

Rethinking breast cancer screening strategies in resource-limited settings

*Galukande M¹, Kiguli-Malwadde E²

1. Department of Surgery, College of Health Sciences, Makerere University
2. Department of Radiology, College of Health Sciences, Makerere University

Abstract

The incidence of breast cancer in sub-Saharan nations is increasing. There is a worsening scarcity of Human Resource for Health in Uganda in particular and Sub Saharan Africa in general. Resources available for health care are predominantly spent on infectious disease care such as (HIV/AIDS, Tuberculosis and Malaria). These factors and more make the future of breast cancer care including screening in Sub Saharan African grim.

Although mass breast cancer screening by mammography has been proved to be efficacious in the developed nations of the world, this has not been replicated in the developing nations because mass screening is not yet possible for the reasons stated. This paper proposes an alternative to mammography mass screening.

Breast health programs for the most part are adhoc or non-existent in Uganda. The challenge of mass screening is not only limited to less readily available mammogram machines and trained human resources but also to the fact that the targeted population is of relatively young women in their 30s, implying that screening should commence earlier than it is practiced in nations where breast cancer peaks among women in their 50s. Mammography is not efficacious in young women with dense breast tissue. Ultra sound scans are not only up to 10 fold more available than mammography machines but are half the cost per examination.

Although using ultra sound Scan for screening for non-palpable lumps is not up to par with standard breast cancer care mammography. It may be better than nothing, may be beneficial in aiding early cancer diagnosis. This concept is akin to the 'task shifting' advocated by WHO. It is worth investigating use of ultra sound scan for mass screening for breast cancer in resource-limited environments. This is not in any way lowering standards of oncologic diagnosis but filling the otherwise unattended to gap, the unmet need.

Key words: breast, cancer, screening

African Health Sciences 2010; 10 (1): 89 - 92

Introduction

The use of mammogram in screening for breast cancer is a well established efficacious practice that is responsible for significant reduction of late presentation of breast cancers in women in the developed nations of the world^{1, 2}.

This practice of mass screening has been so effective that it is been strongly recommended for use in the developing nations and many work and look forwards to the day this will be possible. However it is difficult to predict when exactly this will be, yet there is an urgent need to screen as evidenced by increasing rates of up to 5% per year^{3, 4}.

This opinion piece advocates for mass use of cheaper, more widely available alternatives to mammography; Breast Self Examination and US Scan could be practical substitutes for mass screening in low resourced settings until such a time when we can afford mammograms and have more human resources for health. Currently the human resource for health ratios stand at 0.8 health workers per 1000 population at least 2.5/1000 is required to ease the human resource for health bottle neck⁵.

* Correspondence author

Dr. Moses Galukande
Department of Surgery
College of Health Sciences
Makerere University
P. O. Box 7072
Kampala, Uganda
Email: mosesg@img.co.ug

Table 1: Accuracy of Breast imaging modalities

Modality	Sensitivity	Specificity	Positive Predictive Value	Indications
Mammography	63-95% (>95% palpable, 50% impalpable, 83-92% in women older than 50 y) (decreases to 35% in dense breasts)	14-90% 90% palpable)	10-50% (94% palpable)	Initial investigation for symptomatic breast in women older than 35 years and for screening; investigation of choice for micro calcification
Ultrasonography	68-97% (palpable)	74-94% (palpable)	92% (palpable)	Initial investigation for palpable lesions in women younger than 35 years
MRI	86-100%	21-97% (<40%) primary cancer)	52%	Scarred breast, implants, multifocal lesions, and borderline lesions for breast conservation
Scintigraphy	76-95% (palpable) 52-91% (impalpable)	62-94% (94% impalpable)	70-83% (83% palpable, 79% impalpable)	Lesions larger than 1 cm and axilla assessment; may help predict drug resistance
Positron emission tomography (PET)	96% (90% axillary metastases)	100%		Axilla assessment, scarred breast, and multifocal lesions

Adopted from e-medicine website

The emerging picture

Breast cancer in sub-Saharan Africa runs a very aggressive course and has higher fatality rates compared to those in the western world; breast cancer occurs 10-15 years earlier in black women compared to their white counterparts^{6,7,8} a similar story is found among UK's Black women⁹. The resources available for Health Care in general are far less than the \$ 15 per person per year recommended to governments by WHO as the minimum. Overall projections indicate a worsening picture of cancer

epidemiology for the developing nations of the world, in terms of incidence and mortality¹⁰.

In a recent Ugandan study¹¹ the peak age for patients with breast cancer in Uganda is 30 -39 years, the majority of patients 77% presented late as stage III and IV. The incidence rate is going up in Uganda; it has tripled in the past three decades from 11 per 100,000 to 39.2 per 100,000¹² while it has levelled out and now mortality is decreasing in North America and Europe.

