

INVENTORY OF SOME ENVIRONMENTAL COMPONENTS IN THE TERRESTRIAL AND AQUATIC ECOSYSTEMS OF THE INTEGRATED WASTE TREATMENT FACILITY MAKURDI BENUE STATE, NIGERIA.

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ABSTRACT

*A survey of some environmental components in the terrestrial and aquatic ecosystems in the Integrated Waste Treatment Facility in Makurdi was carried out to obtain baseline information on the area. Inventory, semi-structured interviews and field observations/walks were carried out to obtain information on useful plants, their abundance and distribution within the facility. The micro and macro flora of the facility was sampled and analyzed using the serial dilution method. 77 plant species consisting of 23 species of trees, 6 shrubs, 34 herbs, 3 woody climbers, 3 climbers, 2 woody herbs, bryophytes, mushrooms and lichens were identified and recorded. 17 animal species consisting of 9 mammals, 4 rodents and 4 fish species have being seen and caught within the facility and in Jamo stream. The microbial status of the water indicated that total coliforms ranged from 3.50×10^5 to 9.00×10^5 cfu/ml faecal coliforms ranged between 4.5×10^4 to 2.75×10^5 cfu/ml. These indicate a high level of contamination of the water from animal and organic matter and may harbour human pathogenic microorganisms. The organisms identified were *Escherichia coli*, *Aerobacter aeroginesa*, *Faecal streptococcus*, *Bacillus aureus*. Others were *Escherichia ferundii*, *Salmonella typhosa*, *Pseudomonas aerogenosa*, *Aspergillus spp*, *Penicillium spp* and yeasts.*

INTRODUCTION

Environmental Impact Assessments (EIA) are carried out to determine whether statutory requirements will be met or whether a development project will be deemed environmentally acceptable when fully operational (Harris *et al* 1996). The EIA of the proposed Integrated Waste Treatment Facility (IWTF) in Makurdi was carried out as an integral part of the design, planning and approval process by the Technical Working Group (TWG) of the Department For International Development (DFID) funded State and local Government Programme (SLGP). The total area of the IWTF was found to be 4.42ha with the lowest contour occurring at 96m while the highest contour occurred at 109m and a mean annual precipitation of 1093mm. The disposal of refuse by microorganisms is one of the oldest biological waste treatment systems. Man has buried

unwanted refuse for centuries and the microorganisms have broken the complex organic compounds into simple compounds which have been reused by higher plants. Today, there are two classes of refuse disposal by microorganisms; sanitary landfill and composting. In either case, the operation relies partly on microbial activities to transform the waste materials. One major problem in handling municipal waste is that microbial activities are well advanced before the materials reach the composting area. Thus, the waste materials constitute a potential source of pathogenic and non-pathogenic organisms. When rain water hits the ground, a portion of it runs off below and above the ground surface. It picks up many substances as it flows into rivers, streams, or ponds, microorganisms, organic matter and minerals. Being rich in nutrients, it becomes a perfect medium for the growth of

microorganisms. Water has long served as a mode of transmission of diseases. The common human pathogenic bacteria and protozoa found in water include *Salmonella* spp, *Shigella dysenteriae*, *Vibrio cholerae*, *Entamoeba histolytica* and viruses. The main objective of the study was to identify and understand the major environmental components in the proposed facility in order to provide information for effective impact analysis and mitigation.

METHODOLOGY

Species inventory, abundance and distribution

Investigation of the floristic composition was carried out to generate information on plant distribution, abundance, economic importance/values, among others of the proposed site and surrounding area. Eight quadrates made up of a pair of "reference" plots each were situated within and outside the proposed project site to identify plant species within the reference plots and the number of plants per species in each plot. The reference plots were marked with iron plates indicating the plot number and planted at the edges of the plots. The quadrate size was determined and species area curve for several quadrates sizes were plotted and 10m² was adjudged the best. The method of Onyekwelu and Okafor (1979) was employed in laying the quadrates. In each quadrate, all the plant species were identified and counted. Those plant species difficult to identify on the spot were taken to the Herbarium of the Department of Biological Sciences, University of Agriculture Makurdi for identification and checked with Agishi (2004), Keay (1989) and determination keys provided by Arbonnier (2004). Inventory of species, semi-structured interviews and field

observations (Balemie and Kebebew, 2006) were conducted with local residents around the study area to obtain information on plants and animals utilized within the area. Active users of wild plants made up of herbalists, hunters, house wives, young people and the aged in the area participated in the interview. The people made a collection of all useful wild plants in the facility and assembled them for identification. The plants were identified and classified according to use categories.

