
- Hong J, Wang Y, Kumagai H, Tamaki H (2007). Construction of thermotolerant yeast expressing thermostable cellulase genes. *J. Biotechnol.* 130(2): 114-123.
- Hurst PL, Sullivan PA, Shepherd MG (1978). Substrate specificity and mode of action of a cellulase from *Aspergillus niger*. *Biochem. J.* 169(2): 389-395.
- Ito S (1997). Alkaline cellulases from alkaliphilic *Bacillus*: enzymatic properties, genetics, and application to detergents. *Extremophiles*, 1(2): 61-66.
- Jagtap S, Rao M (2005). Purification and properties of a low molecular weight 1,4-beta-d-glucan glucohydrolase having one active site for carboxymethyl cellulose and xylan from an *alkalothermophilic Thermomonospora sp.* *Biochem. Biophys. Res. Commun.* 329(1): 111-116.
- Jeffries TW, Jin YS (2000). Ethanol and thermotolerance in the bioconversion of xylose by yeasts. *Adv. Appl. Microbiol.* 47: 221-268.
- Ju LK, Afolabi OA (1999). Wastepaper hydrolysate as soluble inducing substrate for cellulase production in continuous culture of *trichoderma reesei*. *Biotechnol. Prog.* 15(1): 91-97.
- Jung H, Wilson DB, Walker LP (2002). Binding mechanisms for *Thermobifida fusca* Cel5A, Cel6B, and Cel48A cellulose-binding modules on bacterial microcrystalline cellulose. *Biotechnol. Bioeng.* 80(4): 380-392.
- Jung H, Wilson DB, Walker LP (2003). Binding and reversibility of *Thermobifida fusca* Cel5A, Cel6B, and Cel48A and their respective catalytic domains to bacterial microcrystalline cellulose. *Biotechnol. Bioeng.* 84(2): 151-159.
- Kanda T, Wakabayashi K, Nisizawa K (1976). Purification and properties of an endo-cellulase of avicelase type from *Irpex lacteus* (Polyporus tulipiferae). *J. Biochem.* 79(5): 977-988.
- Kashima Y, Udaka S (2004). High-level production of hyperthermophilic cellulase in the *Bacillus brevis* expression and secretion system. *Biosci. Biotechnol. Biochem.* 68(1): 235-237.
- Kim KC, Seung-Soo Y, Oh Young A, Seong-Jun K (2003). Isolation and characteristics of *Trichoderma harzianum* FJ1 producing cellulases and xylanase. *J. Microbiol. Biotechnol.* 13: 1-8.
- Kim JY, Hur SH, Hong JH (2005). Purification and characterization of an alkaline cellulase from a newly isolated alkalophilic *Bacillus sp.* HSH-810. *Biotechnol. Lett.* 27(5): 313-316.
- Kobayashi Y, Taquchi H, Goto TN, Koike S, Ohmiya K (2003). Expression and export of a *Ruminococcus albus* cellulase in *Butyrivibrio fibrisolvens* through the use of an alternative gene promoter and signal sequence. *Can. J. Microbiol.* 49(6): 375-382.
- Kumakura M, Kaetsu I, Nisizawa K (1984). Cellulase production from immobilized growing cell composites prepared by radiation polymerization. *Biotechnol. Bioeng.* 26(1): 17-21.
- Lee SJ, Lee KS, Kim SR, Gui ZZ, Kim YS, Yoon HJ, Kim I, Kang PD, Sohn HD, Jin BR (2005). A novel cellulase gene from the mulberry longicorn beetle, *Apriona germari*: gene structure, expression, and enzymatic activity. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* 140(4): 551-260.
- Leschine SB (1995). Cellulose degradation in anaerobic environments. *Annu. Rev. Microbiol.* 49: 399-426.
- Li YH, Guo R, Yin QY, Ding M, Zhang SL, Xu GJ, Zhao FG (2005). Purification and characterization of two endo-beta-1,4-glucanases from mollusca, *Ampullaria crosseana*. *Acta Biochem. Biophys. Sin (Shanghai)*, 37(10): 702-708.
- Liaw ET, Penner MH (1990). Substrate-velocity relationships for the *Trichoderma viride* cellulase-catalyzed hydrolysis of cellulose. *Appl. Environ. Microbiol.* 56(8): 2311-2318.
- Lin Y, Tanaka S (2006). Ethanol fermentation from biomass resources: current state and prospects. *Appl. Microbiol. Biotechnol.* 69(6): 627-642.
- Lee YJ, Kim BK, Lee BH, Jo KI, Lee NK, Chung CH, Lee YC, Lee JW (2008). Purification and characterization of cellulase produced by *Bacillus amyloquelaceus* DL-3 utilizing rice hull. *Bioresour. Technol.* 99(2): 378-386.