Table 2: Early detection and access to care

Level of resources	Detection method(s)	Evaluation goal
Basic	Breast health awareness (education ± self-examination) Clinical breast examination (clinician education)	Baseline assessment and repeated survey
Limited	Targeted outreach/education encouraging CBE for at-risk groups Diagnostic ultrasound ± diagnostic mammography	Down staging of symptomatic disease
Enhanced	Diagnostic mammography Opportunistic mammographic screening	Opportunistic screening of asymptomatic patients
Maximal	Population-based mammographic screening Other imaging technologies as appropriate: high-risk groups, unique imaging challenges	Population-based screening of asymptomatic patients

Constraints

Mammogram screening may not be wholly appropriate, since close to half of Ugandan women who need screening are 30 years and below (see figure 1). The average age for Ugandans is 15 years¹³. Mammography is generally not recommended for women below 35 years because women of this age and younger tend to have denser breasts making it more difficult to distinguish abnormal from normal tissue on the x-ray film⁸. For this reason mass screening is not entirely possible with mammogram use alone since nearly half of the eligible women would be left out because they are below 35 years. Only four mammogram machines exist for a population of 6 to 7 million eligible women (see table 3)^{11, 13}. Three of which are privately owned and attract a fee of \$ 25 per examination, a cost unaffordable by the average Ugandan woman or the government for that matter. The cost of a breast ultrasound scan is a little less than half the cost of a mammogram in the private sector. We may need to use the task-shifting concept⁵, can we then have USScan take on the task for screening, and can we take it away from the radiologist to the sonographer (non physician health worker)? Sonographers are less expensive, more likely to accept deployment out of the capital city, even though additional training for breast screening may be necessary. There are at least 30 radiologists in Uganda making a radiologist to patient ratio of (1:300,000) and at least 60 sonographers making a ratio of 1:150,000¹⁴.

USScan may have a relatively low sensitivity and specificity (see table 1) but this would be better than no screening at all. Isn't it prudent then to use it in the interim until such a time as when mammography will be widely available? But even if mammography were available to all women in Uganda, what about the half for which it may not be appropriate? (Until proven otherwise by research) Limited access to standard screening is a scenario not only unique to Uganda but common within the African sub continent.

Table 3: Population composition in relation to breast cancer screening

Population of selected group	Number	%
Women of child bearing age (15-49years)	5,476,435	2.4%
Women past child bearing age (50+ years)	1,006,547	4.1

Continuation of table 3

Population of selected group	Number	%
Women needing breast screening	6,483,082	26.5
Women 18 – 30 years	2,736,000	11.2
Secondary school age* (13 – 19 years)	3,995,884	16.3

* On average 50.1% are female

Source: 2002 Uganda Population Census - UBOS

Global initiatives

The Breast Health Global Initiative (BHGI)¹⁵ (Anderson et al, 2006) strives to develop evidence based, economically feasible and culturally appropriate guidelines that can be used in nations with limited health care resources to improve breast cancer outcomes. Table 2 highlights the proposed framework to fit the level of resources available to the different nations.

Early breast cancer detection improves outcome in a cost effective fashion assuming treatment is available. The BHGI group recommends future research to better determine the best way to implement guidelines in limited resources settings.

Possible country specific initiatives

Uganda has close to 2,000,000 girls in the 13-19 age bracket who are in school. The figure shown in table 3 for secondary school age 13-19years is for both sexes. A little more than half of which are female (50.1%) if these two million girls were to be screened it would not be with mammography.

Taking away the burden of imaging from the four available mammography machines to at least 60 US scans that exist in the country and mostly situated in or near district hospitals, is plausible and it is not meant to lower standards of oncological diagnosis but narrow the gap that exists.

Using US Scan for screening in this scenario is the major stopgap measure this opinion paper emphasizes. School campaigns for BSE (Breast Self Examination), for all girls are the other possibility¹⁶. The feasibility and subsequent impact of this ought to be investigated and documented. Table 1 indicates how the different investigating modalities compare in terms of sensitivity and specifically in places where they are available for use (a western nation).

Gathering accurate data about breast cancer in Uganda is critical for problem characterization and subsequent evidence based solutions as indeed recommended by BHGI.

Conclusion

Breast Cancer screening, as we know is mostly by the use of mammograms; mammograms are few in Uganda as in many sub Saharan poor countries and wont be enough in the foreseeable future to cover the unmet screening need. In the meantime the next best options include use of USScan and Breast Self Examination (though little evidence to support efficaciousness for both is lacking). Other unexplored options in resource limited settings are, task shifting by the involvement of non-physician cadre in Breast Health Care. There is therefore a need to investigate efficacy of ultrasound in breast cancer screening in resource-limited environments as well as a lower cut off age for mammography screening.