Microbial Evaluation of Soil and Water in the study area

Soil samples were taken within the project site and at the reference plots/quadrates. Each of the samples was wrapped in aluminum foil and analyzed in the laboratory by serial dilution method. The samples were inoculated with nutrient agar and potato-dextrose agar for bacteria and fungi respectively. The emerging colonies after three days of incubation were counted and recorded. The colonies were also examined individually by biochemical tests to identify the species of the organisms. Water samples were collected from Jamo stream in five different locations at 100m intervals, thoroughly homogenized by mixing and tested immediately to avoid contamination. The multiple tube fermentation method using the Most Portable Number (MPN) for coliform bacteria was adopted in the analyses. Prior to the sample collection, a sterile broth was prepared using MacCoukey broth in fermentation tubes. The tubes were then inoculated with the samples and incubated at 35°C for 48 hrs for total coliform bacteria. These were incubated at 44°C in the incubator to obtain results for faecal coliform bacteria count. Soil samples were taken from the same quadrates used for flora evaluation. Samples were also taken

within the project site and reference plots/quadrates referred to as Eco-Control. Each of the samples were wrapped in an aluminum foil, taken to the laboratory and treated within 2 hours after collection (Oshode *et al*, 2008) and analyzed by serial dilution of the samples in accordance with Paul and Clark (1988). The samples were inoculated on nutrient agar and potato-dextrose agar for bacteria and fungi respectively. The emerging colonies after three days of incubation were counted and recorded. The pure cultures of the colonies were also examined by biochemical tests to identify the species of the organisms.

RESULTS

Table 1: Species abundance and distribution in the study area

Species	PLOT I		PLOT II		PLOT III		PLOT IV		TOTAL
	WITHIN	CONTROL	WITHIN	CONTROL	WITHIN	CONTROL	WITHIN	CONTROL	
<i>Asystasia calycina</i> , H	-	-	-	-	-	-	-	10	10
<i>Lannea schimperi</i> , T	1	-	2	-	-	-	1	-	4
<i>Annona senegalensis</i> , S	1	7	-	-	-	-	2	-	10
<i>Voacanga Africana</i> , T	-	-	-	1	-	-	-	1	2
<i>Spathodea campanulate</i> ,T	-	1	-	-	-	4	-	-	5
<i>Stereospermum kunthianum</i> ,T	6	-	-	-	-	-	1	-	7
<i>Chamaecrista mimosoides</i> , H	40	-	100	10	30	-	10	-	190
<i>Maytenus senegalensis</i> ,	1	1	-	-	-	-	-	10	12
<i>Anogeissus leiocarpus</i> ,T	-	1	5	-	-	-	-	2	8
<i>Combretum nigricans</i> , T	3	-	-	-	2	-	-	1	6
<i>Terminalia schimperiana</i> , T	-	-	-	-	-	-	-	-	-
<i>Aspilia helianthoides</i> , H	-	100	100	-	-	200	-	-	400
<i>Chromolaena odorata</i> , WH	-	50	-	-	4	1	-	50	105
<i>Tridax procumbens</i> , S	-	-	-	-	300	-	-	-	300
<i>Vernonia amygdalina</i> , S	-	-	-	-	1	-	-	-	1
<i>Antidesma venosum</i> , T	-	-	-	-	-	1	-	-	1
<i>Bridelia ferruginae</i> , T	10	1	-	-	2	-	-	-	13
<i>Bridelia scleroneura</i> , H	-	-	-	-	-	-	1	-	1
<i>Euphorbia heterophylla</i> , T	-	-	-	-	200	-	-	-	200
<i>Fluggea virosa</i> , T	1	2	-	2	2	-	-	-	7
<i>Phyllanthus amarus</i> , H	-	-	-	-	-	1	-	-	1
<i>Calopogonium mucunoides</i> , H	-	-	-	1	-	-	-	-	1
<i>Crotalaria retusa</i> , H	80	-	-	1	-	-	-	-	81
<i>Desmodium velutinum</i> , H	-	-	-	-	-	-	3	10	13
<i>Eriosema psoraleoides</i> , WH	-	3	-	-	-	-	-	-	3
<i>Indigofera dendroides</i> ,	-	-	-	-	4	-	-	-	4
<i>Indigofera heudelotii</i> ,	-	-	-	-	-	1	-	-	1
<i>Mucuna poggei</i> , H	-	-	-	-	80	-	-	-	80
<i>Tephrosia bracteolata</i> , H	100	100	200	10	300	-	50	-	760
<i>Hymenocardia acida</i> , T	3	-	-	-	-	-	-	-	3
<i>Spigelia anthelmia</i> , H	-	-	-	2	-	-	-	-	2
<i>Hibiscus aspera</i> , H	100	-	-	-	-	-	-	-	100
<i>Sida rhombifolia</i> , H	-	-	-	-	-	-	-	-	-
<i>Azadirachta indica</i> , T	1	10	-	-	-	1	-	10	22
<i>Khaya senegalensis</i> , T	1	-	-	-	-	-	-	-	1
<i>Pseudoecedrela kotchyi</i> , T	-	-	-	-	-	-	30	-	30
<i>Trichilia emetic</i> , T	6	-	-	-	-	-	-	-	6