- Lowe SE, Theodorou MK, Trinci AP (1987). Cellulases and xylanase of an anaerobic rumen fungus grown on wheat straw, wheat straw holocellulose, cellulose, and xylan. *Appl. Environ. Microbiol.* 53(6): 1216-1223.
- Lynd LR, Weimer PJ, van Zyl WH, Pretorius IS (2002). Microbial cellulose utilization: fundamentals and biotechnology. *Microbiol. Mol. Biol. Rev.* 66(3): 506-577.
- Lynd LR, van Zyl WH, McBride JE, Laser M (2005). Consolidated bioprocessing of cellulosic biomass: an update. *Curr. Opin. Biotechnol.* 16(5): 577-583.
- Mae A, Heikinheimo R, Palva ET (1995). Structure and regulation of the *Erwinia carotovora* subspecies *carotovora* SCC3193 cellulase gene *celV1* and the role of cellulase in phytopathogenicity. *Mol. Gen. Genet.* 247(1): 17-26.
- Mandels M (1985). Applications of cellulases. *Biochem. Soc. Trans.* 13(2): 414-416.
- Mori Y (1992). Purification and characterization of an endoglucanase from the cellulosomes (multicomponent cellulase complexes) of *Clostridium thermocellum*. *Biosci. Biotechnol. Biochem.* 56(8): 1198-1203.
- Murashima K, Nishimura T, Nakamura Y, Koga J, Moriya T, Sumida N, Yaguchi T, Kono T (2002). Purification and characterization of new endo-1, 4-β-D-glucanases from *Rhizopus oryzae*. *Enzyme Microb. Technol.* 30: 319-326.
- Murray PG, Collins CM, Grassick A Tuohy MG (2003). Molecular cloning, transcriptional, and expression analysis of the first cellulase gene (*cbh2*), encoding cellobiohydrolase II, from the moderately thermophilic fungus *Talaromyces emersonii* and structure prediction of the gene product. *Biochem. Biophys. Res. Commun.* 301(2): 280-286.

- Ogura J, Toyoda A, Kurosawa T, Chong AL, Chohnan S, Masaki T (2006). Purification, characterization, and gene analysis of cellulase (Cel8A) from *Lysobacter* sp. IB-9374. *Biosci. Biotechnol. Biochem.* 70(10): 2420-2428.
- Ohmiya K, Sakka K, Karita S, Kimura T (1997). Structure of cellulases and their applications. *Biotechnol. Genet. Eng. Rev.* 14: 365-414.
- Ohtoko K, Ohkuma M, Moriya S, Inoue T, Usami R, Kudo T (2000). Diverse genes of cellulase homologues of glycosyl hydrolase family 45 from the symbiotic protists in the hindgut of the termite *Reticulitermes speratus*. *Extremophiles*, 4(6): 343-349.
- Oikawa T, Tsukagawa Y, Soda K (1998). Endo-beta-glucanase secreted by a psychrotrophic yeast: purification and characterization. *Biosci. Biotechnol. Biochem.* 62(9): 1751-1756.
- Okamoto T, Yamano S, Ikaqa H, Nakamura K (1994). Cloning of the *Acetobacter xylinum* cellulase gene and its expression in *Escherichia coli* and *Zymomonas mobilis*. *Appl. Microbiol. Biotechnol.* 42(4): 563-568.
- Penttila M, Lehtovaara P, Nevalainen H, Bhikhabhai R, Knowles J (1986). Homology between cellulase genes of *Trichoderma reesei*: complete nucleotide sequence of the endoglucanase I gene. *Gene*, 45(3): 253-263.
- Pilz I, Schwarz E, Kilburn DG, Miller Jr RC, Warren RA, Gilkes NR (1990). The tertiary structure of a bacterial cellulase determined by small-angle X-ray-scattering analysis. *Biochem. J.* 271(1): 277-280.
- Pinto R, Amaral AL, Carvalho J, Ferreira EC, Mota M, Gama M (2007). Development of a method using image analysis for the measurement of cellulose-binding domains adsorbed onto cellulose fibers. *Biotechnol. Progr.* 23(6): 1492-1497.
- Sahasrabudhe NA, Ranjekar PK (1990). Cloning of the cellulase gene from *Penicillium funiculosum* and its expression in *Escherichia coli*. *FEMS Microbiol. Lett.* 54(1-3): 291-293.
- Saito K, Kawamura Y, Oda Y (2003). Role of the pectinolytic enzyme in the lactic acid fermentation of potato pulp by *Rhizopus oryzae*. *J. Ind. Microbiol. Biotechnol.* 30(7): 440-444.
- Selby K, Maitland CC (1967). The cellulase of *Trichoderma viride*. Separation of the components involved in the solubilization of cotton. *Biochem. J.* 104(3): 716-724.