References

1. Moller B, Weedon-Fekjaer, H, Hakulinen, T, Tryggvadottir, L et al. The influence of mammographic screening on national trends in breast cancer incidence. *European Journal of Cancer Prevention*. April 2005; 14(2): 117-128.
2. Gøtzsche PC, Nielsen M. Screening for breast cancer with mammography. *Cochrane Database Syst Rev*. 2006 Oct 18; (4): CD001877.
3. Wabinga HR, Parkin DM, Wabwire-Mangen F, Namboozee S. Trends in cancer incidence in Kyadondo county, Uganda, 1960-1997. *British Journal of Cancer*. 2000; 82 :1585-1592.
4. Stewart B, Weihs PE. World Cancer Report. Lyon France: IARC Press, 2003.
5. WHO Report. Task shifting to tackle health worker shortages. 2007.3
http://www.who.int/healthsystems/task_shifting_booklet.pdf
6. Ellis P. Current issues in Cancer Management of carcinomas of the upper gastrointestinal tract. *BMJ*. 1994; 308:834-838.
7. Fregene A, Newman LA (2005) Breast cancer in sub-Saharan Africa: how does it relate to breast cancer in African-American women? *Cancer*. 103 (8) : 1540–1550.
8. Gakwaya A, Galukande M, Jombwe J, Fualal J, Lwanga A et al. Breast Cancer guidelines for Uganda (2nd Edition 2007) *African Health Sciences*. 2008; 8 (2): 126-132.
9. Bowen RL, Dutty SW, Ryan DA, Hart IR, Jones JL. Early onset of breast cancer in a group of British black women. *British Journal of Cancer*. 2008; 98: 277-281
10. WHO Report. Emro – Non-communicable diseases. 2007. <http://www.emro.who.int/ncd/>
11. Gakwaya A, Kigula-Mugambe JB, Kavuma A, Luwaga A, Fualal J, Jombwe J, Galukande M and Kanyike D. Cancer of the breast: 5-year survival in a tertiary hospital in Uganda. *British Journal of Cancer*. 2008; 99: 63 – 67.
12. International Agency for Research on Cancer (IARC). Efficacy of Screening by Self Examination in Hand book of Cancer Prevention. 2002; Vol 7. Breast Cancer Screening, Vainio H, Bianchini Freds. Lyon F France. IARC.
13. Uganda Bureau of Statistics (UBOS). The 2002 Uganda Population and housing Census – main report: March, Kampala: UBOS. <http://www.ubos.org/2002%20census%20Fianl%20Reportdoc.pdf>
14. Kawooya M. Imaging burdens, coverage and utilization of facilities. A case study of selected Hospitals in Uganda. PhD thesis, 2008.
15. Anderson BO, Shyyan R, Eniu A, Smith RA, Yip CH, Bese NS, et al. Breast cancer in limited-resource countries: an overview of the breast health global initiative 2005 guidelines. *Breast J* 2006; 12 Suppl 1:S3-S15
16. Hackshaw AK, Paul EA.. Breast Self Examination and death from breast cancer: a Meta-analysis. *British Journal of Cancer*. 2003; 88: 1047 – 1053.

Hepatocellular carcinoma and the underlying mechanisms

*Oyagbemi AA, Azeez OI, Saba AB

Department of Veterinary Physiology, Biochemistry and Pharmacology, Faculty of Veterinary Medicine, University of Ibadan, Oyo State, Nigeria.

Abstract

The incidence of hepatocellular carcinoma is increasing worldwide as well as the associated risk factors, some of which include exposure to aflatoxin B1, Hepatitis B (HBV) virus and hepatitis C (HCV) virus. Mutation of tumour suppressor gene p53 at codon 249^{ser} at exon 7 has been found to contribute significantly to replication of damaged DNA and subsequent tumour progression. The x gene of HBV (HBx) is the most common open reading frame integrated into the host genome in hepatocellular carcinoma and the integrated HBx is frequently mutated in hepatocellular carcinoma. Mutant HBx proteins still retain their ability to bind to p53 thereby attenuating DNA repair and p53-mediated apoptosis.

Keywords: hepatocellular carcinoma, aflatoxin B1, HBV, HCV, p53

African Health Sciences 2010; 10 (1): 93 - 98

Introduction

Hepatocellular carcinoma (HCC) is one of the most common malignancies worldwide. It is the fourth leading cause of cancer-related death in the world.¹ The major risk factors include chronic infections with the hepatitis B (HBV) or C (HCV) virus and exposure to dietary AFB1 or alcohol consumption. A link based on circumstantial evidence has been divulged between high exposure to AFBI and mutation at the 3rd nucleotide base of codon 249, which is located on the 7th exon of p53 gene of cells of primary liver cancer from patients in tropical countries of the world and activation of the WNT signal transduction pathway.²⁻⁶ AFB1 frequently induces G: C to T: A transversions at the third base in codon 249. Interestingly, mutant DNA in plasma is a biomarker of both AFBI exposure and potential risk factor for HCC with subsequent p53 mutation.⁷ The tumour suppressor gene *p53* is the most commonly mutated gene in human cancers.⁸

Chronic infections with HBV and HCV viruses and oxyradical disorders including hemochromatosis also generate reactive oxygen/nitrogen species that both damage DNA and mutate cancer - related genes such as tumour suppressor gene p53.⁹ The p53

biological network is a key responder to this oxidative and nitrosative stress. Depending on the extent of the DNA damage, p53 regulate transcription of protective antioxidant genes and the extent of DNA damage that ultimately trans-activates pro-oxidant genes which eventually contribute to apoptosis. The x gene of HBV (HBx) is the most common open reading frame integrated into the host genome in HCC and the integrated HBx is frequently mutated. Mutant HBx proteins still retain their ability to bind to p53 and attenuate DNA repair and p53-mediated apoptosis. Hence, both viruses and chemicals (especially vinyl chloride) are implicated in the etiology of p53 mutation during the molecular pathogenesis of HCC.