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<i>Cissampelos mucronata</i> , C	-	-	-	2	-	-	-	-	2
<i>Acacia polyacantha</i> , T	-	-	-	1	-	-	-	-	1
<i>Parkia biglobosa</i> , T	1	4	-	-	-	-	1	1	7
<i>Prosopis Africana</i> , T	1	1	-	-	-	-	1	-	3
<i>Ficus sur</i> , T	-	1	-	-	-	-	-	-	1
<i>Elaeis guineensis</i> , T	-	-	-	-	1	2	-	-	3
<i>Borreria ocymoides</i> , H	-	-	-	-	10	-	-	-	10
<i>Gardenia ternifolia</i> , T	-	-	1	-	-	-	-	-	1
<i>Sarcocephalus latifolius</i> , S	-	-	2	-	1	-	-	-	3
<i>Spermacoce ruelliae</i> , H	80	4	100	-	-	-	30	-	214
<i>Allophylus africanus</i> , T	-	-	-	-	-	-	-	10	10
<i>Sterculia setigera</i> , T	-	1	-	-	-	-	-	2	3
<i>Watheria indica</i> , H	60	-	-	3	-	-	-	-	63
<i>Triumfetta rhombodea</i> , H	-	20	-	-	-	-	-	-	20
<i>Triumfetta tomentosa</i> , H	-	-	-	-	-	4	-	-	4
<i>Clerodendrum capitatum</i> , WC	-	-	-	-	-	-	-	20	20
<i>Lippa mutifloar</i> , WH	-	2	-	-	-	-	-	-	2
<i>Vitex doniana</i> , T	1	-	-	-	-	-	-	-	1
<i>Cissus ibuensis</i> , C	-	2	-	-	-	-	-	-	2
<i>Cissus populnea</i> , WC	-	-	-	1	-	-	-	-	1
<i>Cissus rufescens</i> , WC	-	-	100	2	-	-	2	10	114
<i>Scadoxus multiflorus</i> , H	-	-	30	-	20	-	-	-	50
<i>Anchomanes difformis</i> , H	-	-	-	-	-	-	-	10	10
<i>Commelina henahalensis</i> ,	2	-	-	2	-	-	-	300	304
<i>Cyperus dilatatus</i> , H	90	-	-	-	10	-	-	-	100
<i>Mariscus alternifolius</i> , H	-	4	-	5	-	-	-	-	90
<i>Dioscorea bulbifera</i> , H	-	-	-	-	-	-	-	10	10
<i>Dioscorea dumentorum</i> , H	-	1	-	-	-	-	-	4	5
<i>Acroceras zizanioides</i> , H	-	-	-	-	-	-	-	100	100
<i>Andropogon gayanus</i> , H	-	-	-	80	-	-	-	-	80
<i>Digitaria horizontalis</i> , H	200	-	-	-	-	100	-	-	300
<i>Imperata cylindrical</i> , H	-	100	-	100	400	-	-	-	600
<i>Rottbellia cochinchinensis</i> , H	100	300	300	100	300	200	-	10	1310
<i>Setaria barbata</i> , H	200	-	-	40	-	100	-	-	340
<i>Setaria anceps</i> ,	-	300	100	100	200	200	300	-	1200
<i>Sorghum bicolor</i> , H	-	-	-	-	4	-	-	-	4
<i>Viteveria fluvibarbis</i> , H	-	-	-	-	-	100	-	-	100
<i>Tacca involucrate</i> , H	-	1	-	2	-	2	1	1	7
<i>Daniellia oliveri</i> , T	25	1	-	2	-	-	-	-	28
<i>Byrsocarpus coccineus</i> , H	60	-	-	-	-	-	-	-	60
Total									7303