- Scharf ME, Wu-Scharf D, Pittendrigh BR, Bennett GW (2003). Caste- and development-associated gene expression in a lower termite. *Genome Biol.* 4(10): p. 62.
- Stenberg K, Bollok M, Reczey K, Galbe M, Zacchi G (2000). Effect of substrate and cellulase concentration on simultaneous saccharification and fermentation of steam-pretreated softwood for ethanol production. *Biotechnol. Bioeng.* 68(2): 204-210.
- Subramaniyan S, Prema P (2000). Cellulase-free xylanases from *Bacillus* and other microorganisms. *FEMS Microbiol. Lett.* 183(1): 1-7.
- Thongekkaew J, Ikeda H, Masaki K, Iefuji H (2008). An acidic and thermostable carboxymethyl cellulase from the yeast *Cryptococcus* sp. S-2: purification, characterization and improvement of its recombinant enzyme production by high cell-density fermentation of *Pichia pastoris*. *Protein Exp. Purif.* 60(2): 140-146.
- Tokuda G, Lo N, Watanabe H, Slaytor M, Matsumoto T, Noda H (1999). Metazoan cellulase genes from termites: intron/exon structures and sites of expression. *Biochem. Biophys. Acta*, 1447(2-3): 146-159.
- Tomme P, Warren RA, Gilkes NR (1995). Cellulose hydrolysis by bacteria and fungi. *Adv. Microb. Physiol.* 37: 1-81.
- Tomme P, Van Tilbeurgh H, Pettersson G, VanDamme J, Vandekerckhove J, Knowles J, Teeri T, Claeysens M (1988). Studies of the cellulolytic system of *Trichoderma reesei* QM 9414. Analysis of domain function in two cellobiohydrolases by limited proteolysis. *Euro. J. Biochem.* 170(3): 575-581.
- Tong CC, Cole AL, Shepherd MG (1980). Purification and properties of the cellulases from the thermophilic fungus *Thermoascus aurantiacus*. *Biochem. J.* 191(1): 83-94.
- Toyama N, Ogawa K (1975). Sugar production from agricultural woody wastes by saccharification with *Trichoderma viride* cellulase. *Biotechnol. Bioeng. Symp.* (5): 225-244.
- Uhlrig H (1998). Industrial enzymes and their applications. New York: John Wiley & Sons, Inc., p. 435.
- Villena GK, Gutierrez-Correa M (2006). Production of cellulase by *Aspergillus niger* biofilms developed on polyester cloth. *Lett. Appl. Microbiol.* 43(3): 262-268.
- Violot S, Aghajari N, Czjzek M, Feller G, Sonan GK, Gouet P, Gerday C, Haser R, Receveur-Brechot V (2005). Structure of a full length psychrophilic cellulase from *Pseudoalteromonas haloplanktis* revealed by X-ray diffraction and small angle X-ray scattering. *J. Mol. Biol.* 348(5): 1211-1224.
- Voragen AGJ, Heutink R, Pilnik W (1980). Solubilization of apple cell walls with polysaccharide degrading enzymes. *J. Appl. Biochem.* 2: 452-468.
- Warren RA (1996). Microbial hydrolysis of polysaccharides. *Annu Rev. Microbiol.* 50: 183-212.
- Watanabe H, Noda H, Tokuda G, Lo N (1998). A cellulase gene of termite origin. *Nat.* 394(6691): 330-331.
- Watanabe H, Tokuda G (2001). Animal cellulases. *Cell. Mol. Life Sci.* 58(9): 1167-1178.
- Wen Z, Liao W, Chen S (2005). Production of cellulase by *Trichoderma reesei* from dairy manure. *Bioresour. Technol.* 96(4): 491-499.
- Whitaker DR (1951). Purification of the cellulase of *Myrothecium verrucaria*. *Nat.* 168(4288): 1070-1071.
- Wilson DB, Irwin DC (1999). Genetics and properties of cellulases. *Advances in Biochemical Engineering/Biotechnology: Recent Progress in Bioconversion*, 65: 1-21.
- Ye XY, Ng TB, Cheng KJ (2001). Purification and characterization of a cellulase from the ruminal fungus *Orpinomyces joyonii* cloned in *Escherichia coli*. *Int. J. Biochem. Cell. Biol.* 33(1): 87-94.
- Zaldivar J, Nielsen J, Olsson L (2001). Fuel ethanol production from lignocellulose: a challenge for metabolic engineering and process integration. *Appl. Microbiol. Biotechnol.* 56(1-2): 17-34.
- Zhang YH, Lynd LR (2004). Toward an aggregated understanding of enzymatic hydrolysis of cellulose: noncomplexed cellulase systems. *Biotechnol. Bioeng.* 88(7): 797-824.