HCC is a major cause of cancer morbidity and mortality in many parts of the world, including Asia and Sub Saharan Africa, where there are >500,000 new cases each year and >200,000 deaths annually in the People's Republic of China (P.R.C) alone.¹⁰ The major etiological factors associated with development of HCC in these regions are infection with HBV and or HCV and long time exposure to high levels of AFBI in the diet.¹¹⁻¹²

Mechanisms underlying Hepatocarcinogenesis

The biology, mode of transmission, and epidemiology of HBV continue to be actively investigated and have been recently reviewed.¹³ A mutation in the HBV genome can alter the expression of multiple proteins. In many cases of HCC in China and Africa, a double mutation in the HBV genome, an adenine-to-thymine transversion at nucleotide 1762 and a guanine-to-adenine transition at nucleotide 1764 (1762^T/1764^A) has been found in tumours.¹⁴⁻¹⁵

*Corresponding author:

Oyagbemi A.A
Department of Veterinary Physiology, Biochemistry and Pharmacology
Faculty of Veterinary Medicine, University of Ibadan,
Oyo State, Nigeria.
E-mail: ademolaoyagbemi@yahoo.com,
aa.oyagbemi@mail.ui.edu.ng
Phone: +2348033639776
Fax: 02-8103043

This segment of the HBV genome contains an overlapping sequence for the base core promoter region and the HBx gene; therefore, the double mutation in codon 130 and 131 of the HBx gene reported in human HCC is identical to the 1762 and 1764 nucleotide changes.¹⁶ The onset of these mutations was shown to be associated with the increasing severity of the HBV infection and cirrhosis.¹⁴⁻¹⁵ HBx in transformed hepatocyte has been demonstrated to inhibit the repair of damaged hepatocyte DNA. This effect may be mediated by interaction with p53 or through binding to the damaged DNA binding protein (DDB), which plays an accessory role in nucleotide excision repair.¹³ In addition, HBx activates cell signalling cascades involving mitogen-activated protein kinase (MAPK) and Janus family tyrosine kinases (JAK)/signal transducer and activators of transcription (STAT) pathways.¹³ The process by which tumour DNA is released into circulating blood is unclear but may result from accelerated necrosis, apoptosis, or other processes.¹⁷ A specific codon 249 p53 mutation detectable in plasma samples at the time of HCC diagnosis, can be measured in some individual at least 5 years before diagnosis.¹⁸

Heterogeneity in etiological factors of HCC

The frequency of HCC is particularly high in Asia and Africa due to the high frequency of viral hepatitis infections and to Aflatoxin B1 exposure (AFB1). Over the last 10 years, the incidence of HCC has noticeably increased in United Kingdom, France and United States. This is probably linked to viral hepatitis C infections. Etiological factors that are associated with the development of hepatic tumours are well known in these regions. They include infection with the hepatitis B virus (HBV) or hepatitis C virus (HCV), heavy alcohol intake, prolonged dietary exposure to AFB1 or vinyl chloride and primary hemochromatosis. In 90% of the HCC cases, at least one of these risk factors can be identified either alone or in combination with another factor. The presence of each risk factor among patients varies according to the geographical origin of the patients. Globally, exposure to HCV, HBV and AFB1 are responsible for about 80% of all HCC in humans' worldwide but the principal risk factor varies between countries. In Japan almost all HCC are linked to HCV infection, whereas in Africa HBV infections are predominant.¹⁹ In France, HBV and HCV infections and alcohol intake are identified with approximately equal frequency. Exposure to AFB1 is commonly found

in sub-tropical countries where humid heat can lead to the development" of *Aspergillus flavus* in improperly stored foods such as cereals and peanuts. This mycotoxin is strongly hepatocarcinogenic in experimental animal models and acts synergistically with HBV infection to increase the risk of HCC.²⁰ Tobacco exposure is the leading carcinogen associated with multiple solid tumours²¹. Several investigators have previously reported an association between tobacco and HCC with odds ratios ranging from 1.5 to 6.8.²²⁻²³ However, other studies found no association between tobacco and HCC.²⁴