Note: T-Trees, H- Herbs, S- Shrubs, WC- Woody Climber, WH- Woody Herbs

Table 2 : Medicinal plants in study area

Botanical name	Vernacular name (Tiv)	Plant part used	Cure
<i>Ageratum conyzoides</i>	Ngo-kwase	Whole plant ground and applied on affected area.	Body swelling
<i>Strychnos spinosa</i>	Maku	Fresh leaves squeezed	Blood clotting
<i>Hyptis spigera</i>	Eem	Squeezed leaves inhaled	Headache
<i>Laggera aurita</i>	Ijondugh	Squeeze leaves and rub	Rheumatism
<i>Trema orientalis</i>	Chiese	Bath with whole plant boiled	Small pox
<i>Chromolenea odorata</i>	Gbokpai	Whole plant steamed and inhaled	Fever
<i>Coclospermum linktorum</i>	Kpavande	Make a solution of ground roots	Yellow fever and Typhoid
<i>Parkia biglobosa</i>	Nune	Grind bark	Diarrhoea
<i>Cassia spectabilis</i>	Ngaji	Boiled roots and drink	Yellow fever
<i>Hymenocardia acida</i>	Iikwar	Boil leaves/stem, drink	Stomach ache
<i>Boerhavia repens</i>	Mgbera-yongo	Grind underground bulb, eat	Stomach ache, penis erection
<i>Elusine indica</i>	Kangeraka	Grind whole plant	Dislocation, cuts
<i>Tylophora systrica</i>			
<i>Khaya senegalensis</i>	Haa	Squeeze leaves, rub	Headache
<i>Byrsocarpus coccineus</i>	Hwer-baa	Grind leaves	Waist pain
<i>Anona senegalensis</i>	Ahur	Boil whole plant	Reptile & sake bite
<i>Pandiaka inolucrata</i>	Ahambe-akwator	Squeeze whole plant	Catarrh
<i>Azadirachta indica</i>	Dogonyaro	Boil leaves	Fever
<i>Bridelia ferruginea</i>	Ikpine	Boil leaves, bark and root	Hepatitis
<i>Hyptis suaveolens</i>	Human	Boil whole plant	Mosquito
<i>Vitellaria paradoxa</i>	Chamegh	Seed oil	Repellant relaxer
<i>Lonchocopus laxiflorus</i>	Gbagbongum	Ground leaves and stem	Rheumatism
<i>Prosopis africana</i>	Gbaaye	Ground bark	Skin burn
<i>Syzygium guineense</i>	Alom	Boil root	Hernia
<i>Parinari polyora</i>	Ibua	Boil root, leaves, and bark	Stomach and tooth ache

<i>Triculia emetica</i>	Gbur	Boil root	Snake bite
<i>Imperata cylindrica</i>	Ihila	Boil root	Worm infection
<i>Sarcocephalus latifolius</i>	Ikura	Cold bath with ground leaves	Measles
<i>Stereospermum kunthianum</i>	Anema-atumba	Cut stem in water	Stomach ache
<i>Uraria picta</i>	Gyaase	Ground leaves	Heart burn
<i>Cynastrum nodiflora</i>	Mundu-kunankwa	Ground bulb, rub	Skin burn
<i>Vernonia amygdalina</i>	Ituna	Squeeze whole plant, drink	Fever
<i>Pericopsis laxiflora</i>	Giragba	Boil root, drink	Stimulant
<i>Daniela oliveri</i>	Chiha	Ground tender leaves, drink	Diarrhea
<i>Ipomia batatas</i>	Atsaka	Soak whole plant	Anemia
<i>Piliostigma thonningi</i>	Nyihar	Boil whole plant	Stomach ache
<i>Cissus ibuesis</i>	Dedooko	Boil whole plant	Stomach ache
<i>Ficis sur</i>	Tur	Soak bark	Diarrhea
<i>Cissus refescens</i>	Ikpoor	Ground bulb	Stomach ache
<i>Psedocedrela kotchyi</i>	Kpamegh	Boil bark	HIV/AIDS