Hepatocarcinogenesis

The different risk factors of HCC include chronic lesions in the liver with associated inflammation, necrosis of hepatocytes and fibrosis. Overall, HCC development is closely associated with cirrhosis and more than 80% of the tumours are found in a chronic hepatitis or a cirrhotic background.²⁵ Dysplastic nodules and macroregenerative nodules have long been considered to be the likely precursors of HCC because of their frequent association with the HCC occurrence.²⁶ Chromosome aberrations occur in HCC and these may already contain genetic aberrations. However, in rare cases (less than 10% of the cases), HCC are observed in non-cirrhotic liver and even without inflammatory lesions. The HCC which develop in an otherwise normal liver are usually found in patients without well-established risk factors. Some of these cases may correspond to the malignant transformation of liver adenoma that are rare benign hepatocellular tumours sometimes found in young women taking oral contraceptives.²⁷

HBV infection

The incidence of HCC has been shown to vary widely worldwide.²⁸ Among males, the highest incidence rates are found in eastern Asia, particularly in China where HCC was reported to be the third most common cause of cancer death²⁹. Chronic infection with the HBV has been reported by various authors as the strongest risk factor for HCC worldwide.^{28, 30-32} However, populations with similar prevalence of HBV infection have different incidence of HCC, suggesting the presence of other important risk factors. Aflatoxins, a group of mycotoxins produced by the common fungi *Aspergillus flavus* and *Aspergillus parasiticus*, are established human hepatocarcinogens and are well-known HCC risk factors when present in foodstuffs.³²⁻³⁵ Some epidemiological and animal

studies have found evidence for an HBV-aflatoxin interaction in hepatocarcinogenesis.³⁵⁻⁴¹

Several mechanisms underlying this principle have been proposed to explain the interaction between HBV and aflatoxin. The increase in cellular proliferation induced by HBV could increase the probability for clonal expansion of an existing aflatoxin induced-*p53* 249^{ser} mutation⁴². An increase in levels of aflatoxin metabolism enzymes (*e.g.*, P450 enzymes in which its activity is associated with increased hepatotoxicity of aflatoxin) has been described for HBV transgenic mice and has been postulated as a mechanism for interaction.⁴³ The HBx protein, which is encoded by HBV interferes with the nucleotide excision repair pathway, a major repair pathway which cells use to repair damaged DNA.⁴⁴ However, the presence of mutant HBx protein could increase the frequency of aflatoxin-induced mutations.⁴⁴ Also, HBV infection was reported to increase oxidative stress, which could lead to an increase in *p53* mutations.⁴⁵

Mechanisms of HBV-mediated hepatocarcinogenesis

HBV infection can promote carcinogenesis by at least 3 different mechanisms. First, integration of the viral DNA in the host genome can induce chromosome instability. Second, insertional mutations of HBV are known to activate endogenous genes of retinoic acid α -receptor, cyclin A and mevalonate kinase which are involved in cell cycle control, cellular proliferation and differentiation. The second mechanism is associated with specific intracellular receptors. Recently, 15 new genes were found to be altered by HBV integration in tumors suggesting that viral integration in the vicinity of genes controlling cell proliferation, viability and differentiation is a mechanism frequently involved in HBV hepatocarcinogenesis. The third mechanism of carcinogenesis linked to HBV infection is based on the expression of viral protein, in particular HBx, to modulate cell proliferation and viability. Moreover, HBx binds to *p53* and inactivates *p53*-dependent activities, including *p53*-mediated apoptosis. Recently, the association between hepatitis B virus and Hepatocellular carcinoma and the molecular mechanism of action that is involved in the hepatocarcinogenesis has been extensively described.^{46,47}

Interaction of AFB1 with DNA and chromatin proteins (histone)

After an exposure to AFB1, accumulations of damaged DNA are found in the liver, as a result of conversion of the AFB1 to its active metabolites. AFB1 is a very potent mutagen and the AFB1 epoxides (active metabolites of aflatoxin) react with guanine in DNA, leading to genetic changes. The most frequent mutation induced is the (guanine-cytosine to thymine-adenine) GC to TA transversion. However, quantitative determination of AFB1 in human aflatoxin albumin adducts has been elucidated.⁴⁸ The mutational pattern of *p53* gene in HCC from regions where AFB1 exposure level is high, revealed (guanine to thymine) G to T transversion at codon 249 in more than 50% of the cases. A more detail study revealed that AFB1 binds preferentially to lysyl amino acid residues in histone proteins.¹⁰ The binding of AFB1 to histone proteins has significant functional implications because histone has been reported to be the packaging material for DNA and histone H1 is the most external of the histone proteins wrapped around DNA.^{10,49} Because of the high content of basic amino acids in histones, it is conjectured that there is a strong electrostatic interaction between them and DNA and that (addition of acetyl group) acetylation of the lysyl sites which is involved in this type of interaction, reduces the net positive charge of the histone and loosens the bonds between histone and DNA. Acetylation is reported to occur at the amino group of lysyl amino acid residues which is the same binding site of AFB1.^{50, 10}

The effect of AFB1 binding to histone is therefore likely to be similar to reaction elicited by Acetylation (a post transcriptional modification), which is the partial loosening of the histone-DNA bond and the consequent degradation of the histone by specific proteases.⁵¹ It is generally accepted that such a partial loosening of the histone DNA bonds always precedes gene expression. This means that it is most likely that it is the binding of AFB1 to lysyl amino acid residues in histone with the consequent loosening of the histone-DNA bond that makes *p53* accessible for damage. It is also likely that the binding of AFB1 to histones with the consequent loosening of histone is primary to its binding to the DNA of *p53* genes even though the binding to DNA subsequently exceeds its binding to histone. Taken together, the binding of AFB1 to DNA is responsible for the inhibition of RNA Synthesis, which is involved in gene expression.

The implication of the above is that it is the binding to chromatin proteins (histone) may be involved in the expression of the mutated p53 gene resulting from the interaction of AFB1 with DNA and chromatin proteins. The p53 gene is reported to be mutated in HCC after exposure to aflatoxin.⁵² Recently, some authors have extensively discussed the association between AFB1 and the associated risk factors involved in HCC.⁵³⁻⁵⁵

Conclusion

The nexus between hepatocellular carcinoma and the associated risk factors cannot be overemphasized. The interaction between aflatoxin and HBV or HCV in hepatocarcinogenesis and multi-stage carcinogenesis is grossly elucidated. Characterization of the genetic alterations associated with HCC tumors is an essential step to increase our knowledge of hepatocarcinogenesis. Systematic search for these alterations in series of tumors including tumour grades, stages, etiologies and the associated pre-neoplastic lesions is therefore necessary to find and identify the pattern of accumulation of the genetic alterations during tumour progression. Microarray analysis and metagenomics may also contribute significantly to identifying new carcinogenetic pathways altered in these tumors. New insight should therefore be geared towards getting a better clinical application, to identify tumour markers that are useful for early detection of tumors, to predict prognosis, or to find new therapeutic targets with their underlying molecular mechanism of action.

References

1. Murugavel KG, Naranatt PP, Shankar *et al.*: Prevalence of aflatoxin B1 in liver biopsies of prove hepatocellular carcinoma in India determined by an in-house immunoperoxidase test. *J Med Microbiol* 2007; 56(Pt 11):1455 [PubMed](#) -9.
2. Mariana CS, David MU, Mimi CYu *et al.*: Hepatitis B, Aflatoxin B₁, and p53 Codon 249 Mutation in Hepatocellular Carcinomas from Guangxi, People's Republic of China, and a Meta-analysis of Existing Studies. *Cancer Epidemiology Biomarkers & Prevention* 2001; 10: 617-625.
3. Bressac B, Kew M, Wands J, Ozturk M: Selective G to T mutations of p53 gene in hepatocellular carcinoma from southern Africa. *Nature (Lond.)*, 1991; 350: 429-431.
4. Hsu IC, Metcalf RA, Sun T *et al.*: Mutational hotspot in the p53 gene in human hepatocellular carcinomas. *Nature (Lond.)*. 1991; 350: 427-428.
5. Heyward N, Walder G, Graham W, Cooksley, E. Hepatocellular carcinoma mutation (Letter) *Nature (Lond.)* 1996; 350: 427-428.
6. Ozturk M, Bressac B, Puisieux *et al.*: P53 mutation in hepatocellular carcinoma after aflatoxin exposure. *Lancet*. 1991; 338: 113-122.
7. Shuang YK, Suree L, Niwat *et al.*: Hepatitis B 1762^T/1764^A Mutations, Hepatitis C Infection, and Codon 249 p53 Mutations in Hepatocellular Carcinomas from Thailand. *Cancer Epidemiology Biomarkers & Prevention* 2005; 14: 380-384.
8. Greenblatt MS, Bennett W P, Hollstein M, Harris C C: Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res*. 1994; 54: 4855-4878.
9. Paradis V, Mathurin P, Kollinger M *et al.*: In situ detection of lipid peroxidation of Chronic hepatitis C: correlation with pathological features. *J Clin Pathol* 1997; 50: 401-406.
10. Li ND, Lu FZ, Zhang SW, Mo R, Sun XD: Trends and prediction in malignant tumors mortality in past 20 years in China.. *Chin. J Oncol* 1997; 19: 3-9.
11. Qian GS, Yu MC, Ross *et al.*: DA follow-up study of urinary markers of aflatoxin exposure and liver cancer risk in Shanghai, P.R.C. *Cancer Epidemiol Biomark Prev* 1994; 3: 3-11.
12. Kensler TW, Qian GS, Chen JG, Groopman JD: Hepatocarcinogenesis as a paradigm for cancer prevention. *Nat Rev Cancer* 2003; 3:321-29.
13. Arbuthnot P, Capovilla A, Kew M: Putative role of hepatitis B virus X protein in hepatocarcinogenesis: effects on apoptosis, DNA repair, mitogen-activated protein kinase and JAK/STAT pathways. *J Gastroenterol Hepatol* 2000; 15(4):339-41
14. Hsu IC, Metcalf RA, Sun T, Welsh JA, Wang NJ, Harris CC: Mutational hotspot in the p53 gene in human hepatocellular carcinomas. *Nature (Lond.)*, 1991; 350: 427-428.
15. Baptista M, Kramvis A, Kew MC: High prevalence of 1762(T) 1764(A) mutations in the basic core promoter of hepatitis B virus isolated from black Africans with hepatocellular