Table 3: Edible plants in study area

Botanical name	Vernacular name (Tiv)	Part(s) eaten	Traditional food type
<i>Moringa oleifera</i>	Jeglegede	Flowers, leaves	Vegetable
<i>Ficur sur</i>	Tur	Tender leaves, ripe fruits	Vegetable, fruit pulp
<i>Tecca leoutopetaloides</i>	Gbache	Underground bulb	Soup
<i>Physalis angulata</i>	Tam kpur	Ripe fruits	Fruit pulp
<i>Grewia venusta</i>	Hwer	Stem fibre	Soup
<i>Vitex doniana</i>	Hulugh	Fruits	Fruit pulp
<i>Elaeis guineensis</i>	Ikye	Fruit, palm wine	Oil, drink
<i>Imperata cylindrica</i>	Ihila	Root rhizome	Sweetener
<i>Parinari polyandra</i>	Ibua	Fruits	Fruit pulp
<i>Hibiscus aspera</i>	Agakpande	Fresh leaves	Leafy vegetable
<i>Prosopis africana</i>	Gbaaye	Seeds	Condiment
<i>Dioscorea rotundata</i>	Yough	Roots	Pounded yam
<i>Vetellaria paradoxa</i>	Chamegh	Ripe fruits	Fruit pulp
<i>Anona senegalensis</i>	Ahur	Ripe fruits	Fruit pulp
<i>Bombax costatum</i>	Kuka	Dry leaves	Vegetables
<i>Parkia biglobosa</i>	Nune	Ripe fruits	Condiment
<i>Cisus polnea</i>	Ager	Fibre	Soup
<i>Strychnos spinosa</i>	Maku	Ripe fruits	Fruit pulp

Table 4 : Fauna species in the study area

Scientific name	Family name	Common name	Vernacular name
<i>Lepus capensis</i>	Hyracoidean	Rabbit	Alom
<i>Xerus erythropus</i>	Rodentia	Ground squirrel	Hinga
<i>Cricetomys gambianus</i>	Rodentia	Giant rat	Ikyoor
<i>Thryonomis swinderianus</i>	Rodentia	Cane rat	Viha
<i>Myomys daltoni</i>	Rodentia	Bush mouse	Mchoko/ Akpev
<i>Cephalophus rufilatus</i>	Artiodactyla	Red flanked duiker	Ikyuran
<i>Qurebia ourebi</i>	Artiodactyla	Oribi	Ihoh
<i>Genetta tigrina</i>	Carnivora	Bush genet	Ishumbe
<i>Alcedo quadribrachys</i>	Akedinidae	Shining blue king fisher	Ayoosu
<i>Ardeola ibis</i>	Ardeidae	Cattle egret	Inyon-bua
<i>Aremopterix leucotis</i>	Alaudidae	Hawk	Itsoo
<i>Tockus nasa</i>	Bucerotidae	Grey hornbill	Ichaankera
<i>Arvicanthis niloticus</i>	Cricetidae	Nile rat	Iyonguv
<i>Clarias gariepinus</i>	Clariidae	Mud fish	Gbaver
<i>Clarias anguillaris</i>	Clariidae	Mud fish	Ashoon
<i>Oreochromis niloticus</i>	Cichlidae	Tilapia	Ikpoo
<i>Barbus occidentalis</i>	Bagridae	Cat fish	Ambi-uya

Table 5: Results of Presumptive Test (Using Mccradys Statistics Table)

Location of sample	Description	Number/100ml of water	Number/100ml of water	Number/100ml of water
		Coliforms	Faecal coliforms	Colonies/plate
A	(Upper part of the River)	350	45	2.36 x 10 ⁵ cfu/ml
B	Opposite A	550	60	2.31x 10 ⁵ cfu/ml
C	(Middle)	200	45	1.29 x10 ⁵ cfu/ml
D	(lower part of River)	550	170	2.40 x 10 ⁵ cfu/ml
E	(Opposite D)	900	275	2.30 x 10 ⁵ cfu/ml

Table 6: Results of standard plate count from soil sample

Location of sample	Number of colonies per plate			
	10^{-1}	10^{-2}	10^{-3}	10^{-4}
Point I	90	86	50	15
Point II	180	162	80	30
Point III	85	50	42	22
Breath I	91	78	38	20
Breath II	81	55	25	18
Breath III	87	63	31	15
Length I	89	70	43	20
Length II	195	98	60	29
Length III	108	93	65	40
Reference point				
Eco Control I	80	50	41	22
Eco Control II	250	120	70	40
Eco Control III	48	30	20	07
Eco Control IV	Too crowded	160	85	60
Plot I within site	Too crowded	170	91	70
Plot II within site	200	108	70	50
Plot III within site	80	41	30	20