- carcinoma compared with asymptomatic carriers. *Hepatology* 1999; 29:946–53.
16. Zhang F, Zhu Y, Sun Z: Universal presence of HBVx gene and its close association with hotspot mutation of p53 gene in hepatocellular carcinoma of prevalent area in China. *Zhonghua Zhongliu Zazhi* 1998; 20:18-21.
 17. Anker P, Mulcahy H, Chen XQ, Stroun M: Detection of circulating tumor DNA in the blood (plasma/serum) of cancer patients. *Cancer Metastasis Rev.* 1999; 18:65–73.
 18. Jackson PE, Kuang SY, Wang J Bet al.: Prospective detection of codon 249 mutations in plasma of hepatocellular carcinoma patients. *Carcinogenesis* (2003; 24:1–7.
 19. Bosch FX, Ribes J, Díaz M, Cléries R: Primary liver cancer: worldwide incidence and trends. *Gastroenterology* 2004; 127(5 Suppl PubMed 1):S5-S16.
 20. Chen CJ, Wang LY, Lu *et al.* Elevated aflatoxin exposure and increased risk of hepatocellular carcinoma. *Hepatology* 1996; 24:38-42.
 21. Adami HO, Hsing AW, McLaughlin *et al.*: Alcoholism and liver cirrhosis in the etiology of primary liver cancer. *Int J Cancer* 1992; 51:898-902.
 22. Hadzlyannis S, Tabor E, Kaklamani *et al.*: A case-control study of hepatitis B and C virus infections in the etiology of hepatocellular carcinoma. *Int J Cancer* 1995; 60:627 -631.
 23. Tzonou A, Trichopoulos D, Kaklamani E *et al.*: Epidemiologic assessment of interactions of hepatitis-C virus with seromarkers of hepatitis-B and D viruses, cirrhosis and tobacco smoking in hepatocellular carcinoma. *Int J Cancer* 1991; 377-380.
 24. Austin H, Deizell E, Grufferman *et al.*: A case-control study of hepatocellular carcinoma and the hepatitis B virus, cigarette smoking and alcohol consumption. *Cancer Res* 1986; 46:962-966.
 25. Yu MW, Hsu FC, Sheen IS *et al.*: Prospective Study of Hepatocellular Carcinoma and Liver Cirrhosis in Asymptomatic Chronic Hepatitis B Virus Carriers. *American Journal of Epidemiology* 1997; 145(11): 1039 PubMed -1047.
 26. Terada T, Veda K, Nakanuma Y: Histopathological and morphometric analysis of atypical adenomatous hyperplasia of human cirrhotic livers. *Virchows Arch A Pathol Anat Histopathol* 1993; 422:381-8.
 27. Ham JM, Stevenson D, Liddelow AG: Hepatocellular carcinoma possibly induced by oral contraceptives. *Digestive Diseases and Sciences* 2005; 1575-2568.
 28. Yeh F-S, Yu MC, Mo C-C, Tong MJ, Henderson BE: Hepatitis B virus, aflatoxins, and hepatocellular carcinoma in Southern Guangxi, China. *Cancer Res* 1989; 49: 2506–2509.
 29. National Cancer Control Office: Nanjing Institute of Geography. Atlas of Cancer Mortality in the People's Republic of China. Shanghai: China Map Press. 1979.
 30. Blumberg BS, and London WT: Hepatitis B virus and the prevention of primary cancer of the liver. *J Natl Cancer Inst* (Bethesda) 1985; 74: 267–273.
 31. London WT, Evans AA, Buetow *et al.*: Molecular and genetic epidemiology of hepatocellular carcinoma: studies in China and Senegal. Princess Takamatsu Symp. 1995; 25: 51–60.
 32. Vainio H, Heseltine E, Wilbourn J: Report on an IARC working group meeting on some naturally occurring substances. *Int J Cancer* 1993; 53: 535–537.
 33. Busby WF, and Wogan GN: Aflatoxins. In: C. E. Searle (ed). *Chemical Carcinogens* 1984; 2:945–1136 PubMed . Washington D. C.: American Chemical Society
 34. Groopman JD: Do aflatoxin-DNA adduct measurements in humans provide accurate data for cancer risk assessment? *IARC Sci Publ* 1988; 89: 55–62.
 35. Qian GS, Yu MC, Ross *et al.*: A follow-up study of urinary markers of aflatoxin exposure and liver cancer risk in Shanghai, P.R.C *Cancer Epidemiol Biomark Prev* 1994; 3:3-11.
 36. Lunn RM, Zhang Y-J, Wang *et al.*: P53 mutations, chronic hepatitis B virus infection, and aflatoxin exposure in hepatocellular carcinoma in Taiwan. *Cancer Res* 1997; 57: 3471–3477.
 37. Ross RK, Yuan JM, Yu *et al.*: Urinary aflatoxin biomarkers and risk of hepatocellular carcinoma. *Lancet* 1992; 339: 943–946.
 38. McGlynn KA, Rosvold EA, Lustbader *et al.*: Susceptibility to hepatocellular carcinoma is associated with genetic variation in the enzymatic detoxification of aflatoxin B1. *Proc Natl Acad Sci USA* 1995; 92: 2384–2387.
 39. Dragani TA, Manenti G, Farza H, Della-Porta G, Tiollais P, and Pourcel C. Transgenic mice

- containing hepatitis B virus sequences are more susceptible to carcinogen-induced hepatocarcinogenesis. *Carcinogenesis* (Lond.) 1990; 11: 953–956.
40. Sell S, Hunt JM, Dunsford HA, Chisari FV: Synergy between hepatitis B virus expression and chemical hepatocarcinogens in transgenic mice. *Cancer Res* 1991; 51: 1278–1285.
 41. Montesano R, Hainaut P, and Wild CP: Hepatocellular carcinoma: from gene to public health. *J. Natl. Cancer Inst. (Bethesda)* 1997; 89: 1844–1851.
 42. Chisari FV, Klopchin K, Moriyama *et al.*: Molecular pathogenesis of hepatocellular carcinoma in hepatitis B virus transgenic mice. *Cell* 1989; 59: 1145–1156.
 43. Kirby GM, Chemin I, Montesano R, Chisari FV, Lang MA, and Wild CP: Induction of specific cytochrome P450s involved in aflatoxin B metabolism in hepatitis B virus transgenic mice. *Mol Carcinog* 1994; 11: 74–80
 44. Jia L, Wang X W, and Harris CC: Hepatitis B virus, X protein inhibits nucleotide excision repair. *Int J Cancer* 1999; 80: 875–879.
 45. Hussain S P, Aguilar F, Amstad P, and Cerutti P: Oxy-radical induced mutagenesis of hotspot codons 248 and 249 of the human *p53* gene. *Oncogene* 1994; 9: 2277–2281.
 46. Liu ZM, Li LQ, Peng *et al.*: Hepatitis B virus infection contributes to oxidative stress in a population exposed to aflatoxin B1 and high-risk for hepatocellular carcinoma. *Cancer Lett* 2008; 263(2):212-22.
 47. Wu HC, Wang Q, Yang *et al.*: Urinary 15-F2t-isoprostane, aflatoxin B1 exposure and hepatitis B virus infection and hepatocellular carcinoma in Taiwan. *Carcinogenesis*. 2008; 29(5):971-6.
 48. McCoy LF, Scholl PF, Sutcliffe *et al.*: Human aflatoxin albumin adducts quantitatively compared by ELISA, HPLC with fluorescence detection, and HPLC with isotope dilution mass spectrometry. *Cancer Epidemiol Biomarkers Prev* (2008; 17(7):1653-7.
 49. Stelolwagen RH, and Cole RD: Chromosomal proteins *Ann Rev Biochem* 1969; 38: 951.
 50. Sung MT, and Dixon GH. Modifications of histones during spermiogenesis in trout: A molecular mechanism or altering histone binding to DNA *proc. Natll. Acad. Sci U.S.A.* 1970; 67: 1616.
 51. Candido EPM, and Dixon GH. Trout testis cells: Acetvlation of histones in different cells types from developing trouts tests. *J Biol Chem* 1972; 247: 5506.
 52. Paget V, Sichel F, Garon D, Lechevrel M: Aflatoxin B1-induced TP53 mutational pattern in normal human cells using the FASAY (Functional Analysis of Separated Alleles in Yeast). *Mutat Res* 2008; 656(1-2):55-61.
 53. Abdel-Wahab M, Mostafa M, Sabry M, el-Farrash M, Yousef T: Aflatoxin as a risk factor for hepatocellular carcinoma in Egypt, Mansoura Gastroenterology Center study. *Hepatogastroenterology* 2008; 55(86-87):1754-9.
 54. Murugavel KG, Naranatt PP, Shankar *et al.*: Prevalence of aflatoxin B1 in liver biopsies of proven hepatocellular carcinoma in India determined by an in-house immunoperoxidase test. *J Med Microbiol* 2007; 56(Pt 11):1455 [PubMed](#) -9.
 55. Long XD, Ma Y, Qu de *et al.*: The polymorphism of XRCC3 codon 241 and AFB1- related hepatocellular carcinoma in Guangxi population, China. *Ann Epidemiol* 2008; 18(7):572- 8.