DISCUSSION

Species inventory, abundance and distribution

A total of 77 plant species were identified. Of this, 23 species were trees, 6 shrubs, 34 herbs, 3 woody climbers, 3 climbers while woody herbs, bryophytes, mushrooms and lichens

had two species each. The results show that herbs were more abundant than trees and shrubs. This is a physiognomic feature of the guinea savanna zone where Makurdi is located. Seven families were outstandingly abundant. These include Poaceae (9 species) followed by Fabaccae (8 species) and

Euphorbiaceae (6 species). Combretaceae, Compositae and Meliaceae had four species each. The most dominant species were *Rottboellia cochinchinensis*, *Sateria ancepts* and *Tephrosia bracteolate* with 1310, 1200 and 760 species respectively. Information from interviews with hunters and fishermen in the neighboring community reveals that 12 families of animals made up of 4 species of Rodentia and 2 species of Artiodactyla while Carnivora, Cricetidae and Akedinidae had one species each as shown in Table 4. Others were four fish species in 3 families made up of Clariidae (2 species) and Chichlidae with Bagridae having one species each. Six tree species *Lannea schimperi*, *Stereospermum kunthianum*, *Euphorbia heterophylla*, *Hymenocardia acida* *Trichilia emetic* and *Vitex doniana* were found only within the facility with none seen within the control plots.

MEDICINAL PLANTS

Forty-one (41) plants were identified to have medicinal importance in the study area. The ailments reported to have been cured by these

plants range from headaches, fever, waist pain, stomach ache, rheumatism, hernia, snake bites, catarrh, small pox, blood clotting and hepatitis among others as shown in Table 2. Although these claims have not being verified, the people affirmed that for decades, they have being using the plants successfully for the treatment of the said ailments as passed unto them by their fore- fathers.

EDIBLE PLANTS

18 plants species in the area are eaten by the people mostly as vegetables, fruits and condiments (Table 3). *Moringa oleifera*,

Ficus sur, *Hibiscus aspera* and *Bombax costatum* are eaten as leafy vegetables while *Ficus sur*, *Physalis angulata*, *Vitex doniana*, *Parinari polyandra*, *Vitellaria paradoxa*, *Anona senegalensis*, *Bombax costatum* and *Parkia biglobosa* are eaten as fruits. Two species, *Prosopis africana* and *Parkia biglobosa* are eaten as soup condiments. It was observed that *Prosopis africana*, *Strychnos spinosus*, *Anona senegalensis* and *Vitellaria paradoxa* have both medicinal and food values as reported by Etkin and Ross, 1982 and Pieroni et al, 2005.

Microbial Evaluation of Soil and Water in the study area

The result of standard plate count from soil samples and presumptive tests are presented in Table 5 and 6. Differences in soil microbial load are a function of soil fertility resulting from decayed organic matter. The higher the soil microbial count, the more fertile the soil and consequently the higher its potential for higher agricultural productivity. Eco control II, plot I and plot III are located within the plains of Jamo stream. The microbial status of the water is an indication of polluted water.

Total coliforms ranged from 3.50×10^5 to 9.00×10^5 colony forming units (cfu/ml) while faecal coliforms ranged between 4.5×10^4 to 2.75×10^5 cfu/ml. These indicates high level of contamination of the water from animal and organic matter and can harbour human pathogenic microorganisms. The organisms identified include *Escherichia coli*, *Aerobacter aerogenes*, *Faecal Streptococcus* and *Bacillus aureus*. Others were *Escherichia ferundii*, *Salmonella typhosa*, *Pseudomonas aerogenosa* *Aspergillus spp*, *Penicillium spp* and yeast.

CONCLUSION

The results of the study present the status of environmental components within the waste treatment facility and can be useful as baseline information for monitoring and evaluation of the effects of the facility on the environmental resources. Species found only within the facility needs to be monitored to ensure that they are not destroyed to prevent their extinction within the facility. Also, there is need to investigate the nutritional profiles and therapeutic values of the edible and medicinal plant species respectively so as to verify the claims of the local residents. The soil within the facility is fertile due to in soil microbial load and the water is highly contaminated and harbours some pathogenic organisms which can cause disease.

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